

GOPEN ACCESS

Citation: Ookawara S, Kaku Y, Ito K, Kizukuri K, Namikawa A, Nakahara S, et al. (2019) Effects of dietary intake and nutritional status on cerebral oxygenation in patients with chronic kidney disease not undergoing dialysis: A cross-sectional study. PLoS ONE 14(10): e0223605. https://doi.org/ 10.1371/journal.pone.0223605

Editor: Tatsuo Shimosawa, International University of Health and Welfare, School of Medicine, JAPAN

Received: July 25, 2019

Accepted: September 24, 2019

Published: October 10, 2019

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: https://doi.org/10.1371/journal.pone.0223605

Copyright: © 2019 Ookawara et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

RESEARCH ARTICLE

Effects of dietary intake and nutritional status on cerebral oxygenation in patients with chronic kidney disease not undergoing dialysis: A cross-sectional study

Susumu Ookawara^{1,2}*, Yoshio Kaku¹, Kiyonori Ito¹, Kanako Kizukuri², Aiko Namikawa², Shinobu Nakahara², Yuko Horiuchi², Nagisa Inose², Mayako Miyahara², Michiko Shiina², Saori Minato¹, Mitsutoshi Shindo¹, Haruhisa Miyazawa¹, Keiji Hirai¹, Taro Hoshino¹, Miho Murakoshi², Kaoru Tabei³, Yoshiyuki Morishita¹

1 Division of Nephrology, First Department of Integrated Medicine, Saitama Medical Center, Jichi Medical University, Saitama, Japan, 2 Department of Nutrition, Saitama Medical Center, Jichi Medical University, Saitama, Japan, 3 Department of Internal Medicine, Minami-uonuma City Hospital, Niigata, Japan

These authors contributed equally to this work.
* su-ooka@hb.tp1.jp

Abstract

Background

Dietary management is highly important for the maintenance of renal function in patients with chronic kidney disease (CKD). Cerebral oxygen saturation (rSO_2) was reportedly associated with the estimated glomerular filtration rate (eGFR) and cognitive function. However, data concerning the association between cerebral rSO_2 and dietary intake of CKD patients is limited.

Methods

This was a single-center observational study. We recruited 67 CKD patients not undergoing dialysis. Cerebral rSO_2 was monitored using the INVOS 5100c oxygen saturation monitor. Energy intake was evaluated by dietitians based on 3-day meal records. Daily protein and salt intakes were calculated from 24-h urine collection.

Results

Multivariable regression analysis showed that cerebral rSO_2 was independently associated with energy intake (standardized coefficient: 0.370) and serum albumin concentration (standardized coefficient: 0.236) in Model 1 using parameters with p < 0.10 in simple linear regression analysis (body mass index, Hb level, serum albumin concentration, salt and energy intake) and confounding factors (eGFR, serum sodium concentration, protein intake), and the energy/salt index (standardized coefficient: 0.343) and Hb level (standardized coefficient: 0.284) in Model 2 using energy/protein index as indicated by energy intake/ protein intake and energy/salt index by energy intake/salt intake in place of salt, protein and energy intake.

Funding: This work was supported by a grant from the Japanese Association of Dialysis Physicians (http://www.touseki-ikai.or.jp/) (JADP Grant 2017-9) and a grant from The Kidney Foundation, Japan (JKFB 17-4) (http://www.jinzouzaidan.or.jp/) to SO. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Conclusions

Cerebral rSO_2 is affected by energy intake, energy/salt index, serum albumin concentration and Hb level. Sufficient energy intake and adequate salt restriction is important to prevent deterioration of cerebral oxygenation, which might contribute to the maintenance of cognitive function in addition to the prevention of renal dysfunction in CKD patients.

Introduction

Diet therapy, including the energy intake management and protein and salt restriction, is a key aspect of chronic kidney disease (CKD) therapy and makes an important contribution to the maintenance of renal function. Several important guidelines have been proposed regarding the dietary intake of CKD patients. In the clinical setting of CKD management in Japan, energy intake is recommended to be within 25–35 kcal/kg ideal body weight (BW) [1–3] and protein intake is recommended to be 0.6–1.0 g/kg ideal BW [1,4–6]. These recommendations differ according to the stage of CKD, and a salt intake of 3–6 g/day is suggested to be ideal [1,7,8]. Low energy intake has been reported to be associated with deterioration of renal function [9,10], and increased salt intake could increase the risk of progression of renal dysfunction in CKD patients [11,12].

Recently, near-infrared spectroscopy (NIRS) has been used as a tool to measure the regional saturation of oxygen (rSO₂), a marker of tissue oxygenation, in order to clarify the influence of CKD progression on cerebral oxygenation in CKD patients receiving hemodialysis (HD) [13–17]. The results of these measurements reflect the status of cognitive impairment because of the relationship of rSO₂ with the Mini-Mental State Examination scores [16] and the Montreal Cognitive Assessment test [17]. Furthermore, cerebral rSO₂ has been shown to decrease with decreasing estimated glomerular filtration rate (eGFR) [17]. Therefore, cerebral rSO₂ may be influenced by the nutritional status of CKD patients, because of the impact of dietary intake on renal function. To date, few reports have investigated the relationship between cerebral oxygenation using NIRS and dietary intake in CKD patients who are not receiving dialysis therapy, and data regarding the association between cerebral rSO₂ and nutritional status of such patients is limited. This study aimed to investigate the influence of dietary intake and nutritional status on the cerebral oxygenation of CKD patients not receiving dialysis therapy.

Materials and methods

Patients

In this single-center observational study, CKD patients who met the following criteria were enrolled: (1) all-stage CKD patients not yet requiring dialysis who were followed up in the Division of Nephrology of our hospital, (2) patients who were older than 20 years, (3) patients who received dietary education and nutritional assessment for CKD management, and (4) patients who underwent 24-hour urine collection for the evaluation of salt and protein intake. Exclusion criteria were the following comorbidities: congestive heart failure, chronic obstructive pulmonary disease, apparent neurological disorder, or chronic hypotension (defined as systolic blood pressure <100 mmHg). Fig 1 shows the flow chart of patient enrollment and analysis.

Sixty-seven patients were included in this study (47 men, 20 women; mean age, 65.6 ± 15.6 years). As shown in Table 1, the numbers of patients at each CKD stage were as follows: G1, 1;



https://doi.org/10.1371/journal.pone.0223605.g001

G2, 1; G3a, 6; G3b, 12; G4, 28; and G5, 19. Causes of chronic renal failure included type 2 diabetes mellitus (32 patients), nephrosclerosis (19 patients), chronic glomerulonephritis (eight patients), and other causes (eight patients). All patients provided written informed consent to participate in this study. This study and its protocols were approved by the Institutional Review Board of Saitama Medical Center, Jichi Medical University, Japan (DAI-RIN 15–104) and conform to the provisions of the Declaration of Helsinki (as revised in Tokyo in 2004).

Evaluation of patient's renal function

For the classification of CKD stages, renal function was evaluated using eGFR based on the serum creatinine concentration (S-Cr), and eGFR was calculated using Eq.1 [18]:

eGFR (mL/min/1.73 m²) =
$$194 \times \text{S}-\text{Cr}^{-1.094} \times \text{age}^{-0.287}$$
 (for men)
eGFR (mL/min/1.73 m²) = $194 \times \text{S}-\text{Cr}^{-1.094} \times \text{age}^{-0.287} \times 0.793$ (for women). (1)

Method of nutritional assessment

Patients included in this study were asked to record the total quantity of food and beverages consumed either by weight or in household measures and to record the methods of food preparation. Energy intake was evaluated by dietitians based on each patient's 3-day meal record using the fifth edition of the Japanese Standard Tables of Food Composition published by the Science and Technology Agency of Japan [19]. Furthermore, 24-h urine collection was performed to enable evaluation of urinary protein excretion (g/day), urinary urea nitrogen

Characteristics	Total patients n = 67
Male/female	47/20 (70/30)
Cerebral rSO ₂ (%)	55.9 ± 6.6
CKD stages G1/2/3a/3b/4/5	1 (1)/1 (1)/6 (9)/12 (18)/28 (42)/19 (28)
Disease	
Diabetes mellitus	32 (48)
Nephrosclerosis	19 (28)
Chronic glomerulonephritis	8 (12)
Others	8 (12)
Antihypertensive medication	
Renin-angiotensin system blocker	41 (61.2)
Calcium channel blocker	41 (61.2)
Beta blocker	22 (32.8)
Diuretics (loop and/or thiazide)	23 (34.3)
Antidiabetic medication	
Insulin agent	9 (13.4)
Dipeptidyl peptidase-4 inhibitor	17 (25.4)
Insulin secretagogue	4 (6.0)
α-glucosidase inhibitor	3 (4.5)
Thiazolidinedione	3 (4.5)
Sodium-glucose cotransporter-2 inhibitor	3 (4.5)
Others	
Vitamin D analog	10 (14.9)
Phosphate binder	6 (9.0)
Statin	21 (31.3)
Antiplatelet agents	19 (28.4)
Erythropoiesis-stimulating agent	19 (28.4)

Table 1. Patient characteristics.

Categorical data are presented as number (%), continuous data are presented as mean ± standard deviation. Abbreviations: CKD, chronic kidney disease; rSO₂, regional oxygen saturation.

https://doi.org/10.1371/journal.pone.0223605.t001

(UUN) excretion, and urinary Na⁺ excretion. The urine collection method was as follows: collection was started in the morning after the first morning urine was discarded. Thereafter, the entire volume of urine was collected in a disposable 3L container. To avoid the possibility of inadequate urine collection, we trained all patients to properly collect their urine samples and emphasized that collection must be initiated at a specific time and completed at the same time the next day. Daily protein and salt intakes were calculated based on the UUN and urinary Na⁺ excretion values obtained from the 24-h urine collection.

Protein intake was calculated using Maroni's equation [20], as described in Eq 2:

Protein intake
$$(g/kg \text{ ideal } BW/day) = (BW (kg) \times 0.031 + UUN (g/day)) \times 6.25 \div \text{ ideal } BW (kg).$$
 (2)

Salt intake was calculated using Eq 3:

Salt intake
$$(g/day) =$$
 urinary Na⁺ excretion $(mEq/day) \div 17$ (3)

Furthermore, dietary education was provided by a dietician according to the protocols for nutritional management for CKD therapy in Japan; specifically, sufficient energy intake (25–35 kcal/kg ideal BW/day), protein restriction (0.6–1.0 g/kg ideal BW/day), and salt restriction (3–6 g/day) [1]. To evaluate the influence of energy intake, protein restriction, and salt restriction on cerebral oxygenation, we calculated the nutritional markers described in Eqs 4 and 5:

$$\frac{\text{Energy/protein index (kcal/kg ideal BW/g protein)} = \text{Energy intake}}{(kcal/kg ideal BW/day) \div \text{Protein intake (g/day)}}$$
(4)

 $\frac{\text{Energy/salt index (kcal/kg ideal BW/g salt)} = \text{Energy intake}}{(kcal/kg ideal BW/day) \div \text{ salt intake (g/day)}}$ (5)

Cerebral oxygenation monitoring and clinical laboratory measurements

Cerebral rSO₂ was monitored using an INVOS 5100c saturation monitor (Covidien Japan, Tokyo, Japan), which utilizes NIRS technology. This instrument uses a light-emitting diode, which transmits near-infrared light at two wavelengths (735 and 810 nm), and two silicon photodiodes, which act as light detectors to measure oxygenated and deoxygenated hemoglobin (Hb). The ratio of the oxygenated to total Hb (i.e., oxygenated Hb + deoxygenated Hb) signal strength was read as a single numerical value that represents rSO_2 [21,22], and all data were immediately and automatically stored in sequence. The inter-observer variance for this instrument; namely, the reproducibility of the rSO₂ measurement, has been reported to be acceptable [23-25]. Therefore, rSO₂ is considered a reliable indicator for the estimation of actual cerebral oxygenation. Furthermore, the light paths leading from the emitter to the different detectors share a common part; the 30-mm detector assesses superficial tissues, while the 40-mm detector is used to assess deep tissues. By analyzing the differential signals recorded by the two detectors, the data for cerebral rSO_2 can be supposed to be obtained from deep tissue, 20-30 mm from the body's surface [26,27]. Before measurement, patients were asked to sit in the chair for at least 5 min, and an rSO₂ measurement sensor was attached to the patient's forehead. Thereafter, rSO₂ was measured at 6-s intervals for 5 min, and the mean value calculated. Blood and urinary samples were also obtained from each patient under ambient conditions. This measurement was performed approximately from 2 h to 4 h after each meal for each patient.

Clinical parameters including Hb, serum creatinine, sodium, potassium, chloride, total protein, serum albumin, urinary protein, urinary urea nitrogen, and urinary sodium concentration were measured in our hospital laboratory.

Statistics

Data are expressed as mean \pm standard deviation or median (interquartile range) as appropriate. Urinary protein excretion did not show normal distribution, and this variable was transformed using the natural log (ln). Correlations between cerebral rSO₂ and each clinical parameter, including nutritional parameters, were evaluated using Pearson's correlation coefficient and linear regression analysis. Variables with a p value below 0.10 in simple linear regression analysis and plausible confounding factors were included in multivariable linear regression analysis to identify factors affecting cerebral rSO₂ in CKD patients. Statistical significance was accepted at p < 0.05. All analyses were performed using SPSS Statistics for Windows, version 19.0 (IBM Corp., NY, USA).

Results

The mean cerebral rSO₂ values of the CKD patients in this study were 55.9 \pm 6.6%, and these were significantly positively correlated with Hb level, serum albumin concentration, energy intake, and energy/salt index. Cerebral rSO₂ was negatively correlated with body mass index (Table 2). Cerebral rSO₂ was negatively correlated with salt intake (r = -0.228, p = 0.064) and positively correlated with energy/protein index (r = 0.203, p = 0.099), although these correlations were not significant. Fig 2 illustrates the significant correlation between cerebral rSO₂ and energy intake (r = 0.388, p = 0.001).

Results of multivariable linear regression analysis are presented in Tables 3 and 4. For Model 1; body mass index, Hb level, serum albumin concentration, salt and energy intake as variables with p values below 0.10, as well as eGFR, serum sodium concentration, and protein intake as confounding factors, were included in multivariable linear regression analysis. As shown in <u>Table 3</u>, cerebral rSO₂ was independently associated with energy intake (standardized coefficient: 0.370) and serum albumin concentration (standardized coefficient: 0.236). The energy/protein index and energy/salt index were included in place of salt, protein, and energy intake as variables in Model 2 to avoid collinearity with Model 1. As a result, energy/

Characteristics	Total patients n = 67	vs. cerebral rS simple linear	O ₂ values in regression
		r	<i>p</i> value
Age (years)	65.6 ± 15.6	-0.119	0.338
Body mass index (kg/m ²)	24.8 ± 5.2	-0.245	0.045 *
Systolic blood pressure (mmHg)	138 ± 18	-0.037	0.764
Diastolic blood pressure (mmHg)	77 ± 14	0.059	0.633
Sat O ₂ (%)	97.9 ± 0.7	-0.006	0.961
Laboratory findings			
Hb (g/dL)	11.9 ± 1.8	0.271	0.027 *
eGFR (mL/min/1.73m ²)	25.5 ± 17.1	0.201	0.104
Na (mEq/L)	139 ± 3	-0.006	0.963
K (mEq/L)	4.7 ± 0.6	0.065	0.602
Cl (mEq/L)	107 ± 4	-0.130	0.296
Total protein (g/dL)	7.0 ± 0.6	-0.010	0.938
Serum albumin (g/dL)	3.9 ± 0.4	0.264	0.031 *
Urinary protein excretion (g/g-Cr)	1.0 (0.2–1.2)		
ln (urinary protein excretion)	-0.8 ± 1.4	-0.125	0.314
Nutritional markers	·		
Energy intake (kcal/kg ideal BW/day)	27.0 ± 4.2	0.388	0.001 *
Protein intake (g/ kg ideal BW/day)	0.8 ± 0.2	-0.036	0.775
Salt intake (g/day)	6.3 ± 2.3	-0.228	0.064
Energy/protein index (kcal/kg ideal BW/g-protein)	0.7 ± 0.2	0.203	0.099
Energy/salt index (kcal/kg ideal BW/g-salt)	4.9 ± 2.1	0.332	0.006 *

Table 2. Correlation between cerebral oxygen saturation and clinical parameters, including dietary intake and nutritional parameters, in simple linear regression analysis.

Continuous data are presented as mean ± standard deviation.

*Statistically significant.

Abbreviations: BW, body weight; eGFR, estimated glomerular filtration rate; Hb, hemoglobin; rSO₂, regional oxygen saturation.

https://doi.org/10.1371/journal.pone.0223605.t002



Energy intake (kcal/kg ideal BW/day)

Fig 2. Correlation between cerebral oxygen saturation and energy intake in advanced chronic kidney disease patients. Equation of trend line (representing cerebral oxygen saturation) = $0.614 \times$ energy intake + 39.1; r = 0.388, p = 0.001. Abbreviations: BW, body weight; rSO₂, regional saturation of oxygen.

https://doi.org/10.1371/journal.pone.0223605.g002

salt index (standardized coefficient: 0.343) and Hb level (standardized coefficient: 0.284) were also identified as factors affecting cerebral rSO_2 in this study (Table 4).

Discussion

The present study focused on the association between cerebral oxygenation and nutritional status including indices of dietary intake in CKD patients who were not receiving dialysis.

Table 3. Multivariable linear regression analysis in Model 1 using variables including salt, protein, and energy			
intake as a nutritional marker: independent factors of cerebral oxygen saturation.			
	Multivariable linear regression		

	Multivariable linear regression		
vs. cerebral rSO ₂	Standardized coefficient	p value	
Body mass index	-0.152	0.201	
Hb	0.205	0.078	
eGFR	0.179	0.118	
Na	0.052	0.659	
Serum albumin	0.236	0.039 *	
Salt intake	-0.166	0.155	
Protein intake	0.011	0.923	
Energy intake	0.370	0.002 *	

*Statistically significant.

Abbreviations: eGFR, estimated glomerular filtration rate

Hb, hemoglobin; rSO₂, regional oxygen saturation.

https://doi.org/10.1371/journal.pone.0223605.t003

	Multivariable linear regression		
vs. cerebral rSO ₂	Standardized coefficient	p value	
Body mass index	-0.144	0.228	
Hb	0.284	0.014 *	
eGFR	0.121	0.417	
Na	0.069	0.560	
Serum albumin	0.191	0.128	
Energy/protein index	0.115	0.409	
Energy/salt index	0.343	0.003 *	

Table 4. Multivariable linear regression analysis in Model 2 using variables including energy/protein index and energy/salt index as a nutritional marker: independent factors of cerebral oxygen saturation.

*Statistically significant.

Abbreviations: eGFR, estimated glomerular filtration rate

Hb, hemoglobin; rSO₂, regional oxygen saturation.

https://doi.org/10.1371/journal.pone.0223605.t004

These results confirmed that cerebral rSO_2 levels are independently associated with energy intake and serum albumin concentration in Model 1 and with energy/salt index and Hb level in Model 2.

It has previously been reported that cerebral rSO_2 values of healthy individuals are nearly 70%, whereas those in patients undergoing HD are lower at around 50% [14,15]. Furthermore, cerebral rSO_2 values have been shown to decrease according to the progression of renal dysfunction [17]. In this study, cerebral rSO_2 values were found to lie between those of healthy individuals and patients undergoing HD, consistent with the previous report [17].

In both models for determination of modifiable factors independently associated with cerebral rSO₂, energy intake was found to be the most important factor. Adequate dietary intake and nutritional status have well-understood impacts on brain functions, and the mechanisms involved in the transfer of energy from foods to neurons are likely to be fundamental to the control of brain function [28]. Therefore, the effect of energy intake on cerebral oxygenation might be explained by the fact that this factor is essential for the maintenance of brain function via the energy supply to brain tissues, including cerebral microcirculation. Furthermore, it has been recently reported that the brain-gut axis is very important in the control of dietary intake [29]. Ghrelin, which is secreted primarily by epithelial cells of the stomach, stimulates food intake and is strongly associated with the regulation of energy homeostasis [30,31]. In addition, beneficial effects on vascular function and cardiovascular disease have been reported in response to ghrelin, via the stimulation of nitric oxide production and prevention of endothelial cell apoptosis [32-35]. Ghrelin might, therefore, play an important role in the maintenance of microcirculation and oxygenation in systemic tissues. The changes that occur in circulating ghrelin levels in the case of CKD and the effects of ghrelin in this context remain controversial [36,37]. However, the administration of ghrelin to patients with advanced CKD undergoing dialysis leads to increased appetite and food intake and consequent changes in energy balance [38,39]. Based on these results, ghrelin might simultaneously influence energy intake and systemic oxygenation status, including that of the brain, via the regulation of energy homeostasis and prevention of microcirculation impairment, even in patients with advanced CKD. The results presented here of the significant and positive association between cerebral rSO₂ and energy intake may therefore reflect the influence of the brain-gut axis, including the effects of ghrelin. However, the effects of ghrelin were not directly investigated in this study; therefore, we cannot comment on the association between cerebral oxygenation, energy intake, and the effects of ghrelin.

Salt intake has previously been reported to be associated with the progression of renal dysfunction [11,12] and cerebrovascular disease including cognitive impairments [40,41]. Recently, studies in mice have shown that high salt diets induce marked cerebral hypoperfusion and deterioration of cerebral microcirculation associated with endothelial dysregulation via the suppression of endothelial nitric oxide. This suppression was dependent on the high salt diet-induced interleukin-17 response [42], and changes in cerebral blood flow that are affected by salt intake are proposed as a new brain-gut axis. Therefore, according to the degree of increase in salt intake, cerebral oxygenation could be expected to worsen due to decreased oxygen supply induced by the deterioration of cerebral microcirculation. In this study, the mean salt intake was found to be 6.3 ± 2.3 g/day (ranging from 2.6-14.0 g/day), even after dietary education was provided, and was negatively correlated with cerebral rSO₂. Furthermore, a significant association between cerebral rSO₂ and energy/salt index was confirmed. Based on this result, salt restriction might be an approach to maintain cerebral oxygenation in addition to sufficient energy intake in the clinical setting. However, this study could not determine a significant relationship between salt intake and cerebral rSO₂ values; therefore, further study is needed to confirm the effect of salt intake on cerebral oxygenation and microcirculation.

Regarding the association between cerebral rSO_2 and nutritional parameters in this study, serum albumin concentration and Hb level were significantly associated with cerebral rSO₂ in multivariate linear regression analysis. Serum albumin concentration, the main determinant of colloid osmotic pressure in vessels, plays an important role in maintaining microcirculation in systemic tissues via the movement of body fluids, mainly between the vessels and interstitium [43]. Furthermore, consistent with the present study, serum albumin concentration has been reported to be significantly associated with cerebral oxygenation in patients with all stages of CKD, as well as patients undergoing HD [15,17]. In addition, Hb is an important factor in oxygen supply to the peripheral tissues and organs, including the brain; therefore, Hb level is expected to be associated with tissue rSO₂. Thus far, in various clinical settings including hematology [44], surgery [45], pediatrics [46–48], and HD therapy [49], cerebral rSO₂ has been shown to significantly increase in line with the increasing Hb levels following blood transfusion. On the other hand, it has been reported that there is no relationship between Hb concentration and cerebral rSO₂ values in HD patients with well-maintained Hb levels [15,17]. In this study, it is likely that Hb levels were well-maintained (the mean value was found to be 11.9 ± 1.8 g/dL); however, the values were widely distributed, from 7.1–16.0 g/dL. This study might, therefore, confirm the association between cerebral rSO₂ and Hb levels, because the wide distribution of cerebral rSO₂ values reflects the wide distribution of Hb levels.

This study had several limitations which should be noted. First, it was limited by its relatively small sample size. Second, examination of the relationship of cerebral oxygenation with cognitive function could be considered to be important. However; in this study, cognitive assessment could not be performed because of the limits of the medical examination time for each patient. Thus, we cannot comment on the association between cerebral oxygenation and cognitive function at the present time. Third, in this study, salt intake was calculated using urinary Na⁺ excretion based on the 24-h urine collection for each patient. These values were positively correlated to those calculated in each patient's 3-day meal record (salt intake based on the 24-h urine collection: 6.3 ± 2.3 g/day vs salt intake based on each patient's 3-day meal record: 6.1 ± 1.6 g/day, r = 0.719, p< 0.001). However, due to fluctuations in daily salt intake, the values based on the 24-h urine collection may not fully reflect the constant daily salt intake for each patient. Finally, no relationships were detected between cerebral oxygenation and markers of renal function, although cerebral rSO₂ has been reportedly to be associated with eGFR in patients with all stages of CKD [17]. The patients included in this study mainly suffered from severe advanced CKD, and those with CKD stage 4 or 5 represented around 70% of

the cohort (47 out of 67 included patients). This proportion is significantly different to that of the previous report (40% of the study population had CKD stage 4 or 5) [17]. This might be one of the reasons for the different observations of cerebral oxygenation with regards to renal function; however, the precise reason remains unclear. Therefore, additional studies are needed to confirm the association between cerebral oxygenation and clinical parameters including dietary intake and nutritional parameters, in addition to the examination of cognitive function.

In conclusion, cerebral rSO_2 is affected by energy intake and the energy/salt index in addition to serum albumin concentrations and Hb levels. Therefore, sufficient energy intake with adequate salt restriction is important to prevent the deterioration of cerebral oxygenation and might contribute to the maintenance of cognitive function in addition to the prevention of renal dysfunction in CKD patients.

Acknowledgments

We would like to thank the study participants and our hospital's staff in the Department of Nutrition.

This work was supported by a grant from the Japanese Association of Dialysis Physicians (JADP Grant 2017–9) and a grant from The Kidney Foundation, Japan (JKFB 17–4) to SO.

Author Contributions

Conceptualization: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito.

Data curation: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito.

Formal analysis: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito.

Funding acquisition: Susumu Ookawara.

- **Investigation:** Susumu Ookawara, Yoshio Kaku, Kiyonori Ito, Kanako Kizukuri, Aiko Namikawa, Shinobu Nakahara, Yuko Horiuchi, Nagisa Inose, Mayako Miyahara, Michiko Shiina, Saori Minato, Mitsutoshi Shindo, Haruhisa Miyazawa, Keiji Hirai, Taro Hoshino.
- Methodology: Susumu Ookawara, Yoshio Kaku.
- Project administration: Susumu Ookawara.
- Supervision: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito, Miho Murakoshi, Kaoru Tabei, Yoshiyuki Morishita.

Validation: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito.

Writing - original draft: Susumu Ookawara, Yoshio Kaku, Kiyonori Ito.

References

- 1. Japanese Society of Nephrology, eds. *Clinical Practice Guidebook for Diagnosis and Treatment of Chronic Kidney Disease 2012.* Tokyo, Japan: Tokyo-Igakusya; 2012.
- Kopple JD. National kidney foundation K/DOQI clinical practice guidelines for nutrition in chronic renal failure. Am J Kidney Dis. 2001; 37(1 Suppl 2): S66–70.
- Academy of Nutrition and Dietetics. Evidence-based nutrition practice guidelines. Chronic kidney disease (CKD). 2010. (https://www.andeal.org/template.cfm?template=guide_summary&key=2410&highlight=CKD%20energy&home=1#supportevidence).
- Fouque D, Laville M, Boissel JP. Low protein diets for chronic kidney disease in non diabetic adults. Cochrane Database Syst Rev. 2009; CD001892. https://doi.org/10.1002/14651858.CD001892.pub3 PMID: 19588328

- Robertson L, Waugh N, Robertson A. Protein restriction for diabetic renal disease. Cochrane Database Syst Rev. 2007; CD002181. https://doi.org/10.1002/14651858.CD002181.pub2 PMID: 17943769
- Kidney disease: Improving global outcomes (KDOQI) CKD work group. KDIGO 2012 Clinical practice guideline for the evaluation and management of chronic kidney disease. Kidney Int Suppl. 2013; 3: 1– 150.
- Swift PA, Markandu ND, Sagnella GA, He FJ, MacGregor GA. Modest salt reduction reduces blood pressure and urine protein excretion in black hypertensives: a randomized control trial. Hypertension. 2005; 46: 308–312. https://doi.org/10.1161/01.HYP.0000172662.12480.7f PMID: 15983240
- Slagman MCJ, Waanders F, Hemmelder MH, Woittiez AJ, Janssen WM, Lambers Heerspink HJ, et al. Moderate dietary sodium restriction added to angiotensin converting enzyme inhibition compared with dual blockade in lowering proteinuria and blood pressure: randomized controlled trial. BMJ. 2011; 343: d4366. https://doi.org/10.1136/bmj.d4366 PMID: 21791491
- Duenhas MR, Draibe SA, Avesani CM, Sesso R, Cuppari L. Influence of renal function on spontaneous dietary intake and on nutritional status of chronic renal insufficiency patients. Eur J Clin Nutr. 2003; 57: 1473–1478. https://doi.org/10.1038/sj.ejcn.1601713 PMID: 14576761
- Huang MC, Chen ME, Hung HC, Chen HC, Chang WT, Lee CH, et al. Inadequate energy and excess protein intakes may be associated with worsening renal function in chronic kidney disease. J Ren Nutr. 2008; 18: 187–194. https://doi.org/10.1053/j.jrn.2007.08.003 PMID: 18267211
- Lin J, Hu FB, Curhan GC. Association of diet with albuminuria and kidney function decline. Clin J Am Soc Nephrol. 2010; 5: 836–843. https://doi.org/10.2215/CJN.08001109 PMID: 20299364
- Vegter S, Perna A, Postma MJ, Navis G, Remuzzi G, Ruggenenti P. Sodium intake, ACE inhibition, and progression to ESRD. J Am Soc Nephrol. 2012; 23: 165–173. https://doi.org/10.1681/ASN. 2011040430 PMID: 22135311
- Prohovnik I, Post J, Uribarri J, Lee H, Sandu O, Langhoff E. Cerebrovascular effects of hemodialysis in chronic kidney disease. J Cereb Blood Flow Metab. 2007; 27: 1861–1869. https://doi.org/10.1038/sj. jcbfm.9600478 PMID: 17406658
- Hoshino T, Ookawara S, Goto S, Miyazawa H, Ito K, Ueda Y, et al. Evaluation of cerebral oxygenation in patients undergoing long-term hemodialysis. Nephron Clin Pract. 2014; 126: 57–61. <u>https://doi.org/ 10.1159/000358432</u> PMID: 24526002
- Ito K, Ookawara S, Ueda Y, Goto S, Miyazawa H, Yamada H, et al. Factors affecting cerebral oxygenation in hemodialysis patients: cerebral oxygenation associates with pH, hemodialysis duration, serum albumin concentration, and diabetes mellitus. PLoS One. 2015; 10: e0117474. <u>https://doi.org/10.1371/</u> journal.pone.0117474 PMID: 25706868
- Kovarova L, Valerianova A, Kmentova T, Lachmanova J, Hladinova Z, Malik J. Low cerebral oxygenation Is associated with cognitive impairment in chronic hemodialysis patients. Nephron. 2018; 139: 113–119. https://doi.org/10.1159/000487092 PMID: 29439251
- Miyazawa H, Ookawara S, Ito K, Ueda Y, Yanai K, Ishii H, et al. Association of cerebral oxygenation with estimated glomerular filtration rate and cognitive function in chronic kidney disease patients without dialysis therapy. PLoS One. 2018; 13: e0199366. https://doi.org/10.1371/journal.pone.0199366 PMID: 29940017
- Matsuo S, Imai E, Horio M, Yasuda Y, Tomino K, Nitta K, et al. Revised equations for estimated GFR from serum creatinine in Japan. Am J Kidney Dis. 2009; 53: 982–992. https://doi.org/10.1053/j.ajkd. 2008.12.034 PMID: 19339088
- **19.** Resource Council, eds. *Standard Tables of Food Composition in Japan*, 5st ed. Tokyo, Japan: Resource Council, Science and Technology Agency; 2012. (in Japanese).
- Maroni BJ, Steinman TI, Mitch WE. A method for estimating nitrogen intake of patients with chronic renal failure. Kidney Int. 1985; 27: 58–65 https://doi.org/10.1038/ki.1985.10 PMID: 3981873
- Tobias JD. Cerebral oxygenation monitoring: near-infrared spectroscopy. Expert Rev Med Devices. 2006; 3: 235–243. https://doi.org/10.1586/17434440.3.2.235 PMID: 16515389
- 22. Ferrari M, Mottola L, Quaresima V. Principles, techniques, and limitations of near infrared spectroscopy. Can J Appl Physiol. 2004; 29: 463–487. PMID: 15328595
- Lemmers PMA, Toet MC, van Bel F. Impact of patent ductus arteriosus and subsequent therapy with indomethacin on cerebral oxygenation in preterm infants. Pediatrics. 2008; 121: 142–147. https://doi. org/10.1542/peds.2007-0925 PMID: 18166568
- Hyttel-Sorensen S, Sorensen LC, Riera J, Greisen G. Tissue oximetry: a comparison of mean values of regional tissue saturation, reproducibility and dynamic range of four NIRS-instruments on the human forearm. Biomed Opt Express. 2011; 2: 3047–3057. https://doi.org/10.1364/BOE.2.003047 PMID: 22076266

- 25. Schmitz J, Pichler G, Schwaberger B, Urlesberger B, Baik N, Binder C. Feasibility of long-term cerebral and peripheral regional tissue oxygen saturation measurements. Physiol Meas. 2014; 35: 1349–1355. https://doi.org/10.1088/0967-3334/35/7/1349 PMID: 24854420
- Hongo K, Kobayashi S, Okudera H, Hokama M, Nakagawa F. Noninvasive cerebral optical spectroscopy: depth-resolved measurements of cerebral haemodynamics using indocyanine green. Neuro Res. 1995; 17: 89–93.
- Maslehaty H, Krause-Tilz U, Petridis AK, Barth H, Mehdorn HM. Continuous measurement of cerebral oxygenation with near-infrared spectroscopy after spontaneous subarachnoid hemorrhage. ISRN Neurol. 2012; 907187. https://doi.org/10.5402/2012/907187 PMID: 23209938
- Gomez-Pinilla F. Brain foods: the effect of nutrients on brain function. Nat Rev Neurosci. 2008; 9: 568– 578. https://doi.org/10.1038/nrn2421 PMID: 18568016
- Konturek SJ, Konturek JW, Pawlik T, Brzozowki T. Brain-gut axis and its role in the control of food intake. J Physiol Pharmacol. 2004; 55: 137–154. PMID: 15082874
- Meier U, Gressner AM. Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin, and resistin. Clin Chem. 2004; 50: 1511–1525. https://doi.org/10.1373/clinchem.2004.032482 PMID: 15265818
- Gualillo O, Laga F, Gomez-Reino J, Casanueva FF, Diguez C. Ghrelin, a widespread hormone: insights into molecular and cellular regulation of its expression and metabolism of action. FEBS Lett. 2003; 552: 105–109. https://doi.org/10.1016/s0014-5793(03)00965-7 PMID: 14527669
- Teauro M, Schinzar F, Caramanti M, Lauro R, Cardillo C. Cardiovascular and metabolic effects of ghrelin. Curr. Diabetes Rev. 2010; 6: 228–235. PMID: 20459393
- Virdis A, Lerman LO, Regoli F, Ghiadoni L, Lerman A. Human ghrelin: A gastric hormone with cardiovascular properties. Curr Pharm Des. 2016; 22: 52–58. <u>https://doi.org/10.2174/</u> 1381612822666151119144458 PMID: 26581223
- Wang Y, Narsinh K, Zhao L, Sun D, Wang D, Zhang Z, et al. Effects and mechanisms of ghrelin on cardiac microvascular endothelial cells in rats. Cell Biol Int. 2011; 35: 135–140. https://doi.org/10.1042/ CBI20100139 PMID: 20843299
- Zhao H, Liu G, Wang Q, Ding L, Cai H, Jiang H, et al. Effect of ghrelin on human endothelial cells apoptosis induced by high glucose. Biochem Biophys Res Commun. 2007; 362: 677–681. https://doi.org/10.1016/j.bbrc.2007.08.021 PMID: 17719561
- 36. Cheung WW, Mak RH. Ghrelin in chronic kidney disease. Int J Pept. 2010; pii: 567343.
- Gupta RK, Kuppusamy TK, Patrie JT, Gaylinn B, Thomer MO, Bolton WK. Association of plasma desacyl ghrelin levels with CKD. Clin J Am Nephrol. 2013; 8: 1098–1105.
- Wynne K, Giannitsopoulou K, Small CJ, Patterson M, Frost G, Ghatei MA, et al. Subcutaneous ghrelin enhances acute food intake in malnourished patients who receive maintenance peritoneal dialysis: A randomized, placebo-controlled trial. J Am Soc Nephrol. 2005; 16: 2111–2118. <u>https://doi.org/10.1681/</u> ASN.2005010039 PMID: 15888560
- Ashby DR, Ford HE, Wynne KJ, Wren AM, Murphy KG, Busbridge M, et al. Sustained appetite improvement in malnourished dialysis patients by daily ghrelin treatment. Kidney Int. 2009; 76: 199–206. https://doi.org/10.1038/ki.2009.114 PMID: 19387475
- Heye AK, Thrippleton MJ, Chappell FM, Hemandez Mdel C, Armitage PA, Makin SD, et al. Blood pressure and sodium: association with MRI markers in cerebral small vessel disease. J Cereb Blood Flow Metab. 2016; 36: 264–274. https://doi.org/10.1038/jcbfm.2015.64 PMID: 25899292
- Strazzullo P, D'Elia L, Kandala NB, Cappucio EP. Salt intake, stroke, and cardiovascular disease: meta-analysis of prospective studies. BMJ. 2009; 339: b4567. https://doi.org/10.1136/bmj.b4567 PMID: 19934192
- Faraco G, Brea D, Garcia-Bonilla L, Wang G, Racchumi G, Chang H, et al. Dietary salt promotes neurovascular and cognitive dysfunction through a gut-initiated TH17 response. Nat Neurosci. 2018; 21: 240–249. https://doi.org/10.1038/s41593-017-0059-z PMID: 29335605
- Ookawara S, Sato H, Takeda H, Tabei K. Methods for approximating colloid osmotic pressure in longterm hemodialysis patients. Ther Apher Dial. 2014; 18: 202–207. https://doi.org/10.1111/1744-9987. 12070 PMID: 24720412
- Yuruk K, Bartels SA, Milstein DM, Bezemer R, Biemond BJ, Ince C. Red blood cell transfusions and tissue oxygenation in anemic hematology outpatients. Transfusion. 2012; 52: 641–646. <u>https://doi.org/ 10.1111/j.1537-2995.2011.03312.x PMID: 21883269</u>
- 45. Torella F, Haynes SL, McCollum CN. Cerebral and peripheral oxygen saturation during red cell transfusion. J Surg Res. 2003; 110: 217–221. https://doi.org/10.1016/s0022-4804(03)00037-4 PMID: 12697404

- 46. van Hoften JC, Verhagen EA, Keating P, ter Horst HJ, Bos AF. Cerebral tissue oxygen saturation and extraction in preterm infants before and after blood transfusion. Arch Dis Child Fetal Neonatal Ed. 2010; 95: F352–F358. https://doi.org/10.1136/adc.2009.163592 PMID: 20466739
- Dhabangi A, Ainomugisha B, Cserti-Gazdewich C, Ddungu H, Kyeyune D, Musisi E, et al. Cerebral oximetry in Ugandan children with severe anemia: Clinical categories and response to transfusion. JAMA Pediatr. 2016; 170: 995–1002. <u>https://doi.org/10.1001/jamapediatrics.2016.1254</u> PMID: 27532507
- Neunhoeffer F, Hofbeck M, Schuhmann MU, Fuchs J, Schlensak C, Esslinger M, et al. Cerebral oxygen metabolism before and after RBC transfusion in infants following major surgical procedures. Pediatr Crit Care Med. 2018; 19: 318–327. https://doi.org/10.1097/PCC.00000000001483 PMID: 29406374
- **49.** Ito K, Ookawara S, Ueda Y, Miyazawa H, Kofuji M, Hayasaka H, et al. Changes in cerebral oxygenation associated with intradialytic blood transfusion in patients with severe anemia undergoing hemodialysis. Nephron Extra. 2017; 7: 42–51. https://doi.org/10.1159/000471812 PMID: 28559914