



# The large-conductance calcium-activated potassium channel holds the key to the conundrum of familial hypokalemic periodic paralysis

June-Bum Kim, MD, PhD<sup>1</sup>, Sung-Jo Kim, PhD<sup>2</sup>, Sun-Yang Kang, MS<sup>2</sup>, Jin Woong Yi, MD<sup>3</sup>, Seung-Min Kim, PhD<sup>4</sup>

<sup>1</sup>Department of Pediatrics, Hallym University Hwang Sacred Heart Hospital, Seoul, <sup>2</sup>Department of Biotechnology, Hoseo University, Asan, <sup>3</sup>Department of Orthopedic Surgery, Konyang University Hospital, Daejeon, <sup>4</sup>Department of Biological Sciences, Korea Advanced Institute of Science and Technology, Daejeon, Korea

**Purpose:** Familial hypokalemic periodic paralysis (HOKPP) is an autosomal dominant channelopathy characterized by episodic attacks of muscle weakness and hypokalemia. Mutations in the calcium channel gene, *CACNA1S*, or the sodium channel gene, *SCN4A*, have been found to be responsible for HOKPP; however, the mechanism that causes hypokalemia remains to be determined. The aim of this study was to improve the understanding of this mechanism by investigating the expression of calcium-activated potassium ( $K_{Ca}$ ) channel genes in HOKPP patients.

**Methods:** We measured the intracellular calcium concentration with fura-2-acetoxymethyl ester in skeletal muscle cells of HOKPP patients and healthy individuals. We examined the mRNA and protein expression of  $KCa$  channel genes (*KCNMA1*, *KCNN1*, *KCNN2*, *KCNN3*, and *KCNN4*) in both cell types.

**Results:** Patient cells exhibited higher cytosolic calcium levels than normal cells. Quantitative reverse transcription polymerase chain reaction analysis showed that the mRNA levels of the  $K_{Ca}$  channel genes did not significantly differ between patient and normal cells. However, western blot analysis showed that protein levels of the *KCNMA1* gene, which encodes  $K_{Ca}1.1$  channels (also called big potassium channels), were significantly lower in the membrane fraction and higher in the cytosolic fraction of patient cells than normal cells. When patient cells were exposed to 50 mM potassium buffer, which was used to induce depolarization, the altered subcellular distribution of BK channels remained unchanged.

**Conclusion:** These findings suggest a novel mechanism for the development of hypokalemia and paralysis in HOKPP and demonstrate a connection between disease-associated mutations in calcium/sodium channels and pathogenic changes in nonmutant potassium channels.

**Corresponding author:** June-Bum Kim, MD, PhD  
Department of Pediatrics, Hallym University Hwang Sacred Heart Hospital, 12 Beodeunaru-ro 7-gil, Yeongdeungpo-gu, Seoul 150-719, Korea  
Tel: +82-2-2639-5200  
Fax: +82-2-2202-3366  
E-mail: hoppdoctor@hanmail.net

Received: 17 January, 2014

Revised: 27 March, 2014

Accepted: 3 June, 2014

**Key words:** Channelopathies, Hypokalemic periodic paralysis, Potassium channels

## Introduction

Familial hypokalemic periodic paralysis (HOKPP, OMIM no. 170400) is an autosomal dominant disorder that features reversible attacks of flaccid paralysis with concomitant hypokalemia<sup>1)</sup>. The first attack can occur any time between infancy and puberty, with the majority occurring around puberty. Paralytic attacks usually occur at night or early in the morning and last for hours and sometimes days. They can be induced by carbohydrate- or sodium-rich meals, strenuous exercise, emotional stress, and exposure to cold<sup>2)</sup>.

The majority of HOKPP cases are caused by mutations in the skeletal muscle voltage-gated calcium channel gene, *CACNA1S*, or in the sodium channel gene, *SCN4A*<sup>1)</sup>. These

Copyright © 2014 by The Korean Pediatric Society

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

mutations have been shown to contribute to an inward cation leakage current (referred to as the gating-pore current), which makes the muscle fibers of patients susceptible to abnormal depolarization in response to low extracellular potassium concentration<sup>3,4</sup>. Alterations in the expression, subcellular localization, and/or kinetics of nonmutated potassium channels, which reduce outward potassium currents, have been implicated in the development of hypokalemia as well as pathological depolarization<sup>5-8</sup>. However, the reason why potassium channels are affected by mutations in the *CACNA1S* or *SCN4A* gene is not yet clear.

A previous study demonstrated that skeletal muscle fibers from HOKPP patients exhibited increased intracellular calcium levels compared with normal cells<sup>6</sup>. An increase in cytoplasmic calcium levels via its entry and/or release from intracellular stores triggers a variety of physiological processes, such as muscle contraction, neuronal excitability, vasoregulation, hormone secretion, and immune responses<sup>9-12</sup>. The elevated intracellular calcium ions, however, also stimulate the activity of calcium-activated potassium (KCa) channels, of which the large-conductance KCa channels (also termed big potassium [BK] channels) primarily serve as negative feedback regulators that repolarize the cell by increasing potassium efflux. This can lead to membrane hyperpolarization and a consequent decrease in cell excitability<sup>13</sup>.

We conducted this study to test the hypothesis that the expression patterns of the K<sub>ca</sub> channel genes (*KCNMA1*, *KCNN1*, *KCNN2*, *KCNN3*, and *KCNN4*) in the skeletal muscle cells of HOKPP patients differ from those in normal cells and to find a mechanistic link between HOKPP mutant ion channels and pathogenic alterations to non-mutant potassium channels.

## Materials and methods

### 1. Subjects

We reviewed the records of 185 patients who were being treated for HOKPP from March 2008 to April 2013 in the clinics of Seoul Children's Hospital and Konyang University Hospital. Each patient underwent genetic testing. All the patients fulfilled the diagnostic criteria for HOKPP, including (1) episodic attacks of muscle weakness with hypokalemia (<3.5 mEq/L); (2) positive family history or genetically confirmed skeletal calcium or sodium channel mutation; and (3) exclusion of secondary causes of hypokalemia, such as thyroid, adrenal, and renal disorders, and drug abuse<sup>2,14</sup>. Mutation screening was performed by sequencing the entire coding region of the *CACNA1S*, *SCN4A*, and *KCNJ2* genes, as described elsewhere<sup>15,16</sup>. The clinical details of some patients have been reported previously<sup>2,16-18</sup>. Of these,

we selected the three who presented with the most obvious symptoms of HOKPP in terms of both severity and frequency of paralytic attacks. These patients had the Arg1239Gly mutation in the *CACNA1S* gene. Three healthy volunteers acted as controls. All participants provided written informed consent, and the study was conducted in compliance with the guidelines of the Institutional Review Board of Konyang University Hospital.

## 2. Methods

### 1) Skeletal muscle tissue sampling and preparation

Skeletal muscle biopsy samples were obtained as described previously<sup>7</sup>. The muscle biopsy samples were rinsed in phosphate buffered saline (PBS) with Ca<sup>2+</sup>-Mg<sup>2+</sup> and dissected into small pieces (diameter, 1–2 mm) with a sterilized scalpel. The tissue fragments were cultured in Dulbecco's modified Eagle's Medium (DMEM; Thermo Scientific, Marietta, OH, USA) supplemented with 5% fetal bovine serum (FBS; Thermo Scientific), 2μM glutamine (Sigma-Aldrich, St. Louis, MO, USA), 1% nonessential amino acids (Thermo Scientific), 0.1mM β-mercaptoethanol (Sigma-Aldrich), 5-ng/mL fibroblast growth factor-basic recombinant human protein (Invitrogen, Carlsbad, CA, USA), and 1% penicillin-streptomycin (Caisson, Logan, UT, USA) for 2 weeks at 37°C in an incubator containing 95% air and 5% CO<sub>2</sub> (Thermo Scientific). Isolation of skeletal muscle cells from the original culture was performed according to a method described previously<sup>19</sup>.

### 2) Preparation of potassium buffers

Potassium buffer (4mM) at pH 7.35 (4mM KCl, 145mM NaCl, 1mM MgCl<sub>2</sub>, 0.5mM CaCl<sub>2</sub>, 5mM glucose, and 10mM 3-(N-morpholino) propanesulfonic acid [MOPS]) was prepared and used for exposing cells to a normal physiological concentration of extracellular potassium. To induce depolarization of skeletal muscle cells via a high concentration of extracellular potassium, 50 mM potassium buffer (50 mM KCl, 145 mM NaCl, 1 mM MgCl<sub>2</sub>, 0.5 mM CaCl<sub>2</sub>, 5 mM glucose, and 10 mM MOPS; pH 7.35) was prepared. The buffers were sterilized before use.

### 3) Culturing of skeletal muscle cells and treatment with potassium buffer solutions

Skeletal muscle cells obtained from HOKPP patients (patient cells) and healthy controls (normal cells) were cultured in DMEM containing 20% FBS (Thermo Scientific) and 1% penicillin-streptomycin at 37°C in an incubator containing 95% air and 5% CO<sub>2</sub>. To induce differentiation, skeletal muscle cells were incubated in DMEM supplemented with 2% horse serum (Thermo Scientific) and 1% penicillin-streptomycin for up to 5 days. The differentiation medium was replaced every 48 hours. Both normal and patient cells were collected for analysis during the 12th passage. The mRNA and protein levels of the KCa channel genes (*KCNMA1*, *KCNN1*, *KCNN2*, *KCNN3*, and *KCNN4*) were analyzed in both types of cells, prior to and at 1 hour after

exposure to the 4 and 50 mM potassium buffers.

#### 4) Measurement of cytosolic calcium levels

Cytosolic calcium levels were analyzed using fura-2-acetoxymethyl ester (Fura-2AM), a membrane-permeable, calcium-sensitive fluorescent dye (Sigma-Aldrich). Both normal and patient cells were loaded with 1  $\mu$ M Fura-2AM diluted in 4mM potassium buffer for 30 minutes in a CO<sub>2</sub> incubator. After washing with potassium buffer to remove residual dye, the cells were treated with trypsin-ethylenediaminetetraacetic acid (Caisson). They were then harvested, washed with Ca<sup>2+</sup>-free PBS, and analyzed by flow cytometry (Millipore, Billerica, MA, USA). All the experiment protocols were done in triplicate.

#### 5) Quantitative reverse transcription polymerase chain reaction analysis

Total RNA from the skeletal muscle cells was extracted using TRIzol reagent (Invitrogen), and 100 ng was converted to cDNA by using SuperScript III reverse transcriptase (Invitrogen) according to the manufacturer's instructions. AccuPower PCR PreMix (Bioneer, Daejun, Korea) was added to the reaction mix, and quantitative reverse transcription polymerase chain reaction (RT-PCR) analysis was performed using primers for *KCNMA1*, *KCNN1*, *KCNN2*, *KCNN3*, and *KCNN4*. Primers were designed with the VectorNTI10 (Invitrogen) and Primer3 software (<http://sourceforge.net/projects/primer3/>). The sequences of these primers are available on request. The PCR conditions consisted of initial denaturation at 95°C for 5 minutes, followed by 30 cycles at 95°C for 30 seconds, 55°C for 30 seconds, and 72°C for 1 minute, with final extension at 72°C for 5 minutes. All the reactions were performed in triplicate. The mRNA expression levels for each gene were normalized to the expression of glyceraldehyde 3-phosphate dehydrogenase (GAPDH).

#### 6) Western blot analysis

Prior to and after exposure to the two different concentrations of potassium buffers for 1 hour, the cytosolic and membranous protein fractions of normal and patient cells were separated and analyzed by western blotting. Detailed methods for subcellular fractionation and western blot analysis have been previously described<sup>8</sup>). To detect the channel proteins, primary antibodies (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for KCa1.1 (MaxiK $\alpha$  [B-1]), K<sub>Ca</sub>2.1 (SK1 [A-13]), K<sub>Ca</sub>2.2 (KCNN2 [U-24]), K<sub>Ca</sub>2.3 (SK3 [H-45]), K<sub>Ca</sub>3.1 (IK1 [D-5]), and  $\beta$ -actin ( $\beta$ -actin [C4]) were used in combination with anti-goat, antimouse, or antirabbit secondary antibodies. The proteins were then quantified using the TINA 2.0 densitometric analytical system (Raytest Isotopenmeßgeräte GmbH, Straubenhardt, Germany) according to the manufacturer's instructions. All the experiments were performed independently at least three times.

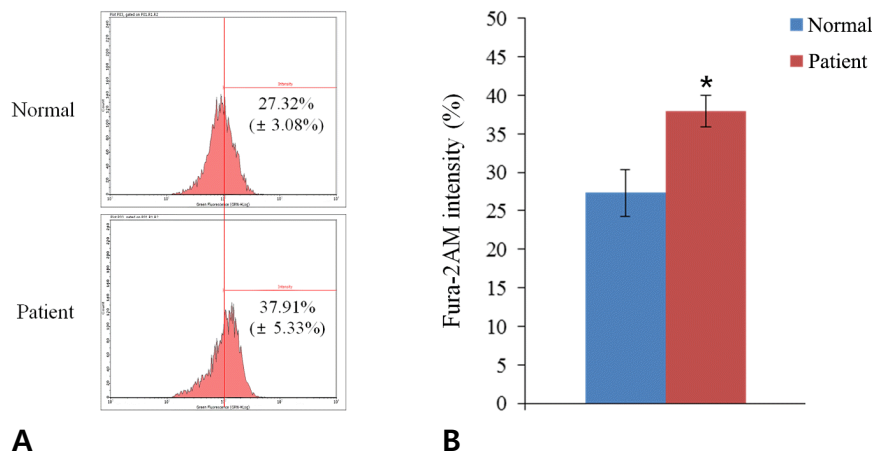
#### 7) Statistical analysis

The IBM SPSS Statistics ver. 19.0 (IBM Co., Armonk, NY, USA) was used for statistical analysis. Data are presented as mean  $\pm$  standard deviation. Comparisons were made using a two-way analysis of variance test with  $P < 0.05$  indicating statistical significance.

## Results

### 1. Cytosolic calcium levels in skeletal muscle cells

We examined cytosolic calcium levels in both patient and normal cells in 4mM potassium buffer using the calcium-sensitive dye Fura-2AM. The percentage of fluorescence was measured by flow cytometry. At least 5,000 cells were analyzed for each sample by using InCite software ver. 2.2.2 (Millipore),



**Fig. 1.** Measurement of cytosolic calcium levels in skeletal muscle cells by fura-2-acetoxymethyl ester staining and flow cytometry. Histograms (A) and bar graph (B) showing that patient cells had a significantly higher concentration of calcium ions in the cytosol than normal cells in 4mM potassium buffer. \* $P < 0.05$  vs. normal samples.

and the threshold was matched at SSC 291. Patient cells exhibited significantly higher levels of cytosolic calcium ions than normal cells ( $37.91\% \pm 5.33\%$  vs.  $27.32\% \pm 3.08\%$ ,  $P < 0.05$ ) (Fig. 1).

## 2. K<sub>Ca</sub> channel mRNA expression

We performed quantitative RT-PCR to compare the mRNA levels for the K<sub>Ca</sub> channel genes (*KCNMA1*, *KCNN1*, *KCNN2*, *KCNN3*, and *KCNN4*) between patient and normal (control) cells in both 4mM and 50mM potassium buffers. No significant differences were found at either potassium concentration (data not shown).

## 3. Western blot analysis to evaluate the expression patterns of K<sub>Ca</sub> channel proteins

We compared the protein expression profiles of the K<sub>Ca</sub> channels in patient versus normal cells, following exposure to 4 and 50mM potassium. Membranous and cytosolic fractions of the cells were separated, and the protein expression levels of the K<sub>Ca</sub> channel genes were evaluated by western blot as mentioned above. No significant difference was observed for K<sub>Ca</sub>2.1, K<sub>Ca</sub>2.2, K<sub>Ca</sub>2.3, and K<sub>Ca</sub>3.1 (data not shown); however, the expression of K<sub>Ca</sub>1.1 (encoded by *KCNMA1*) differed significantly in patient cells: compared with control levels, expression was lower in the membrane fraction ( $P < 0.05$ ) and higher in the cytosolic fraction ( $P < 0.05$ ) for both 4 and 50 mM potassium buffers (Figs. 2, 3).

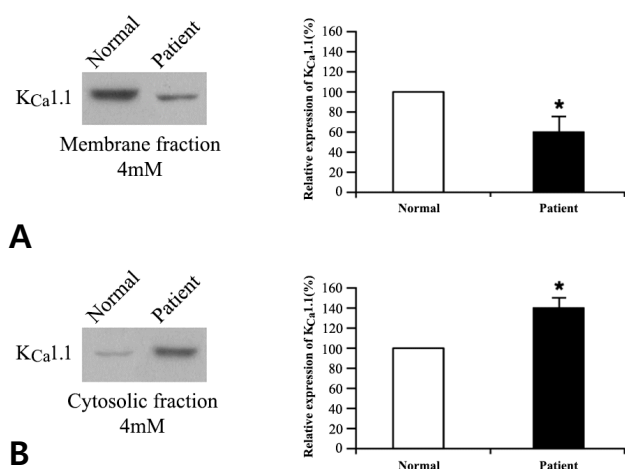
## Discussion

Previous studies have reported alterations in the expression, subcellular localization, and/or kinetics of nonmutant potassium

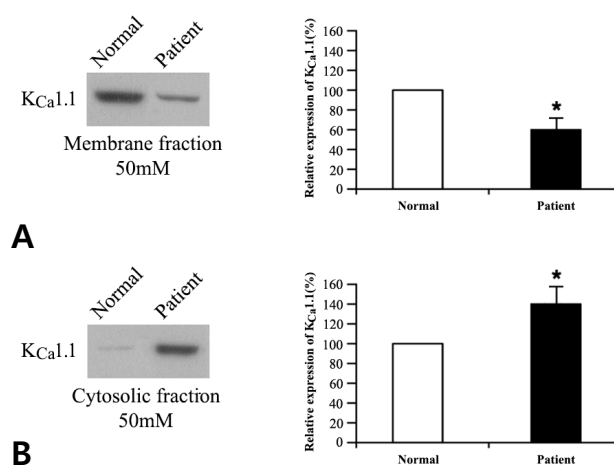
channels (but not BK channels) in HOKPP<sup>5-8</sup>). However, the relationship between mutations in the *CACNA1S* or *SCN4A* gene and the pathological operation of nonmutant potassium channels has not yet been elucidated.

The mutant channels responsible for HOKPP have been shown to generate a nonselective inward cation leakage current through an aberrant gating pore that is open at the resting membrane potential<sup>3,4</sup>). In agreement with the results of Puwanant and Ruff<sup>6</sup>), we observed an increase in the cytoplasmic calcium levels of HOKPP patient muscle cells at a normal physiological concentration of extracellular potassium. Although the precise mechanism responsible for this increase is not known, from the findings of the above studies<sup>3,4</sup>), we can reasonably infer that it occurs via inward calcium leakage through mutant channels and/or calcium release from intracellular stores in response to an inward cation current through the mutant channels at resting potential.

The BK channel exists as a tetrameric structure, composed of four  $\alpha$ -subunits, either alone or associated with  $\beta$ -subunits. The  $\alpha$ -subunit, encoded by *KCNMA1*, is the pore-forming unit, and the  $\beta$ -subunit, encoded by *KCNMB1*, *KCNMB2*, *KCNMB3*, or *KCNMB4*, is the modulatory unit. The different subunit compositions of BK channels give rise to its functional diversity in various tissues; for example, skeletal muscle BK is assembled from  $\alpha$ -subunits alone, whereas vascular BK is composed of  $\alpha$ + $\beta$ 1 and neuronal types are composed of  $\alpha$ + $\beta$ 4 or  $\alpha$ + $\beta$ 2<sup>20-22</sup>). Additional mechanisms involving multiple promoters, alternative splicing, and metabolic regulation also modify BK functional properties, thus producing the diverse range of phenotypes necessary for the normal functioning of many tissues<sup>23-26</sup>). Consistent with the wide distribution of BK channels



**Fig. 2.** Western blot analysis of K<sub>Ca</sub>1.1 protein in the membrane (A) and cytosolic (B) fractions of normal and patient cells in 4mM potassium buffer. Densitometric analysis of K<sub>Ca</sub>1.1 protein is shown on the right. Values are expressed as a percentage of the control (normal cells) level. \* $P < 0.05$  vs. normal samples. K<sub>Ca</sub>, calcium-activated potassium.



**Fig. 3.** Western blot analysis of K<sub>Ca</sub>1.1 protein in the membrane (A) and cytosolic (B) fractions of normal and patient cells in 50 mM potassium buffer. Densitometric analysis of K<sub>Ca</sub>1.1 is shown on the right. Values are expressed as a percentage of the control (normal cells) level. \* $P < 0.05$  vs. normal samples. K<sub>Ca</sub>, calcium-activated potassium.



throughout the human body, malfunction of BK channels has been implicated in various medical conditions, such as epilepsy, psychiatric disorders, deafness, hypertension, asthma, and urinary incontinence<sup>10,27-31</sup>. BK channels play an essential role in regulating both the firing frequency and the action potential repolarization phase of muscle cells. BK channels also serve as molecular targets for drugs used in the treatment of HOKPP: acetazolamide and dichlorphenamide, which act by opening skeletal muscle BK channels<sup>32</sup>.

This is the first study to demonstrate altered subcellular distribution of BK channels in the skeletal muscle cells of patients with HOKPP. Paralysis in HOKPP results from persistent membrane depolarization, and skeletal muscle fibers from patients with HOKPP are extremely susceptible to depolarization-induced inactivation<sup>33</sup>. Given the pivotal role played by BK channels in action potential repolarization by increasing potassium efflux out of cells, the reduced membrane expression of BK channels in patient cells, observed in this study, correlates well with the characteristic pathological conditions of HOKPP, such as hypokalemia and paralysis.

The altered subcellular distribution of BK channels in patient cells in the 4 mM potassium buffer may represent a physiological mechanism developed to compensate for the chronic abnormal increase in cytoplasmic calcium at resting potential. Indeed, several posttranslational modifications have been reported to affect the membrane expression of BK channels<sup>25,26</sup>. On the other hand, it is currently unclear how the altered subcellular distribution of BK channels is maintained in patient cells in the depolarizing 50 mM potassium buffer. Whether alternative splicing of the *KCNMA1* gene or certain posttranslational mechanisms modify the subcellular distribution of BK channels in patient cells remains to be determined.

In conclusion, we observed an abnormal increase in cytoplasmic calcium levels and altered subcellular distribution of BK channels in the skeletal muscle cells of patients with HOKPP. The altered distribution of BK channels represents a novel mechanism linking elevated intracellular calcium, induced by HOKPP-associated mutations, to the hypokalemia and paralysis that is symptomatic of the disease and also demonstrates a connection between HOKPP mutant ion channels and pathogenic changes in nonmutant potassium channels.

## Conflict of interest

No potential conflict of interest relevant to this article was reported.

## Acknowledgments

This work was supported by the 12th Seokcheon Research Grant funded by the Korean Pediatric Society.

## References

1. Kim JB. Channelopathies. Korean J Pediatr 2014;57:1-18.
2. Kim JB, Kim MH, Lee SJ, Kim DJ, Lee BC. The genotype and clinical phenotype of Korean patients with familial hypokalemic periodic paralysis. J Korean Med Sci 2007;22:946-51.
3. Wu F, Mi W, Hernandez-Ochoa EO, Burns DK, Fu Y, Gray HF, et al. A calcium channel mutant mouse model of hypokalemic periodic paralysis. J Clin Invest 2012;122:4580-91.
4. Jurkat-Rott K, Weber MA, Fauler M, Guo XH, Holzherr BD, Paczulla A, et al. K<sup>+</sup>-dependent paradoxical membrane depolarization and Na<sup>+</sup> overload, major and reversible contributors to weakness by ion channel leaks. Proc Natl Acad Sci U S A 2009;106:4036-41.
5. Tricarico D, Mele A, Liss B, Ashcroft FM, Lundquist AL, Desai RR, et al. Reduced expression of Kir6.2/SUR2A subunits explains KATP deficiency in K<sup>+</sup>-depleted rats. Neuromuscul Disord 2008;18:74-80.
6. Puwanant A, Ruff RL. INa and IKir are reduced in Type 1 hypokalemic and thyrotoxic periodic paralysis. Muscle Nerve 2010;42:315-27.
7. Kim SJ, Lee YJ, Kim JB. Reduced expression and abnormal localization of the K(ATP) channel subunit SUR2A in patients with familial hypokalemic periodic paralysis. Biochem Biophys Res Commun 2010;391:974-8.
8. Kim JB, Lee GM, Kim SJ, Yoon DH, Lee YH. Expression patterns of two potassium channel genes in skeletal muscle cells of patients with familial hypokalemic periodic paralysis. Neurol India 2011;59:527-31.
9. Sausbier M, Hu H, Arntz C, Feil S, Kamm S, Adelsberger H, et al. Cerebellar ataxia and Purkinje cell dysfunction caused by Ca<sup>2+</sup>-activated K<sup>+</sup> channel deficiency. Proc Natl Acad Sci U S A 2004;101:9474-8.
10. Brenner R, Perea GJ, Bonev AD, Eckman DM, Kosek JC, Wiler SW, et al. Vasoregulation by the beta1 subunit of the calcium-activated potassium channel. Nature 2000;407:870-6.
11. Van Goor F, Li YX, Stojilkovic SS. Paradoxical role of large-conductance calcium-activated K<sup>+</sup> (BK) channels in controlling action potential-driven Ca<sup>2+</sup> entry in anterior pituitary cells. J Neurosci 2001;21:5902-15.
12. Shaw PJ, Feske S. Physiological and pathophysiological functions of SOCE in the immune system. Front Biosci (Elite Ed) 2012;4:2253-68.
13. Latorre R, Brauchi S. Large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> (BK) channel: activation by Ca<sup>2+</sup> and voltage. Biol Res 2006;39:385-401.
14. Sansone V, Meola G, Links TP, Panzeri M, Rose MR. Treatment for periodic paralysis. Cochrane Database Syst Rev 2008;(1):CD005045.
15. Kim JB, Chung KW. Novel de novo mutation in the KCNJ2 gene in a patient with Andersen-Tawil syndrome. Pediatr Neurol 2009;41:464-6.
16. Park YH, Kim JB. An atypical phenotype of hypokalemic periodic paralysis caused by a mutation in the sodium channel gene SCN4A. Korean J Pediatr 2010;53:909-12.

17. Song YW, Kim SJ, Heo TH, Kim MH, Kim JB. Normokalemic periodic paralysis is not a distinct disease. *Muscle Nerve* 2012;46:914-6.
18. Kil TH, Kim JB. Severe respiratory phenotype caused by a de novo Arg528Gly mutation in the CACNA1S gene in a patient with hypokalemic periodic paralysis. *Eur J Paediatr Neurol* 2010;14:278-81.
19. Dellavalle A, Sampaolesi M, Tonlorenzi R, Tagliafico E, Sacchetti B, Perani L, et al. Pericytes of human skeletal muscle are myogenic precursors distinct from satellite cells. *Nat Cell Biol* 2007;9:255-67.
20. Ghatta S, Nimmagadda D, Xu X, O'Rourke ST. Large-conductance, calcium-activated potassium channels: structural and functional implications. *Pharmacol Ther* 2006;110:103-16.
21. Torres YP, Morera FJ, Carvacho I, Latorre R. A marriage of convenience: beta-subunits and voltage-dependent K<sup>+</sup> channels. *J Biol Chem* 2007;282:24485-9.
22. Dinardo MM, Camerino G, Mele A, Latorre R, Conte Camerino D, Tricarico D. Splicing of the rSlo gene affects the molecular composition and drug response of Ca<sup>2+</sup>-activated K<sup>+</sup> channels in skeletal muscle. *PLoS One* 2012;7:e40235.
23. Bohm RA, Wang B, Brenner R, Atkinson NS. Transcriptional control of Ca<sup>2+</sup>-activated K<sup>+</sup> channel expression: identification of a second, evolutionarily conserved, neuronal promoter. *J Exp Biol* 2000;203(Pt 4):693-704.
24. Adelman JP, Shen KZ, Kavanaugh MP, Warren RA, Wu YN, Lagrutta A, et al. Calcium-activated potassium channels expressed from cloned complementary DNAs. *Neuron* 1992;9:209-16.
25. Jin P, Weiger TM, Levitan IB. Reciprocal modulation between the alpha and beta 4 subunits of hSlo calcium-dependent potassium channels. *J Biol Chem* 2002;277:43724-9.
26. Chen L, Bi D, Tian L, McClafferty H, Steeb F, Ruth P, et al. Palmitoylation of the  $\beta$ 4-subunit regulates surface expression of large conductance calcium-activated potassium channel splice variants. *J Biol Chem* 2013;288:13136-44.
27. Du W, Bautista JF, Yang H, Diez-Sampedro A, You SA, Wang L, et al. Calcium-sensitive potassium channelopathy in human epilepsy and paroxysmal movement disorder. *Nat Genet* 2005;37:733-8.
28. Laumonier F, Roger S, Guerin P, Molinari F, M'rad R, Cahard D, et al. Association of a functional deficit of the BKCa channel, a synaptic regulator of neuronal excitability, with autism and mental retardation. *Am J Psychiatry* 2006;163:1622-9.
29. Pyott SJ, Meredith AL, Fodor AA, Vazquez AE, Yamoah EN, Aldrich RW. Cochlear function in mice lacking the BK channel alpha, beta1, or beta4 subunits. *J Biol Chem* 2007;282:3312-24.
30. Seibold MA, Wang B, Eng C, Kumar G, Beckman KB, Sen S, et al. An african-specific functional polymorphism in KCNMB1 shows sex-specific association with asthma severity. *Hum Mol Genet* 2008;17:2681-90.
31. Meredith AL, Thorneloe KS, Werner ME, Nelson MT, Aldrich RW. Overactive bladder and incontinence in the absence of the BK large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel. *J Biol Chem* 2004;279:36746-52.
32. Tricarico D, Camerino DC. Recent advances in the pathogenesis and drug action in periodic paralyses and related channelopathies. *Front Pharmacol* 2011;2:8.
33. Ruff RL. Insulin acts in hypokalemic periodic paralysis by reducing inward rectifier K<sup>+</sup> current. *Neurology* 1999;53:1556-63.