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CASE SERIES

Post-LASIK exacerbation of granular corneal dystrophy type 2 in members of a chinese family

Abstract

Purpose The post-LASIK exacerbation of corneal dystrophy, otherwise asymptomatic, is almost exclusively associated with the *TGFBI* gene mutations at codon 124 in exon 4 and codon 555 in exon 12. It is our intention to demonstrate that the pre-operative genetic screening for *TGFBI* mutations should be mandatory for refractive surgery candidates.

Patients and Methods In this study, we reviewed the proband's post-LASIK slit-lamp and in vivo confocal microscopy images and genetic testing results, and performed genetic testing on eleven additional members of the family to investigate the penetrance of corneal dystrophy in asymptomatic members who carry the mutation. Results The proband demonstrated a post-LASIK exacerbation of Granular Corneal Dystrophy type 2 (GCD2), identified as a TGFBI R124H mutation. Three of the 11 family members tested positive for the same R124H mutation as the proband. Conclusion The lesson learned from this case is that the genetic screening of TGFBI mutations must be incorporated into the preoperative screening procedures to prevent exacerbation and recurrence, which eventually could lead to the need for a corneal transplant.

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Introduction

As the first report of autosomal-dominant Granular Corneal Dystrophy type 2 (GCD2) in individuals from Avellino, Italy¹ in 1988, many cases of post-laser surgery exacerbation have been reported worldwide.^{1–7} Inherited corneal dystrophy exacerbation is characterized by bilateral opacity in anterior corneal stroma leading to a severe decrease of the best corrected visual acuity (BCVA) and ultimately to surgical treatment.^{8–10}

The Avellino Universal Test examines the five most common *TGFBI* corneal dystrophies, each triggered by different mutations in exons 4 and 12 of the *TGFBI* gene, located on chromosome 5q31.1. Purified DNA is extracted from oral epithelial cells collected by buccal swabs and the genotype of the LASIK candidate obtained by amplifying the targeted DNA point mutations (Table 1).

Herein, we report the results of ophthalmic and genetic examination of an individual with post-LASIK GCD2 and his family from Jiangsu province, China.

Case series

Case 1

The proband is a 29-year-old Chinese male who underwent bilateral LASIK surgery in 2006. The post-operative uncorrected visual acuity (UCVA) in both eyes was 20/20. He had an uneventful post-operative course and did not have regular follow-up examinations after the surgery. In July 2016, 10 years after LASIK surgery, he was referred to Shanghai First People's Hospital for evaluation of dryness, foreign body sensation, and mildly decreased vision acuity in both eyes. The findings were opacities in both of his corneas. Slip-lamp examination was conducted (Figure 1a and b). His UCVA was 16/20 in the right eye and 12/20 in the left eye. The manifest refraction was - 1.00 diopters cylinder

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Table 1	The five most	common TGFBI	corneal	dystrophies	are GDC2,	LCD1,	RBCD,	GCD1,	and TE	3CD
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Corneal dystrophy name	Exon location	Nucleotide change	Sequence change	Amino-acid change
Granular corneal dystrophy, type 2	4	c.418G>A	CGC>CAC	p.R124H
Lattice corneal dystrophy, type 1	4	c.417C>T	CGC>TGC	p.R124C
Reis-Buckler corneal dystrophy	4	c.418G>T	CGC>CTC	p.R124L
Granular corneal dystrophy, type 1	12	c.1663C>T	CGG>TGG	p.R555W
Thiel-Behnke corneal dystrophy	12	c.1664G>A	CGG>CAG	p.R555Q

The respective exon locations, nucleotide changes, and amino-acid changes of their causative mutations are listed (Nucleotide change and Sequence change information is from HGMD by QIAGEN, Hilden, Germany). A sterile Copan buccal swab (Copan, Brescia, Italy) was used for collecting oral epithelial cells. The swabs were then sent to Avellino Labs China for testing using the Universal Test, which detects the five most common TGFBI corneal dystrophies. The laboratory performed a DNA extraction from the swab using the DNA Extract All Reagents kit (Thermo Fisher Scientific Inc., Waltham, MA, USA). The purified DNA was then amplified by the TaqMan GTXpress Master Mix (Thermo Fisher Scientific Inc.) with Life Technology's 7500 FAST real-time PCR system (Thermo Fisher Scientific Inc.). Through a series cycle of denaturing, annealing and extension, the 7500 instrument collects the amplification fluorescence signals of the targets and generates a result from which the genotype may be inferred.

 \times 30 oculus dexter and -0.5 oculus sinister, yielding a BCVA of 20/20 in the right eye and 16/20 in the left eye. Subsequent examination

by *in vivo* confocal microscopy (IVCM) revealed a large quantity of dense white deposits of various shapes and sizes which presented as hyper-reflective extracellular structures located mainly in the anterior stroma (Figure 1c and d).

To uncover the cause of these deposits, the proband underwent genetic testing with the Universal Test (Avellino Labs China, Shanghai, China) which showed that he harbored a heterozygous R124H GCD2 mutation but not the R124C LCD1, R124L RBCD, R555W GCD1 or R555Q TBCD mutations (Table 1).

Case 2

The proband's older sister is a 38-year-old female without any history of ocular surgery, eye complaints, or decreased vision. Following DNA testing, which revealed a heterozygous result for the R124H mutation, she was evaluated by IVCM which demonstrated similar hyperreflective extracellular opacities at the anterior stroma with fewer, lower density deposits (Figure 1e and f).

Case 3

GCD2 is an autosomal-dominant disorder caused by the R124H *TGFBI* mutation. A study of 10 family members, in addition to the proband and his sister, was conducted and a family tree with test results was constructed (Figure 2d). The proband's mother and his 15-year-old nephew (his sister's son) also tested positive. Slit-lamp examination and IVCM were not conducted on these individuals due to limited access to a suitably equipped medical facility. Testing showed that the proband's 7-year-old son had not inherited the mutation. The mutation status of the proband's grandparents was unknown. Since the proband's aunt tested negative and the medical history of the deceased uncle is unknown, it is unclear whether the proband's mother inherited the mutation or it arose *de novo*.

Discussion

Corneal dystrophies contraindicate refractive surgery due to the likelihood of recurrence and exacerbation. As they may be difficult to determine by family history and clinical examination alone, a genetic test to detect TGFBI mutations should be incorporated into standard practice as one of the prescreening tools for refractive surgeries. In Case 1, the patient displayed no clinical symptoms and passed the LASIK surgery prescreening examination. Ten years after surgery, he developed symptoms that affected his vision. A genetic test identified that most likely the cause of the stromal deposits was the TGFBI R124H heterozygous mutation. The proband's 15-year-old nephew inherited the mutation from proband's sister, who tested positive for the mutation without symptoms; however, the examination conducted after the testing revealed that she too had corneal stromal deposits. These two cases demonstrate that a lack of clinical signs does not mean the absence of corneal dystrophy-causing mutations. It is justified to test the asymptomatic refractive surgery candidates to rule out disease-causing mutations.

Corneal dystrophy is a disease with a low prevalence (US: 1:1115,¹¹ Korea: 1:870,¹² China 1:416⁷) and debilitating outcome, ultimately resulting in corneal transplant as a treatment. The recurrent nature of the disease can result in multiple corneal transplants. Therefore, prevention and prescreening with a genetic test to detect the mutations, in addition to a thorough clinical examination is key.



Figure 1 Slit-Lamp (Nikon Corporation, Tokyo, Japan) photographs of the proband revealed dense, fine, white granules in the central corneas of both eyes, located at the interface between the flap and stromal bed on the right (a) and left (b) eyes. *In vivo* confocal microscopy (IVCM) (Rostock Cornea Module of Retina Tomograph (HRT/RCM); Heidelberg Engineering GmbH, Heidelberg, Germany) images of the right (c) and left (d) anterior stroma with hyper-reflective extracellular deposits. *In vivo* confocal microscopy (IVCM) images of the right (e) and left (f) anterior stroma with hyper-reflected extracellular deposits observed in the corneas of the proband's sister.



Figure 2 The amplification plot displays normalized dye fluorescence (Δ Rn) as a function of cycle number. The magenta slope in the amplification plot represents the normal allele (allele 1) and the blue slope represents the mutant allele (allele 2). (a) This plot shows the Proband's sample of heterozygous mutation (blue slope) being amplified, which indicates the presence of mutation on allele 2. (b and c) amplification plots of GCD2 normal genotype and GCD2 homozygous mutation genotype for comparison purposes. (d) Family tree with test result of Heterozygous mutation plot. It is unknown whether the mutation was passed down from the proband's grandmother or grandfather. From the family tree, it is clear that the proband's mother carries the mutation, either inherited from the grandparents or caused by a *de novo* mutation. The proband's mother did pass down the mutation to both of her children. The proband did not pass the mutation to his son. However, his 15-year-old nephew inherited the mutation from the proband's sister.

Summary

What was known before

- Post-LASIK exacerbation of corneal dystrophy is almost exclusively associated with TGFBI gene mutations.
- Some mutation carriers are asymptomatic.

What this study adds

 Genetic screening of TGFBI mutations must be incorporated into the pre-operative screening, especially for those who are asymptomatic.

Conflict of interest

John Marshall is a consultant to Avellino Lab USA, Inc., Frost Professor at Institute of Ophthalmology, University College of London, London, UK. Tara Moore is a consultant to Avellino Lab USA, Inc., Director of the Biomedical Sciences Research Institute and Professor of Personalized Medicine at University of Ulster, Coleraine, Northern Ireland, UK. M Andrew Nesbit is a Senior Lecturer at University of Ulster, Coleraine, Northern Ireland, UK. Rao Me is an Ophthalmologist and BL Ke is Professor of Ophthalmology at Shanghai First People's Hospital, Shanghai Jiaotong University School of Medicine, Shanghai, China. Larry DeDionisio is an employee of Avellino Lab USA, Inc., Menlo Park, CA, USA. Connie Chao-Shern is a PhD student at University of Ulster, Coleraine, Northern Ireland, UK and an employee of Avellino Lab USA, Inc., Menlo Park, CA, USA. The authors declare no conflict of interests. The authors alone are responsible for the content and writing of this article.

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