



Draft Genome Sequences of Two *Pseudomonas aeruginosa* Isolates from the Female Urogenital Tract

Genevieve Johnson,^a Carine R. Mores,^{b,c} Alan J. Wolfe,^b  Catherine Putonti^{a,b,c,d}

^aDepartment of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, Maywood, Illinois, USA

^bDepartment of Biology, Loyola University Chicago, Chicago, Illinois, USA

^cBioinformatics Program, Loyola University Chicago, Chicago, Illinois, USA

^dDepartment of Computer Science, Loyola University Chicago, Chicago, Illinois, USA

ABSTRACT *Pseudomonas aeruginosa* is a Gram-negative bacterium that has the ability to survive in and readily adapt to a variety of environmental conditions. Here, we report 2 genome sequences of *P. aeruginosa* strains, UMB1046 and UMB5686, isolated from the female urogenital tract.

Pseudomonas aeruginosa is an opportunistic pathogen in compromised hosts but is harmless to healthy individuals. *P. aeruginosa* is associated with chronic lung infections in individuals with cystic fibrosis (1), as well as nosocomial urinary tract infections (2). While it is not frequently found within the urogenital microbiota of healthy women (3, 4), strains have been isolated from women with lower urinary tract symptoms (3, 5, 6). Here, we present the genomes of two *P. aeruginosa* strains isolated from two different women. *P. aeruginosa* UMB5686 was isolated from a vaginal swab sample obtained from a woman with overactive bladder (OAB) symptoms after 12 weeks of treatment with a vaginal estrogen cream (5). *P. aeruginosa* UMB1046 was isolated from a catheterized urine sample obtained from a woman with a urinary tract infection (6).

P. aeruginosa UMB1046 and UMB5686 were isolated from prior institutional review board (IRB)-approved studies (5, 6) using the expanded quantitative urinary culture (EQUC) protocol (5). Briefly, vaginal swabs were collected using the BD liquid Amies elution swab (ESwab) collection system and cultured as previously described (5); catheterized urine samples were cultured as previously described (6). The genus and species for these isolates were determined via matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry prior to storage at -80°C . From these freezer stocks, each *P. aeruginosa* isolate was first streaked on an LB agar plate and incubated at 37°C for 24 hours. A single colony was selected from each plate to inoculate LB broth and incubated at 37°C with shaking for 24 hours. DNA was extracted using the Qiagen DNeasy UltraClean microbial kit and quantified using the Qubit fluorometer. DNA was sent to the Microbial Genomic Sequencing Center (MiGS) at the University of Pittsburgh for sequencing, where the DNA was first enzymatically fragmented using an Illumina tagmentation enzyme. Indices were attached using PCR and sequenced using an Illumina NextSeq 500 flow cell, producing 938,702 and 1,641,347 pairs of 151-bp reads for UMB1046 and UMB5686, respectively. Raw reads were trimmed using Sickle v1.33 (<https://github.com/najoshi/sickle>) and assembled using SPAdes v3.13.0 with the “only-assembler” option for k values of 55, 77, 99, and 127 (7). Genome coverage was calculated using BMap v38.47 (<https://sourceforge.net/projects/bbmap/>). The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.8 (8) was used to annotate the genome sequences. Unless previously noted, default parameters were used for each software tool.

Citation Johnson G, Mores CR, Wolfe AJ, Putonti C. 2020. Draft genome sequences of two *Pseudomonas aeruginosa* isolates from the female urogenital tract. Microbiol Resour Announc 9:e01378-19. <https://doi.org/10.1128/MRA.01378-19>.

Editor David A. Baltus, University of Arizona

Copyright © 2020 Johnson et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Catherine Putonti, cputonti@luc.edu.

Received 3 November 2019

Accepted 14 November 2019

Published 2 January 2020

The *P. aeruginosa* UMB1046 genome is 6,513,817 bp long in 161 contigs with a GC content of 64%, genome coverage of 34.03 \times , and an N_{50} score of 64,232 bp. The *P. aeruginosa* UMB5686 genome has a similar size, 6,684,697 bp, with a GC content of 63%. The UMB5686 assembly includes 98 contigs with a coverage of 58.48 \times and an N_{50} score of 120,243 bp. PGAP annotation identified 6,257 and 6,351 protein-coding genes for UMB1046 and UMB5686, respectively. The strains vary in their number of rRNA operons and tRNAs; UMB1046 carries 3 rRNA operons and 58 tRNAs, whereas UMB5686 carries 4 rRNA operons and 59 tRNAs. Future analyses of these strains and genomes will further our understanding of this opportunistic pathogen within the female urogenital tract.

Data availability. This whole-genome shotgun (WGS) project has been deposited in GenBank under the accession numbers [WHVN00000000](#) and [WHVM00000000](#) for *P. aeruginosa* UMB1046 and UMB5686, respectively. The raw sequence reads have been deposited under accession numbers [SRR10336114](#) and [SRR10336113](#) for *P. aeruginosa* UMB1046 and UMB5686, respectively. The WGS and SRA records are associated with BioProject number [PRJNA316969](#).

ACKNOWLEDGMENTS

We thank the authors of the prior studies that isolated the bacteria. For prior patient recruitment, we acknowledge the Loyola Urinary Education and Research Collaborative (LUEREC), specifically Mary Tulke, Linda Brubaker, Elizabeth Mueller, Cynthia Brincat, Susanne Taege, and Tanaka Dune, and the patients who provided the samples for this study.

G.J. is partially supported by a Mulcahy research fellowship from Loyola University Chicago.

REFERENCES

1. Faure E, Kwong K, Nguyen D. 2018. *Pseudomonas aeruginosa* in chronic lung infections: how to adapt within the host? *Front Immunol* 9:2416. <https://doi.org/10.3389/fimmu.2018.02416>.
2. Djordjevic Z, Folic MM, Zivic Z, Markovic V, Jankovic SM. 2013. Nosocomial urinary tract infections caused by *Pseudomonas aeruginosa* and *Acinetobacter* species: sensitivity to antibiotics and risk factors. *Am J Infect Control* 41:1182–1187. <https://doi.org/10.1016/j.ajic.2013.02.018>.
3. Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. *J Clin Microbiol* 52:871–876. <https://doi.org/10.1128/JCM.02876-13>.
4. Ma B, Forney LJ, Ravel J. 2012. The vaginal microbiome: rethinking health and diseases. *Annu Rev Microbiol* 66:371–389. <https://doi.org/10.1146/annurev-micro-092611-150157>.
5. Thomas-White K, Forster SC, Kumar N, Van Kuiken M, Putonti C, Stares MD, Hilt EE, Price TK, Wolfe AJ, Lawley TD. 2018. Culturing of female bladder bacteria reveals an interconnected urogenital microbiota. *Nat Commun* 9:1557. <https://doi.org/10.1038/s41467-018-03968-5>.
6. Price TK, Dune T, Hilt EE, Thomas-White KJ, Kliethermes S, Brincat C, Brubaker L, Wolfe AJ, Mueller ER, Schreckenberger PC. 2016. The clinical urine culture: enhanced techniques improve detection of clinically relevant microorganisms. *J Clin Microbiol* 54:1216–1222. <https://doi.org/10.1128/JCM.00044-16>.
7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
8. Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.