



Review

Effects of Probiotic Supplementation during Pregnancy on the Future Maternal Risk of Metabolic Syndrome

Aleksandra Obuchowska, Kamila Gorczyca * , Arkadiusz Standyło * , Karolina Obuchowska, Żaneta Kimber-Trojnar , Magdalena Wierzchowska-Opoka and Bożena Leszczyńska-Gorzelał

Chair and Department of Obstetrics and Perinatology, Medical University of Lublin, 20-090 Lublin, Poland; aobuchowska12@gmail.com (A.O.); karolinaobuchowska99@gmail.com (K.O.); zkimber@poczta.onet.pl (Ż.K.-T.); magdaopoka11@gmail.com (M.W.-O.); b.leszczynska@umlub.pl (B.L.-G.)

* Correspondence: kamila.gorczyca@o2.pl (K.G.); a.standylo@gmail.com (A.S.)

Abstract: Probiotics are live microorganisms that induce health benefits in the host. Taking probiotics is generally safe and well tolerated by pregnant women and their children. Consumption of probiotics can result in both prophylactic and therapeutic effects. In healthy adult humans, the gut microbiome is stable at the level of the dominant taxa: *Bacteroidetes*, *Firmicutes* and *Actinobacteria*, and has a higher presence of *Verrucomicrobia*. During pregnancy, an increase in the number of *Proteobacteria* and *Actinobacteria* phyla and a decrease in the beneficial species *Roseburia intestinalis* and *Faecalibacterium prausnitzii* are observed. Pregnancy is a “window” to the mother’s future health. The aim of this paper is to review studies assessing the potentially beneficial effects of probiotics in preventing the development of diseases that appear during pregnancy, which are currently considered as risk factors for the development of metabolic syndrome, and consequently, reducing the risk of developing maternal metabolic syndrome in the future. The use of probiotics in gestational diabetes mellitus, preeclampsia and excessive gestational weight gain is reviewed. Probiotics are a relatively new intervention that can prevent the development of these disorders during pregnancy, and thus, would reduce the risk of metabolic syndrome resulting from these disorders in the mother’s future.

Keywords: probiotic supplementation; gestational diabetes mellitus; metabolic syndrome; preeclampsia; microbiome; obesity; pregnant women



Citation: Obuchowska, A.; Gorczyca, K.; Standyło, A.; Obuchowska, K.; Kimber-Trojnar, Ż.; Wierzchowska-Opoka, M.; Leszczyńska-Gorzelał, B. Effects of Probiotic Supplementation during Pregnancy on the Future Maternal Risk of Metabolic Syndrome. *Int. J. Mol. Sci.* **2022**, *23*, 8253. <https://doi.org/10.3390/ijms23158253>

Academic Editors: María P. Portillo and Iñaki Milton-Laskibar

Received: 4 July 2022

Accepted: 23 July 2022

Published: 26 July 2022

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Probiotics are live microorganisms that induce health benefits to the host [1]. Prebiotics, such as inulin and fructooligosaccharides, are nondigestible food components that promote the growth of beneficial bacteria in the colon [2], whereas synbiotics are a mixture of live microorganisms with substrates that are selectively utilized by host which can provide even more benefits than prebiotics alone [3].

Recommendations regarding probiotics are complex and varied by country. There is a distinction between probiotics found in food and probiotics that are pharmaceuticals. The scientific criteria regarding the health benefits of probiotics are also becoming increasingly stringent [4]. Taking probiotics is generally safe and well tolerated by pregnant women and their children [5–9]. Nevertheless, side effects such as systemic infections, mild gastrointestinal disturbances and skin complications have been reported [5,10]. According to a study by Homayouni et al., food is a better carrier of probiotics than supplements [11]. Consumption of probiotics can result in both prophylactic and therapeutic effects. They may help to prevent infections, decrease cholesterol levels, promote the synthesis of vitamins and cytokines, and inhibit cancer progression [12]. Probiotics can normalize the composition of the microbiota of the gastrointestinal tract, genitourinary system, and prevent antibiotic-associated diarrhea and travelers’ diarrhea [13,14]. The intestinal microflora can be affected by various factors, and the use of probiotics during an infection or antibiotic therapy can improve it [15].

The human gut, mainly in the distal colon, contains approximately 100 trillion gut microflora, including bacteria, archaea, viruses, and eukaryotic microbes [16,17]. The gut microbiota maintains interaction with the host through digestion, metabolism, nutrient extraction, vitamin synthesis, and pathogen protection [17–20]. Moreover, it has been found that the role of the human microbiome goes beyond the digestive tract, and the brain-gut-microbiota axis has been described [21,22]. It is generally accepted that in healthy adult humans, the gut microbiome is stable at the level of the dominant taxa: *Bacteroidetes*, *Firmicutes* and *Actinobacteria* and a higher presence of *Verrucomicrobia* (*Akkermansia muciniphila*) [21,23,24]. A healthy gut microflora was linked to an increase in the diversity and abundance of *Bacteroidetes*, and a higher ratio of *Akkermansia muciniphila* to *Ruminococcus gnavus* [21].

During pregnancy, an increase in the number of *Proteobacteria* and *Actinobacteria* phyla and a decrease in the beneficial species *Roseburia intestinalis* and *Faecalibacterium prausnitzii* were observed [25,26]. Changes in the composition of the microbiota in pregnancy may be associated with an increase in adipose mass, blood glucose levels, insulin resistance (IR), and the circulation of pro-inflammatory cytokines in the pregnant woman [27].

During pregnancy, women undergo significant anatomical and physiological changes related to the development of the fetus and the preparation for delivery. The diseases that appear during pregnancy usually concern endothelial dysfunction and endocrine disorders. These diseases put the mother at high risk of developing diseases such as type 2 diabetes mellitus (T2DM), cardiovascular disease (CVD) and possibly the development of the metabolic syndrome (MS) as a consequence.

MS is defined as a group of related risk factors that increase the risk of overall mortality and morbidity, as well as the cost and burden of healthcare. MS is an independent risk factor for CVD including microvascular dysfunction, coronary atherosclerosis and calcification, cardiac dysfunction, myocardial infarction, and heart failure [28,29]. The criteria for diagnosing MS in women vary depending on the recommendation, and any three of the five criteria constitute a diagnosis of MS [30,31] (Table 1).

Table 1. The criteria for the diagnosis of MS.

Measure	NCEP ATP3 2005	IDF 2009
Elevated waist circumference	≥88 cm (≥34.6 inches)	≥80 cm (≥31.5 inches)
Elevated triglycerides (TG)	≥150 mg/dL (1.7 mmol/L) or drug treatment for elevated TG	≥150 mg/dL (1.7 mmol/L) or drug treatment for high TG
Reduced high-density lipoprotein (HDL) cholesterol	<50 mg/dL (1.3 mmol/L) or drug treatment for low HDL cholesterol	<50 mg/dL (1.3 mmol/L) or drug treatment for low HDL cholesterol
Elevated blood pressure (BP)	≥130 mmHg systolic BP or ≥85 mmHg diastolic BP or drug treatment for hypertension	≥130 mmHg systolic BP or ≥85 mmHg diastolic BP or drug treatment for hypertension
Elevated fasting glucose	≥100 mg/dL (≥5.6 mmol/L) or drug treatment for elevated blood glucose	≥100 mg/dL (≥5.6 mmol/L) or diagnosed diabetes

The aim of the research was to review studies on the assessment of the potentially beneficial effects of probiotics in preventing the development of diseases that appear during pregnancy, which are currently considered as risk factors for the development of MS, and consequently, the reduction of the risk of developing maternal MS in the future. The use of probiotics in gestational diabetes mellitus (GDM), preeclampsia (PE) and excessive gestational weight gain (EGWG) has been reviewed.

2. Pregnancy as a Window for the Development of Metabolic Diseases

Pregnancy is a “window” to the mother’s future health. We focused on the diseases occurring during gestation that have implications for long-term maternal complications and, therefore, their programming towards metabolic syndrome.

The scientific literature defines GDM as a state of hyperglycemia developing in pregnancy as a result of IR or reduced insulin production, which resolves following delivery [32–36]. GDM is the most common metabolic disease during pregnancy and it can affect 3–31% of women. In Europe, the incidence of GDM during pregnancy ranges from 2 to 6% [33,37–41]. The prevalence is particularly increased in high-income countries, paralleling excessive weight and obesity, multiple gestations, and delayed childbearing age [42].

Due to placental hormones such as human placental lactogen, progesterone, cortisol, growth hormone, and prolactin, pregnancy is linked to physiological IR, especially in the second and third trimester of pregnancy [43,44]. Recently, the role of leptin stimulating IR and the inhibitory role of adiponectin have been emphasized [45–50]. The pathogenesis of GDM also takes into account the participation of less known molecules such as galectins, growth differentiation factor-15, chemerin, omentin-1, osteocalcin, resistin, visfatin, vaspin, irisin, apelin, Fatty Acid-Binding Protein 4 (FABP4), fibroblast growth factor 21, lipocalin-2, fetuin-A and zonulin [43,44,46,51–55]. IR is seen in GDM patients, as well as in impaired insulin secretion due to a defect in pancreatic-cell function [33,37,56]. Chronic inflammatory reaction increases IR, disrupts the function of pancreatic β cells and insulin secretion. Vitamin D deficiency can also worsen the function of β cells in the pancreas [57–60].

It is observed that IR rises distinctly in pregnant women with pre-pregnancy obesity and excessive weight compared with those with a healthy pre-pregnancy weight [61]. It is well known that a key factor in limiting the action of insulin is the increased inflammatory response associated with excess body fat [62–64]. The risk factors for GDM are pre-pregnancy obesity and excess weight, EGWG, a family history of T2DM, advanced maternal age, multiparity, GDM during previous pregnancies, giving birth to a baby with a birth weight greater than 4000 g in previous pregnancies, and polycystic ovary syndrome [65,66]. It should be noted that diabetes often coexists with obesity.

GDM is gaining increasing attention from researchers due to the serious risks and adverse health effects for the mother and her offspring. The main health consequences include an increased risk of PE, macrosomia, polyhydramnios, stillbirth and the development of metabolic diseases in the offspring in the future [67–73].

Women with a history of GDM have an increased risk of recurrence of GDM in subsequent pregnancies. Evidence suggests that GDM is a precursor to T2DM in predisposed women [74]. In women with GDM, the risk of developing T2DM is 7–10 times higher than in women without GDM [75–79]. Furthermore, the development of T2DM in patients with GDM has been associated with waist circumference, BMI, early gestational age at the time of diagnosis and gestational insulin use [80,81]. Women with GDM are at increased risk of developing CVD postpartum, which in combination with T2DM, suggests an increased incidence of MS in the future [79,82]. GDM cannot be absolutely prevented; however, early diagnosis and prompt therapy meaningfully improve the development of the fetus, the pregnancy course, delivery, and the postpartum period, as well as the development of civilization diseases in the future. Furthermore, GDM is a long-term risk factor for the development of T2DM, CVD, MS, malignant neoplasms, maternal ophthalmic, mental and renal diseases and non-alcoholic fatty liver disease [69,75,83].

MS comprises central obesity, increased plasma glucose levels, dyslipidemia and hypertension, and its incidence is increasing due to changes in lifestyle and an incorrect diet in recent years. Between 28% and 60% of women with GDM will develop MS later in life, excluding women with GDM who already have parallel MS (with central obesity, hypertension and high level of TG and/or low level of HDL cholesterol) [80,84–86]. In women with GDM, the risk of developing MS is 3–6 times higher than in women with normoglycemia during pregnancy [27,29,30]. Women with GDM who deliver large for gestational age (LGA) neonates have a higher prevalence of MS later in life than women with normoglycemia and LGA offspring [87].

In recent decades, the worldwide prevalence of obesity and the related metabolic disorders has increased dramatically. Obese or overweight people face a 10-fold increased

risk of developing MS [88]. Current and pregestational obesity and excess weight were found to be important predictors of MS in some studies, especially when combined with a fasting glycemia [80,84,89]. Compared with women with manifest diabetes, the rate of MS is delayed by more than ten years in women with glucose intolerance [57]. Obese people's microbiomes are physically and functionally diverse from their healthier counterparts, according to research [90]. Ferrer et al. found that in the gut of obese people, *Firmicutes* (94.6%) were more numerous than *Bacteroidetes* (3.2%), whereas the gut of lean people showed a change towards increased *Bacteroidetes* (18.9%) [91]. Additionally, the intestinal microbiome of an obese person is less diverse than that of lean person [92,93]. This strongly suggests the possibility of using the microbiome in the therapy of obesity.

The extended low-grade inflammation which accompanies MS and obesity is associated with the growth of adipose tissue. Currently, adipose tissue is considered as a potent immune and endocrine organ [94]. It consists of endothelial cells, preadipocytes, adipocytes, fibroblasts, and immune cells, including mainly T cells, B cells, macrophages, and neutrophils [95–97]. Excessive caloric intake, body weight and diet control the cellular composition of adipose tissue, especially the composition of the immune cells. In response to these stimuli, immune cells of the stromal vascular fraction change from anti-inflammatory subtypes to more pro-inflammatory subtypes [98]. It is known that adipokines secreted by adipose tissue are involved in inflammatory processes. Patients with pre-existing obesity have a greater risk of subclinical inflammation, IR and endothelial dysfunction leading to the development of MS [80]. Some evidence suggests that MS and GDM may have a common genetic basis [99]. Furthermore, GDM and MS appear to have similar pathophysiologic pathways, as evidenced by a link between the risk gene variants TCF7L2 and FTO [100].

Women at a higher risk of developing MS should be identified using antenatal glucose measurements and BMI [85]. Effective treatment of GDM may be a way to avoid the development of cardiovascular complications in the future [85]. The aim of GDM therapy is to maintain normoglycemia and prevent excessive weight gain to reduce maternal and fetal complications. Exercise and nutritional therapy are examples of lifestyle changes. To avoid postprandial hyperglycemia and reduce IR, caloric restriction combined with a low glycemic index diet is recommended [101]. It is important to look for other methods to prevent or support the treatment of the GDM in order to prevent the onset of MS in the future.

3. Therapeutic Applications of Probiotics

3.1. The Use of Probiotics in GDM and EGWG

Preventing GDM, rather than treating it, can have a number of benefits, both health and economic. Homayouni et al. suggest that probiotics are a relatively new intervention that can lower glucose levels, prevent GDM, and reduce maternal and fetal complications resulting from it [102]. Several studies have shown that the gut microflora is significantly altered in women with GDM and is similar to that of adults with T2DM [103–105]. An increased number of gram-negative bacteria, such as *Parabacteroides*, *Prevotella*, *Haemophilus*, and *Desulfovibrio*, has been found in the intestines of GDM patients [104,106–108]. However, a study by Mokkalá et al. showed that the presence or absence of a specific bacterial species or function did not predict the onset of GDM, nor did it differ depending on the severity of GDM [109], although in the group of women with GDM, a higher number of *Ruminococcus obeum* was found in late pregnancy [109].

The roles of probiotics in modulating the composition of the intestinal microbiota and reducing the adherence of pathobionts, regulating the permeability of the intestinal epithelium and reducing the inflammatory process has been observed [77]. A randomized study of probiotics (*Lactobacillus rhamnosus* GG and *Bifidobacterium lactis*) showed a reduction of more than 60% in GDM, with an incidence of 13% in the probiotic group compared with 34% in the control group [110]. However, some studies found no significant differences between the probiotic and placebo groups in terms of glycemic control and antioxidant

capability [111,112]. Nevertheless, the study by Sahhaf Ebrahimi et al. found that the use of probiotic yogurt improved blood glucose levels as well as glycated hemoglobin (HbA1c) levels [112]. Probiotic supplementation may improve blood glucose control during the third trimester, according to a study conducted in healthy pregnant women [113]. Supplementing with probiotics reduced fasting plasma glucose, serum insulin levels, and IR while increasing insulin sensitivity [7,114,115]. However, other studies have shown no benefit of taking probiotics in reducing the risk of GDM [116–120] or improving glucose metabolism in overweight and obese women [109,119,120]. Studies in women with GDM found no significant differences in fasting glucose levels between a group receiving supplementation of *Lactobacillus salivarius* and a placebo group [121]. In contrast, a reduction in insulin resistance (HOMA-IR) and β -cell function (HOMA-B) was observed after probiotic administration [7,115,122–124]. Other studies have shown that taking probiotics containing *Lactobacillus acidophilus*, *Lactobacillus casei* and *Bifidobacterium bifidum* in GDM patients had a beneficial effect on glycemic control, TG levels and very low-density lipoprotein (VLDL) [114,122].

The potential positive effects of using probiotics may also result from other mechanisms. Reduction of oxidative stress and increased secretion of incretin are considered as potential mechanisms by which probiotics improve glucose metabolism [125–128]. *Bifidobacterium* and *Lactobacillus* are among the most common, non-pathogenic living microorganisms used as probiotics [129]. They have been identified as probiotics that reduce systemic inflammation, regulate immune function, improve intestinal mucosa permeability, and ultimately reduce IR [130–133]. *Bifidobacteria* are commensals of humans and animals, they belong to the phylum of *Actinobacteria* [4]. *Lactic acid bacteria* are gram-positive bacteria that are traditionally used in the production of yogurt and other fermented dairy products [4]. Similar results were presented by Badehnoosh et al., who showed that taking a probiotic capsule containing *Lactobacillus acidophilus*, *Lactobacillus casei*, and *Bifidobacterium bifidum* (2×10^9 CFU/g each) for 6 weeks improved the glycemic response and markers of inflammation but did not affect pregnancy outcomes [134].

In addition, recent studies indicate that plasma vitamin C levels negatively correlate with the development of MS. It has been suggested that vitamin C supplementation may help reduce oxidative stress and postpone the chronic inflammation associated with the development of MS.

The relative abundance of short-chain fatty acids (SCFA) producing bacteria from the genera *Faecalibacterium*, *Ruminococcus*, *Roseburia*, *Coprococcus*, *Akkermansia*, *Phascolarctobacterium*, and *Eubacterium* was found to be lower in GDM women, obese and T2DM [104–106,108,135–141]. SCFA affect the activity of cells of the immune system, as well as their migration to the site of inflammation, showing a significant anti-inflammatory potential [142]. The results of many animal studies show that supplementation with the probiotic *Lactobacillus* spp. induces the production of SCFA by modulating the intestinal microbiome [141,143–146]. Manipulating the composition of the gut microbiome, and thus the level of SCFA, may prove to be a promising method in the treatment of inflammatory diseases [142].

In the peripheral blood of patients with GDM, probiotic intake increases the expression of peroxisome proliferator-activated receptor gamma (PPAR- γ), transform growth factor beta (TGF- β), and vascular endothelial growth factor (VEGF) and decreases the expression of tumor necrosis factor alpha (TNF- α) [114]. This suggests that probiotics alleviate IR and chronic inflammation through the PPAR pathway [147].

The supplementation of probiotics also results in a considerable decrease in plasma malondialdehyde (MDA) and a significant increase in plasma nitric oxide (NO) and total antioxidant capacity [114]. MDA has cytotoxic, mutagenic and carcinogenic properties [148]. It can also inhibit enzymes related to the cell's defence against oxidative stress [148]. NO is the main factor in endothelial cells responsible for maintaining vascular homeostasis [149].

The studies found that there was no significant difference in reducing GWG in overweight or obese pregnant women between the probiotic group (*Lactobacillus rhamnosus* and *Bifidobacterium lactis*) and the placebo group [119,150,151].

3.2. The Use of Probiotics in PE

PE is a complicated disorder in pregnancy which occurs after 20 weeks of gestation and affects 2–8% of all pregnancies in the world [152–154]. PE is a vascular pregnancy complication with high fetal and maternal mortality and morbidity rates [155,156]. The presence of PE in the past increases the risk of hypertension, ischemic heart disease, venous thromboembolism, kidney disease and CVD, including myocardial infarction, stroke, and heart failure [152,157–161]. PE is associated with a 4-fold increased risk of future stroke [162]. PE is associated with a 2–7 times higher risk of CVD, especially if PE occurs before 34 weeks of gestation [157,163–166]. Research has shown a significant relationship between the occurrence of PE and MS later in life [167–172]. PE is connected to MS risk factors, which are modifiable CVD risk factors [164,173,174]. According to research by Heidema et al., obesity, IR, and arterial hypertension are all common within the first year following a pregnancy affected by PE [159]. Moreover, Hooijschuur et al. showed that the incidence of MS was higher in the subgroup of women with PE and small-for-gestational-age (SGA) neonate than in women without SGA neonate [164]. It is important to look for new interventions that can reduce the risk of developing PE and consequently, complications for the mother and offspring.

Brantsæter et al. showed that consumption of milk-based probiotic products was associated with a reduced risk of overall PE, with the association most prominent in severe PE [175]. Probiotic consumption has been linked to lower blood pressure in non-pregnant women, according to clinical intervention trials using milk-based probiotic supplements [176,177]. The results of a study by Nordqvist et al. showed that consumption of probiotic milk in late pregnancy was associated with a reduced risk of PE, and consumption in early pregnancy was associated with a reduced risk of preterm delivery [178]. On the other hand, a 2018 Cochrane review of maternal oral probiotic supplementation did not find appreciable benefit or harm to neonates as a result of supplementation of pregnant women at low risk of preterm birth [179]. Furthermore, the study by Yeganegi et al. showed that *Lactobacillus rhamnosus* influences the lipopolysaccharide (LPS) response in placental trophoblast cells, which may affect the inflammatory response important in the pathophysiology of PE development [180]. However, a 2021 Cochrane review showed that probiotics can be harmful by increasing the risk of hypertensive disorders in pregnancy including PE [116]. The authors report that the best variables to predict the occurrence of MS in women with prior PE are early-onset PE, SGA newborn and measurement of systolic blood pressure [164].

3.3. The Use of Probiotics in Obesity and Lipid Disorders

The influence of obesity on the development of the MS is unquestionable, and the prevention of excessive weight gain may reduce the risk of complications in the future. Diets high in saturated fat and fructose affect the composition of the gut flora [181]. The resulting dysbiosis leads to a cascade of events including increased intestinal barrier permeability, bacterial translocation, and activation of hepatic receptor-induced inflammation [182]. One proposed mechanism pertains to the production of endogenous alcohol and acetaldehyde [183–185].

Probiotics containing *Lactobacillus paracasei* can impact adipose tissue mass by modulating the activity of angiotensin-like protein 4, a circulating lipoprotein lipase (LPL) inhibitor that controls TG deposition into adipocytes, which can help prevent obesity and metabolic disorders [186].

The beneficial effect of weight reduction was obtained during *Lactobacillus gasseri* (SBT2055) supplementation in overweight and obese people [187,188]. Daily consumption of 200 g of yogurt containing *Lactobacillus gasseri* (108 CFU/g) for 12 weeks significantly reduced abdominal obesity. BMI, waist and hip circumferences, and body fat mass were also significantly decreased from the baseline, but discontinuation of the probiotic for 4 weeks weakened these effects [187]. Additionally, in studies conducted by Ilmonen

et al., supplementation with probiotics *Lactobacillus rhamnosus* GG and *Bifidobacterium lactis* showed a reduction in maternal central adiposity at 6 months postpartum [189].

Lactobacillus plantarum (PL62) as a probiotic has been shown in two mouse trials to produce conjugated linoleic acid (CLA), which has been linked to weight loss [190,191]. In a study on mice by Bagarolli et al. the effects of probiotics (*Lactobacillus rhamnosus*, *Lactobacillus acidophilus* and *Bifidobacterium bifidum*) on the intestinal microflora, changes in insulin permeability, sensitivity and signaling in a high-fat diet (HFD) were investigated [192,193]. Probiotic-treated mice receiving a HFD gained significantly less weight and had reduced food consumption and also reduced fasting blood glucose and serum insulin compared with animals that did not receive probiotics [28]. The administration of probiotics in mice fed HFD improved leptin sensitivity [192,194,195]. The study showed that the administration of probiotics to obese animals was able to reduce Toll-like receptor 4 (TLR4) activation, downstream c-Jun N-terminal kinases (JNK) phosphorylation and the subsequent insulin receptor substrate-1 (IRS1Ser307) phosphorylation [192]. The probiotic administration had no effect on TLR4 signalling in mice with a normal body weight [192]. TLR4 is a LPS receptor that plays an important role in the regulation of immunological responses to infection [196]. As one of the components of the outer membrane of Gram-negative bacteria, LPS is considered an endotoxin that may contribute to the development of inflammation and IR [197,198]. The relative levels of TNF- α and Interleukin 6 (IL-6) transcripts in the liver, muscle and blood of probiotic-treated mice were significantly lower than in the control group [192]. When compared to untreated mice, treatment with probiotics boosted the predominance of *Firmicutes* and *Actinobacteria* while decreasing the presence of *Bacteroidetes* [192]. Obese animals also had a higher frequency of *Bacteroidetes* and a lower number of *Firmicutes* and *Actinobacteria* [192]. In obese animals, probiotic therapy resulted in the continuing presence of *Bacteroidetes*, an increase in the prevalence of *Actinobacteria*, and a decrease in *Firmicutes* compared with the control group [192]. An increased number of bacteria from the taxonomic family *Lachnospiraceae* has been associated with the development of diabetes in obese mice [18,199]. Probiotic administration has beneficial effects on the host, including increased adipose tissue lipolysis. Anorexigenic peptides, such as glucagon-like peptide-1 (GLP-1) and peptide YY (PYY), are secreted as a result, leading to an enhanced glucose tolerance and greater energy utilization [200–202]. In addition, treatment with probiotics decreased the expression of the main molecules involved in intestinal microflora inflammation and bacterial translocation, nucleotide-binding oligomerization domain-containing protein 1 (NOD-1) and CD-14 [192]. Treatment with probiotics significantly reduced the adipocyte surface area in obese mice, but the values were still outside the norm [192].

A study by Fabersani et al. showed that the *Lactobacillus fermentum* (CRL1446) strain induced a decrease in the production of leptin in adipose tissue [94]. In animal studies, this strain has antioxidant, hypoglycemic and hypocholesterolemic properties, making it an interesting alternative in the treatment of obesity, which is characterized by elevated levels of leptin in the serum [203,204].

A study by Le Barz et al. showed that mice fed the high-fat high-sucrose (HFHS) diet and treated with *Lactobacillus plantarum* (Lb38), *Lactobacillus rhamnosus* (Lbl02) or *Bifidobacterium animalis* ssp. *lactis* (Bfl41) exhibited a significant decrease in weight gain compared with the control group [205]. Lbl02 and Bfl41 significantly reduced visceral obesity, and improved insulin sensitivity and glucose tolerance in HFHS-fed mice [205]. The applied probiotics reduced the concentration of pro-inflammatory chemokines (MCP-1 and RANTES) [205]. Long-term administration of Lbl02 and Lb38 significantly lowered the plasma leptin concentration compared with the control group [205]. Lbl02 and Bfl41 showed a tendency to reduce HFHS-induced lipid accumulation in the liver [205]. The gene expression of zonula occludens 1 (zo-1) and occludin, which code for key tight-junction proteins controlling epithelial integrity, was greatly elevated after Lbl02 therapy. Mucins 2 and 3 (muc2 and -3) gene expression was also raised following Lbl02 therapy, implying that the gut barrier could be strengthened by increasing mucus-layer thickness [205].

Thiennimitr et al. showed that probiotics *Lactobacillus paracasei* (HII01), prebiotic xylooligosaccharide (XOS), and synbiotics reduce intestinal dysbiosis and enteritis, leading to improved metabolic dysfunction in obese insulin-resistant rats [206].

Numerous studies have shown that dysbiosis, small intestinal bacterial overgrowth and increased intestinal permeability have a role in the development of non-alcoholic fatty liver disease (NAFLD) and non-alcoholic steatohepatitis (NASH), both of which are closely linked to MS [207–210]. Dysbiosis was manifested in these disorders by an increase in *Enterobacteriaceae* and *Proteobacteria* and a decrease in *Bacteroidetes* [208]. The specific composition of gut microbiota may play a role in both the inflammatory and fibrosis responses in patients with NAFLD [211]. In patients with NASH, an increase in the numbers of *Bacteroides* has been shown, and a higher degree of fibrosis has been observed in patients with an increased amount of *Ruminococcus* [212,213].

A meta-analysis showed that treatment with *Lactobacillus acidophilus*, a mixture of *Lactobacillus acidophilus* and *Bifidobacterium lactis*, and *Lactobacillus plantarum* for 3 to 12 weeks lowered total and low-density lipoprotein (LDL) cholesterol concentrations compared with a placebo [214–216]. On the other hand, no effect of *Lactobacillus helveticus* and *Enterococcus faecium* on cholesterol concentrations was demonstrated [214]. Studies found no significant differences in LDL cholesterol in women with GDM between a group with supplementation of *Lactobacillus salivarius* and a placebo group [120]. In a research review by Okesene-Gafa et al. it was observed that taking probiotics may be associated with a slight reduction in TG and total cholesterol [115]. In a study by Babadi et al., probiotic capsules containing *Lactobacillus acidophilus*, *Lactobacillus casei*, *Bifidobacterium bifidum* and *Lactobacillus fermentum* reduced the concentration of TG, VLDL cholesterol, and the total/HDL cholesterol ratio while increasing HDL cholesterol levels [114].

NAFLD is strongly associated with obesity and thus closely related to the elements of MS (abdominal fat distribution, IR, diabetes, dyslipidemia, and hypertension) [217–222]. Preventing the development of NAFLD and NASH may contribute to reducing the risk of developing MS in the future.

The table below summarizes the studies conducted in pregnant women on the effects of probiotics on metabolic disorders (Table 2).

Table 2. The use of probiotics in the prevention of metabolic disorders in pregnant women.

Reference	Strain	Dosage	Treatment Duration	Population	Results
Kijmanawat et al. (2019) [7]	<i>Lactobacillus acidophilus</i> and <i>Bifidobacterium bifidum</i>	10 ⁹ CFU/capsule	4 weeks	30 patients with GDM	- decreased fasting plasma glucose ($p = 0.034$), - decreased fasting plasma insulin ($p = 0.001$) - decreased HOMA-IR ($p = 0.001$)
Nabhani et al. (2018) [111]	Synbiotic capsule consisting of <i>Lactobacillus acidophilus</i> , <i>Lactobacillus plantarum</i> , <i>Lactobacillus fermentum</i> , <i>Lactobacillus gasseri</i> with fructooligosaccharide (38.5 mg)	1.5–7.0 × 10 ^{9–10} CFU/g	6 weeks	45 patients with GDM	- increased HDL-C and TAC levels ($p < 0.05$) - decreased systolic and diastolic blood pressure ($p < 0.05$) - increased LDL-C in the placebo group ($p < 0.05$)
Sahhaf Ebrahimi et al. (2019) [112]	<i>Lactobacillus acidophilus</i> and <i>Bifidobacterium lactis</i>	10 ⁶ (300 mg of probiotic yoghurt)	8 weeks	42 patients with GDM	- decreased fasting and post prandial blood glucose ($p < 0.05$) - decrease in the level of HbA1c ($p < 0.05$)

Table 2. Cont.

Reference	Strain	Dosage	Treatment Duration	Population	Results
Babadi et al. (2019) [114]	<i>Lactobacillus acidophilus</i> , <i>Lactobacillus casei</i> , <i>Bifidobacterium bifidum</i> and <i>Lactobacillus fermentum</i>	2×10^9 CFU/g	6 weeks	24 patients with GDM	<ul style="list-style-type: none"> - upregulated PPAR-γ ($p = 0.01$), - upregulated TGF-β ($p = 0.002$), - upregulated VEGF ($p = 0.006$), - downregulated TNF-α ($p = 0.03$), - decreased fasting plasma glucose ($p = 0.02$), - decreased serum insulin levels ($p = 0.001$), - decreased insulin resistance ($p = 0.001$), - increased insulin sensitivity ($p = 0.001$), - decreased TG ($p = 0.02$), - decreased VLDL-C ($p = 0.02$), - decreased total-/HDL-C ratio ($p = 0.006$), - increased HDL-C ($p = 0.03$), - reduction in plasma MDA ($p < 0.001$), - elevation in plasma NO ($p = 0.01$), - elevation in total antioxidant capacity ($p = 0.01$)
Pellonperä et al. (2019) [119]	<i>Lactobacillus rhamnosus</i> and <i>Bifidobacterium animalis ssp. lactis</i>	10^{10} CFU/capsule	throughout the pregnancy, and until 6 months postpartum	439 overweight or obese pregnant women	<ul style="list-style-type: none"> - no differences in the maternal pregnancy outcomes ($p > 0.05$), - no change in the glucose, insulin, or HOMA2-IR ($p > 0.11$)
Callaway et al. (2019) [120]	<i>Lactobacillus rhamnosus</i> and <i>Bifidobacterium animalis subspecies lactis</i>	10^9 CFU/capsule	throughout pregnancy from the first half of the second trimester	207 overweight and obese women prevent GDM	<ul style="list-style-type: none"> - no effect of probiotics on carbohydrate metabolism
Badehnoosh et al. (2018) [134]	<i>Lactobacillus acidophilus</i> , <i>Lactobacillus casei</i> and <i>Bifidobacterium bifidum</i>	2×10^9 CFU/g	6 weeks	60 patients with GDM	<ul style="list-style-type: none"> - decreased fasting plasma glucose ($p = 0.01$) - decreased serum CRP ($p < 0.001$) - decreased plasma MDA ($p = 0.03$) - increased TAC levels ($p = 0.002$) - decreased MDA/TAC ratio ($p = 0.004$)
Okesene-Gafa et al. (2019) [150]	<i>Lactobacillus rhamnosus GG</i> and <i>Bifidobacterium lactis BB12</i>	6.5×10^9 CFU/capsule	throughout the pregnancy	230 obese pregnant women	<ul style="list-style-type: none"> - no significant difference in total maternal weight gain
Brantsæter et al. (2011) [175]	<i>Lactobacillus acidophilus</i> , <i>Bifidobacterium lactis</i> and <i>Lactobacillus rhamnosus</i>	10^8 CFU/mL	the first half of pregnancy	33,399 primiparous women	<ul style="list-style-type: none"> - reduced risk of all PE (OR = 0.80, 95% CI: 0.66, 0.96) - reduced risk of severe PE (OR = 0.61, 95% CI: 0.43, 0.89)

Table 2. Cont.

Reference	Strain	Dosage	Treatment Duration	Population	Results
Nordqvist et. al. (2018) [178]	<i>Lactobacillus acidophilus</i> , <i>Bifidobacterium lactis</i> and <i>Lactobacillus rhamnosus</i>	10 ⁸ CFU/mL	Early pregnancy or late pregnancy	37,050 primiparous women	- reduced risk of PE if taken in late pregnancy (<i>p</i> = 0.007) - reduced risk of preterm delivery if taken in early pregnancy (<i>p</i> = 0.03)

HOMA-IR (Homeostatic Model Assessment—Insulin Resistance); GDM (gestational diabetes mellitus); HDL-C (high density lipoprotein cholesterol); TAC (total antioxidant capacity); LDL-C (low density lipoprotein cholesterol); PPAR- γ (peroxisome proliferator-activated receptor gamma); TGF- β (transforming growth factor β); VEGF (vascular endothelial growth factor); TNF- α (tumor necrosis factor α); TG (triglycerides); VLDL-C (very-low-density lipoprotein cholesterol); MDA (Malondialdehyde); NO (nitric oxide); CRP (C-reactive protein); PE (preeclampsia).

3.4. Probiotics and the Prevention of the Development of MS

The studies suggest that the intestinal microbiota is a key player in the development of a chronic low-grade inflammatory state associated with MS [206]. The relationship between gut microbiota and the onset of metabolic inflammation related to obesity, IR and T2DM has been demonstrated [223,224]. Metabolic endotoxemia, which is caused primarily by the Gram-negative bacterial membrane component—LPS, is a critical event in the development of these conditions [224,225]. Through metabolic pathways, LPS leads to pro-inflammatory changes (increases in TNF- α , IL-1 β and IL-6, leptin and resistin, plasminogen activator inhibitor-1 and C-reactive protein) and induces IR [226].

It has been noted that specific probiotics may, apart from their immunomodulatory and metabolic effects, modulate the intestinal microflora [12]. For these reasons, probiotics may play an important role in immunomodulation to help prevent the low-grade chronic inflammation associated with MS [4,94,226].

Inflammatory reactions in the gut can occur through activation of the TLR pathway, degradation of the I κ B kinase, and release of nuclear-kappa B factor (NF- κ B), which activates the pro-inflammatory cascade [227,228]. Several probiotic strains such as *Lactobacillus rhamnosus* or *Lactobacillus casei* have been shown to be effective in preventing I κ B breakdown and thus reducing the release of pro-inflammatory molecules [229,230].

SCFAs such as acetate, propionate, and butyrate can be catabolized by probiotics from complex polysaccharides from the diet [231]. These substances are thought to help with metabolic disorders associated with MS. SCFAs show significant anti-inflammatory potential by reducing IR, and increasing the secretion of the protective GLP-1, which stimulates insulin release and improves β -cell function [142]. A study by Yadav et al. showed that acetate can suppress insulin signaling in adipocytes, inhibiting fat accumulation in adipose tissue [126].

It is important not only to take probiotics, but also other substances which affect probiotics bioavailability. Therefore, further work should pay attention to additional aspects including plasma concentrations of substances such as beta glucans and curcuminoids in pregnant women, which may affect probiotic absorption and increase the well-being of the gut microflora [232,233]. Vitamin C taken with probiotics may multiply their beneficial effect by reducing oxidative stress and postponing the chronic inflammation associated with the development of chronic diseases, including MS [234]. In addition, a proper diet rich in whole grains should be beneficial and could impact the microbiota profile in these patients [232].

Our study has some limitations. This work is not a systematic review and publications covering animal studies were included.

4. Conclusions

Pregnancy is said to be “a window to future health” as the occurrence of characteristic complications during pregnancy may trigger a vascular or metabolic risk of developing civilization diseases in the future life of the mother.

Even modest reductions in maternal glucose levels in nondiabetic women, especially those at high risk of GDM, may decrease the risk of developing maternal MS in the future. Effective postpartum follow-up of patients diagnosed with GDM is essential since GDM may progress to T2DM and MS, both of which are major public health problems. Pregestational obesity is predictive of progression to MS, while patients with high FPG and insulin requirements during pregnancy are at an increased risk of developing T2DM. Targeting and identifying high-risk individuals might delay and possibly prevent MS and T2DM. Knowledge about eventual interactions between the gut microflora in GDM and the host could be a potential therapeutic approach to improve health outcomes in women with GDM. A synergistic approach involving both probiotic supplementation and lifestyle modification (diet and exercise) may be a new way to prevent the development of MS in women with GDM. Probiotic supplements can help people with metabolic disorders maintain bacterial diversity and homeostasis because certain microorganisms in the gastrointestinal tract can have a positive effect on host metabolism (Figure 1).

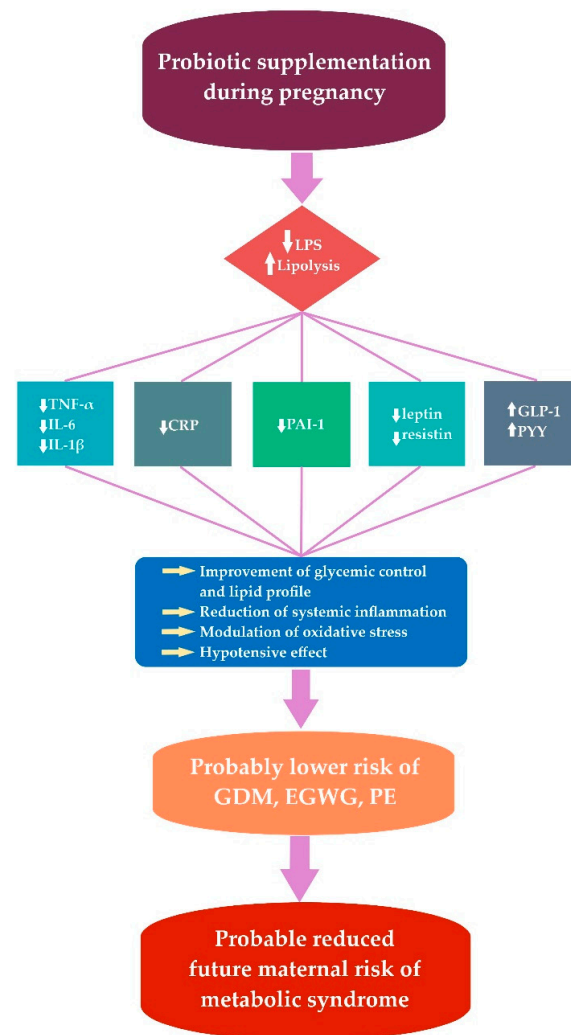


Figure 1. The effect of probiotic supplementation during pregnancy. LPS (lipopolysaccharide); TNF- α (tumor necrosis factor α); IL-6 (interleukin 6); IL-1 β (interleukin 1 beta); CRP (C-reactive protein); PAI-1 (plasminogen activator inhibitor-1); GLP-1 (glucagon-like peptide 1); PYY (peptide YY); GDM (gestational diabetes mellitus); EGWG (excessive gestational weight gain); PE (preeclampsia).

At the present time, there is no consensus regarding the effectiveness of symbiotic or probiotic supplements in GDM management due to limited evidence. Further high-quality studies of longer duration are required to determine the optimal dose, safety and the optimal composition of probiotics used in the group of patients at risk of developing MS. Additional research on the efficacy of probiotics for lipid management is also warranted. The development of obesity, NASH and NAFLD may contribute to the onset of MS in the mother in the future.

The occurrence of PE during pregnancy may be an early warning sign of metabolic complications, described in the literature as “a window to future health”. Women who have been diagnosed with metabolic disorders in pregnancy should be monitored in the puerperium and later in life for the development of these disorders. In patients with a history of PE, it is worth recommending a lifestyle change, reduction of excess body weight and regular medical examinations, which can prevent them from developing MS. It can be hypothesized that probiotics may reduce the risk of PE by modulating blood pressure and reducing inflammation, and more research is needed to elucidate their mechanisms of action and their safety for mother and child. Attention should be paid to reports on the possibility of increasing the risk of PE by taking probiotics and conducting research with due care.

Author Contributions: Conceptualization, A.O.; methodology, A.O.; data curation, A.O.; writing—original draft preparation, A.O.; writing—review and editing, A.O., K.G., K.O. and M.W.-O.; visualization, A.O. and A.S.; supervision, Ž.K.-T. and B.L.-G.; project administration, A.O.; funding acquisition, Ž.K.-T. and B.L.-G. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Medical University of Lublin, grant number 332,336.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Reid, G.; Food and Agricultural Organization of the United Nation and the WHO. The Importance of Guidelines in the Development and Application of Probiotics. *Curr. Pharm. Des.* **2005**, *11*, 11–16. [[CrossRef](#)] [[PubMed](#)]
2. Sanders, M.E.; Merenstein, D.J.; Reid, G.; Gibson, G.R.; Rastall, R.A. Probiotics and Prebiotics in Intestinal Health and Disease: From Biology to the Clinic. *Nat. Rev. Gastroenterol. Hepatol.* **2019**, *16*, 605–616. [[CrossRef](#)] [[PubMed](#)]
3. Li, H.Y.; Zhou, D.D.; Gan, R.Y.; Huang, S.Y.; Zhao, C.N.; Shang, A.; Xu, X.Y.; Li, H.B. Effects and Mechanisms of Probiotics, Prebiotics, Synbiotics, and Postbiotics on Metabolic Diseases Targeting Gut Microbiota: A Narrative Review. *Nutrients* **2021**, *13*, 3211. [[CrossRef](#)] [[PubMed](#)]
4. Zoumpopoulou, G.; Pot, B.; Tsakalidou, E.; Papadimitriou, K. Dairy Probiotics: Beyond the Role of Promoting Gut and Immune Health. *Int. Dairy J.* **2017**, *67*, 46–60. [[CrossRef](#)]
5. Didari, T.; Solki, S.; Mozaffari, S.; Nikfar, S.; Abdollahi, M. A Systematic Review of the Safety of Probiotics. *Expert Opin. Drug Saf.* **2014**, *13*, 227–239. [[CrossRef](#)]
6. Dugoua, J.J.; Machado, M.; Zhu, X.; Chen, X.; Koren, G.; Einarson, T.R. Probiotic Safety in Pregnancy: A Systematic Review and Meta-Analysis of Randomized Controlled Trials of Lactobacillus, Bifidobacterium, and Saccharomyces spp. *JOGC* **2009**, *31*, 542–552. [[CrossRef](#)]
7. Kijmanawat, A.; Panburana, P.; Reutrakul, S.; Tangshewinsirikul, C. Effects of Probiotic Supplements on Insulin Resistance in Gestational Diabetes Mellitus: A Double-Blind Randomized Controlled Trial. *J. Diabetes Investig.* **2019**, *10*, 163–170. [[CrossRef](#)]
8. Allen, S.J.; Jordan, S.; Storey, M.; Thornton, C.A.; Gravenor, M.; Garaiova, I.; Plummer, S.F.; Wang, D.; Morgan, G. Dietary Supplementation with Lactobacilli and Bifidobacteria Is Well Tolerated and Not Associated with Adverse Events during Late Pregnancy and Early Infancy. *J. Nutr.* **2010**, *140*, 483–488. [[CrossRef](#)]
9. Elias, J.; Bozzo, P.; Einarson, A. Are Probiotics Safe for Use during Pregnancy and Lactation? *Can. Fam. Physician* **2011**, *57*, 299–301.
10. Sotoudegan, F.; Daniali, M.; Hassani, S.; Nikfar, S.; Abdollahi, M. Reappraisal of Probiotics’ Safety in Human. *Food Chem. Toxicol.* **2019**, *129*, 22–29. [[CrossRef](#)]
11. Homayoni, A.; Mehrabany, E.V.; Alipoor, B.; Mehrabany, L.V.; Javadi, M. Do probiotics act more efficiently in foods than in supplements? *Nutrition* **2012**, *28*, 733–736. [[CrossRef](#)] [[PubMed](#)]

12. He, M.; Shi, B. Gut Microbiota as a Potential Target of Metabolic Syndrome: The Role of Probiotics and Prebiotics. *Cell Biosci.* **2017**, *7*, 54. [[CrossRef](#)]
13. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. The International Scientific Association for Probiotics and Prebiotics Consensus Statement on the Scope and Appropriate Use of the Term Probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [[CrossRef](#)] [[PubMed](#)]
14. Guarino, A.; Guandalini, S.; Lo Vecchio, A. Probiotics for Prevention and Treatment of Diarrhea. *J. Clin. Gastroenterol.* **2015**, *49*, 37. [[CrossRef](#)] [[PubMed](#)]
15. Kim, S.K.; Guevarra, R.B.; Kim, Y.T.; Kwon, J.; Kim, H.; Cho, J.H.; Kim, H.B.; Lee, J.H. Role of Probiotics in Human Gut Microbiome-Associated Diseases. *J. Microbiol. Biotechnol.* **2019**, *29*, 1335–1340. [[CrossRef](#)]
16. Bäckhed, F.; Ley, R.E.; Sonnenburg, J.L.; Peterson, D.A.; Gordon, J.I. Host-Bacterial Mutualism in the Human Intestine. *Science* **2005**, *307*, 1915–1920. [[CrossRef](#)]
17. Qin, J.; Li, R.; Raes, J.; Arumugam, M.; Burgdorf, K.S.; Manichanh, C.; Nielsen, T.; Pons, N.; Levenez, F.; Yamada, T.; et al. Human Gut Microbial Gene Catalogue Established by Metagenomic Sequencing. *Nature* **2010**, *464*, 59–65. [[CrossRef](#)]
18. Qin, J.; Li, Y.; Cai, Z.; Li, S.; Zhu, J.; Zhang, F.; Liang, S.; Zhang, W.; Guan, Y.; Shen, D.; et al. Metagenome-Wide Association Study of Gut Microbiota in Type 2 Diabetes. *Nature* **2012**, *490*, 55–60. [[CrossRef](#)]
19. Grenham, S.; Clarke, G.; Cryan, J.F.; Dinan, T.G. Brain-Gut-Microbe Communication in Health and Disease. *Front. Physiol.* **2011**, *2*, 94. [[CrossRef](#)] [[PubMed](#)]
20. The Human Microbiome Project Consortium. Structure, Function and Diversity of the Healthy Human Microbiome. *Nature* **2012**, *486*, 207–214. [[CrossRef](#)]
21. Torres-Fuentes, C.; Schellekens, H.; Dinan, T.G.; Cryan, J.F. The Microbiota-Gut-Brain Axis in Obesity. *Lancet Gastroenterol. Hepatol.* **2017**, *2*, 747–756. [[CrossRef](#)]
22. Dinan, T.G.; Cryan, J.F. Brain-Gut-Microbiota Axis-Mood, Metabolism and Behaviour. *Nat. Rev. Gastroenterol. Hepatol.* **2017**, *14*, 69–70. [[CrossRef](#)] [[PubMed](#)]
23. Lloyd-Price, J.; Abu-Ali, G.; Huttenhower, C. The Healthy Human Microbiome. *Genome Med.* **2016**, *8*, 51. [[CrossRef](#)] [[PubMed](#)]
24. Cani, P.D.; Delzenne, N.M. The Gut Microbiome as Therapeutic Target. *Pharmacol. Ther.* **2011**, *130*, 202–212. [[CrossRef](#)] [[PubMed](#)]
25. Koren, O.; Goodrich, J.K.; Cullender, T.C.; Spor, A.; Laitinen, K.; Bäckhed, H.K.; Gonzalez, A.; Werner, J.J.; Angenent, L.T.; Knight, R.; et al. Host Remodeling of the Gut Microbiome and Metabolic Changes during Pregnancy. *Cell* **2012**, *150*, 470–480. [[CrossRef](#)]
26. Tilg, H.; Moschen, A.R. Food, Immunity, and the Microbiome. *Gastroenterology* **2015**, *148*, 1107–1119. [[CrossRef](#)]
27. Gohir, W.; Whelan, F.J.; Surette, M.G.; Moore, C.; Schertzer, J.D.; Sloboda, D.M. Pregnancy-Related Changes in the Maternal Gut Microbiota Are Dependent upon the Mother’s Periconceptional Diet. *Gut Microbes* **2015**, *6*, 310–320. [[CrossRef](#)]
28. Tune, J.D.; Goodwill, A.G.; Sassoon, D.J.; Mather, K.J. Cardiovascular Consequences of Metabolic Syndrome. *Transl. Res.* **2017**, *183*, 57–70. [[CrossRef](#)]
29. Tran, V.; De Silva, T.M.; Sobey, C.G.; Lim, K.; Drummond, G.R.; Vinh, A.; Jelinic, M. The Vascular Consequences of Metabolic Syndrome: Rodent Models, Endothelial Dysfunction, and Current Therapies. *Front. Pharmacol.* **2020**, *11*, 148. [[CrossRef](#)]
30. Grundy, S.M.; Cleeman, J.I.; Daniels, S.R.; Donato, K.A.; Eckel, R.H.; Franklin, B.A.; Gordon, D.J.; Krauss, R.M.; Savage, P.J.; Smith, S.C.; et al. Diagnosis and Management of the Metabolic Syndrome. *Circulation* **2005**, *112*, 285–290.
31. Alberti, K.; Eckel, R.H.; Grundy, S.M.; Zimmet, P.Z.; Cleeman, J.I.; Donato, K.A.; Fruchart, J.C.; James, W.P.T.; Loria, C.M.; Smith, S.C. Harmonizing the Metabolic Syndrome. *Circulation* **2009**, *120*, 1640–1645. [[CrossRef](#)] [[PubMed](#)]
32. Thanawala, U.; Divakar, H.; Jain, R.; Agarwal, M.M. Negotiating Gestational Diabetes Mellitus in India: A National Approach. *Medicina* **2021**, *57*, 942. [[CrossRef](#)] [[PubMed](#)]
33. Alejandro, E.U.; Mamerto, T.P.; Chung, G.; Villavieja, A.; Gaus, N.L.; Morgan, E.; Pineda-Cortel, M.R.B. Gestational Diabetes Mellitus: A Harbinger of the Vicious Cycle of Diabetes. *Int. J. Mol. Sci.* **2020**, *21*, 5003. [[CrossRef](#)] [[PubMed](#)]
34. Skórzyńska-Dziduszko, K.E.; Kimber-Trojnar, Ż.; Patro-Małyśza, J.; Stenzel-Bembenek, A.; Oleszczuk, J.; Leszczyńska-Gorzela, B. Heat Shock Proteins as a Potential Therapeutic Target in the Treatment of Gestational Diabetes Mellitus: What We Know so Far. *Int. J. Mol. Sci.* **2018**, *19*, 3205. [[CrossRef](#)] [[PubMed](#)]
35. De Mendonça, E.L.S.S.; Fragoso, M.B.T.; de Oliveira, J.M.; Xavier, J.A.; Goulart, M.O.F.; de Oliveira, A.C.M. Gestational Diabetes Mellitus: The Crosslink among Inflammation, Nitroxidative Stress, Intestinal Microbiota and Alternative Therapies. *Antioxidants* **2022**, *11*, 129. [[CrossRef](#)]
36. Kampmann, U.; Knorr, S.; Fuglsang, J.; Ovesen, P. Determinants of Maternal Insulin Resistance during Pregnancy: An Updated Overview. *J. Diabetes Res.* **2019**, *2019*, 5320156. [[CrossRef](#)]
37. Plows, J.F.; Stanley, J.L.; Baker, P.N.; Reynolds, C.M.; Vickers, M.H. The Pathophysiology of Gestational Diabetes Mellitus. *Int. J. Mol. Sci.* **2018**, *19*, 3342. [[CrossRef](#)]
38. Anonymous. ACOG Practice Bulletin No. 190: Gestational Diabetes Mellitus. *Obstet. Gynecol.* **2018**, *131*, e49–e64. [[CrossRef](#)]
39. Juan, J.; Yang, H. Prevalence, Prevention, and Lifestyle Intervention of Gestational Diabetes Mellitus in China. *Int. J. Environ. Res. Res.* **2020**, *17*, 9517. [[CrossRef](#)]
40. Nguyen, C.L.; Pham, N.M.; Binns, C.W.; Duong, D.V.; Lee, A.H. Prevalence of Gestational Diabetes Mellitus in Eastern and Southeastern Asia: A Systematic Review and Meta-Analysis. *J. Diabetes Res.* **2018**, *2018*, 6536974. [[CrossRef](#)]
41. Dalfrà, M.G.; Burlina, S.; Del Vecovo, G.G.; Lapolla, A. Genetics and Epigenetics: New Insight on Gestational Diabetes Mellitus. *Front. Endocrinol.* **2020**, *11*, 602477. [[CrossRef](#)] [[PubMed](#)]

42. Tranidou, A.; Dagklis, T.; Tsakiridis, I.; Siargkas, A.; Apostolopoulou, A.; Mamopoulos, A.; Goulis, D.G.; Chourdakis, M. Risk of Developing Metabolic Syndrome after Gestational Diabetes Mellitus—A Systematic Review and Meta-Analysis. *J. Endocrinol. Investig.* **2021**, *44*, 1139–1149. [[CrossRef](#)] [[PubMed](#)]
43. Ruszała, M.; Niebrzydowska, M.; Pilszyk, A.; Kimber-Trojnar, Ż.; Trojnar, M.; Leszczyńska-Gorzelać, B. Novel Biomolecules in the Pathogenesis of Gestational Diabetes Mellitus. *Int. J. Mol. Sci.* **2021**, *22*, 11578. [[CrossRef](#)]
44. Trojnar, M.; Patro-Małyśza, J.; Kimber-Trojnar, Ż.; Leszczyńska-Gorzelać, B.; Mosiewicz, J. Associations between Fatty Acid-Binding Protein 4—A Proinflammatory Adipokine and Insulin Resistance, Gestational and Type 2 Diabetes Mellitus. *Cells* **2019**, *8*, 227. [[CrossRef](#)]
45. Patro-Małyśza, J.; Trojnar, M.; Skórzyńska-Dziduszko, K.E.; Kimber-Trojnar, Ż.; Darmochwał-Kolarz, D.; Czuba, M.; Leszczyńska-Gorzelać, B. Leptin and Ghrelin in Excessive Gestational Weight Gain—Association between Mothers and Offspring. *Int. J. Mol. Sci.* **2019**, *20*, 2398. [[CrossRef](#)]
46. Kimber-Trojnar, Ż.; Patro-Małyśza, J.; Trojnar, M.; Skórzyńska-Dziduszko, K.E.; Bartosiewicz, J.; Oleszczuk, J.; Leszczyńska-Gorzelać, B. Fatty Acid-Binding Protein 4—An “Inauspicious” Adipokine—In Serum and Urine of Post-Partum Women with Excessive Gestational Weight Gain and Gestational Diabetes Mellitus. *J. Clin. Med.* **2018**, *7*, 505. [[CrossRef](#)]
47. Florian, A.R.; Cruciat, G.; Pop, R.M.; Staicu, A.; Daniel, M.; Florin, S. Predictive Role of Altered Leptin, Adiponectin and 3-Carboxy-4-Methyl-5-Propyl-2-Furanpropanoic Acid Secretion in Gestational Diabetes Mellitus. *Exp. Ther. Med.* **2021**, *21*, 520. [[CrossRef](#)]
48. Kapustin, R.V.; Chepanov, S.V.; Babakov, V.N.; Rogovskaya, N.Y.; Kopteeva, E.V.; Alekseenkova, E.N.; Arzhanova, O.N. Maternal serum leptin, adiponectin, resistin and monocyte chemoattractant protein-1 levels in different types of diabetes mellitus. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2020**, *254*, 284–291. [[CrossRef](#)]
49. Aslfalah, H.; Jamilian, M.; Khosrowbeygi, A. Elevation of the Adiponectin/Leptin Ratio in Women with Gestational Diabetes Mellitus after Supplementation with Alpha-Lipoic Acid. *Gynecol. Endocrinol.* **2019**, *35*, 271–275. [[CrossRef](#)]
50. Meriç, P.; Özçaka, Ö.; Ceyhan-Öztürk, B.; Akcal, A.; Nalbantsoy, A.; Buduneli, N. Salivary Adiponectin and Leptin Levels Are Increased in Women with Gestational Diabetes Mellitus and Gingival Inflammation. *Oral Health Prev. Dent.* **2018**, *16*, 541–547.
51. Ruszała, M.; Pilszyk, A.; Niebrzydowska, M.; Kimber-Trojnar, Ż.; Trojnar, M.; Leszczyńska-Gorzelać, B. Novel Biomolecules in the Pathogenesis of Gestational Diabetes Mellitus 2.0. *Int. J. Mol. Sci.* **2022**, *23*, 4364. [[CrossRef](#)] [[PubMed](#)]
52. Pérez-Pérez, A.; Vilariño-García, T.; Guadix, P.; Dueñas, J.L.; Sánchez-Margalet, V. Leptin and Nutrition in Gestational Diabetes. *Nutrients* **2020**, *12*, 1970. [[CrossRef](#)] [[PubMed](#)]
53. Pheiffer, C.; Dias, S.; Jack, B.; Malaza, N.; Adam, S. Adiponectin as a Potential Biomarker for Pregnancy Disorders. *Int. J. Mol. Sci.* **2021**, *22*, 1326. [[CrossRef](#)] [[PubMed](#)]
54. Patro-Małyśza, J.; Trojnar, M.; Kimber-Trojnar, Ż.; Mierzyński, R.; Bartosiewicz, J.; Oleszczuk, J.; Leszczyńska-Gorzelać, B. FABP4 in Gestational Diabetes—Association between Mothers and Offspring. *J. Clin. Med.* **2019**, *8*, 285. [[CrossRef](#)] [[PubMed](#)]
55. Lorenzo-Almorós, A.; Hang, T.; Peiró, C.; Soriano-Guillén, L.; Egido, J.; Tuñón, J.; Lorenzo, Ó. Predictive and Diagnostic Biomarkers for Gestational Diabetes and Its Associated Metabolic and Cardiovascular Diseases. *Cardiovasc. Diabetol.* **2019**, *18*, 140. [[CrossRef](#)]
56. Nguyen-Ngo, C.; Jayabalan, N.; Salomon, C.; Lappas, M. Molecular Pathways Disrupted by Gestational Diabetes Mellitus. *J. Mol. Endocrinol.* **2019**, *63*, 51–72. [[CrossRef](#)]
57. Szymczak-Pajor, I.; Śliwińska, A. Analysis of Association between Vitamin D Deficiency and Insulin Resistance. *Nutrients* **2019**, *11*, 794. [[CrossRef](#)]
58. Wimalawansa, S.J. Associations of Vitamin D with Insulin Resistance, Obesity, Type 2 Diabetes, and Metabolic Syndrome. *J. Steroid. Biochem. Mol. Biol.* **2018**, *175*, 177–189. [[CrossRef](#)]
59. Said, J.; Lagat, D.; Kimaina, A.; Oduor, C. Beta Cell Function, Insulin Resistance and Vitamin D Status among Type 2 Diabetes Patients in Western Kenya. *Sci. Rep.* **2021**, *11*, 4084. [[CrossRef](#)]
60. Ebadi, S.A.; Sharifi, L.; Rashidi, E.; Ebadi, S.S.; Khalili, S.; Sadeghi, S.; Afzali, N.; Shiri, S.M. Supplementation with Vitamin D and Insulin Homeostasis in Healthy Overweight and Obese Adults: A Randomized Clinical Trial. *Obes. Res. Clin. Pract.* **2021**, *15*, 256–261. [[CrossRef](#)]
61. Skórzyńska-Dziduszko, K.E.; Kimber-Trojnar, Ż.; Patro-Małyśza, J.; Olszewska, A.; Zaborowski, T.; Małecka-Massalska, T. An Interplay between Obesity and Inflammation in Gestational Diabetes Mellitus. *Curr. Pharm. Biotechnol.* **2016**, *17*, 603–613. [[CrossRef](#)] [[PubMed](#)]
62. Ornoy, A.; Becker, M.; Weinstein-Fudim, L.; Ergaz, Z. Diabetes during Pregnancy: A Maternal Disease Complicating the Course of Pregnancy with Long-Term Deleterious Effects on the Offspring. A Clinical Review. *Int. J. Mol. Sci.* **2021**, *22*, 2965. [[CrossRef](#)] [[PubMed](#)]
63. Patro-Małyśza, J.; Kimber-Trojnar, Ż.; Skorzynska-Dziduszko, K.; Marciniak, B.; Darmochwał-Kolarz, D.; Bartosiewicz, J.; Leszczyńska-Gorzelać, B.; Oleszczuk, J. The Impact of Substance P on the Pathogenesis of Insulin Resistance Leading to Gestational Diabetes. *Curr. Pharm. Biotechnol.* **2014**, *15*, 32–37. [[CrossRef](#)] [[PubMed](#)]
64. Trojnar, M.; Patro-Małyśza, J.; Kimber-Trojnar, Ż.; Czuba, M.; Mosiewicz, J.; Leszczyńska-Gorzelać, B. Vaspin in Serum and Urine of Post-Partum Women with Excessive Gestational Weight Gain. *Medicina* **2019**, *55*, 76. [[CrossRef](#)]
65. Zhang, Y.; Xiao, C.M.; Zhang, Y.; Chen, Q.; Zhang, X.Q.; Li, X.F.; Shao, R.Y.; Gao, Y.M. Factors Associated with Gestational Diabetes Mellitus: A Meta-Analysis. *J. Diabetes Res.* **2021**, *2021*, 6692695. [[CrossRef](#)]

66. Davidson, K.W.; Barry, M.J.; Mangione, C.M.; Cabana, M.; Caughey, A.B.; Davis, E.M.; Donahue, K.E.; Doubeni, C.A.; Kubik, M.; Li, L.; et al. Screening for Gestational Diabetes: US Preventive Services Task Force Recommendation Statement. *JAMA* **2021**, *326*, 531–538.
67. Johns, E.C.; Denison, F.C.; Norman, J.E.; Reynolds, R.M. Gestational Diabetes Mellitus: Mechanisms, Treatment, and Complications. *Trends Endocrinol. Metab.* **2018**, *29*, 743–754. [[CrossRef](#)]
68. Franzago, M.; Fraticelli, F.; Stuppia, L.; Vitacolonna, E. Nutrigenetics, Epigenetics and Gestational Diabetes: Consequences in Mother and Child. *Epigenetics* **2019**, *14*, 215–235. [[CrossRef](#)]
69. Farahvar, S.; Walfisch, A.; Sheiner, E. Gestational Diabetes Risk Factors and Long-Term Consequences for Both Mother and Offspring: A Literature Review. *Expert Rev. Endocrinol. Metab.* **2019**, *14*, 63–74. [[CrossRef](#)]
70. Wacker-Gussmann, A.; Schopen, J.; Engelhard, J.; Sitzberger, C.; Lienert, N.; Ewert, P.; Müller, A.; Schmidt, G.; Oberhoffer-Fritz, R.; Lobmaier, S.M. The Impact of Gestational Diabetes in Pregnancy on the Cardiovascular System of Children at One Year of Age. *J. Clin. Med.* **2021**, *10*, 5839. [[CrossRef](#)]
71. Wu, P.; Gulati, M.; Kwok, C.S.; Wong, C.W.; Narain, A.; O'Brien, S.; Chew-Graham, C.A.; Verma, G.; Kadam, U.T.; Mamas, M.A. Preterm Delivery and Future Risk of Maternal Cardiovascular Disease: A Systematic Review and Meta-Analysis. *J. Am. Heart Assoc.* **2018**, *7*, 007809. [[CrossRef](#)] [[PubMed](#)]
72. Nijs, H.; Benhalima, K. Gestational Diabetes Mellitus and the Long-Term Risk for Glucose Intolerance and Overweight in the Offspring: A Narrative Review. *J. Clin. Med.* **2020**, *9*, 599. [[CrossRef](#)] [[PubMed](#)]
73. Smith, G.N.; Louis, J.M.; Saade, G.R. Pregnancy and the Postpartum Period as an Opportunity for Cardiovascular Risk Identification and Management. *Obstet. Gynecol.* **2019**, *134*, 851–862. [[CrossRef](#)]
74. Bentley-Lewis, R. Gestational Diabetes Mellitus: An Opportunity of a Lifetime. *Lancet* **2009**, *373*, 1738–1740. [[CrossRef](#)]
75. Hanna, F.W.; Duff, C.J.; Shelley-Hitchen, A.; Hodgson, E.; Fryer, A.A. Diagnosing Gestational Diabetes Mellitus: Implications of Recent Changes in Diagnostic Criteria and Role of Glycated Haemoglobin (HbA1c). *Clin. Med.* **2017**, *17*, 108–113. [[CrossRef](#)] [[PubMed](#)]
76. Herath, H.; Herath, R.; Wickremasinghe, R. Gestational Diabetes Mellitus and Risk of Type 2 Diabetes 10 Years after the Index Pregnancy in Sri Lankan Women—A Community Based Retrospective Cohort Study. *PLoS ONE* **2017**, *12*, e0179647. [[CrossRef](#)] [[PubMed](#)]
77. Hasain, Z.; Mokhtar, N.M.; Kamaruddin, N.A.; Mohamed Ismail, N.A.; Razalli, N.H.; Gnanou, J.V.; Raja Ali, R.A. Gut Microbiota and Gestational Diabetes Mellitus: A Review of Host-Gut Microbiota Interactions and Their Therapeutic Potential. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 188. [[CrossRef](#)] [[PubMed](#)]
78. Ehrlich, S.F.; Hedderson, M.M.; Feng, J.; Davenport, E.R.; Gunderson, E.P.; Ferrara, A. Change in Body Mass Index between Pregnancies and the Risk of Gestational Diabetes in a Second Pregnancy. *Obstet. Gynecol.* **2011**, *117*, 1323–1330. [[CrossRef](#)]
79. Bellamy, L.; Casas, J.P.; Hingorani, A.D.; Williams, D. Type 2 Diabetes Mellitus after Gestational Diabetes: A Systematic Review and Meta-Analysis. *Lancet* **2009**, *373*, 1773–1779. [[CrossRef](#)]
80. Can, B.; Çiftçi, S.; Yenidünya, G.; Dinççağ, N. Risk factors predicting the development of diabetes mellitus and metabolic syndrome following gestational diabetes mellitus. *Turk. J. Med. Sci.* **2021**, *51*, 595–603. [[CrossRef](#)]
81. Barker, J.; Su, F.; Alwan, N.A. Risk Factors for Type 2 Diabetes after Gestational Diabetes: A Population-Based Cohort Study. *Lancet* **2017**, *390*, 21. [[CrossRef](#)]
82. Gunderson, E.P.; Chiang, V.; Pletcher, M.J.; Jacobs, D.R.; Quesenberry, C.P.; Sidney, S.; Lewis, C.E. History of Gestational Diabetes Mellitus and Future Risk of Atherosclerosis in Mid-life: The Coronary Artery Risk Development in Young Adults Study. *J. Am. Heart Assoc.* **2014**, *3*, 000490. [[CrossRef](#)] [[PubMed](#)]
83. Retnakaran, R.; Shah, B.R. Role of Type 2 Diabetes in Determining Retinal, Renal, and Cardiovascular Outcomes in Women With Previous Gestational Diabetes Mellitus. *Diabetes Care* **2017**, *40*, 101–108. [[CrossRef](#)] [[PubMed](#)]
84. Puhkala, J.; Raitanen, J.; Kolu, P.; Tuominen, P.; Husu, P.; Luoto, R. Metabolic Syndrome in Finnish Women 7 Years after a Gestational Diabetes Prevention Trial. *BMJ Open* **2017**, *7*, 014565. [[CrossRef](#)] [[PubMed](#)]
85. Kaiser, K.; Nielsen, M.F.; Kallfa, E.; Dubietyte, G.; Lauszus, F.F. Metabolic Syndrome in Women with Previous Gestational Diabetes. *Sci. Rep.* **2021**, *11*, 11558. [[CrossRef](#)]
86. Pathirana, M.M.; Lassi, Z.S.; Ali, A.; Arstall, M.A.; Roberts, C.T.; Andraweera, P.H. Association between Metabolic Syndrome and Gestational Diabetes Mellitus in Women and Their Children: A Systematic Review and Meta-Analysis. *Endocrine* **2021**, *71*, 310–320. [[CrossRef](#)]
87. Hakkarainen, H.; Huopio, H.; Cederberg, H.; Voutilainen, R.; Heinonen, S. Future Risk of Metabolic Syndrome in Women with a Previous LGA Delivery Stratified by Gestational Glucose Tolerance: A Prospective Cohort Study. *BMC Pregnancy Childbirth* **2018**, *18*, 326. [[CrossRef](#)]
88. Bo, S.; Menato, G.; Gallo, M.L.; Bardelli, C.; Lezo, A.; Signorile, A.; Gambino, R.; Cassader, M.; Massobrio, M.; Pagano, G. Mild Gestational Hyperglycemia, the Metabolic Syndrome and Adverse Neonatal Outcomes. *Acta Obstet. Gynecol. Scand.* **2004**, *83*, 335–340. [[CrossRef](#)]
89. Barquiel, B.; Herranz, L.; Hillman, N.; Burgos, M.Á.; Pallardo, L.F. Prepregnancy Body Mass Index and Prenatal Fasting Glucose Are Effective Predictors of Early Postpartum Metabolic Syndrome in Spanish Mothers with Gestational Diabetes. *Metab. Syndr. Relat. Disord.* **2014**, *12*, 457–463. [[CrossRef](#)]

90. Green, M.; Arora, K.; Prakash, S. Microbial Medicine: Prebiotic and Probiotic Functional Foods to Target Obesity and Metabolic Syndrome. *Int. J. Mol. Sci.* **2020**, *21*, 2890. [[CrossRef](#)]
91. Ferrer, M.; Ruiz, A.; Lanza, F.; Haange, S.B.; Oberbach, A.; Till, H.; Bargiela, R.; Campoy, C.; Segura, M.T.; Richter, M.; et al. Microbiota from the Distal Guts of Lean and Obese Adolescents Exhibit Partial Functional Redundancy besides Clear Differences in Community Structure. *Environ. Microbiol.* **2013**, *15*, 211–226. [[CrossRef](#)] [[PubMed](#)]
92. Sanz, Y.; Rastmanesh, R.; Agostoni, C.; Agostonic, C. Understanding the Role of Gut Microbes and Probiotics in Obesity: How Far Are We? *Pharmacol. Res.* **2013**, *69*, 144–155. [[CrossRef](#)] [[PubMed](#)]
93. Raoult, D. Probiotics and Obesity: A Link? *Nat. Rev. Microbiol.* **2009**, *7*, 616. [[CrossRef](#)] [[PubMed](#)]
94. Fabersani, E.; Abeijon-Mukdsi, M.C.; Ross, R.; Medina, R.; González, S.; Gauffin-Cano, P. Specific Strains of Lactic Acid Bacteria Differentially Modulate the Profile of Adipokines In Vitro. *Front. Immunol.* **2017**, *8*, 266. [[CrossRef](#)] [[PubMed](#)]
95. Frigolet, M.E.; Gutiérrez-Aguilar, R. The Colors of Adipose Tissue. *Gac. Med. Mex.* **2020**, *156*, 142–149. [[CrossRef](#)] [[PubMed](#)]
96. Kawai, T.; Autieri, M.V.; Scalia, R. Adipose Tissue Inflammation and Metabolic Dysfunction in Obesity. *Am. J. Physiol. Cell Physiol.* **2021**, *320*, 375–391. [[CrossRef](#)]
97. Röszer, T. Adipose Tissue Immunometabolism and Apoptotic Cell Clearance. *Cells* **2021**, *10*, 2288. [[CrossRef](#)]
98. Grant, R.W.; Dixit, V.D. Adipose Tissue as an Immunological Organ. *Obesity* **2015**, *23*, 512–518. [[CrossRef](#)]
99. Xu, Y.; Shen, S.; Sun, L.; Yang, H.; Jin, B.; Cao, X. Metabolic Syndrome Risk after Gestational Diabetes: A Systematic Review and Meta-Analysis. *PLoS ONE* **2014**, *9*, e87863. [[CrossRef](#)]
100. Huopio, H.; Cederberg, H.; Vangipurapu, J.; Hakkarainen, H.; Pääkkönen, M.; Kuulasmaa, T.; Heinonen, S.; Laakso, M. Association of Risk Variants for Type 2 Diabetes and Hyperglycemia with Gestational Diabetes. *Eur. J. Endocrinol.* **2013**, *169*, 291–297. [[CrossRef](#)]
101. Oskovi-Kaplan, Z.A.; Ozgu-Erdinc, A.S. Management of Gestational Diabetes Mellitus. *Adv. Exp. Med. Biol.* **2021**, *1307*, 257–272. [[PubMed](#)]
102. Homayouni, A.; Bagheri, N.; Mohammad-Alizadeh-Charandabi, S.; Kashani, N.; Mobaraki-Asl, N.; Mirghafurvand, M.; Asgharian, H.; Ansari, F.; Pourjafar, H. Prevention of Gestational Diabetes Mellitus (GDM) and Probiotics: Mechanism of Action: A Review. *Curr. Diabetes Rev.* **2020**, *16*, 538–545. [[CrossRef](#)]
103. Ferrocino, I.; Ponzio, V.; Gambino, R.; Zarovska, A.; Leone, F.; Monzeglio, C.; Goitre, I.; Rosato, R.; Romano, A.; Grassi, G.; et al. Changes in the Gut Microbiota Composition during Pregnancy in Patients with Gestational Diabetes Mellitus (GDM). *Sci. Rep.* **2018**, *8*, 12216. [[CrossRef](#)] [[PubMed](#)]
104. Crusell, M.K.W.; Hansen, T.H.; Nielsen, T.; Allin, K.H.; Rühlemann, M.C.; Damm, P.; Vestergaard, H.; Rørbye, C.; Jørgensen, N.R.; Christiansen, O.B.; et al. Gestational Diabetes Is Associated with Change in the Gut Microbiota Composition in Third Trimester of Pregnancy and Postpartum. *Microbiome* **2018**, *6*, 89. [[CrossRef](#)]
105. Kuang, Y.S.; Lu, J.H.; Li, S.H.; Li, J.H.; Yuan, M.Y.; He, J.R.; Chen, N.N.; Xiao, W.Q.; Shen, S.Y.; Qiu, L.; et al. Connections between the Human Gut Microbiome and Gestational Diabetes Mellitus. *Gigascience* **2017**, *6*, 1–12. [[CrossRef](#)] [[PubMed](#)]
106. Liu, H.; Pan, L.L.; Lv, S.; Yang, Q.; Zhang, H.; Chen, W.; Lv, Z.; Sun, J. Alterations of Gut Microbiota and Blood Lipidome in Gestational Diabetes Mellitus with Hyperlipidemia. *Front. Physiol.* **2019**, *10*, 1015. [[CrossRef](#)] [[PubMed](#)]
107. Xu, Y.; Zhang, M.; Zhang, J.; Sun, Z.; Ran, L.; Ban, Y.; Wang, B.; Hou, X.; Zhai, S.; Ren, L.; et al. Differential Intestinal and Oral Microbiota Features Associated with Gestational Diabetes and Maternal Inflammation. *Am. J. Physiol. Endocrinol. Metab.* **2020**, *319*, 247–253. [[CrossRef](#)] [[PubMed](#)]
108. Cortez, R.V.; Taddei, C.R.; Sparvoli, L.G.; Ângelo, A.G.S.; Padilha, M.; Mattar, R.; Daher, S. Microbiome and Its Relation to Gestational Diabetes. *Endocrine* **2019**, *64*, 254–264. [[CrossRef](#)] [[PubMed](#)]
109. Mokkalá, K.; Paulin, N.; Houttu, N.; Koivuniemi, E.; Pellonperä, O.; Khan, S.; Pietilä, S.; Tertti, K.; Elo, L.L.; Laitinen, K. Metagenomics Analysis of Gut Microbiota in Response to Diet Intervention and Gestational Diabetes in Overweight and Obese Women: A Randomised, Double-Blind, Placebo-Controlled Clinical Trial. *Gut* **2021**, *70*, 309–318. [[CrossRef](#)] [[PubMed](#)]
110. Luoto, R.; Laitinen, K.; Nermes, M.; Isolauri, E. Impact of Maternal Probiotic-Supplemented Dietary Counselling on Pregnancy Outcome and Prenatal and Postnatal Growth: A Double-Blind, Placebo-Controlled Study. *Br. J. Nutr.* **2010**, *103*, 1792–1799. [[CrossRef](#)] [[PubMed](#)]
111. Nabhani, Z.; Hezaveh, S.J.G.; Razmpoosh, E.; Asghari-Jafarabadi, M.; Gargari, B.P. The Effects of Synbiotic Supplementation on Insulin Resistance/Sensitivity, Lipid Profile and Total Antioxidant Capacity in Women with Gestational Diabetes Mellitus: A Randomized Double Blind Placebo Controlled Clinical Trial. *Diabetes Res. Clin. Pract.* **2018**, *138*, 149–157. [[CrossRef](#)] [[PubMed](#)]
112. Sahhaf Ebrahimi, F.; Homayouni Rad, A.; Mosen, M.; Abbasalizadeh, F.; Tabrizi, A.; Khalili, L. Effect of *L. Acidophilus* and *B. Lactis* on Blood Glucose in Women with Gestational Diabetes Mellitus: A Randomized Placebo-Controlled Trial. *Diabetol. Metab. Syndr.* **2019**, *11*, 75. [[CrossRef](#)] [[PubMed](#)]
113. Laitinen, K.; Poussa, T.; Isolauri, E. Probiotics and Dietary Counselling Contribute to Glucose Regulation during and after Pregnancy: A Randomised Controlled Trial. *Br. J. Nutr.* **2008**, *101*, 1679–1687. [[CrossRef](#)] [[PubMed](#)]
114. Babadi, M.; Khorshidi, A.; Aghadavood, E.; Samimi, M.; Kavossian, E.; Bahmani, F.; Mafi, A.; Shafabakhsh, R.; Satari, M.; Asemi, Z. The Effects of Probiotic Supplementation on Genetic and Metabolic Profiles in Patients with Gestational Diabetes Mellitus: A Randomized, Double-Blind, Placebo-Controlled Trial. *Probiotics Antimicrob. Proteins* **2019**, *11*, 1227–1235. [[CrossRef](#)] [[PubMed](#)]
115. Okesene-Gafa, K.A.; Moore, A.E.; Jordan, V.; McCowan, L.; Crowther, C.A. Probiotic Treatment for Women with Gestational Diabetes to Improve Maternal and Infant Health and Well-being. *Cochrane Database Syst. Rev.* **2020**, *2020*, 012970.

116. Davidson, S.J.; Barrett, H.L.; Price, S.A.; Callaway, L.K.; Dekker Nitert, M. Probiotics for Preventing Gestational Diabetes. *Cochrane Database Syst. Rev.* **2021**, *4*, 9951.
117. Taylor, B.L.; Woodfall, G.E.; Sheedy, K.E.; O'Riley, M.L.; Rainbow, K.A.; Bramwell, E.L.; Kellow, N.J. Effect of Probiotics on Metabolic Outcomes in Pregnant Women with Gestational Diabetes: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Nutrients* **2017**, *9*, 461. [[CrossRef](#)] [[PubMed](#)]
118. Masulli, M.; Vitacolonna, E.; Fraticelli, F.; Della Pepa, G.; Mannucci, E.; Monami, M. Effects of Probiotic Supplementation during Pregnancy on Metabolic Outcomes: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Diabetes Res. Clin. Pract.* **2020**, *162*, 108111. [[CrossRef](#)]
119. Pellonperä, O.; Mokka, K.; Houttu, N.; Vahlberg, T.; Koivuniemi, E.; Tertti, K.; Rönnemaa, T.; Laitinen, K. Efficacy of Fish Oil and/or Probiotic Intervention on the Incidence of Gestational Diabetes Mellitus in an At-Risk Group of Overweight and Obese Women: A Randomized, Placebo-Controlled, Double-Blind Clinical Trial. *Diabetes Care* **2019**, *42*, 1009–1017. [[CrossRef](#)]
120. Callaway, L.K.; McIntyre, H.D.; Barrett, H.L.; Foxcroft, K.; Tremellen, A.; Lingwood, B.E.; Tobin, J.M.; Wilkinson, S.; Kothari, A.; Morrison, M.; et al. Probiotics for the Prevention of Gestational Diabetes Mellitus in Overweight and Obese Women: Findings From the SPRING Double-Blind Randomized Controlled Trial. *Diabetes Care* **2019**, *42*, 364–371. [[CrossRef](#)]
121. Lindsay, K.L.; Brennan, L.; Kennelly, M.A.; Maguire, O.C.; Smith, T.; Curran, S.; Coffey, M.; Foley, M.E.; Hatunic, M.; Shanahan, F.; et al. Impact of Probiotics in Women with Gestational Diabetes Mellitus on Metabolic Health: A Randomized Controlled Trial. *Am. J. Obstet. Gynecol.* **2015**, *212*, 496. [[PubMed](#)]
122. Karamali, M.; Dadkhah, F.; Sadrkhanlou, M.; Jamilian, M.; Ahmadi, S.; Tajabadi-Ebrahimi, M.; Jafari, P.; Asemi, Z. Effects of Probiotic Supplementation on Glycaemic Control and Lipid Profiles in Gestational Diabetes: A Randomized, Double-Blind, Placebo-Controlled Trial. *Diabetes Metab.* **2016**, *42*, 234–241. [[CrossRef](#)] [[PubMed](#)]
123. Jafarnejad, S.; Saremi, S.; Jafarnejad, F.; Arab, A. Effects of a Multispecies Probiotic Mixture on Glycemic Control and Inflammatory Status in Women with Gestational Diabetes: A Randomized Controlled Clinical Trial. *J. Nutr. Metab.* **2016**, *2016*, 5190846. [[CrossRef](#)]
124. Dolatkhan, N.; Hajifaraji, M.; Abbasalizadeh, F.; Aghamohammadzadeh, N.; Mehrabi, Y.; Abbasi, M.M. Is There a Value for Probiotic Supplements in Gestational Diabetes Mellitus? A Randomized Clinical Trial. *J. Health Popul. Nutr.* **2015**, *33*, 25. [[CrossRef](#)] [[PubMed](#)]
125. Paszti-Gere, E.; Szeker, K.; Csibrik-Nemeth, E.; Csizinszky, R.; Marosi, A.; Palocz, O.; Farkas, O.; Galfi, P. Metabolites of *Lactobacillus Plantarum* 2142 Prevent Oxidative Stress-Induced Overexpression of Proinflammatory Cytokines in IPEC-J2 Cell Line. *Inflammation* **2012**, *35*, 1487–1499. [[CrossRef](#)]
126. Yadav, H.; Lee, J.H.; Lloyd, J.; Walter, P.; Rane, S.G. Beneficial Metabolic Effects of a Probiotic via Butyrate-Induced GLP-1 Hormone Secretion. *J. Biol. Chem.* **2013**, *288*, 25088–25097. [[CrossRef](#)] [[PubMed](#)]
127. Ejtahed, H.S.; Mohtadi-Nia, J.; Homayouni-Rad, A.; Niafar, M.; Asghari-Jafarabadi, M.; Mofid, V. Probiotic Yogurt Improves Antioxidant Status in Type 2 Diabetic Patients. *Nutrition* **2012**, *28*, 539–543. [[CrossRef](#)]
128. Kim, Y.A.; Keogh, J.B.; Clifton, P.M. Probiotics, Prebiotics, Synbiotics and Insulin Sensitivity. *Nutr. Res. Rev.* **2018**, *31*, 35–51. [[CrossRef](#)]
129. Gomes, A.C.; Bueno, A.A.; de Souza, R.G.M.; Mota, J.F. Gut Microbiota, Probiotics and Diabetes. *Nutr. J.* **2014**, *13*, 60. [[CrossRef](#)]
130. Sichert, M.; De Marco, S.; Pagiotti, R.; Traina, G.; Pietrella, D. Anti-Inflammatory Effect of Multistrain Probiotic Formulation (*L. rhamnosus*, *B. lactis*, and *B. longum*). *Nutrition* **2018**, *53*, 95–102. [[CrossRef](#)] [[PubMed](#)]
131. Wang, G.; Li, X.; Zhao, J.; Zhang, H.; Chen, W. *Lactobacillus Casei* CCFM419 Attenuates Type 2 Diabetes via a Gut Microbiota Dependent Mechanism. *Food Funct.* **2017**, *8*, 3155–3164. [[CrossRef](#)]
132. Tamtaji, O.R.; Kouchaki, E.; Salami, M.; Aghadavod, E.; Akbari, E.; Tajabadi-Ebrahimi, M.; Asemi, Z. The Effects of Probiotic Supplementation on Gene Expression Related to Inflammation, Insulin, and Lipids in Patients With Multiple Sclerosis: A Randomized, Double-Blind, Placebo-Controlled Trial. *J. Am. Coll. Nutr.* **2017**, *36*, 660–665. [[CrossRef](#)]
133. Krumbeck, J.A.; Rasmussen, H.E.; Hutkins, R.W.; Clarke, J.; Shawron, K.; Keshavarzian, A.; Walter, J. Probiotic Bifidobacterium Strains and Galactooligosaccharides Improve Intestinal Barrier Function in Obese Adults but Show No Synergism When Used Together as Synbiotics. *Microbiome* **2018**, *6*, 121. [[CrossRef](#)]
134. Badehnoosh, B.; Karamali, M.; Zarrati, M.; Jamilian, M.; Bahmani, F.; Tajabadi-Ebrahimi, M.; Jafari, P.; Rahmani, E.; Asemi, Z. The Effects of Probiotic Supplementation on Biomarkers of Inflammation, Oxidative Stress and Pregnancy Outcomes in Gestational Diabetes. *J. Matern. Fetal Neonatal Med.* **2018**, *31*, 1128–1136. [[CrossRef](#)]
135. Wang, J.; Zheng, J.; Shi, W.; Du, N.; Xu, X.; Zhang, Y.; Ji, P.; Zhang, F.; Jia, Z.; Wang, Y.; et al. Dysbiosis of Maternal and Neonatal Microbiota Associated with Gestational Diabetes Mellitus. *Gut* **2018**, *67*, 1614–1625. [[CrossRef](#)]
136. Liu, Y.; Qin, S.; Feng, Y.; Song, Y.; Lv, N.; Liu, F.; Zhang, X.; Wang, S.; Wei, Y.; Li, S.; et al. Perturbations of Gut Microbiota in Gestational Diabetes Mellitus Patients Induce Hyperglycemia in Germ-Free Mice. *J. Dev. Orig. Health Dis.* **2020**, *11*, 580–588. [[CrossRef](#)]
137. Ye, G.; Zhang, L.; Wang, M.; Chen, Y.; Gu, S.; Wang, K.; Leng, J.; Gu, Y.; Xie, X. The Gut Microbiota in Women Suffering from Gestational Diabetes Mellitus with the Failure of Glycemic Control by Lifestyle Modification. *J. Diabetes Res.* **2019**, *2019*, 6081248. [[CrossRef](#)]
138. Zheng, W.; Xu, Q.; Huang, W.; Yan, Q.; Chen, Y.; Zhang, L.; Tian, Z.; Liu, T.; Yuan, X.; Liu, C.; et al. Gestational Diabetes Mellitus Is Associated with Reduced Dynamics of Gut Microbiota during the First Half of Pregnancy. *mSystems* **2020**, *5*, 109–120. [[CrossRef](#)]

139. Sun, M.; Wu, W.; Liu, Z.; Cong, Y. Microbiota Metabolite Short Chain Fatty Acids, GCPR, and Inflammatory Bowel Diseases. *J. Gastroenterol.* **2017**, *52*, 1–8. [[CrossRef](#)]
140. Hu, J.; Lin, S.; Zheng, B.; Cheung, P.C.K. Short-Chain Fatty Acids in Control of Energy Metabolism. *Crit. Rev. Food Sci. Nutr.* **2018**, *58*, 1243–1249. [[CrossRef](#)]
141. Nagpal, R.; Wang, S.; Ahmadi, S.; Hayes, J.; Gagliano, J.; Subashchandrabose, S.; Kitzman, D.W.; Becton, T.; Read, R.; Yadav, H. Human-Origin Probiotic Cocktail Increases Short-Chain Fatty Acid Production via Modulation of Mice and Human Gut Microbiome. *Sci. Rep.* **2018**, *8*, 12649. [[CrossRef](#)]
142. Czajkowska, A.; Szponar, B. Short Chain Fatty Acids (SCFA), the Products of Gut Bacteria Metabolism and Their Role in the Host. *Postepy Hig. Med. Dosw.* **2018**, *72*, 131–142. [[CrossRef](#)]
143. Markowiak-Kopeć, P.; Ślizewska, K. The Effect of Probiotics on the Production of Short-Chain Fatty Acids by Human Intestinal Microbiome. *Nutrients* **2020**, *12*, 1107. [[CrossRef](#)] [[PubMed](#)]
144. Wang, Y.; Guo, Y.; Chen, H.; Wei, H.; Wan, C. Potential of *Lactobacillus Plantarum* ZDY2013 and *Bifidobacterium Bifidum* WBIN03 in Relieving Colitis by Gut Microbiota, Immune, and Anti-Oxidative Stress. *Can. J. Microbiol.* **2018**, *64*, 327–337. [[CrossRef](#)]
145. Huang, Y.C.; Wu, B.H.; Chu, Y.L.; Chang, W.C.; Wu, M.C. Effects of Tempeh Fermentation with *Lactobacillus Plantarum* and *Rhizopus Oligosporus* on Streptozotocin-Induced Type II Diabetes Mellitus in Rats. *Nutrients* **2018**, *10*, 1143. [[CrossRef](#)]
146. Mu, W.C.; VanHoosier, E.; Elks, C.M.; Grant, R.W. Long-Term Effects of Dietary Protein and Branched-Chain Amino Acids on Metabolism and Inflammation in Mice. *Nutrients* **2018**, *10*, 918. [[CrossRef](#)]
147. Huang, L.; Thonusin, C.; Chattipakorn, N.; Chattipakorn, S.C. Impacts of Gut Microbiota on Gestational Diabetes Mellitus: A Comprehensive Review. *Eur. J. Nutr.* **2021**, *60*, 2343–2360. [[CrossRef](#)]
148. Całyniuk, B.; Grochowska-Niedworok, E.; Walkiewicz, K.W.; Kawecka, S.; Popiołek, E.; Fatyga, E. Dialdehyd Malonowy-Produkt Peroksydacji Lipidów Jako Marker Zaburzeń Homeostazy i Wieku. *Ann. Acad. Med. Silesiensis* **2016**, *70*, 224–228. [[CrossRef](#)]
149. Incalza, M.A.; D’Oria, R.; Natalicchio, A.; Perrini, S.; Laviola, L.; Giorgino, F. Oxidative Stress and Reactive Oxygen Species in Endothelial Dysfunction Associated with Cardiovascular and Metabolic Diseases. *Vascul. Pharmacol.* **2018**, *100*, 1–19. [[CrossRef](#)]
150. Okesene-Gafa, K.A.M.; Li, M.; McKinlay, C.J.D.; Taylor, R.S.; Rush, E.C.; Wall, C.R.; Wilson, J.; Murphy, R.; Taylor, R.; Thompson, J.M.D.; et al. Effect of Antenatal Dietary Interventions in Maternal Obesity on Pregnancy Weight-Gain and Birthweight: Healthy Mums and Babies (HUMBA) Randomized Trial. *AJOG* **2019**, *221*, 152. [[CrossRef](#)]
151. Aung, W.; Saw, L.; Sweet, L. An Integrative Review of Interventions for Limiting Gestational Weight Gain in Pregnant Women Who Are Overweight or Obese. *Women Birth* **2022**, *35*, 108–126. [[CrossRef](#)]
152. Rafeenia, A.; Tabandeh, A.; Khajeniazi, S.; Marjani, A. Metabolic Syndrome in Preeclampsia Women in Gorgan. *Open Biochem. J.* **2014**, *8*, 94–99.
153. Steegers, E.A.; Daddelen, P.; Duvekot, J.J.; Pijnenborg, R. Pre-Eclampsia. *Lancet* **2010**, *376*, 631–644. [[CrossRef](#)]
154. Dildy, G.A.; Belfort, M.A.; Smulian, J.C. Preeclampsia Recurrence and Prevention. *Semin. Perinatol.* **2007**, *31*, 135–141. [[CrossRef](#)]
155. Roberts, J.M.; Pearson, G.; Cutler, J.; Lindheimer, M. Summary of the NHLBI Working Group on Research on Hypertension During Pregnancy. *Hypertension* **2003**, *41*, 437–445. [[CrossRef](#)] [[PubMed](#)]
156. Duley, L. The Global Impact of Pre-Eclampsia and Eclampsia. *Semin. Perinatol.* **2009**, *33*, 130–137. [[CrossRef](#)] [[PubMed](#)]
157. Bellamy, L.; Casas, J.P.; Hingorani, A.D.; Williams, D.J. Pre-Eclampsia and Risk of Cardiovascular Disease and Cancer in Later Life: Systematic Review and Meta-Analysis. *BMJ* **2007**, *335*, 974. [[CrossRef](#)] [[PubMed](#)]
158. Romundstad, P.R.; Magnussen, E.B.; Smith, G.D.; Vatten, L.J. Hypertension in Pregnancy and Later Cardiovascular Risk: Common Antecedents? *Circulation* **2010**, *122*, 579–584. [[CrossRef](#)]
159. Heidema, W.M.; Scholten, R.R.; van Drongelen, J.; Spaanderman, M.E.A. Metabolic Syndrome after Preeclamptic Pregnancy: A Longitudinal Cohort Study. *J. Womens Health* **2019**, *28*, 357–362. [[CrossRef](#)]
160. Rangaswami, J.; Naranjo, M.; McCullough, P.A. Preeclampsia as a Form of Type 5 Cardiorenal Syndrome: An Underrecognized Entity in Women’s Cardiovascular Health. *Cardiorenal Med.* **2018**, *8*, 160–172. [[CrossRef](#)]
161. Ahmed, R.; Dunford, J.; Mehran, R.; Robson, S.; Kunadian, V. Pre-Eclampsia and Future Cardiovascular Risk among Women: A Review. *J. Am. Coll. Cardiol.* **2014**, *63*, 1815–1822. [[CrossRef](#)] [[PubMed](#)]
162. James, A.H.; Bushnell, C.D.; Jamison, M.G.; Myers, E.R. Incidence and Risk Factors for Stroke in Pregnancy and the Puerperium. *Obstet. Gynecol.* **2005**, *106*, 509–516. [[CrossRef](#)] [[PubMed](#)]
163. Ray, J.G.; Vermeulen, M.J.; Schull, M.J.; Redelmeier, D.A. Cardiovascular Health after Maternal Placental Syndromes (CHAMPS): Population-Based Retrospective Cohort Study. *Lancet* **2005**, *366*, 1797–1803. [[CrossRef](#)]
164. Hooijschuur, M.C.E.; Ghossein-Doha, C.; Kroon, A.A.; De Leeuw, P.W.; Zandbergen, A.M.; Van Kuijk, S.M.J.; Spaanderman, M.E.A. Metabolic Syndrome and Pre-Eclampsia. *Ultrasound Obstet. Gynecol.* **2019**, *54*, 64–71. [[CrossRef](#)]
165. Mosca, L.; Benjamin, E.J.; Berra, K.; Bezanson, J.L.; Dolor, R.J.; Lloyd-Jones, D.M.; Newby, L.K.; Piña, I.L.; Roger, V.L.; Shaw, L.J.; et al. American Heart Association. Effectiveness-Based Guidelines for the Prevention of Cardiovascular Disease in Women-2011 Update: A Guideline from the American Heart Association. *J. Am. Coll. Cardiol.* **2011**, *57*, 1404–1423. [[CrossRef](#)]
166. Powe, C.E.; Levine, R.J.; Karumanchi, S.A. Preeclampsia, a Disease of the Maternal Endothelium: The Role of Antiangiogenic Factors and Implications for Later Cardiovascular Disease. *Circulation* **2011**, *123*, 2856–2869. [[CrossRef](#)]
167. Smith, G.N.; Walker, M.C.; Liu, A.; Wen, S.W.; Swansburg, M.; Ramshaw, H.; White, R.R.; Roddy, M.; Hladunewich, M.; Pre-Eclampsia New Emerging Team (PE-NET). A History of Preeclampsia Identifies Women Who Have Underlying Cardiovascular Risk Factors. *Am. J. Obstet. Gynecol.* **2009**, *200*, 58. [[CrossRef](#)]

168. Yang, J.J.; Lee, S.A.; Choi, J.Y.; Song, M.; Han, S.; Yoon, H.S.; Lee, Y.; Oh, J.; Lee, J.K.; Kang, D. Subsequent Risk of Metabolic Syndrome in Women with a History of Preeclampsia: Data from the Health Examinees Study. *J. Epidemiol.* **2015**, *25*, 281–288. [[CrossRef](#)]
169. Jenabi, E.; Afshari, M.; Khazaei, S. The Association between Preeclampsia and the Risk of Metabolic Syndrome after Delivery: A Meta-Analysis. *J. Matern. Fetal Neonatal Med.* **2021**, *34*, 3253–3258. [[CrossRef](#)]
170. Smith, G.N.; Pudwell, J.; Walker, M.; Wen, S.W. Risk Estimation of Metabolic Syndrome at One and Three Years after a Pregnancy Complicated by Preeclampsia. *J. Obstet. Gynaecol. Can.* **2012**, *34*, 836–841. [[CrossRef](#)]
171. Forest, J.C.; Girouard, J.; Massé, J.; Moutquin, J.M.; Kharfi, A.; Ness, R.B.; Roberts, J.M.; Giguère, Y. Early Occurrence of Metabolic Syndrome after Hypertension in Pregnancy. *Obstet. Gynecol.* **2005**, *105*, 1373–1380. [[CrossRef](#)]
172. Cho, G.J.; Jung, U.S.; Sim, J.Y.; Lee, Y.J.; Bae, N.Y.; Choi, H.J.; Park, J.H.; Kim, H.J.; Oh, M.J. Is Preeclampsia Itself a Risk Factor for the Development of Metabolic Syndrome after Delivery? *Obstet. Gynecol. Sci.* **2019**, *62*, 233–241. [[CrossRef](#)]
173. Veerbeek, J.H.W.; Hermes, W.; Breimer, A.Y.; van Rijn, B.B.; Koenen, S.V.; Mol, B.W.; Franx, A.; de Groot, C.J.M.; Koster, M.P.H. Cardiovascular Disease Risk Factors after Early-Onset Preeclampsia, Late-Onset Preeclampsia, and Pregnancy-Induced Hypertension. *Hypertension* **2015**, *65*, 600–606. [[CrossRef](#)]
174. Stekkinger, E.; Zandstra, M.; Peeters, L.L.H.; Spaanderman, M.E.A. Early-Onset Preeclampsia and the Prevalence of Postpartum Metabolic Syndrome. *Obstet. Gynecol.* **2009**, *114*, 1076–1084. [[CrossRef](#)]
175. Brantsæter, A.L.; Myhre, R.; Haugen, M.; Myking, S.; Sengpiel, V.; Magnus, P.; Jacobsson, B.; Meltzer, H.M. Intake of Probiotic Food and Risk of Preeclampsia in Primiparous Women. *Am. J. Epidemiol.* **2011**, *174*, 807–815. [[CrossRef](#)]
176. Agerholm-Larsen, L.; Raben, A.; Haulrik, N.; Hansen, A.S.; Manders, M.; Astrup, A. Effect of 8 Week Intake of Probiotic Milk Products on Risk Factors for Cardiovascular Diseases. *Eur. J. Clin. Nutr.* **2000**, *54*, 288–297. [[CrossRef](#)]
177. Aihara, K.; Kajimoto, O.; Hirata, H.; Takahashi, R.; Nakamura, Y. Effect of Powdered Fermented Milk with *Lactobacillus Helveticus* on Subjects with High-Normal Blood Pressure or Mild Hypertension. *J. Am. Coll. Nutr.* **2005**, *24*, 257–265. [[CrossRef](#)]
178. Nordqvist, M.; Jacobsson, B.; Brantsæter, A.L.; Myhre, R.; Nilsson, S.; Sengpiel, V. Timing of Probiotic Milk Consumption during Pregnancy and Effects on the Incidence of Preeclampsia and Preterm Delivery: A Prospective Observational Cohort Study in Norway. *BMJ Open* **2018**, *8*, 18021. [[CrossRef](#)]
179. Grev, J.; Berg, M.; Soll, R. Maternal Probiotic Supplementation for Prevention of Morbidity and Mortality in Preterm Infants. *Cochrane Database Syst. Rev.* **2018**, *12*, 12519. [[CrossRef](#)]
180. Yeganegi, M.; Watson, C.S.; Martins, A.; Kim, S.O.; Reid, G.; Challis, J.R.G.; Bocking, A.D. Effect of *Lactobacillus Rhamnosus* GR-1 Supernatant and Fetal Sex on Lipopolysaccharide-Induced Cytokine and Prostaglandin-Regulating Enzymes in Human Placental Trophoblast Cells: Implications for Treatment of Bacterial Vaginosis and Prevention of Preterm Labor. *Am. J. Obstet. Gynecol.* **2009**, *200*, 532.
181. Rahman, K.; Desai, C.; Iyer, S.S.; Thorn, N.E.; Kumar, P.; Liu, Y.; Smith, T.; Neish, A.S.; Li, H.; Tan, S.; et al. Loss of Junctional Adhesion Molecule A Promotes Severe Steatohepatitis in Mice on a Diet High in Saturated Fat, Fructose, and Cholesterol. *Gastroenterology* **2016**, *151*, 733–746. [[CrossRef](#)]
182. Maslennikov, R.; Ivashkin, V.; Efremova, I.; Poluektova, E.; Shirokova, E. Gut-Liver Axis in Cirrhosis: Are Hemodynamic Changes a Missing Link? *World J. Clin. Cases* **2021**, *9*, 9320–9332. [[CrossRef](#)]
183. Cope, K.; Risby, T.; Diehl, A.M. Increased Gastrointestinal Ethanol Production in Obese Mice: Implications for Fatty Liver Disease Pathogenesis. *Gastroenterology* **2000**, *119*, 1340–1347. [[CrossRef](#)]
184. Salaspuro, M. Bacteriocolonial Pathway for Ethanol Oxidation: Characteristics and Implications. *Ann. Med.* **1996**, *28*, 195–200. [[CrossRef](#)]
185. Nair, S.; Cope, K.; Risby, T.H.; Diehl, A.M.; Terence, R.H. Obesity and Female Gender Increase Breath Ethanol Concentration: Potential Implications for the Pathogenesis of Nonalcoholic Steatohepatitis. *Am. J. Gastroenterol.* **2001**, *96*, 1200–1204. [[CrossRef](#)] [[PubMed](#)]
186. Aronsson, L.; Huang, Y.; Parini, P.; Korach-André, M.; Håkansson, J.; Gustafsson, J.Å.; Pettersson, S.; Arulampalam, V.; Rafter, J. Decreased Fat Storage by *Lactobacillus Paracasei* Is Associated with Increased Levels of Angiopoietin-Like 4 Protein (ANGPTL4). *PLoS ONE* **2010**, *5*, e13087. [[CrossRef](#)]
187. Kadooka, Y.; Sato, M.; Ogawa, A.; Miyoshi, M.; Uenishi, H.; Ogawa, H.; Ikuyama, K.; Kagoshima, M.; Tsuchida, T. Effect of *Lactobacillus Gasseri* SBT2055 in Fermented Milk on Abdominal Adiposity in Adults in a Randomised Controlled Trial. *Br. J. Nutr.* **2013**, *110*, 1696–1703. [[CrossRef](#)] [[PubMed](#)]
188. Wiciński, M.; Gębalski, J.; Gołębiewski, J.; Malinowski, B. Probiotics for the Treatment of Overweight and Obesity in Humans—A Review of Clinical Trials. *Microorganisms* **2020**, *8*, 1148. [[CrossRef](#)]
189. Ilmonen, J.; Isolauri, E.; Poussa, T.; Laitinen, K. Impact of Dietary Counselling and Probiotic Intervention on Maternal Anthropometric Measurements during and after Pregnancy: A Randomized Placebo-Controlled Trial. *Clin. Nutr.* **2011**, *30*, 156–164. [[CrossRef](#)] [[PubMed](#)]
190. Lee, K.; Paek, K.; Lee, H.Y.; Park, J.H.; Lee, Y. Antiobesity Effect of Trans-10,Cis-12-Conjugated Linoleic Acid-Producing *Lactobacillus Plantarum* PL62 on Diet-Induced Obese Mice. *J. Appl. Microbiol.* **2007**, *103*, 1140–1146. [[CrossRef](#)]
191. Kennedy, A.; Martinez, K.; Schmidt, S.; Mandrup, S.; LaPoint, K.; McIntosh, M. Antiobesity Mechanisms of Action of Conjugated Linoleic Acid. *J. Nutr. Biochem.* **2010**, *21*, 171–179. [[CrossRef](#)]

192. Bagarolli, R.A.; Tobar, N.; Oliveira, A.G.; Araújo, T.G.; Carvalho, B.M.; Rocha, G.Z.; Vecina, J.F.; Calisto, K.; Guadagnini, D.; Prada, P.O.; et al. Probiotics Modulate Gut Microbiota and Improve Insulin Sensitivity in DIO Mice. *J. Nutr. Biochem.* **2017**, *50*, 16–25. [[CrossRef](#)] [[PubMed](#)]
193. McIntyre, H.D.; Catalano, P.; Zhang, C.; Desoye, G.; Mathiesen, E.R.; Damm, P. Gestational Diabetes Mellitus. *Nat. Rev. Dis. Primers* **2019**, *5*, 47. [[CrossRef](#)] [[PubMed](#)]
194. Cano, P.G.; Santacruz, A.; Trejo, F.M.; Sanz, Y. Bifidobacterium CECT 7765 Improves Metabolic and Immunological Alterations Associated with Obesity in High-Fat Diet-Fed Mice. *Obesity* **2013**, *21*, 2310–2321. [[CrossRef](#)] [[PubMed](#)]
195. Moya-Pérez, A.; Neef, A.; Sanz, Y. Bifidobacterium Pseudocatenulatum CECT 7765 Reduces Obesity-Associated Inflammation by Restoring the Lymphocyte-Macrophage Balance and Gut Microbiota Structure in High-Fat Diet-Fed Mice. *PLoS ONE* **2015**, *10*, e0126976. [[CrossRef](#)]
196. Fawzy El-Sayed, K.M.; Klingebiel, P.; Dörfer, C.E. Toll-like Receptor Expression Profile of Human Dental Pulp Stem/Progenitor Cells. *J. Endod.* **2016**, *42*, 413–417. [[CrossRef](#)] [[PubMed](#)]
197. Cardona Gloria, Y.; Latz, E.; De Nardo, D. Generation of Innate Immune Reporter Cells Using Retroviral Transduction. *Methods Mol. Biol.* **2018**, *1714*, 97–117.
198. Rosadini, C.V.; Kagan, J.C. Early Innate Immune Responses to Bacterial LPS. *Curr. Opin. Immunol.* **2017**, *44*, 14–19. [[CrossRef](#)] [[PubMed](#)]
199. Cho, I.; Yamanishi, S.; Cox, L.; Methé, B.A.; Zavadil, J.; Li, K.; Gao, Z.; Mahana, D.; Raju, K.; Teitler, I.; et al. Antibiotics in Early Life Alter the Murine Colonic Microbiome and Adiposity. *Nature* **2012**, *488*, 621–626. [[CrossRef](#)] [[PubMed](#)]
200. Martínez, I.; Wallace, G.; Zhang, C.; Legge, R.; Benson, A.K.; Carr, T.P.; Moriyama, E.N.; Walter, J. Diet-Induced Metabolic Improvements in a Hamster Model of Hypercholesterolemia Are Strongly Linked to Alterations of the Gut Microbiota. *Appl. Environ. Microbiol.* **2009**, *75*, 4175–4184. [[CrossRef](#)]
201. Round, J.L.; Mazmanian, S.K. The Gut Microbiota Shapes Intestinal Immune Responses during Health and Disease. *Nat. Rev. Immunol.* **2009**, *9*, 313–323. [[CrossRef](#)]
202. Mazloom, K.; Siddiqi, I.; Covasa, M. Probiotics: How Effective Are They in the Fight against Obesity? *Nutrients* **2019**, *11*, 258. [[CrossRef](#)] [[PubMed](#)]
203. Mukdsi, M.C.A.; Cano, M.P.G.; González, S.N.; Medina, R.B. Administration of Lactobacillus Fermentum CRL1446 Increases Intestinal Feruloyl Esterase Activity in Mice. *Letts. Appl. Microbiol.* **2012**, *54*, 18–25. [[CrossRef](#)] [[PubMed](#)]
204. Russo, M.; Fabersani, E.; Abeijón-Mukdsi, M.C.; Ross, R.; Fontana, C.; Benítez-Páez, A.; Gauffin-Cano, P.; Medina, R.B. Lactobacillus Fermentum CRL1446 Ameliorates Oxidative and Metabolic Parameters by Increasing Intestinal Feruloyl Esterase Activity and Modulating Microbiota in Caloric-Restricted Mice. *Nutrients* **2016**, *8*, 415. [[CrossRef](#)]
205. Le Barz, M.; Daniel, N.; Varin, T.V.; Naimi, S.; Demers-Mathieu, V.; Pilon, G.; Audy, J.; Laurin, É.; Roy, D.; Urdaci, M.C.; et al. In Vivo Screening of Multiple Bacterial Strains Identifies Lactobacillus Rhamnosus Lb102 and Bifidobacterium Animalis Ssp. Lactis Bf141 as Probiotics That Improve Metabolic Disorders in a Mouse Model of Obesity. *FASEB J.* **2019**, *33*, 4921–4935. [[CrossRef](#)] [[PubMed](#)]
206. Thiennimitr, P.; Yasom, S.; Tunapong, W.; Chunchai, T.; Wanchai, K.; Pongchaidecha, A.; Lungkaphin, A.; Sirilun, S.; Chaiyasut, C.; Chattipakorn, N.; et al. Lactobacillus Paracasei HIII01, Xylooligosaccharides, and Synbiotics Reduce Gut Disturbance in Obese Rats. *Nutrition* **2018**, *54*, 40–47. [[CrossRef](#)]
207. Borrelli, A.; Bonelli, P.; Tuccillo, F.M.; Goldfine, I.D.; Evans, J.L.; Buonaguro, F.M.; Mancini, A. Role of Gut Microbiota and Oxidative Stress in the Progression of Non-Alcoholic Fatty Liver Disease to Hepatocarcinoma: Current and Innovative Therapeutic Approaches. *Redox Biol.* **2018**, *15*, 467–479. [[CrossRef](#)]
208. Brandi, G.; De Lorenzo, S.; Candela, M.; Pantaleo, M.A.; Bellentani, S.; Tovoli, F.; Saccoccio, G.; Biasco, G. Microbiota, NASH, HCC and the Potential Role of Probiotics. *Carcinogenesis* **2017**, *38*, 231–240. [[CrossRef](#)]
209. Wigg, A.J.; Roberts-Thomson, I.C.; Dymock, R.B.; McCarthy, P.J.; Grose, R.H.; Cummins, A.G. The Role of Small Intestinal Bacterial Overgrowth, Intestinal Permeability, Endotoxaemia, and Tumour Necrosis Factor Alpha in the Pathogenesis of Non-Alcoholic Steatohepatitis. *Gut* **2001**, *48*, 206–211. [[CrossRef](#)]
210. Miele, L.; Valenza, V.; La Torre, G.; Montalto, M.; Cammarota, G.; Ricci, R.; Mascianà, R.; Forgione, A.; Gabrieli, M.L.; Perotti, G.; et al. Increased Intestinal Permeability and Tight Junction Alterations in Nonalcoholic Fatty Liver Disease. *Hepatology* **2009**, *49*, 1877–1887. [[CrossRef](#)] [[PubMed](#)]
211. Boursier, J.; Mueller, O.; Barret, M.; Machado, M.; Fizanne, L.; Araujo-Perez, F.; Guy, C.D.; Seed, P.C.; Rawls, J.F.; David, L.A.; et al. The Severity of Nonalcoholic Fatty Liver Disease Is Associated with Gut Dysbiosis and Shift in the Metabolic Function of the Gut Microbiota. *Hepatology* **2016**, *63*, 764–775. [[CrossRef](#)]
212. Caussy, C.; Hsu, C.; Lo, M.T.; Liu, A.; Bettencourt, R.; Ajmera, V.H.; Bassirian, S.; Hooker, J.; Sy, E.; Richards, L.; et al. Genetics of NAFLD in Twins Consortium. Link between Gut-Microbiome Derived Metabolite and Shared Gene-Effects with Hepatic Steatosis and Fibrosis in NAFLD. *Hepatology* **2018**, *68*, 918–932. [[CrossRef](#)] [[PubMed](#)]
213. Loomba, R.; Seguritan, V.; Li, W.; Long, T.; Klitgord, N.; Bhatt, A.; Dulai, P.S.; Caussy, C.; Bettencourt, R.; Highlander, S.K.; et al. Gut Microbiome-Based Metagenomic Signature for Non-Invasive Detection of Advanced Fibrosis in Human Nonalcoholic Fatty Liver Disease. *Cell Metab.* **2017**, *25*, 1054–1062. [[CrossRef](#)] [[PubMed](#)]
214. Cho, Y.A.; Kim, J. Effect of Probiotics on Blood Lipid Concentrations: A Meta-Analysis of Randomized Controlled Trials. *Medicine* **2015**, *94*, 1714. [[CrossRef](#)] [[PubMed](#)]

215. Shimizu, M.; Hashiguchi, M.; Shiga, T.; Tamura, H.; Mochizuki, M. Meta-Analysis: Effects of Probiotic Supplementation on Lipid Profiles in Normal to Mildly Hypercholesterolemic Individuals. *PLoS ONE* **2015**, *10*, e139795.
216. Companys, J.; Pla-Pagà, L.; Calderón-Pérez, L.; Llauroadó, E.; Solà, R.; Pedret, A.; Valls, R.M. Fermented Dairy Products, Probiotic Supplementation, and Cardiometabolic Diseases: A Systematic Review and Meta-Analysis. *Adv. Nutr.* **2020**, *11*, 834–863. [[CrossRef](#)] [[PubMed](#)]
217. Speiser, P.W.; Rudolf, M.C.J.; Anhalt, H.; Camacho-Hubner, C.; Chiarelli, F.; Eliakim, A.; Freemark, M.; Gruters, A.; HersHKovitz, E.; Iughetti, L.; et al. Childhood Obesity. *J. Clin. Endocrinol. Metab.* **2005**, *90*, 1871–1887. [[CrossRef](#)] [[PubMed](#)]
218. Matteoni, C.A.; Younossi, Z.M.; Gramlich, T.; Boparai, N.; Liu, Y.C.; McCullough, A.J. Nonalcoholic Fatty Liver Disease: A Spectrum of Clinical and Pathological Severity. *Gastroenterology* **1999**, *116*, 1413–1419. [[CrossRef](#)]
219. Mandato, C.; Lucariello, S.; Licenziati, M.R.; Franzese, A.; Spagnuolo, M.I.; Ficarella, R.; Pacilio, M.; Amitrano, M.; Capuano, G.; Meli, R.; et al. Metabolic, Hormonal, Oxidative, and Inflammatory Factors in Pediatric Obesity-Related Liver Disease. *J. Pediatr.* **2005**, *147*, 62–66. [[CrossRef](#)]
220. Schwimmer, J.B.; Zepeda, A.; Newton, K.P.; Xanthakos, S.A.; Behling, C.; Hallinan, E.K.; Donithan, M.; Tonascia, J.; Nonalcoholic Steatohepatitis Clinical Research Network. Longitudinal Assessment of High Blood Pressure in Children with Nonalcoholic Fatty Liver Disease. *PLoS ONE* **2014**, *9*, e112569. [[CrossRef](#)]
221. Newton, K.P.; Hou, J.; Crimmins, N.A.; Lavine, J.E.; Barlow, S.E.; Xanthakos, S.A.; Africa, J.; Behling, C.; Donithan, M.; Clark, J.M.; et al. Nonalcoholic Steatohepatitis Clinical Research Network. Prevalence of Prediabetes and Type 2 Diabetes in Children with Nonalcoholic Fatty Liver Disease. *JAMA Pediatr.* **2016**, *170*, 161971. [[CrossRef](#)] [[PubMed](#)]
222. Schwimmer, J.B.; Deutsch, R.; Rauch, J.B.; Behling, C.; Newbury, R.; Lavine, J.E. Obesity, Insulin Resistance, and Other Clinicopathological Correlates of Pediatric Nonalcoholic Fatty Liver Disease. *J. Pediatr.* **2003**, *143*, 500–505. [[CrossRef](#)]
223. Cani, P.D.; Amar, J.; Iglesias, M.A.; Poggi, M.; Knauf, C.; Bastelica, D.; Neyrinck, A.M.; Fava, F.; Tuohy, K.M.; Chabo, C.; et al. Metabolic Endotoxemia Initiates Obesity and Insulin Resistance. *Diabetes* **2007**, *56*, 1761–1772. [[CrossRef](#)] [[PubMed](#)]
224. Rastelli, M.; Knauf, C.; Cani, P.D. Gut Microbes and Health: A Focus on the Mechanisms Linking Microbes, Obesity, and Related Disorders. *Obesity* **2018**, *26*, 792–800. [[CrossRef](#)] [[PubMed](#)]
225. Kirsch, R.; Clarkson, V.; Verdonk, R.C.; Marais, A.D.; Shephard, E.G.; Ryffel, B.; Hall, P. Rodent Nutritional Model of Steatohepatitis: Effects of Endotoxin (Lipopolysaccharide) and Tumor Necrosis Factor Alpha Deficiency. *J. Gastroenterol. Hepatol.* **2006**, *21*, 174–182. [[CrossRef](#)] [[PubMed](#)]
226. Torres, S.; Fabersani, E.; Marquez, A.; Gauffin-Cano, P. Adipose Tissue Inflammation and Metabolic Syndrome. The Proactive Role of Probiotics. *Eur. J. Nutr.* **2019**, *58*, 27–43. [[CrossRef](#)] [[PubMed](#)]
227. Kelly, J.R.; Clarke, G.; Cryan, J.F.; Dinan, T.G. Brain-Gut-Microbiota Axis: Challenges for Translation in Psychiatry. *Ann. Epidemiol.* **2016**, *26*, 366–372. [[CrossRef](#)]
228. Kemgang, T.; Kapila, S.; Shanmugam, V.; Kapila, R. Cross-Talk between Probiotic Lactobacilli and Host Immune System. *J. Appl. Microbiol.* **2014**, *117*, 303–319. [[CrossRef](#)]
229. Zhang, L.; Li, N.; Caicedo, R.; Neu, J. Alive and Dead Lactobacillus Rhamnosus GG Decrease Tumor Necrosis Factor- α -Induced Interleukin-8 Production in Caco-2 Cells. *J. Nutr.* **2005**, *135*, 1752–1756. [[CrossRef](#)] [[PubMed](#)]
230. Tien, M.T.; Girardin, S.E.; Regnault, B.; Bourhis, L.L.; Dillies, M.A.; Coppée, J.Y.; Bourdet-Sicard, R.; Sansonetti, P.J.; Pédrón, T. Anti-Inflammatory Effect of Lactobacillus Casei on Shigella-Infected Human Intestinal Epithelial Cells. *J. Immunol.* **2006**, *176*, 1228–1237. [[CrossRef](#)]
231. Martin-Gallausiaux, C.; Marinelli, L.; Blottière, H.M.; Larraufie, P.; Lapaque, N. SCFA: Mechanisms and Functional Importance in the Gut. *Proc. Nutr. Soc.* **2021**, *80*, 37–49. [[CrossRef](#)] [[PubMed](#)]
232. Rossi, P.; Difrancia, R.; Quagliariello, V.; Savino, E.; Tralongo, P.; Randazzo, C.; Berretta, M. B-glucans from Grifola frondosa and Ganoderma lucidum in breast cancer: An example of complementary and integrative medicine. *Oncotarget* **2018**, *9*, 24837–24856. [[CrossRef](#)] [[PubMed](#)]
233. Yucel, C.; Quagliariello, V.; Iaffaioli, R.; Ferrari, G.; Donsi, F. Submicron complex lipid carriers for curcumin delivery to intestinal epithelial cells: Effect of different emulsifiers on bioaccessibility and cell uptake. *Int. J. Pharm.* **2015**, *15*, 357–369. [[CrossRef](#)] [[PubMed](#)]
234. Berretta, M.; Quagliariello, V.; Maurea, N.; Di Francia, R.; Sharifi, S.; Facchini, G.; Rinaldi, L.; Piezzo, M.; Manuela, C.; Nunnari, G.; et al. Multiple Effects of Ascorbic Acid against Chronic Diseases: Updated Evidence from Preclinical and Clinical Studies. *Antioxidants* **2020**, *9*, 1182. [[CrossRef](#)] [[PubMed](#)]