

Research article

Open Access

Intrathecal levels of matrix metalloproteinases in systemic lupus erythematosus with central nervous system engagement

Estelle Trysberg¹, Kaj Blennow², Olof Zachrisson² and Andrej Tarkowski¹¹Department of Rheumatology and Inflammation Research, Göteborg University, Sahlgrenska University Hospital, Göteborg, Sweden²Institute of Clinical Neuroscience, Göteborg University, Sahlgrenska University Hospital, Göteborg, SwedenCorresponding author: Estelle Trysberg, Estelle@immuno.gu.se

Received: 11 Apr 2004 Revisions requested: 19 May 2004 Revisions received: 15 Jun 2004 Accepted: 23 Jul 2004 Published: 23 Sep 2004

Arthritis Res Ther 2004, **6**:R551-R556 (DOI 10.1186/ar1228)© 2004 Trysberg *et al.*, licensee BioMed Central Ltd.This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is cited.

Abstract

Symptoms originating from the central nervous system (CNS) occur frequently in patients with systemic lupus erythematosus (SLE), and CNS involvement in lupus is associated with increased morbidity and mortality. We recently showed that neurones and astrocytes are continuously damaged during the course of CNS lupus. The matrix metalloproteinases (MMPs) are a group of tissue degrading enzymes that may be involved in this ongoing brain destruction. The aim of this study was to examine endogenous levels of free, enzymatically active MMP-2 and MMP-9 in cerebrospinal fluid from patients with SLE. A total of 123 patients with SLE were evaluated clinically, with magnetic resonance imaging of brain and cerebrospinal fluid (CSF) analyses. Levels of free MMP-2 and MMP-9 were determined in CSF using an enzymatic activity assay. CSF samples from

another 22 cerebrally healthy individuals were used as a control. Intrathecal MMP-9 levels were significantly increased in patients with neuropsychiatric SLE as compared with SLE patients without CNS involvement ($P < 0.05$) and healthy control individuals ($P = 0.0012$). Interestingly, significant correlations between MMP-9 and intrathecal levels of neuronal and glial degradation products were noted, indicating ongoing intrathecal degeneration in the brains of lupus patients expressing MMP-9. In addition, intrathecal levels of IL-6 and IL-8 – two cytokines that are known to upregulate MMP-9 – both exhibited significant correlation with MMP-9 levels in CSF ($P < 0.0001$), suggesting a potential MMP-9 activation pathway. Our findings suggest that proinflammatory cytokine induced MMP-9 production leads to brain damage in patients with CNS lupus.

Keywords: cerebrospinal fluid, magnetic resonance imaging of the brain, matrix metalloproteinase-2, matrix metalloproteinase-9, neuropsychiatric involvement in systemic lupus erythematosus

Introduction

Central nervous system (CNS) involvement has been reported to occur in 14–75% of all systemic lupus erythematosus (SLE) patients [1-3]. However, frequency rates vary considerably, depending on the diagnostic criteria applied. CNS lupus can occur at any time during the course of SLE, and its symptoms are diverse. The features of this condition can include seizures, stroke, depression, psychoses and disordered mentation. Neuropsychiatric involvement in SLE (NPSLE) has been shown to predict a high frequency of flares, and it is considered a major cause of long-standing functional impairment as well as a cause of mortality [4]. Over the past decade CNS lupus has been treated with cytotoxic drugs, which improve disease outcome [5,6].

Because of the multiple pathogenic mechanisms that cause manifestations of CNS lupus, no single confirmatory diagnostic test is available. Several clinical, laboratory and radiographic test findings are reported to be abnormal in some but not all patients with CNS lupus. Magnetic resonance imaging (MRI) of brain has been shown to be valuable in detecting even minor lesions caused by CNS lupus, and these correlate with CNS manifestations in SLE [7]. Pleocytosis and elevated protein levels are found in some but not all patients with CNS lupus [8,9]. Elevated concentrations of IgG in cerebrospinal fluid (CSF), IgG:albumin ratio and IgG index, and the presence of oligoclonal bands have all been described with varying frequencies in patients with NPSLE [10-12]. Few studies have demonstrated elevated IL-6 levels in CSF from patients with CNS lupus [13-

17]. Some other reports have described increased levels of IL-1 [13], IL-8 [15] and interferon- γ [18] in CSF from patients with CNS lupus. We recently reported neuronal damage, astrogliosis and formation of toxic metabolic products such as A β 42 in patients with NPSLE [19].

One of the prototypical destructive events in the human brain, initiated by the release of inflammatory cytokines and ending with tissue destruction, is production of matrix metalloproteinases (MMPs). The MMPs are a family of endopeptidases produced by a variety of inflammatory cells [20]. All of the cell types that exist in the CNS are potential sources of MMPs. *In vitro*, neurones, astrocytes, microglia [21,22] and oligodendrocytes [23] express various MMP family members, and production of MMPs by neuronal cells can be upregulated by several inflammatory cytokines. MMPs have a multitude of regulatory functions, including control of influx of inflammatory mononuclear cells into the CNS, participation in myelin destruction and disruption of the integrity of the blood–brain barrier [24]. With respect to MMP-9, it was recently shown that CSF levels of this enzyme increase during bacterial meningitis and that it is associated with brain damage [25,26]. The aim of the present study was to measure levels of free active MMP-2 and MMP-9 in CSF of SLE patients with CNS lupus, and to relate these data to clinical and laboratory measures of brain parenchymal degradation. Our results suggest that MMP-9 but not MMP-2 actively participates in brain destruction in CNS lupus.

Methods

Participants

A total of 122 patients fulfilled four or more of the American Rheumatism Association 1987 revised criteria for the classification of SLE [27]. The patients (106 females and 16 males, aged 17–91 years [mean age \pm standard deviation 42 ± 14 years]) were being treated at the Department of Rheumatology at Sahlgrenska University Hospital. Disease duration varied between 0 and 49 years (mean duration 8 ± 9 years). The patients were consecutively included in the study. They underwent a thorough clinical examination by experienced staff rheumatologist, neurologist and neuropsychologist. Examination of CNS signs and symptoms included lumbar puncture, neuropsychological tests and MRI of the brain.

The proposed definition of CNS lupus in the American Rheumatism Association criteria for SLE [27] appears inadequate, given that only two elements – psychosis and seizures – are included. As previously described [28], we defined CNS lupus as the presence of at least two of the following seven items in association with clinical evidence of disease progression: recent onset psychosis, transverse myelitis, aseptic meningitis, seizures, pathological brain MRI, severely abnormal neuropsychiatric test findings [29]

and oligoclonal IgG bands in CSF. The pathogenesis of antiphospholipid antibody mediated brain damage is a thrombotic rather than inflammatory complication of SLE, and so we decided to exclude this condition from the definition of CNS lupus. Non-SLE causes of neurological events (e.g. cerebral infections) were also ruled out. Based on the criteria above, patients were divided into three distinct groups: patients with CNS lupus ($n = 43$); patients with SLE but without any signs of CNS involvement ($n = 71$); and patients with SLE complicated by antiphospholipid syndrome or patients who met the CNS lupus criteria but were ruled out because of non-SLE origin of neurological events ($n = 9$). The frequencies of various CNS manifestations and patient treatments in the SLE patients are summarized in Tables 1 and 2, respectively. The study was approved by the ethical committee of the University of Göteborg.

Control individuals

CSF from 22 healthy individuals (mean age \pm standard deviation: 38 ± 11 years; 12 females and 10 males), without previous history of neurological disorder and with normal neurological status, served as control individuals. There were no significant differences between males and females with respect to intrathecal levels of MMP-9 (0.17 ± 0.17 pg/ml versus 0.30 ± 0.30 pg/ml; not significant) or intrathecal levels of MMP-2 (388 ± 48 pg/ml versus 379 ± 48 pg/ml; not significant).

Cerebrospinal fluid analyses

Levels of MMP-2 and MMP-9 were determined using an activity assay system (Amersham Pharmacia Biotech, Buckinghamshire, UK), which was constructed to measure enzymatically active forms of MMP-2 and MMP-9. The detection level was 190 pg/ml for MMP-2 and 125 pg/ml for MMP-9. All values below the detection levels were considered negative. Paired serum and CSF samples were analyzed for albumin and IgG levels using nephelometry. As an indicator of blood–brain barrier function, the quotient of CSF albumin $\times 10^3$ /serum albumin was analyzed (normal values <6.8 [<45 years of age] and <10.2 [>45 years of age]) [30]. The CSF/serum IgG index was used as a measure of intrathecal IgG production and calculated by using the following formula (normal value <0.7): $(\text{CSF IgG} \times 10^3)/([\text{CSF albumin} \times 10^3]/\text{serum albumin})$. All CSF samples were also analyzed by isoelectric focusing to permit detection of oligoclonal IgG bands.

Magnetic resonance imaging analyses

Neuroimaging was performed to evaluate the extent and localization of brain lesions. The neuroimaging technique used was multiplanar MRI. The MRI examinations (Philips Gyroscan T5-II, Eindhoven, The Netherlands) were performed with axial proton density and T₂-weighted images of the brain. MRI abnormalities were seen in 72% of patients

Table 1**Clinical central nervous system manifestations in systemic lupus erythematosus patients included in the study**

CNS manifestations	NPSLE (n = 43)	No NPSLE (n = 71)	aPL syndrome (n = 9)
Acute confusional state	1	1	0
Anxiety disorder	0	5	0
Aseptic meningitis	1	0	0
Cerebrovascular disease	6	8	4
Cognitive dysfunction	9	19	1
Demyelinating syndrome	4	1	0
Headache	8	21	4
Mood disorders	5	11	1
Movement disorder	2	0	3
Myasthenia gravis	0	2	0
Myelopathy	1	1	0
Polyneuropathy	0	1	0
Mononeuropathy	0	1	0
Neuropathy	0	1	0
Psychosis	7	1	0
Seizure disorders	6	2	0
Tiredness	0	8	0

Each patient may have had multiple clinical manifestations of central nervous system (CNS) involvement. aPL, antiphospholipid; NPSLE, neuropsychiatric involvement in systemic lupus erythematosus.

with CNS lupus and in 33% of SLE cases classified as cerebrally healthy.

Statistical analysis

Statistical comparisons were made using the nonparametric Mann–Whitney U-test or, in case of follow-up data, the Wilcoxon's test for paired data. Results are presented as means \pm standard error of the mean. $P < 0.05$ was considered statistically significant. Spearman-rank correlation was used for calculation of correlation. The statistical analyses were conducted using the Statview® (SAS, Cary, NC, USA) program.

Results

A total of 123 patients met criteria for SLE. Forty-three patients were found to have NPSLE (in accordance with the criteria presented in the Methods section above), and nine patients either were found to have phospholipid antibody syndrome or met the CNS lupus criteria but were excluded because of non-SLE origin of neurological events. The remaining 71 SLE patients were considered to be cerebrally healthy.

Table 2**Pharmacological treatment of patients with systemic lupus erythematosus included in the study at the time the lumbar puncture was performed**

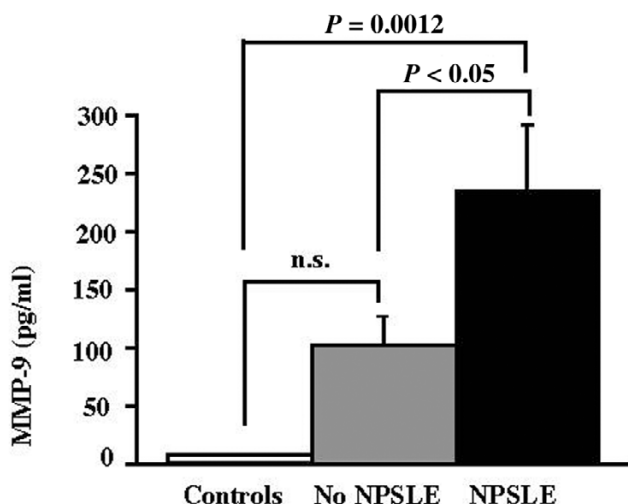
Treatment	NPSLE (n = 43)	No NPSLE (n = 71)	aPL syndrome (n = 9)
Prednisolone (≤ 10 mg)	18	37	3
Prednisolone (> 10 mg)	7	8	0
No prednisolone	18	23	5
Antimalarials	1	8	0
Azathioprine	5	10	3
Azathioprine + cyclosporin A	2	3	0
Azathioprine + antimalarials	0	0	0
Methotrexate	5	1	0
Cyclosporin A	0	6	1
Cyclophosphamide	14	10	0
Cyclophosphamide + cyclosporin A	3	0	0
Cyclophosphamide + antimalarials	0	1	0
No cytotoxic drug	11	30	4
Antihypertensive treatment	14	15	2
Low-dose aspirin	8	19	3
Warfarin	3	2	2

aPL, antiphospholipid; NPSLE, neuropsychiatric involvement in systemic lupus erythematosus.

Mild pleocytosis was seen in patients with CNS lupus ($9 \times 10^6 \pm 6 \times 10^6$ cells/l) as compared with cerebrally healthy SLE patients ($2 \times 10^6 \pm 0.5 \times 10^6$ cells/l; not significant). As previously validated [31], we found an increased number of oligoclonal bands in CSF from the CNS lupus group (1.7 ± 0.4) as compared with SLE patients without CNS involvement (0.5 ± 0.1 ; $P < 0.05$). The mean level of CSF:serum albumin ratio was not increased in patients with NPSLE (6.1 ± 0.7 mg/dl) as compared with cerebrally healthy SLE patients (5.2 ± 0.3 mg/dl; not significant). There were no significant differences in levels of serum antibodies specific for native DNA or in complement levels (C3 and C4) between SLE patients with and those without CNS involvement.

Intrathecal MMP-9 levels were significantly increased in all SLE patients as compared with cerebrally healthy control individuals (153 ± 27 versus 0.28 ± 0.16 pg/ml; $P = 0.016$). In CNS lupus patients MMP-9 levels were significantly increased, both compared with cerebrally healthy SLE patients (240 ± 60 versus 100 ± 20 pg/ml; $P < 0.05$; Fig. 1) and compared with cerebrally healthy control individuals (240 ± 60 versus 0.28 ± 0.16 pg/ml; $P = 0.0012$).

Figure 1



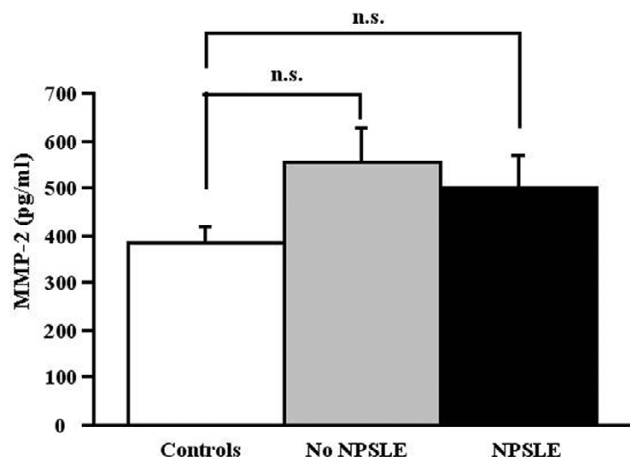
Cerebrospinal fluid expression of active matrix metalloproteinase (MMP)-9 in patients with systemic lupus erythematosus with (NPSLE) and without (No NPSLE) central nervous system involvement, and in cerebrally healthy control individuals.

On stratification of all SLE patients into two groups with respect to the presence or absence of brain MRI pathology, we found increased CSF MMP-9 levels in SLE patients with MRI pathology as compared with those without (160 ± 50 versus 140 ± 30 pg/ml; not significant).

There were no statistically significant differences in CSF MMP-2 levels between NPSLE and patients without CNS lupus (499 ± 70 versus 555 ± 70 pg/ml; not significant) or compared with cerebrally healthy control individuals (499 ± 70 versus 384 ± 33 pg/ml; not significant; Fig. 2).

CSF levels of IL-6 (47 ± 25 pg/ml versus 15 ± 3 pg/ml; $P < 0.008$) and IL-8 (91 ± 23 pg/ml versus 45 ± 6 pg/ml; $P < 0.05$) were both significantly increased in CSF from patients with CNS lupus as compared with cerebrally healthy SLE patients, supporting our previous findings [15,19]. Importantly, intrathecal levels of IL-6 and IL-8 significantly correlated with those of MMP-9 ($r = 0.30$ [$P < 0.002$] and $r = 0.47$ [$P < 0.0001$], respectively; Table 3 and Fig. 3). A neuronal degeneration marker (protein tau) and an astrocytic degeneration marker (glial fibrillary acidic protein) were both significantly increased in CSF from NPSLE patients as compared with CSF from SLE patients who were clinically free from CNS involvement (311 ± 78 pg/ml versus 178 ± 16 pg/ml [$P < 0.05$] and 1288 ± 708 pg/ml versus 396 ± 30 pg/ml [$P < 0.009$]), in concordance with previous findings [28,32]. Importantly, a significant correlation was noted between intrathecal MMP-9 and levels of tau and glial fibrillary acidic protein ($P < 0.05$ in both cases; Table 3).

Figure 2



Cerebrospinal fluid expression of active matrix metalloproteinase (MMP)-2 in patients with systemic lupus erythematosus with (NPSLE) and without (No NPSLE) central nervous system involvement, and in cerebrally healthy control individuals.

Table 3

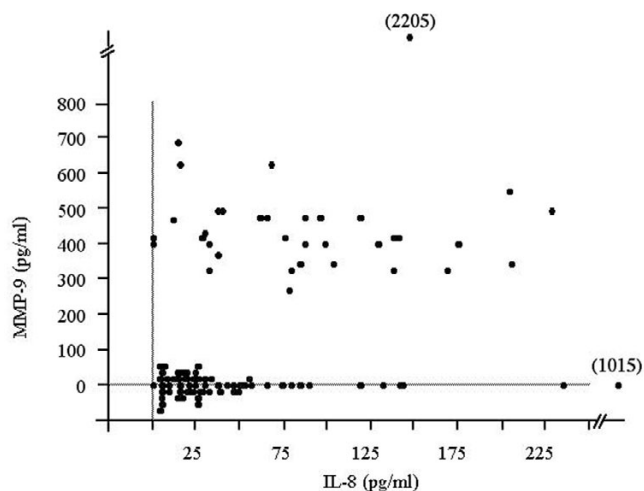
Relationship between intrathecal expression of matrix metalloproteinase-9 and inflammatory cytokines, and astrocytic and neuronal degradation products in 119 patients with systemic lupus erythematosus

	MMP-9	
	r	P
MMP-2	0.40	<0.0001
IL-6	0.30	0.002
IL-8	0.47	<0.0001
Tau	0.26	0.007
GFAP	0.33	<0.04

GFAP, glial fibrillary acidic protein; MMP, matrix metalloproteinase.

Discussion

We previously reported that patients with CNS lupus express increased intrathecal levels of proinflammatory cytokines IL-6, IL-8 and interferon- γ . Furthermore, we recently demonstrated that inflammation in CNS during SLE leads to neurodegeneration, which manifests as increased levels of neuronal and astrocytic degradation products [28,32]. However, the mechanisms responsible for the brain damage occurring during CNS lupus have never been clarified. It was previously demonstrated [33,34] that proinflammatory cytokines are able to trigger production and release of tissue-derived MMPs. It is also established that most of the resident cells in human brain have the capacity to produce MMP-2 and MMP-9 – enzymes that are known to participate in brain damage (e.g. in case of infectious meningitis) [35-37].

Figure 3

Scattergram showing relationship between IL-8 and matrix metalloproteinase (MMP)-9 in cerebrospinal fluid of all patients with systemic lupus erythematosus ($r = 0.47$; $P < 0.0001$).

In the present study we found significantly higher levels of MMP-9 in CSF from NPSLE patients than in CSF from SLE patients without CNS lupus and healthy control individuals. This finding may be of clinical significance because of correlation between MMP-9 and levels of neuronal/astrocytic degradation products in CSF, reflecting the potential of MMP-9 to damage brain parenchyma. Findings of a relationship between MMP-9 and brain damage in SLE are new and should be scrutinized critically. We believe that effort should be invested in analyzing free (i.e. non-tissue inhibitor of metalloproteinase bound) and metabolically active enzyme to ascertain the validity of our conclusions. The method used in the present study fulfils both of these requirements.

MMP-9 plays an essential role in the breakdown of extracellular matrix molecules at the blood–brain barrier. This, together with locally elevated levels of the chemokine IL-8, greatly facilitates the transmigration of activated inflammatory cells across the endothelium. Indeed, a significant correlation was observed between the intrathecal levels of MMP-9 and IL-8.

A broad range of cytokines that are expressed in viral meningitis are able to regulate the production of MMPs and tissue inhibitors of metalloproteinase [38,39]. IL-1 α and IL-1 β , as well as IL-6, induce or upregulate the transcription of MMP genes [40]. This, along with increased IL-6 levels in CSF of patients with CNS lupus, may provide an explanation for the local synthesis of MMP-9.

CSF levels of enzymatically active MMP-2 were not increased in NPSLE patients as compared with SLE

patients without CNS involvement or compared with cerebrally healthy control individuals. This finding is consistent with a previous study of viral meningitis, in which increased expression of MMP-9 but constitutive expression of MMP-2 was found [37]. The reason for this discrepancy, seen in the present study as well as in the previous one, might be differential regulation of gene promoters for MMP-9 and MMP-2. Indeed, whereas the promoter regions of MMP-9, which is regulated by cytokines, harbor a nuclear factor- κ B responsive element, those of MMP-2 lack a TATA box and contain two SP-1 sites, both characteristic of house keeping genes [40,41].

Conclusion

Our present and previous findings indicate that the following chain of events takes place during NPSLE: intrathecal release of inflammatory cytokines, leading to synthesis and release of MMP-9, potentially culminating in insult to brain parenchyma, resulting in release of neuronal and astrocytic degradation products and culminating in MRI verifiable lesions and clinical states of brain deficiency. Interestingly, a recent study [42] indicated that IgG dose-dependently downregulates secretion of MMP-9 by macrophages, suggesting a possible early therapeutic manipulation in NPSLE.

Competing interests

None declared.

Acknowledgement

This work was supported by the Göteborg Medical Society, Swedish Association against Rheumatism, King Gustaf V: s Foundation, Swedish Medical Research Council, Nanna Svartz' Foundation, Börje Dahlin's Foundation, Swedish National Inflammation Network, Swedish National Infection and Vaccination Network, AME Wolff Foundation, and the University of Göteborg.

References

1. McCune WJ, Golbus J: **Neuropsychiatric lupus.** *Rheum Dis Clin North Am* 1988, **14**:149-167.
2. Hanly JG, Liang MH: **Cognitive disorders in systemic lupus erythematosus. Epidemiologic and clinical issues.** *Ann NY Acad Sci* 1997, **823**:60-68.
3. Feinglass EJ, Arnett FC, Dorsch CA, Zizic TM, Stevens MB: **Neuropsychiatric manifestations of systemic lupus erythematosus: diagnosis, clinical spectrum, and relationship to other features of the disease.** *Medicine (Baltimore)* 1976, **55**:323-339.
4. Jonsson H, Nived O, Sturfelt G: **Outcome in systemic lupus erythematosus: a prospective study of patients from a defined population.** *Medicine (Baltimore)* 1989, **68**:141-150.
5. McCune WJ, Golbus J, Zeldes W, Bohlke P, Dunne R, Fox DA: **Clinical and immunologic effects of monthly administration of intravenous cyclophosphamide in severe systemic lupus erythematosus.** *N Engl J Med* 1988, **318**:1423-1431.
6. Boumpas DT, Yamada H, Patronas NJ, Scott D, Klippel JH, Balow JE: **Pulse cyclophosphamide for severe neuropsychiatric lupus.** *Q J Med* 1991, **81**:975-984.
7. Oku K, Atsumi T, Furukawa S, Horita T, Sakai Y, Jodo S, Amasaki Y, Ichikawa K, Amengual O, Koike T: **Cerebral imaging by magnetic resonance imaging and single photon emission computed tomography in systemic lupus erythematosus with**

- central nervous system involvement. *Rheumatology (Oxford)* 2003, **42**:773-777.
8. Sergent JS, Lockshin MD: **Editorial treatment of central nervous system lupus erythematosus.** *Ann Intern Med* 1974, **80**:413-414.
 9. Abel T, Gladman DD, Urowitz MB: **Neuropsychiatric lupus.** *J Rheumatol* 1980, **7**:325-333.
 10. Ernerudh J, Olsson T, Lindstrom F, Skogh T: **Cerebrospinal fluid immunoglobulin abnormalities in systemic lupus erythematosus.** *J Neurol Neurosurg Psychiatry* 1985, **48**:807-813.
 11. Winfield JB, Shaw M, Silverman LM, Eisenberg RA, Wilson HAD, Koffler D: **Intrathecal IgG synthesis and blood-brain barrier impairment in patients with systemic lupus erythematosus and central nervous system dysfunction.** *Am J Med* 1983, **74**:837-844.
 12. Hirohata S, Hirose S, Miyamoto T: **Cerebrospinal fluid IgM, IgA, and IgG indexes in systemic lupus erythematosus. Their use as estimates of central nervous system disease activity.** *Arch Intern Med* 1985, **145**:1843-1846.
 13. Alcocer-Varela J, Aleman-Hoey D, Alarcon-Segovia D: **Interleukin-1 and interleukin-6 activities are increased in the cerebrospinal fluid of patients with CNS lupus erythematosus and correlate with local late T-cell activation markers.** *Lupus* 1992, **1**:111-117.
 14. Hirohata S, Tanimoto K, Ito K: **Elevation of cerebrospinal fluid interleukin-6 activity in patients with vasculitides and central nervous system involvement.** *Clin Immunol Immunopathol* 1993, **66**:225-229.
 15. Trysberg E, Carlsten H, Tarkowski A: **Intrathecal cytokines in systemic lupus erythematosus with central nervous system involvement.** *Lupus* 2000, **9**:498-503.
 16. Tsai CY, Wu TH, Tsai ST, Chen KH, Thajeb P, Lin WM, Yu HS, Yu CL: **Cerebrospinal fluid interleukin-6, prostaglandin E2 and autoantibodies in patients with neuropsychiatric systemic lupus erythematosus and central nervous system infections.** *Scand J Rheumatol* 1994, **23**:57-63.
 17. Yeh TS, Wang CR, Jeng GW, Lee GL, Chen MY, Wang GR, Lin KT, Chuang CY, Chen CY: **The study of anticardiolipin antibodies and interleukin-6 in cerebrospinal fluid and blood of Chinese patients with systemic lupus erythematosus and central nervous system involvement.** *Autoimmunity* 1994, **18**:169-175.
 18. Svenungsson E, Andersson M, Brundin L, van Vollenhoven R, Khademi M, Tarkowski A, Greitz D, Dahlstrom M, Lundberg I, Klareskog L, et al.: **Increased levels of proinflammatory cytokines and nitric oxide metabolites in neuropsychiatric lupus erythematosus.** *Ann Rheum Dis* 2001, **60**:372-379.
 19. Trysberg E, Nylén K, Rosengren L, Tarkowski A: **Neuronal and astrocytic damage in systemic lupus erythematosus patients with central nervous system involvement.** *Arthritis Rheum* 2003, **48**:2881-2887.
 20. Leppert D, Lindberg RL, Kappos L, Leib SL: **Matrix metalloproteinases: multifunctional effectors of inflammation in multiple sclerosis and bacterial meningitis.** *Brain Res Brain Res Rev* 2001, **36**:249-257.
 21. Gottschall PE, Deb S: **Regulation of matrix metalloproteinase expressions in astrocytes, microglia and neurons.** *Neuroimmunomodulation* 1996, **3**:69-75.
 22. Apodaca G, Rutka JT, Bouhana K, Berens ME, Giblin JR, Rosenblum ML, McKerrow JH, Banda MJ: **Expression of metalloproteinases and metalloproteinase inhibitors by fetal astrocytes and glioma cells.** *Cancer Res* 1990, **50**:2322-2329.
 23. Uhm JH, Dooley NP, Oh LY, Yong VW: **Oligodendrocytes utilize a matrix metalloproteinase, MMP-9, to extend processes along an astrocyte extracellular matrix.** *Glia* 1998, **22**:53-63.
 24. Gijbels K, Masure S, Carton H, Opdenakker G: **Gelatinase in the cerebrospinal fluid of patients with multiple sclerosis and other inflammatory neurological disorders.** *J Neuroimmunol* 1992, **41**:29-34.
 25. Leib SL, Clements JM, Lindberg RL, Heimgartner C, Loeffler JM, Pfister LA, Tauber MG, Leppert D: **Inhibition of matrix metalloproteinases and tumour necrosis factor alpha converting enzyme as adjuvant therapy in pneumococcal meningitis.** *Brain* 2001, **124**:1734-1742.
 26. Leib SL, Kim YS, Chow LL, Sheldon RA, Tauber MG: **Reactive oxygen intermediates contribute to necrotic and apoptotic neuronal injury in an infant rat model of bacterial meningitis due to group B streptococci.** *J Clin Invest* 1996, **98**:2632-2639.
 27. Hochberg MC: **Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus [letter].** *Arthritis Rheum* 1997, **40**:1725.
 28. Trysberg E, Nylén K, Rosengren LE, Tarkowski A: **Neuronal and astrocytic damage in systemic lupus erythematosus patients with central nervous system involvement.** *Arthritis Rheum* 2003, **48**:2881-2887.
 29. Breitbach SA, Alexander RW, Daltroy LH, Liang MH, Boll TJ, Karlson EW, Partiridge AJ, Roberts WN, Stern SH, Wacholtz MC, et al.: **Determinants of cognitive performance in systemic lupus erythematosus.** *J Clin Exp Neuropsychol* 1998, **20**:157-166.
 30. Blennow K, Fredman P, Wallin A, Gottfries CG, Karlsson I, Langstrom G, Skoog I, Svennerholm L, Wikkelso C: **Protein analysis in cerebrospinal fluid. II. Reference values derived from healthy individuals 18-88 years of age.** *Eur Neurol* 1993, **33**:129-133.
 31. Moore BW: **A soluble protein characteristic of the nervous system.** *Biochem Biophys Res Commun* 1965, **19**:739-744.
 32. Trysberg E, Höglund K, Svenungsson E, Blennow K, Tarkowski A: **Decreased levels of soluble amyloid β -protein precursor and β -amyloid protein in cerebrospinal fluid of patients with systemic lupus erythematosus.** *Arthritis Res Ther* 2004, **6**:R129-R136.
 33. Okada Y, Tsuchiya H, Shimizu H, Tomita K, Nakanishi I, Sato H, Seiki M, Yamashita K, Hayakawa T: **Induction and stimulation of 92-kDa gelatinase/type IV collagenase production in osteosarcoma and fibrosarcoma cell lines by tumor necrosis factor alpha.** *Biochem Biophys Res Commun* 1990, **171**:610-617.
 34. Lotz M, Guerne PA: **Interleukin-6 induces the synthesis of tissue inhibitor of metalloproteinases-1/erythroid potentiating activity (TIMP-1/EPA).** *J Biol Chem* 1991, **266**:2017-2020.
 35. Matsuura E, Umehara F, Hashiguchi T, Fujimoto N, Okada Y, Osame M: **Marked increase of matrix metalloproteinase 9 in cerebrospinal fluid of patients with fungal or tuberculous meningoencephalitis.** *J Neurol Sci* 2000, **173**:45-52.
 36. Shapиро S, Miller A, Lahat N, Sobel E, Lerner A: **Expression of matrix metalloproteinases, sICAM-1 and IL-8 in CSF from children with meningitis.** *J Neurol Sci* 2003, **206**:43-48.
 37. Kolb SA, Lahrtz F, Paul R, Leppert D, Nadal D, Pfister HW, Fontana A: **Matrix metalloproteinases and tissue inhibitors of metalloproteinases in viral meningitis: upregulation of MMP-9 and TIMP-1 in cerebrospinal fluid.** *J Neuroimmunol* 1998, **84**:143-150.
 38. Goetzl EJ, Banda MJ, Leppert D: **Matrix metalloproteinases in immunity.** *J Immunol* 1996, **156**:1-4.
 39. Johnatty RN, Taub DD, Reeder SP, Turcovski-Corrales SM, Cottam DW, Stephenson TJ, Rees RC: **Cytokine and chemokine regulation of proMMP-9 and TIMP-1 production by human peripheral blood lymphocytes.** *J Immunol* 1997, **158**:2327-2333.
 40. Mauviel A: **Cytokine regulation of metalloproteinase gene expression.** *J Cell Biochem* 1993, **53**:288-295.
 41. Benbow U, Brinckerhoff CE: **The AP-1 site and MMP gene regulation: what is all the fuss about?** *Matrix Biol* 1997, **15**:519-526.
 42. Shapиро S, Shoenfeld Y, Gilburd B, Sobel E, Lahat N: **Intravenous gamma globulin inhibits the production of matrix metalloproteinase-9 in macrophages.** *Cancer* 2002, **95**:2032-2037.