

Research Article

Combination Treatment with PPAR γ Ligand and Its Specific Inhibitor GW9662 Downregulates BIS and 14-3-3 Gamma, Inhibiting Stem-Like Properties in Glioblastoma Cells

Chang-Nim Im

Department of Psychiatry, Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul 06591, Republic of Korea

Correspondence should be addressed to Chang-Nim Im; milybud@gmail.com

Received 15 February 2017; Revised 12 April 2017; Accepted 24 April 2017; Published 31 May 2017

Academic Editor: Marta M. Alonso

Copyright © 2017 Chang-Nim Im. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

PPAR γ is a nuclear receptor that regulates differentiation and proliferation and is highly expressed in many cancer cells. Its synthetic ligands, such as rosiglitazone and ciglitazone, and its inhibitor GW9662, were shown to induce cellular differentiation, inhibit proliferation, and lead to apoptosis. Glioblastoma is a common brain tumor with poor survival prospects. Recently, glioblastoma stem cells (GSCs) have been examined as a potential target for anticancer therapy; however, little is known about the combined effect of various agents on GSCs. In this study, we found that cotreatment with PPAR γ ligands and GW9662 inhibited stem-like properties in GSC-like spheres, which significantly express SOX2. In addition, this treatment decreased the activation of STAT3 and AKT and decreased the amounts of 14-3-3 gamma and BIS proteins. Moreover, combined administration of small-interfering RNA (siRNA) transfection with PPAR γ ligands induced downregulation of SOX2 and MMP2 activity together with inhibition of sphere-forming activity regardless of poly(ADP-ribose) polymerase (PARP) cleavage. Taken together, our findings suggest that a combination therapy using PPAR γ ligands and its inhibitor could be a potential therapeutic strategy targeting GSCs.

1. Introduction

Glioblastoma multiform (GBM, also known as glioblastoma) is a common and lethal malignant brain tumor [1]. Glioblastoma stem cells (GSCs) are at the root of tumor recurrence and are regarded as a potential target for anticancer therapy [2, 3].

PPAR γ belongs to a nuclear receptor family, along with PPAR α and β/δ , and is highly expressed not only in adipocytes during differentiation [4] but also in various cancer cells [5]. PPAR γ ligands such as thiazolidinedione and rosiglitazone induce differentiation and apoptosis in various human glioblastoma cells [6–11]. For instance, Kato et al. showed that 95% of glioma tissue expressed PPAR γ mRNA and that a PPAR γ ligand, troglitazone, inhibited growth in both SK-MG-1 and NB-1 cell lines [5]. Morosetti et al. reported that human glioblastoma cell lines, such as A172 and U87-MG, also express high levels of PPAR γ , which rosiglitazone inhibited proliferation of those cell lines by G2/M

arrest and apoptosis, and that the growth inhibitory effect was partially reversed by the PPAR γ antagonist GW9662 [9], suggesting that it may work through PPAR γ -dependent and PPAR γ -independent pathways. PPAR γ antagonists, such as GW9662, enhance PPAR γ ligand-induced apoptosis [6, 12]. This suggests that both PPAR γ ligands [13–16] and GW9662 may be potential agents for glioblastoma therapy specifically targeting GSCs [17, 18].

14-3-3 is an adaptor protein that binds a variety of proteins via a p-Ser/Thr-containing motif. Seven 14-3-3 isoforms (beta, epsilon, zeta, eta, theta, gamma, and sigma) have been identified in mammalian cells. These isoforms are broadly and differentially expressed in almost all tissues and in brain tumors, such as glioblastoma and astrocytoma [19–21]. 14-3-3 beta and sigma are well described, suggesting that these isoforms regulate proliferation in astrocytoma and stem cells, respectively [22, 23]. Jin et al. demonstrated that 14-3-3 gamma interacts with diverse proteins and that this interaction is strengthened by AKT [24], suggesting that AKT

may act upstream of 14-3-3 and that they are connected to cancer progression [25]. The antiapoptotic actions of ligand-activated PPAR γ are mediated through enhanced binding of PPAR to the promoter of 14-3-3 epsilon and upregulation of 14-3-3 epsilon expression, suggesting that the PPAR to 14-3-3 transcriptional axis plays an important role protecting cell and tissue integrity and may be a possible target for drug discovery [26, 27].

Bcl2-interacting cell death suppressor (BIS) [28], also known as BAG3 [29], has antiapoptotic functions and controls cellular protein quality [30, 31], and it is overexpressed in human glioblastoma tissue [32]. Our recent report demonstrated that BIS is linked to glioblastoma stemness by stabilization of the signal transducer and activator of transcription 3 (STAT3) [33]. These findings provide reliable evidence that BIS is a potential target for therapy. Although PPAR γ ligands and/or its antagonists induce apoptosis in cancer cells, including glioblastoma cells [9, 10, 12], the link between 14-3-3 gamma and BIS in GSC-like spheres is not well defined.

Previously, we established a GSC-like sphere culture system in which SOX2 was expressed at significant levels [34] and hypothesized that PPAR γ ligands may affect cancer stemness and induce apoptosis in GBM [35]. In this study, we describe the effect of a combination treatment with PPAR γ ligands and its inhibitor GW9662 on spheres of glioblastoma cells through downregulation of BIS and 14-3-3 gamma levels, as well as inhibition of SOX2, MMP2 activity, and sphere-forming activity without enhancing the levels of cleaved poly(ADP-ribose) polymerase (PARP).

2. Materials and Methods

2.1. Cell Culture. A172 and U87-MG (U87) human glioblastoma cells were purchased from American Type Culture Collection (ATCC, Manassas, VA, USA) and maintained in DMEM (HyClone, Logan, UT, USA) contained with 10% heat-inactivated fetal bovine serum (FBS), 100 units/mL penicillin, and 100 mg/mL streptomycin at 37°C in 5% CO₂ atmosphere. For sphere culture, cells (1×10^5 cells per well) were cultured on ultralow attachment 6-well plate (Corning, Tewksbury, MA) for 72 hours in serum-free glioblastoma sphere medium containing epidermal growth factor (EGF, 20 ng/mL, R&D Systems, Minneapolis, MN) and basic fibroblast growth factor (bFGF, 20 ng/mL, R&D Systems). For morphological examination, spheres per field were counted and pictures were taken under the inverted microscope.

2.2. Sphere-Formation Assay. Spheres cultivated in serum-free glioblastoma sphere medium containing EGF and bFGF were attached to standard culture plates in media containing 5% FBS stained with crystal violet solution (Sigma-Aldrich, St. Louis, MO, USA). For morphological examination, spheres per field were counted and pictures were taken under the inverted microscope.

2.3. Cell Viability Assay. Cell proliferation was assessed as a function of metabolic activity using an EZ-Cytox Cell Viability Assay Kit (ItsBio, Seoul, Korea). The assay is based

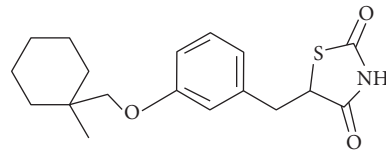
on reduction of tetrazolium chloride to the water-soluble formazan by succinate-tetrazolium reductase, which forms part of the mitochondrial respiratory chain. After treatment with 20 μ L per well, cells were incubated for 2 hours at 37°C in a 5% CO₂ atmosphere. Absorbance was measured on a microplate reader (Bio-Rad 680; Bio-Rad, Hercules, CA, USA) at 450 nm. After subtraction of the background, the viability was determined as the ratio relative to the control and reported as the mean \pm standard error (SE).

2.4. Western Blot. Cells were lysed with RIPA buffer (150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate, 0.1% SDS, 50 mM Tris-HCl pH 8.0) with protease inhibitor (Roche Diagnostics, Mannheim, Germany) on ice for 30 m. Equal amounts of protein were separated on 10% sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to nitrocellulose membranes (GE Healthcare Life Sciences, Buckinghamshire, UK). The membranes were incubated for 1 h with 5% dry skim milk in TBST (20 mM Tris, 137 mM NaCl, and 0.1% Tween 20) buffer and then incubated with antibodies against BIS [28], p-STAT3 (Y705), p-AKT (S473), and PPAR γ (Cell Signaling, Danvers, Massachusetts, USA), SOX2 (Santa Cruz Biotechnology), cleaved PARP (Abcam, Cambridge, UK), or beta-actin (Sigma-Aldrich, St. Louis, MO, USA). After incubation with horseradish peroxidase-conjugated anti-mouse or anti-rabbit IgG (1:5,000; Santa Cruz Biotechnology), the immunoreactive bands were visualized by an enhanced chemiluminescence substrate (ThermoFisher Scientific). Quantification of the intensities of each band was carried out using ImageJ software (National Institutes of Health, Bethesda, MD, USA).

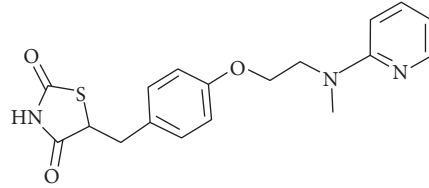
2.5. Small-Interfering RNA (siRNA) Transfection. Knock-down was performed by transfection of specific siRNA targeted with G-fectin (Genolution Pharmaceuticals, Seoul, Korea) according to manufacturer's instruction. siRNA for control (5'-CCUACGCCACCAAUUUCGU-3') and BIS (5'-AAGGUUCAGACCAUCUUGGAA-3') were purchased from Bioneer (Daejeon, Korea). Si-14-3-3 γ (5'-GCGAGC-AACUGGUGCAGAA-3') was purchased from Genolution Pharmaceuticals (Seoul, Korea).

2.6. Gelatin Zymography. Gelatin zymography was performed using supernatants from sphere media cultivated in sphere-forming media on ultralow attachment plate as previously described [33]. Briefly, conditioned media was separated by 10% SDS-PAGE containing 0.2% gelatin. The gel was renatured with 2.5% Triton X-100 buffer for 1 h and incubated with developing solution (50 mM Tris, pH 7.5, and 10 mM CaCl₂) for 18–20 h at 37°C. The gel was then stained with 0.5% Coomassie Brilliant Blue (Sigma-Aldrich) in 30% methanol and 10% glacial acetic acid and destained with the same solution without dye.

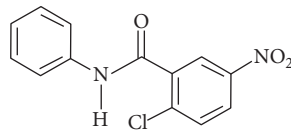
2.7. Statistics. The Student *t*-test was used to compare the differences between two groups. Each experiment was repeated at least three times and *p* values of <0.05 were considered as statistically significant.



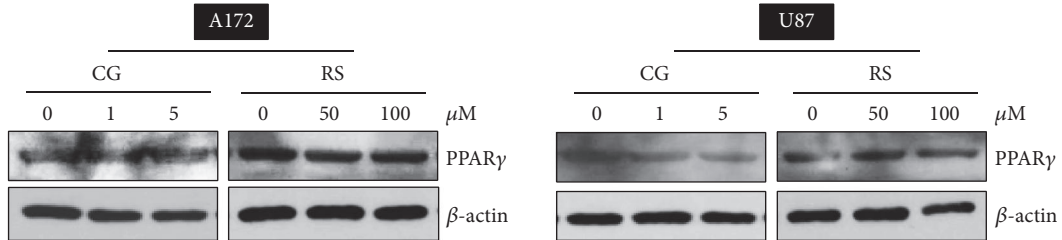
(a)



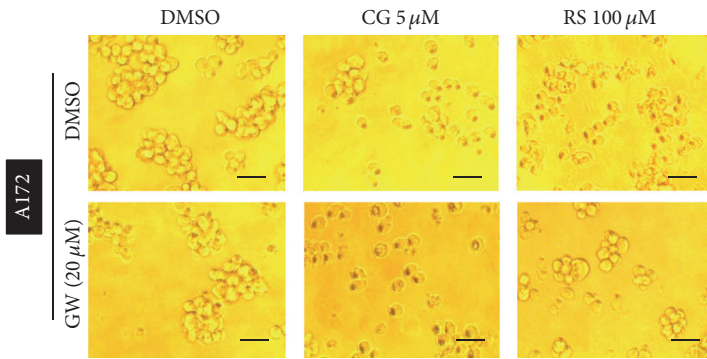
(b)



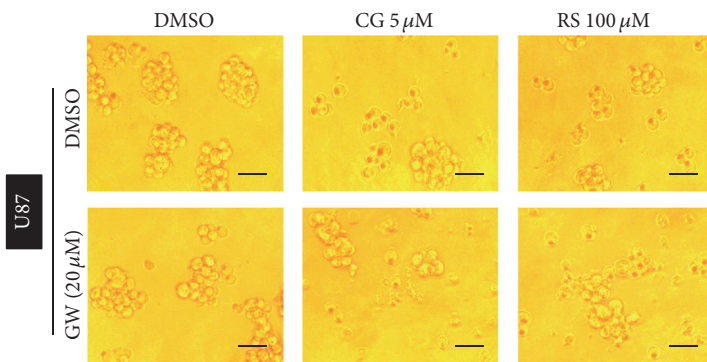
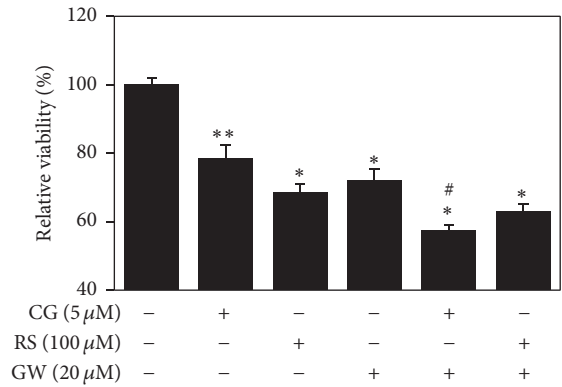
(c)



(d)



(e)



(f)

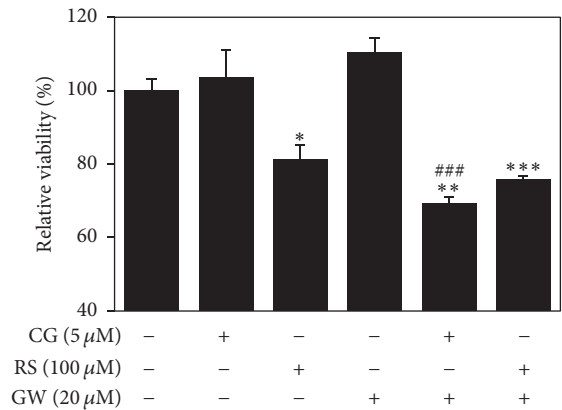


FIGURE 1: Continued.

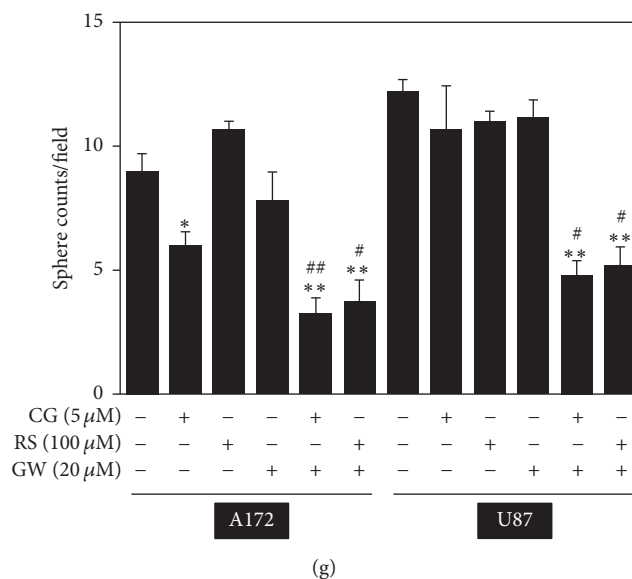


FIGURE 1: Structures of chemicals. (a) Ciglitazone (CG), $C_{18}H_{19}N_3O_3S$, and MW 357.428. (b) Rosiglitazone (RS), $C_{18}H_{23}NO_3S$, and MW 333.44. (c) GW9662 (GW), $C_{13}H_9ClN_2O_3$, and MW 276.7. (d) Effects of CG and RS on protein levels of PPAR γ . (e) and (f) combined treatments with PPAR γ ligand and GW9662 decreased sphere-forming activity and viability in A172 and U87 spheres. After treatment with PPAR γ ligand (CG or RS) with or without GW at the indicated concentrations for 48 h, spheres were photographed under an inverted microscope, cellular viability was measured using an assay kit and (g) spheres per field were counted as described in Materials and Methods. Scale bars: 100 μ m. * p < 0.05, ** p < 0.01, and *** p < 0.005 versus control group without treatment. # p < 0.05, ## p < 0.01, and ### p < 0.005 versus only GW treated group.

3. Results

3.1. Effects of Single or Cotreatment with PPAR γ Ligand and GW9662 on Glioblastoma Spheres. We verified that glioblastoma A172 and U87 cells express PPAR γ and single treatment with ciglitazone (CG) or rosiglitazone (RS) at a concentration tested in this study did not affect protein levels of PPAR γ as determined by western blotting (Figure 1(d)). To test our hypothesis that combined treatment with PPAR γ ligands and the PPAR γ inhibitor GW9662 (GW) yields a more favorable antiproliferative effect in glioblastoma spheres compared to single-agent treatments, we first examined its effects on cellular viability. To address this question, we utilized an in vitro sphere culture system [16] in which glioblastoma cells exhibited increased levels of SOX2, a representative stemness-related marker [34]. After treatment of a monolayer or spheres cultivated for 48 h with these agents, a WST-1 assay was performed as described in Materials and Methods. Combined treatment with PPAR γ ligands and GW9662 (Figure 1(a)) for 48 h resulted in a moderate cooperative antiproliferative effect on A172 and U87 spheres compared with single treatment (Figures 1(e) and 1(f)). The morphology of monolayers was not significantly altered (data not shown), whereas sphere-forming ability was reduced upon combined treatment with PPAR γ ligand and GW9662 compared with single-agent treatment (Figure 1(g)).

3.2. Cotreatment with PPAR γ Ligand and GW9662 Decreased BIS, 14-3-3 Gamma, and SOX2 Together with Reduced Phosphorylation of STAT3 and AKT. Since STAT3 and AKT are well-known regulators of proliferation and stemness, we

analyzed their protein levels in a monolayer and in spheres after single or combinatorial treatment via western blotting. Combined treatment decreased SOX2 protein levels in spheres compared with a monolayer (Figures 2(a) and 2(b)). Similarly, both BIS and 14-3-3 gamma were moderately reduced upon cotreatment with PPAR γ ligand and GW9662. Moreover, the combined treatment partially inhibited phosphorylation of both STAT and AKT in A172 and U87 spheres, indicating that the inhibitory effect on proliferation is mediated through the cellular survival pathway.

3.3. Effect of BIS or 14-3-3 γ Knockdown on Cellular Viability, MMP2, and SOX2. To address whether downregulation of BIS and 14-3-3 gamma by combined treatment with PPAR γ ligands and GW9662 sensitizes glioblastoma cells towards inhibition of proliferation, we performed knockdown experiments silencing BIS and 14-3-3 gamma. Cells were treated with a PPAR γ ligand agent for 48 h and a WST-1 assay was performed. Single treatment with si-BIS or si-14-3-3 γ siRNAs induced morphological changes in spheres compared with the si-CTL-treated control group (Figure 3(a)). Furthermore, cotreatment with siRNA and PPAR γ ligand significantly enhanced the inhibitory effect of WST activity (Figure 3(b)). Since cancer stem cells display a high potential for epithelial mesenchymal transition (EMT), which is subsequently linked to an invasive phenotype, we examined matrix metalloproteinase (MMP) activity as a critical agent for EMT [36, 37]. Data from zymography showed that combined treatment with 14-3-3 gamma depletion and CG or RS significantly inhibited MMP2 activity, while combination treatment with BIS depletion and RS did not (Figures 3(c) and 3(d)).

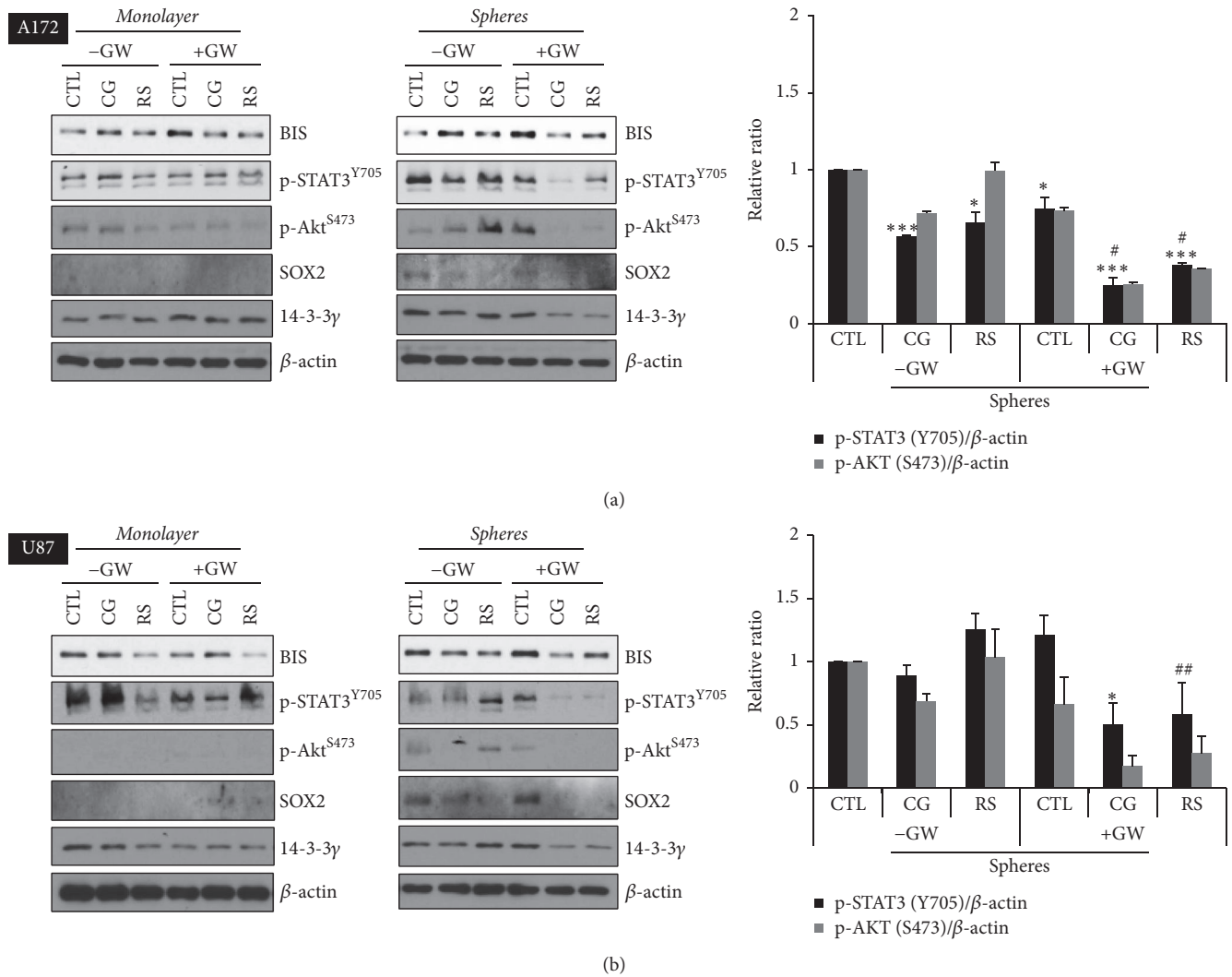


FIGURE 2: Combined treatment with PPAR γ ligand and GW9662 downregulated survival-related signaling molecules and stemness-related marker protein SOX2. After treatment with PPAR γ ligands (CG, 10 μ M ciglitazone; RS, 100 μ M rosiglitazone) with or without 20 μ M GW9662 (GW) for 48 h, monolayer cells or spheres of A172 (a) or U87 (b) were analyzed via western blotting with specific antibodies and the intensity of each protein per β -actin was measured using ImageJ software (right panels) as described in Materials and Methods. * $p < 0.05$ and *** $p < 0.005$ versus control group without treatment. # $p < 0.05$ and ## $p < 0.01$, versus only GW treated group.

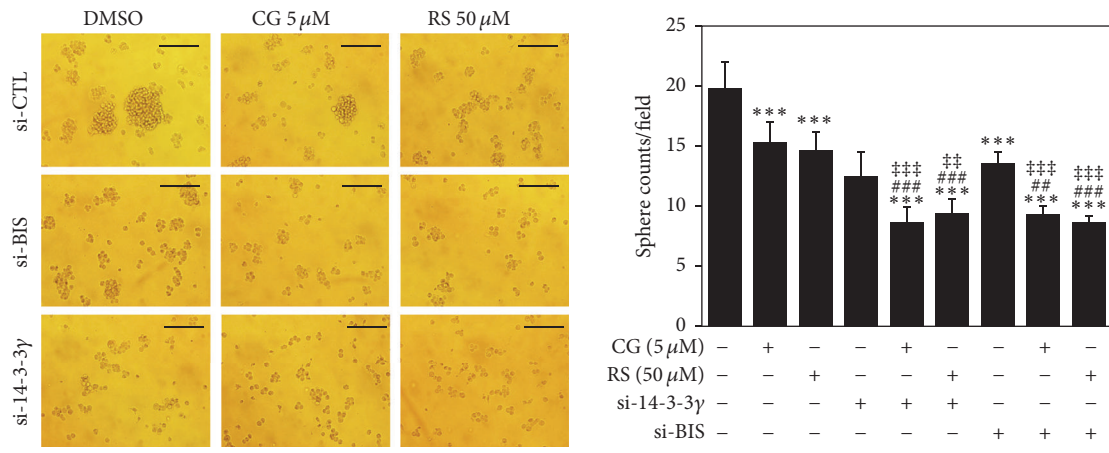
To examine whether these combination treatments might affect AKT, stemness, and apoptosis, we performed western blotting assays using specific antibodies. Single treatment with si-14-3- γ or si-BIS decreased phosphorylation of AKT and SOX2 levels without enhancing poly(ADP-ribose) polymerase (PARP) cleavage (Figure 3(e) and right panel). Finally, we examined whether 14-3-3 gamma or BIS depletion using siRNA affects sphere-forming activity. Single treatment with si-14-3-3 γ or si-BIS decreased sphere-forming activity and cotreatment with CG or RS moderately inhibited sphere-forming activity (Figures 3(f) and 3(g)).

4. Discussion

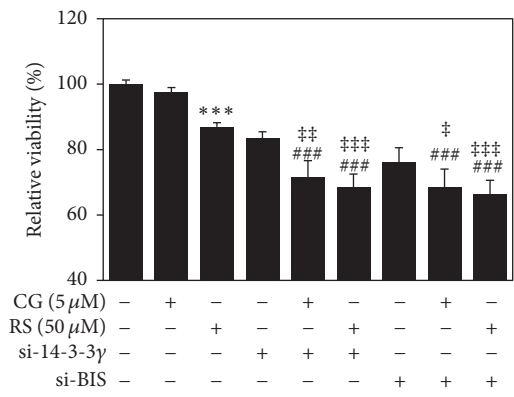
In this study, we demonstrated that a combined treatment with PPAR γ ligand and its inhibitor GW9662 downregulated

BIS and 14-3-3 gamma expression and decreased phosphorylation of STAT3 and AKT, leading to inhibition of GSC-like spheres and SOX2 expression (Figure 4). Combined administration of PPAR γ ligand and its antagonist targeted 14-3-3 gamma and BIS, inhibiting stem cell-like properties in GSC-like spheres expressing SOX2. Our findings provide additional evidence that BIS may be a potential target in glioblastoma [38].

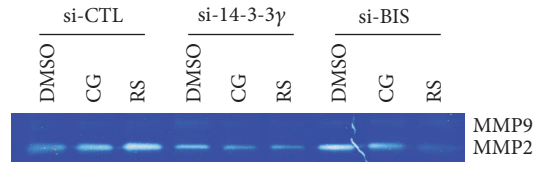
Emerging studies are focusing on combination therapy, such as chemo- and radiotherapy, to manage resistance from anticancer drug administration. We also found that A172 and U87 glioblastoma cells express PPAR γ using western blotting analysis. Hence, its ligands have been suggested as potential targets for glioblastoma stem cells [35]. Our findings provide a BIS- and 14-3-3 gamma-mediated inhibitory mechanism for the GSC-like sphere system suggesting the potential of



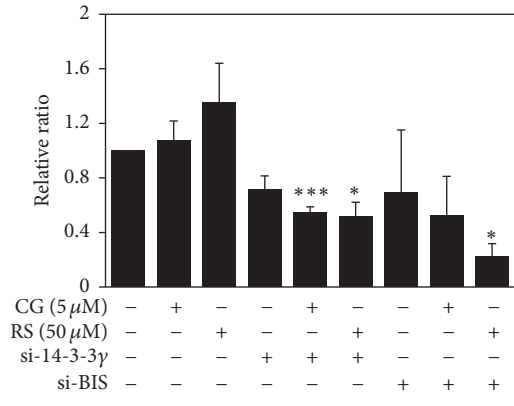
(a)



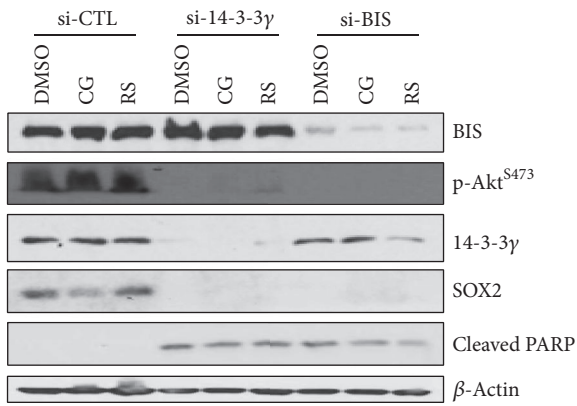
(b)



(c)



(d)



(e)

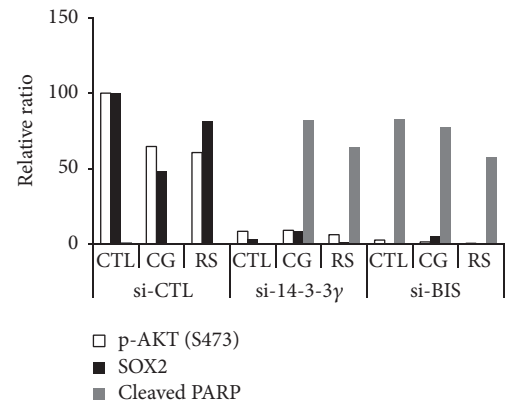


FIGURE 3: Continued.

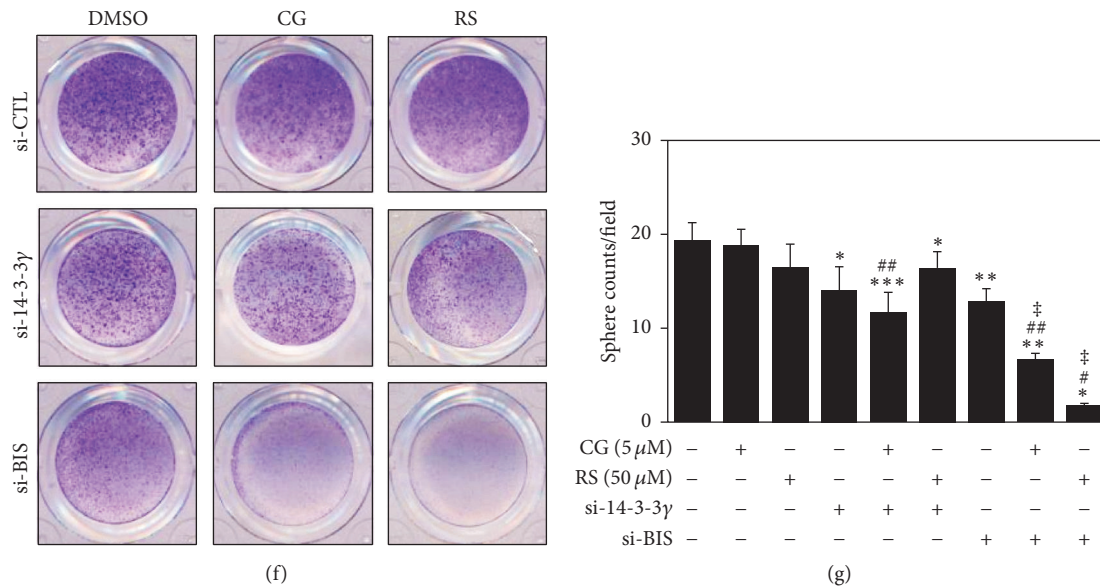


FIGURE 3: Downregulation using siRNA-enhanced PPAR γ ligand-induced cell death in A172 spheres together with inhibition of matrix metalloprotease 2 (MMP2) activity. After transfection with 100 nM siRNA (si-CTL, si-14-3-3 γ , or si-BIS), cells were transferred to sphere-forming conditions with or without PPAR γ ligands (CG, 5 μ M ciglitazone; RS, 50 μ M rosiglitazone) for 48 h. Morphological spheres (a) were photographed under an inverted microscope and spheres per field (right panel) were counted, followed by viability (b), zymography (c and d), and western blotting (e) and spheres per field were counted (g) after crystal violet staining (f) as described in Materials and Methods. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.005$ versus control group without treatment. # $p < 0.05$ and ## $p < 0.01$ versus only CG- or RS-treated group. † $p < 0.05$ versus only si-BIS or si-14-3-3 γ treated group. ### $p < 0.005$ versus only CG- or RS-treated group. ** $p < 0.01$ and *** $p < 0.005$ versus only si-BIS or si-14-3-3 γ treated group.

combination therapy targeting GSCs via the manipulation of the BIS and/or 14-3-3 gamma/SOX2 axis in which STAT3 and AKT function upstream. It should be noted that 14-3-3 γ or BIS silencing itself resulted in the considerable decrease in SOX2 levels as well as increase in PARP cleavage without synergistic and additive effect of being combined with CG or RS.

Since it has been reported that invasion is the hallmark of malignant glioblastoma and MMP [37, 39] is considered as an important player and a potential target, zymography was performed using supernatant of spheres after respective siRNA transfection to A172 cells. Single treatment with PPAR γ ligands did not significantly inhibit MMP2 activity but cotreatment with si-BIS or si-14-3-3 γ inhibited MMP2 activity suggesting that these molecules might play a role in regulating MMP2 activity. It was consistent with findings from previous studies that BIS inhibits MMP2 activity [33, 40]. Meanwhile, Kim et al. [41] reported that CG increased the expression of MMP-2, and GW9662 attenuated the CG-induced PPAR γ activation but it did not affect the pro-MMP2 activation in a fibrosarcoma cell line HT1080. Although there is difference in cellular context, significant inhibition of MMP2 activity by CG or RG single treatment in this study was not observed.

We tested whether si-14-3-3 beta and gamma affect sphere-forming activity and found that single treatment with si-14-3-3 gamma led to moderately reduced sphere-forming activity compared to si-14-3-3 beta (data not shown), meaning that 14-3-3 gamma may play a significant role in

glioblastoma stemness. Each isoform appears to differentially modulate cellular progression. 14-3-3 beta negatively regulates senescence in glioblastoma cells through the ERK pathway [42] and 14-3-3 gamma controls stem cell-like properties, as described in this study. We observed that 14-3-3 gamma or BIS deletion downregulated the SOX2 expression level without enhancing PARP cleavage, suggesting that combined treatment with PPAR ligand and its inhibitor may modulate the stemness-related pathway through AKT and STAT3 rather than the apoptosis-related pathway. Frasson et al. reported that the PI3K/AKT pathway is closely related to the stemness of medulloblastoma cancer stem cells [43], supporting our findings of combined administration in the downregulation of SOX2 in glioblastoma cells. We previously reported that BIS depletion degrades STAT3 protein leading to inhibition of sphere-forming activity [33].

As an important survival signaling kinase, AKT phosphorylates BAD at Ser136 promoting binding of BAD to 14-3-3 proteins, preventing an association between BAD and BCL-2 and BCL-xL [44]. 14-3-3 sequesters BAD in the cytoplasm under physiologically normal conditions. When cells are stressed, BAD is phosphorylated and this phosphorylated BAD (p-BAD) translocates from the cytoplasm into mitochondria leading to apoptosis. Although BIS forms an immunoreactive complex with 14-3-3 zeta [45], suggesting that BIS may affect BAD directly or indirectly through 14-3-3, we did not find that BIS or 14-3-3 gamma depletion enhanced PPAR γ ligand-induced PARP cleavage. Rather, single treatment significantly downregulated SOX2 expression levels.

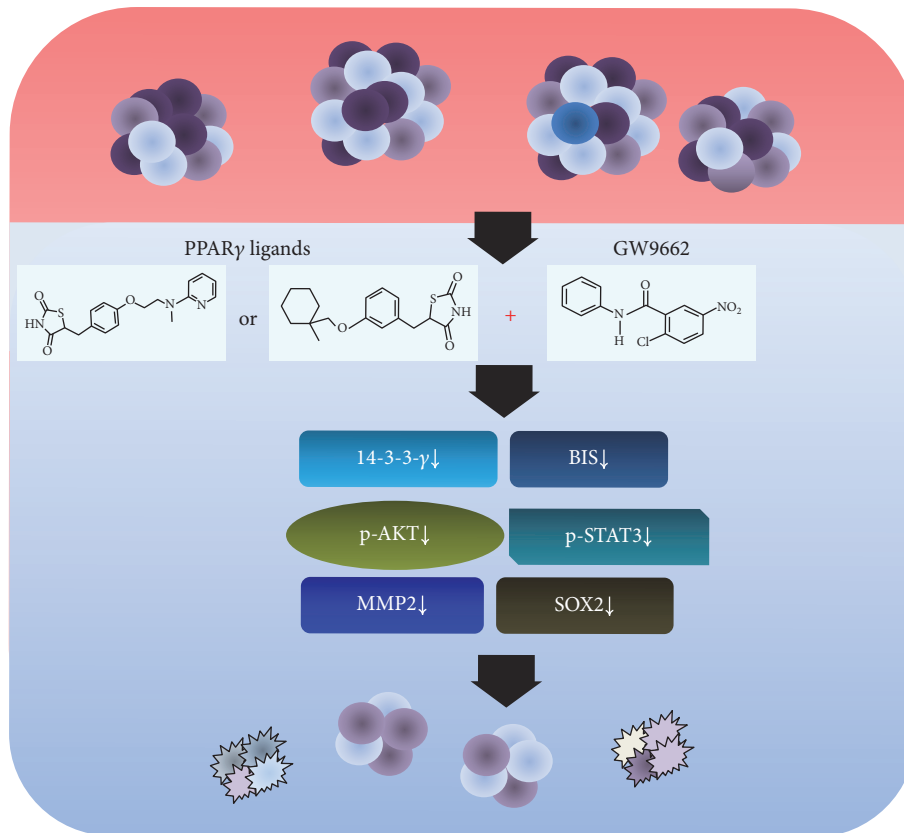


FIGURE 4: A scheme of the combined effects of PPAR γ ligand and GW9662 on human glioblastoma cells under sphere-forming conditions. The schematic diagram demonstrates that combination treatment with PPAR γ ligand (ciglitazone or rosiglitazone) and GW9662 downregulates BIS, 14-3-3 gamma, and activation of survival signaling molecules (p-AKT, p-STAT3) together with suppression of SOX2 and MMP2 activity, inhibiting stem cell-like properties and leading to apoptosis.

Hence, they may act directly or indirectly upon SOX2. The underlying mechanism by which BIS or 14-3-3 beta regulates SOX2 expression requires further investigation.

5. Conclusions

Combination treatment of GCSs with PPAR γ ligand and its inhibitor GW9662 inhibited stem-like properties via downregulation of BIS and 14-3-3 gamma levels together with decreased SOX2 and MMP2 activity without enhancing PARP cleavage.

Disclosure

The English in this document has been checked by at least two professional editors, both native speakers of English. For a certificate, please see <http://www.textcheck.com/certificate/JSHuCk>.

Conflicts of Interest

The author declares that there are no conflicts of interest regarding the publication of this paper.

Authors' Contributions

Chang-Nim Im conceived the concept; Chang-Nim Im designed and performed the experiments, analyzed the data, and wrote the paper.

Acknowledgments

This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT and future Planning (NRF-2014R1A1A1006961 and NRF-2017R1A1A1A05000839). The author is grateful to Jung-Sook Im for her inspiration and support.

References

- [1] J. G. Gurney and N. Kadan-Lottick, "Brain and other central nervous system tumors: rates, trends, and epidemiology," *Current Opinion in Oncology*, vol. 13, no. 3, pp. 160–166, 2001.
- [2] X. Zhang, W. Zhang, X. Mao, H. Zhen, W. Cao, and S. Hu, "Targeting role of glioma stem cells for glioblastoma multiforme," *Current Medicinal Chemistry*, vol. 20, no. 15, pp. 1974–1984, 2013.
- [3] M. Jackson, F. Hassiotou, and A. Nowak, "Glioblastoma stem-like cells: at the root of tumor recurrence and a therapeutic target," *Carcinogenesis*, vol. 36, no. 2, pp. 177–185, 2015.

- [4] A. Chawla, E. J. Schwarz, D. D. Dimaculangan, and M. A. Lazar, "Peroxisome proliferator-activated receptor (PPAR) γ : adipose-predominant expression and induction early in adipocyte differentiation," *Endocrinology*, vol. 135, no. 2, pp. 798–800, 1994.
- [5] M. Kato, T. Nagaya, M. Fujieda, K. Saito, J. Yoshida, and H. Seo, "Expression of PPAR γ and its ligand-dependent growth inhibition in human brain tumor cell lines," *Japanese Journal of Cancer Research*, vol. 93, no. 6, pp. 660–666, 2002.
- [6] M. W. Lee, D. S. Kim, H. R. Kim et al., "Cell death is induced by ciglitazone, a peroxisome proliferator-activated receptor gamma (PPARgamma) agonist, independently of PPARgamma in human glioma cells," *Biophysical Research Communications*, vol. 417, no. 1, pp. 552–557, 2012.
- [7] D. W. Kang, C. H. Choi, J. Y. Park, S. K. Kang, and Y. K. Kim, "Ciglitazone induces caspase-independent apoptosis through down-regulation of XIAP and survivin in human glioma cells," *Neurochemical Research*, vol. 33, no. 3, pp. 551–561, 2008.
- [8] D. C. Liu, C. B. Zang, H. Y. Liu, K. Possinger, S. G. Fan, and E. Elstner, "A novel PPAR alpha/gamma dual agonist inhibits cell growth and induces apoptosis in human glioblastoma T98G cells," *Acta Pharmacologica Sinica*, vol. 25, no. 10, pp. 1312–1319, 2004.
- [9] R. Morosetti, T. Servidei, M. Mirabella et al., "The PPAR γ ligands PGJ2 and rosiglitazone show a differential ability to inhibit proliferation and to induce apoptosis and differentiation of human glioblastoma cell lines," *International Journal of Oncology*, vol. 25, no. 2, pp. 493–502, 2004.
- [10] N. Strakova, J. Ehrmann, P. Dzubak, J. Bouchal, and Z. Kolar, "The synthetic ligand of peroxisome proliferator-activated receptor- γ ciglitazone affects human glioblastoma cell lines," *Journal of Pharmacology and Experimental Therapeutics*, vol. 309, no. 3, pp. 1239–1247, 2004.
- [11] N. Strakova, J. Ehrmann, J. Bartos, J. Malikova, J. Dolezel, and Z. Kolar, "Peroxisome proliferator-activated receptors (PPAR) agonists affect cell viability, apoptosis and expression of cell cycle related proteins in cell lines of glial brain tumors," *Neoplasma*, vol. 52, no. 2, pp. 126–136, 2005.
- [12] K. L. Schaefer, K. Wada, H. Takahashi et al., "Peroxisome proliferator-activated receptor γ inhibition prevents adhesion to the extracellular matrix and induces anoikis in hepatocellular carcinoma cells," *Cancer Research*, vol. 65, no. 6, pp. 2251–2259, 2005.
- [13] H. P. Ellis and K. M. Kurian, "Biological rationale for the use of PPARgamma agonists in glioblastoma," *Frontiers in Oncology*, vol. 4, p. 52, 2014.
- [14] W. Chearwae and J. J. Bright, "PPAR γ agonists inhibit growth and expansion of CD133+ brain tumour stem cells," *British Journal of Cancer*, vol. 99, no. 12, pp. 2044–2053, 2008.
- [15] C. Grommes, D. S. Conway, A. Alshekhlee, and J. S. Barnholtz-Sloan, "Inverse association of PPAR γ agonists use and high grade glioma development," *Journal of Neuro-Oncology*, vol. 100, no. 2, pp. 233–239, 2010.
- [16] E. Pestereva, S. Kanakasabai, and J. J. Bright, "PPAR γ agonists regulate the expression of stemness and differentiation genes in brain tumour stem cells," *British Journal of Cancer*, vol. 106, no. 10, pp. 1702–1712, 2012.
- [17] G. Karpel-Massler, M. A. Westhoff, S. Zhou et al., "Combined inhibition of HER1/EGFR and RAC1 results in a synergistic antiproliferative effect on established and primary cultured human glioblastoma cells," *Molecular Cancer Therapeutics*, vol. 12, no. 9, pp. 1783–1795, 2013.
- [18] G. Karpel-Massler, C. Shu, L. Chau et al., "Combined inhibition of Bcl-2/Bcl-xL and Usp9X/Bag3 overcomes apoptotic resistance in glioblastoma in vitro and in vivo," *Oncotarget*, vol. 6, pp. 14507–14521, 2015.
- [19] S. Liang, G. Shen, Q. Liu et al., "Isoform-specific expression and characterization of 14-3-3 proteins in human glioma tissues discovered by stable isotope labeling with amino acids in cell culture-based proteomic analysis," *Proteomics–Clinical Applications*, vol. 3, no. 6, pp. 743–753, 2009.
- [20] X. Yang, W. Cao, H. Lin et al., "Isoform-specific expression of 14-3-3 proteins in human astrocytoma," *Journal of the Neurological Sciences*, vol. 276, no. 1-2, pp. 54–59, 2009.
- [21] L. Cao, W. Cao, W. Zhang et al., "Identification of 14-3-3 protein isoforms in human astrocytoma by immunohistochemistry," *Neuroscience Letters*, vol. 432, no. 2, pp. 94–99, 2008.
- [22] F. H. Gong, G. L. Wang, J. Ye, T. D. Li, H. M. Bai, and W. M. Wang, "14-3-3 beta regulates the proliferation of glioma cells through the GSK3 beta/beta-catenin signaling pathway," *Oncology Reports*, vol. 30, no. 6, pp. 2976–2982, 2013.
- [23] T. C. Chang, C. C. Liu, E. W. Hsing et al., "14-3-3 sigma regulates beta-catenin-mediated mouse embryonic stem cell proliferation by sequestering GSK-3 beta," *PLoS ONE*, vol. 7, no. 6, 2012.
- [24] Y. H. Jin, Y. J. Kim, D. W. Kim et al., "Sirt2 interacts with 14-3-3 beta/gamma and down-regulates the activity of p53," *Biochemical and Biophysical Research Communications*, vol. 368, no. 3, pp. 690–695, 2008.
- [25] H. Hermeking, "The 14-3-3 cancer connection," *Nature Reviews Cancer*, vol. 3, no. 12, pp. 931–943, 2003.
- [26] K. K. Wu, "Peroxisome proliferator-activated receptors protect against apoptosis via 14-3-3," *PPAR Research*, vol. 2010, Article ID 417646, 7 pages, 2010.
- [27] K. K. Wu and J. Y. Liou, "Cyclooxygenase inhibitors induce colon cancer cell apoptosis via PPARdelta \rightarrow 14-3-3epsilon pathway," *Methods in Molecular Biology*, vol. 512, pp. 295–307, 2009.
- [28] J. Lee, T. Takahashi, N. Yasuhara, J. Inazawa, S. Kamada, and Y. Tsujimoto, "Bis, a Bcl-2-binding protein that synergizes with Bcl-2 in preventing cell death," *Oncogene*, vol. 18, no. 46, pp. 6183–6190.
- [29] S. Takayama, Z. Xie, and J. C. Reed, "An evolutionarily conserved family of Hsp70/Hsc70 molecular chaperone regulators," *Journal of Biological Chemistry*, vol. 274, no. 2, pp. 781–786, 1999.
- [30] H. Doong, K. Rizzo, S. Fang, V. Kulpa, A. M. Weissman, and E. C. Kohn, "CAIR-1/BAG-3 abrogates heat shock protein-70 chaperone complex-mediated protein degradation," *Journal of Biological Chemistry*, vol. 278, no. 31, pp. 28490–28500, 2003.
- [31] A. Gentilella and K. Khalili, "BAG3 expression in glioblastoma cells promotes accumulation of ubiquitinated clients in an Hsp70-dependent manner," *Journal of Biological Chemistry*, vol. 286, no. 11, pp. 9205–9215, 2011.
- [32] M. Festa, L. Del Valle, K. Khalili et al., "BAG3 protein is overexpressed in human glioblastoma and is a potential target for therapy," *The American Journal of Pathology*, vol. 178, no. 6, pp. 2504–2512, 2011.
- [33] C. N. Im, H. HyeonYun, B. Song et al., "BIS-mediated STAT3 stabilization regulates glioblastoma stem cell-like phenotypes," *Oncotarget*, vol. 7, no. 23, pp. 35056–35070, 2016.
- [34] C. N. Im, H. H. Yun, H. J. Yoo, M. J. Park, and J. H. Lee, "Enhancement of SOX-2 expression and ROS accumulation by culture of A172 glioblastoma cells under non-adherent culture conditions," *Oncology Reports*, vol. 34, no. 2, pp. 920–928, 2015.

- [35] C. N. Im, "Targeting glioblastoma stem cells (GSCs) with peroxisome proliferator-activated receptor gamma (PPAR γ) ligands," *IUBMB Life*, vol. 68, no. 3, pp. 173–177, 2016.
- [36] S. A. Mani, W. Guo, and M.-J. Liao, "The epithelial-mesenchymal transition generates cells with properties of stem cells," *Cell*, vol. 133, no. 4, pp. 704–715, 2008.
- [37] P. Gabelloni, E. Da Pozzo, S. Bendinelli et al., "Inhibition of metalloproteinases derived from tumours: new insights in the treatment of human glioblastoma," *Neuroscience*, vol. 168, no. 2, pp. 514–522, 2010.
- [38] S. An, D. He, E. Wagner, and C. Jiang, "Peptide-like polymers exerting effective glioma-targeted siRNA delivery and release for therapeutic application," *Small*, vol. 11, no. 38, pp. 5142–5150, 2015.
- [39] K. Yasui, M. Shimamura, N. Mitsutake, and Y. Nagayama, "SNAIL induces epithelial-to-mesenchymal transition and cancer stem cell-like properties in aldehyde dehydrogenase-negative thyroid cancer cells," *Thyroid*, vol. 23, no. 8, pp. 989–996, 2013.
- [40] C. N. Im, H. H. Yun, and J. H. Lee, "Heat shock factor 1 depletion sensitizes A172 glioblastoma cells to temozolomide via suppression of cancer stem cell-like properties," *International Journal of Molecular Sciences*, vol. 18, no. 2, p. 468, 2017.
- [41] K. H. Kim, Y. S. Cho, J. Park, S. Yoon, K. Kim, and A. Chung, "Pro-MMP-2 activation by the PPAR γ agonist, ciglitazone, induces cell invasion through the generation of ROS and the activation of ERK," *FEBS Letters*, vol. 581, no. 17, pp. 3303–3310, 2007.
- [42] S. B. Seo, J. Lee, H. H. Yun et al., "14-3-3 β depletion drives a senescence program in glioblastoma cells through the ERK/SKP2/p27 pathway," *Molecular Neurobiology*.
- [43] C. Frasson, E. Rampazzo, B. Accordi et al., "Inhibition of PI3K signalling selectively affects medulloblastoma cancer stem cells," *BioMed Research International*, vol. 2015, Article ID 973912, 11 pages, 2015.
- [44] S. R. Datta, H. Dudek, X. Tao et al., "Akt phosphorylation of BAD couples survival signals to the cell- intrinsic death machinery," *Cell*, vol. 91, no. 2, pp. 231–241, 1997.
- [45] J. J. Lee, J. S. Lee, M. N. Cui et al., "BIS targeting induces cellular senescence through the regulation of 14-3-3 zeta/STAT3/SKP2/p27 in glioblastoma cells," *Cell Death and Disease*, vol. 5, no. 11, p. e1537, 2014.