Contents lists available at ScienceDirect



Molecular Genetics and Metabolism Reports

journal homepage: www.elsevier.com/locate/ymgmr



# Genetic and clinical characteristics of Filipino patients with Gaucher disease



Mary Anne D. Chiong<sup>a,b,c,\*</sup>, Marie Julianne C. Racoma<sup>d</sup>, Mary Ann R. Abacan<sup>a,b</sup>

<sup>a</sup> Institute of Human Genetics, National Institutes of Health University of the Philippines Manila, Philippines

<sup>b</sup> Department of Pediatrics, Philippine General Hospital, Manila, Philippines

<sup>c</sup> University of Santo Tomas, Manila, Philippines

<sup>d</sup> Newborn Screening Reference Center, National Institutes of Health, University of the Philippines, Manila, Philippines

#### ARTICLE INFO ABSTRACT Gaucher disease (GD) is a lysosomal storage disorder caused by the deficiency of the $\beta$ -glucocerebrosidase en-Keywords: Gaucher disease zyme due to disease causing mutations in the GBA1 (glucosidase beta acid) gene, leading to the abnormal Filipinos accumulation of the lipid glucocerebroside in lysosomal macrophages. This is a review of the clinical features β glucocerebrosidase and molecular profiles of 14 Filipino patients with GD. Five patients presented with type 1 disease, two had type GBA1 mutations 2 GD and seven had type 3 GD. The age of onset for all types was between 1 and 2 years of age but there was a Enzyme replacement therapy delay of 2.2 years from the time of symptom onset to confirmation of diagnosis. Hepatosplenomegaly, anemia Severe phenotype and thrombocytopenia were present in most of the patients. Stunting was seen in 64.3% and bone abnormalities were present in 63.6%. The most common mutant allele detected in this cohort was L483P (previously L444P), followed by F252I, P358A and G241R. IVS2+1 G > A, N409S and G416S mutations were reported singularly. There were 3 patients who were found to have N131S mutations and one patient with D257V mutation, mutant alleles that have only been reported among the Filipinos to date. Except for N409S, the mutations found in this cohort were generally severe and were congruent with the severe phenotypes found in most patients. Of the 14 patients, only 6 were able to undergo enzyme replacement therapy which significantly improved the hematologic parameters and decreased the sizes of the liver and spleen but did not consistently improve the growth and skeletal abnormalities nor alleviate the neurological manifestations of our patients with GD. Improved monitoring through recommended modalities for assessments and tools for evaluation should be implemented in order to fully appreciate the severity of the disease and accuracy of the response to treatment.

# 1. Introduction

Gaucher disease (GD) is a multisystemic disorder that results from the accumulation of undegraded glucocerebroside in lysosomes of tissue macrophages due to the deficiency of the enzyme  $\beta$ -glucocerebrosidase (EC 3.2.1.45) [1]. The deficiency of the enzyme is caused by autosomal recessive mutations in the *GBA1* (glucosidase beta acid) gene located on chromosome 1 (1q21) [2].

Gaucher disease is traditionally classified into three broad phenotypic categories: Type 1, a non-neuronopathic disease ranging from asymptomatic subjects to those who display childhood-onset disease. Type 2 GD is an acute neuronopathic form characterized by the early appearance of visceral signs, oculomotor abnormalities, brainstem involvement and limited survival to the first two or three years of life. Type 3 GD is a chronic neuronopathic disease characterized by neurologic symptoms that appear later in life. The appreciation that patients with type 1 may develop late-onset neurologic manifestations suggest that GD is a continuum of disease states [3,4]. The incidence is 1/40,000 to 1/60,000 births in the general population, but it can reach 1/800 births in the Ashkenazi Jewish population [5]. The diagnosis relies on the measurement of  $\beta$ -glucocerebrosidase activity in leukocytes, fibroblasts or the recently developed screening tool using dry blood spots. Almost 300 mutations and polymorphisms in *GBA1* gene have been identified [4]. Treatment with macrophage-targeted mannose-terminated glucocerebrosidase enzyme replacement therapy (ERT) is the standard of care for patients with type 1 and 3 Gaucher disease [6]. Oral substrate reduction therapy in the form of Eliglustat has likewise been found to produce significant improvements in spleen and liver sizes, hematologic parameters, bone mineral density and disease biomarkers among adult patients with Gaucher disease type 1 [7].

In this review, we describe the clinical features, laboratory parameters and molecular characteristics of 14 Filipino patients with Gaucher disease, as well as the hematologic, skeletal, growth and neurologic responses of 6 patients who have undergone ERT.

\* Corresponding author at: Institute of Human Genetics, National Institutes of Health University of the Philippines Manila, Philippines. *E-mail addresses*: mdchiong1@up.edu.ph (M.A.D. Chiong), mcracoma@up.edu.ph (M.J.C. Racoma), mrabacan@up.edu.ph (M.A.R. Abacan).

https://doi.org/10.1016/j.ymgmr.2018.03.010

Received 24 January 2018; Received in revised form 23 March 2018; Accepted 23 March 2018

2214-4269/ © 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/BY-NC-ND/4.0/).

### 2. Methods

## 2.1. Subjects

This study involved chart reviews of 14 Filipino patients confirmed to have Gaucher disease from 2003 to July 2017. The diagnosis was based on clinical findings and confirmed by low β-glucocerebrosidase activity in peripheral leukocytes or dried blood spots. The enzyme measurement was done at the National Taiwan University Hospital from 2004 to 2017. The first 2 patients diagnosed in 2003 had enzyme assays done at the National Referral Laboratory of the Women's and Children's Hospital in Adelaide Australia. The data during initial and follow up evaluations were gathered up to the latest month/year of consult. This study was approved by the institution's ethical review board (2017-265-01).

# 2.2. Clinical characteristics

Data on the patients' anthrophometrics, date of birth, date of diagnosis, date of the first symptoms and clinical features such as neurologic signs and organ measurements were obtained from the medical records of the Philippine General Hospital and the Institute of Human Genetics, National Institutes of Health-Manila (IHG-NIH). Hematologic, biochemical parameters and results of the ancillary investigations were also reviewed.

# 2.3. Mutation studies

Twelve patients who are enrolled in RegistryNXT (https://www. registrynxt.com/) for Gaucher disease had the opportunity to have mutation testing done at the Molecular Development Laboratory of the University of Washington, USA after the families have undergone an informed consent process and pre genetic counseling at IHG-NIH. Sanger dideoxy sequencing of the entire coding region of the glucocerebrosidase (GBA1) gene was the reported method used for the identification of mutations. Post genetic counseling was given by geneticists at IHG-NIH upon receipt of the mutation results.

# 2.4. Method of data analysis

Quantitative patient characteristics are summarized by means and standard deviations (SDs). Qualitative characteristics such as clinical features and skeletal findings are presented as a frequency distribution.

# 3. Results

# 3.1. Demographic, clinical and laboratory characteristics

There were 14 patients with 3 sibling pairs from non-consanguineous pedigrees. There were 3 males and 11 females with ages that ranged from 1 month to 21 years. All were Filipinos except for Patient 12 and Patient 14. Five had type 1 disease who had no neurological involvement at diagnosis and latest follow up. Two patients (Patients 13 and 14) with type 2 GD presented with severe signs and symptoms early. Seven patients who manifested with seizures, developmental delay or abnormal eye movements were classified as type 3 (Table 1). The mean age of onset of clinical manifestations for type 1 was 1.12 years (SD 0.56; 0.5-2 years) and the mean age for the confirmation of diagnosis was 3.35 years (SD 2.1; 0.25-5.5 years). For type 3, the mean age of onset of the signs and symptoms was 1.7 years (SD1.19; 1-4 years) and the diagnosis was confirmed at a mean of 3.9 years (SD 1.5; 2-6 years). Overall, it took an average of 2.2 years from the onset of clinical manifestations to confirmation of diagnosis for all types.

Hepatosplenomegaly as an initial presentation was seen in 12 patients (85.7%) while 2 patients presented with splenomegaly alone Table 1

Summary of clinical, biochemical and molecular characteristics

Pt #	Age at Dx	Sex	Sex Ethnicity	Presenting clinical S/Sx	β- gluco cerebrosidase Genotype	Genotype	Type	Type On/had ERT	Outcome
1	3.5y	F	Filipino	$\mathrm{HS}^{\mathrm{a}}$ and bleeding	390 <sup>b</sup>	p.L483P/p.P358A	3	Started ERT at 3.7 yrs. old. Ongoing for 14 yrs.	Alive
2	4.5y	н	Filipino	HS <sup>a</sup>	$0.88^{\circ}$	p.L483P/p.P358A	з	Started ERT at 5.5 yrs. old. Ongoing for 12 yrs	Alive
с	5y	н	Filipino	HS <sup>a</sup>	1.55 <sup>c</sup>	p.F2521/p.F2521	1	Started ERT at 5.3 yrs. old. Ongoing for 6.6 yrs	Alive
4	2y	ц	Filipino	HS <sup>a</sup> and Seizures	3.89 <sup>c</sup>	p.N131S/p.P358A	3	Started ERT at 3.5 yrs. old and lasted for 2.7 years	Died
ъ	2y	ц	Filipino	HS <sup>a</sup> and Developmental delay	3.26 <sup>c</sup>	p.L483P/p.G241R/p.G241R	3	Started ERT at 2.6 yrs. old and lasted for 3.3 yrs	Died
9	3y	Μ	Filipino	HS <sup>a</sup>	$1.23^{\circ}$	Not available	1	Started ERT at 4 yrs. old and lasted for a month	Died
7	3 m	ц	Filipino	HS <sup>a</sup>	$0.04^{d}$	p.F2521/p.F2521	1	No	Alive
8	6y	Μ	Filipino	HS <sup>a</sup> and Esotropia	0.36 <sup>d</sup>	p.N131S/p.G416S	З	No	Alive
6	5y	ц	Filipino	Splenomegaly, Pallor, Esotropia and Seizures	$1.32^{d}$	p.N131S/p.G416S	e	No	Alive
10	5.5y	ц	Filipino	HS <sup>a</sup>	0.63 <sup>c</sup>	p.L483P/p.L483P	1	No	Alive
11	4y	ц	Filipino	HS <sup>a</sup>	310 <sup>b</sup>	Not available	1	No	Died
12	3y	ц	Indian	HS <sup>a</sup>	2.97 <sup>c</sup>	p.N409S/p.L438P	1	No	Lost to follow up
13	2y	ц	Filipino	HS <sup>a</sup> and Ophthalmoplegia	0.01 <sup>c</sup>	IVS2 + 1 G > A/p.F252I	2	No	Died
14	$1 \mathrm{m}$	Μ	Filipino/Japanese	Splenomegaly and Icthyosis	0.12 <sup>d</sup>	p.R519W/p.D257V	2	No	Died
<sup>a</sup> Hej <sup>b</sup> Nor	<sup>a</sup> Hepatosplenomegaly. <sup>b</sup> Normal Value: 600–3	tegaly. 600–32	200 pmole/min/mg	<sup>a</sup> Hepatosplenomegaly. <sup>b</sup> Normal Value: 600–3200 pmole/min/mg protein: National Referral Laboratory. Adelaide: done in peripheral blood leukocytes.	aide: done in periphera	l blood leukocytes.			

Normal Value: > 5.1 nmol/mg/prot/h; National Taiwan University Hospital; done in peripheral blood leukocytes.

υ Ψ

Normal Value: 1.8 umoles/L/h; National Taiwan University Hospital; done from blood spot aliquots.

#### Table 2

Distribution of signs and symptoms according to the type of GD.

Signs and symptoms		Types of Gaucher dis			
		Type 1 (n = 5)	Type 2(n = 2)	Type 3 (n = 7)	Total
Hepatosplenomegaly		5	1	6	12 (85.7%)
Splenomegaly alone		0	1	1	2 (14.2%)
Hematological	Pallor	5	2	6	13 (92.8%)
	Bleeding	2	1	3	6 (42.8)
Eye findings	Strabismus	0	1	3	4 (28.5%)
	Gaze Initiation Defect	0	1	4	5 (35.7%)
Neurologic	Seizures	0	0	5	5 (35.7%)
	Developmental delay	0	1	5	6 (42.8%)
	Intellectual disability	0	0	4	4 (28.5)
Failure to thrive		1	0	4	5 (35.7%)
Fatigue		2	0	4	6 (42.8)
Bone Fracture		0	0	1	1 (7.14%)
Bone Pain		0	0	1	1 (7.14%)
Radiologic Bone Abnormalities <sup>a</sup>		3	0	4	7 (63.6%)
	Erlenmeyer flask deformity	3		2	5 (45.4%)
	Osteopenia	1		1	2 (18.2%)
	Lytic changes			1	1 (9.09%)
	Fracture			1	1 (9.09%)

<sup>a</sup> Two patients with type 2 GD and 1 with type 3 GD had no skeletal survey done.

(Patients 9 and 14). Hepatosplenomegaly was accompanied by bleeding in 1 patient (7.14%), pallor in one (7.14%) and neurologic symptoms in 3 patients which manifested as either developmental delay, esotropia or seizures (21.4%). Of the type 2 patients, patient 13 presented with ocular abnormalities, hyperextension of the neck, developmental regression, microcephaly, spasticity, and tonic clonic seizures apart from hepatosplenomegaly. She succumbed at 2.5 years of age. Patient 14 had collodion ichthyosis at birth accompanied by hepatosplenomegaly, anemia and thrombocytopenia. He died at 2 months of age due to massive bleeding. At baseline, 9 (64.3%) patients were stunted (height for age z score -2) and 5 (35.7%) were underweight (weight for age z score -2). Patients 9 and 10 were splenectomized before the confirmation of diagnosis. The distribution of the other clinical manifestations are in Table 2.

The initial mean hemoglobin for all types was 85.3 g/L (SD 35; range 10.9–16.4) and mean platelets were at  $93 \times 10^9$ /L (SD 75; range  $9-228 \times 10^9$ /L). Type 2 patients in this cohort were initially found to have normal hemoglobin and white blood cell count. Types 1 and 3 presented with anemia, thrombocytopenia and low WBC at baseline. AST that was minimally elevated in 10 of 12 patients with available data (83.3%) may not be liver specific. ALT was slightly elevated in 2 of the 12 (16.7%) (Table 3). None were tested for hepatitis virus infection. Liver and spleen volumes were assessed by physical examination and ultrasound. Hepatomegaly was present in 12 of the 14 patients with a mean of 10 cm below the right costal margin (SD 6.7 cm; range 5-30 cm). On ultrasound, the liver span had an average of 11.9 cm at the mid-clavicular line (SD 2.4; 8.5-14.6 cm) and were all abnormal for age and sex. Splenomegaly affected all patients with a mean of 17.2 cm (SD 5.13; 10-27) below the left costal margin. Ultrasound splenic volume showed a mean of 504 mL at baseline (SD 232.6; range 170-897.5 mL) which were all above the upper limit based on weight [8,9]. Eleven patients had skeletal survey done and 7 were found to have bone abnormalities. Erlenmeyer flask deformity was seen in 5 (45.5%), osteopenia in 2 (18.2%), lytic changes in 1 (9%) and fracture of the femoral bone in 1 (9%).

All had low  $\beta$ -glucocerebrosidase enzyme activity (Table 1). Seven of the 14 patients had a bone marrow biopsy done which showed the characteristic "Gaucher cells". Chitotriosidase was elevated in 8 patients but 4 were noted to have deficient levels.

Only 6 of the 14 patients underwent compassionate enzyme replacement therapy (ERT). Two had type 1 GD and 4 had type 3 GD There was a delay of 1.2 years from the time of diagnosis to initiation of

# Table 3 Summary of baseline laboratory findings

Laboratory Parameters	Frequency	Mean	SD	Min	Max
Hemoglobin (g/L)	14	85.3	35.0	53.0	164.0
Type 1	5	84.2	15.9	66.1	109.0
Type 2	2	144.0	28.3	124.0	164.0
Туре 3	7	83.3	19.3	53.0	108.0
Platelet ( $\times 10^9$ /L)	14	92.9	75.4	9.0	228.0
Type 1	5	121.6	96.7	9.0	226.0
Type 2	2	71.0	37.0	45.0	97.0
Туре 3	7	78.6	69.2	31.0	228.0
WBC ( $\times 10^9$ /L)	14	8.0	4.9	2.7	19.7
Type 1	5	9.2	7.0	3.2	19.7
Type 2	2	12.6	0.4	12.3	12.9
Туре 3	7	5.9	2.8	2.7	10.1
AST (IU/L)	12	103.2	130.9	24.0	513.0
Type 1	3	89.9	5.3	84.0	94.0
Type 2	2	291.0	313.9	69.0	513.0
Туре З	7	55.1	21.3	24.0	91.0
ALT (IU/L)	12	32.3	9.2	22.1	50.0
Type 1	4	32.3	11.9	24.0	50.0
Type 2	1	24.0	-	-	-
Туре З	7	33.5	8.5	22.1	49.0
Chitotriosidase nmol/ml/ h <sup>a</sup>	8	11,138.5	11,185.0	322.5	27,128.9
Type 1	3	17,238.8	14,448.1	658.9	27,128.9
Type 2	2	10,888.0	14,942.2	322.5	21,454.0
Туре 3	3	5204.1	2959.4	2344.4	8253.9

<sup>a</sup> Chitotriosidase done in 12 patients but 4 were noted to have CHIT gene duplication.

ERT. The mean age of starting ERT was 4.1 years (SD 1.1; 2.6–5.5 years) with a mean dose of 58.3 units/kg (SD 4.1). The mean duration of ERT at the time of this report was 6.45 years (SD 5.52; 0.08–14 years). Three patients (Patients 4, 5 and 6) died of pneumonia while on the 3rd year, 4th year and 1st month of ERT respectively. Two of the 6 patients, Patients 1 and 2, achieved normal growth parameters during the 2nd and 1st years of ERT respectively. One patient had normal height at the outset (Patient 5). Patient 3 who is on the 6th year of treatment continues to be stunted as of this report. Patients 4 and 6, who died while receiving treatment, did not achieve normal height. Normal hemoglobin [10] was achieved in Patients 1–5 at a mean of 7.8 months after initiation had no normalization of hemoglobin. Platelet count increased by 1.5% in patients 1–5 at a mean of 6.2 months (SD 5.8), but normal

levels  $(150-350 \times 10^9/L)$  were achieved in only 4 of them at a mean of 18 months (SD 5.03). Patients 5 and 6 remained to have low platelet counts at the time of their demise. Patients 1 and 2 achieved normal splenic volumes at a mean of 4.2 years (SD 2.2). Patient 3 had a 50% decrease in spleen volume after 3.75 years of treatment, however she remains to have an enlarged spleen even while on the 6th year of treatment. Patient 4 and 5 had enlarged spleen at the time of demise. All five patients achieved normal liver size by physical examination and ultrasound at a mean of 4.9 years after starting ERT (SD 2.8). Patients 1, 2 and 5 developed thoracic scoliosis while on the 14th, 12th, and 5th year of treatment respectively. Patients 3 and 4 still showed Erlenmeyer flask deformity after 6 and 2 years of ERT respectively.

There was note of increase in the frequency of seizures in two patients on ERT (Patients 4 and 5), while stabilization was seen in two (Patients 1 and 2). There was evolution of seizures to myoclonic jerks for Patient 4 on the 7th month of ERT. Patient 2 had cholelithiasis on the 11th year of treatment.

#### 3.2. Mutation analysis of the GBA gene

Using the current nomenclature for GBA1 mutations from the Human Genome Variation Society (http://varnomen.hgvs.org/), the most common mutant allele was L483P (previously L444), followed by F252I, P358A and G241R. IVS2 + 1 G > A, N409S, R159W and G416S mutations were reported singularly. There were 3 patients who had the N131S mutation, a mutant allele that has only been reported among the Filipinos. Another novel mutation D257V was seen in one patient. The reported severe mutations L483P, F252I, G241R, G416S, R159W and IVS2 + 1G > A were seen in patients with Types 2 and 3 disease (Table 1). However, the 2 patients homozygous for F252I (Patients 3 and 7) are clinically presenting with type 1 disease. N409S was found in patient 12 with Type 1 disease. The novel mutations N131S and D257V were found in patients who had Types 2 and 3 disease. Two patients (Patients 6 and 11) were not able to undergo mutation testing. They were classified to have type 1 disease; however, the possibility that they had severe mutations could not be discounted as their early demise and severe manifestations implied a possible type 3 GD.

## 4. Discussion

This is the first review done among Filipino patients with Gaucher disease. The age of onset of clinical manifestations is in keeping with what has been reported in the literature. Most patients with type 1 disease have onset during the first decade of life, while patients with type 3 present at around 12 months old [11]. Despite early manifestations, diagnostic delay is one of the most significant observations in this study. Majority had severe manifestations early but the delay in diagnosis was most probably due to the failure to include GD in the differential diagnosis because of lack of awareness. Another reason could be the lack of local facilities for patients with rare diseases that can comprehensively assess these patients.

Hepatosplenomegaly was seen as the most common finding across all types in association with hematologic and bone abnormalities and is congruent with what have been reported in previous studies [12,13]. Many of our patients presented with near normal liver function. The neurologic, ocular and other manifestations of our patients with types 2 and 3 GD are likewise similar with other registry reports [11,14,15].

The stunting in our patients could be due to the large mass of Gaucher's cells in the liver and spleen which exert an excessive metabolic burden leading to higher resting energy expenditure and increase in caloric requirements which manifested as a height-for-age z score less than zero [16]. The reported bone abnormalities such as osteopenia and lytic lesions are related to the accumulation of Gaucher cells in the bone marrow which lead to vascular occlusion and localized bone defects [1,4].

The results of  $\beta$  glucocerebrosidase assay did not reliably enable

prediction of disease severity or subtype [17]. Some of our patients who were classified as having a milder disease even had lower levels than patients with severe types. Our patients who had elevated chitotriosidase reflect the presence of lipid-laden macrophages due to GD [18]. However due to limited resources, this was not utilized to monitor the response to therapy. Four patients who were deficient in chitotriosidase were assumed to have the common 24 bp duplication polymorphism in the *CHIT* gene which is reported to be highest among Asians [19]. It was observed from the laboratory parameters of the 14 patients that hemoglobin levels for types 1 and 3 were almost similar, but platelets and WBC were higher in type 1. There is usually a significant correlation between the degree of splenomegaly and cytopenias but this proved to be clinically negligible [20]. It is difficult to make observations for other parameters according to type because of the small number of affected patients.

Only 6 of our patients were able to avail of compassionate ERT. Normal hemoglobin was achieved at an average of 7.8 months and is in keeping with a reported collaborative study that anemic patients usually normalized within 6-12 months [21]. All patients who underwent ERT had severe thrombocytopenia (< 120,000/µL) at baseline which improved 1.5 fold at a mean of 6 months. It is expected that platelet counts increase to 1.5-fold by year 1, and continue during the next 2 to 5 years, however restoration to normal is not usually expected [22]. The 2 patients who did not achieve normal platelets had significant splenomegaly at baseline. Although liver and spleen volumes were not routinely done during treatment monitoring, the liver sizes for 5 patients fell within the observed timeline for improvement of 5 years [23]. The decrease in spleen sizes have not been uniform among our patients on ERT. Three patients had slower response which could be due to prior splenic infarctions or fibrosis [23]. None had bone crisis or bone pain among our patients on ERT. However, 3 patients developed thoracic scoliosis and 2 were still found to have Erlenmeyer flask deformity years after initiating ERT. It has been reported that after 2 to 4 years of ERT, bone involvement improved in 30% to 40% of pediatric patients while there was progression in < 5% [22]. The scoliosis could be linked to abnormal bone density [24].

Cholelithiasis seen in Patient 2 has been reported to have increased prevalence in GD due to bile acid composition abnormalities [15]. The neurologic symptoms of our patients, as with other reports [25–27] were not reversed by ERT because the enzyme does not cross the blood-brain barrier.

The most common mutation seen in our population, L483P, is a mutation known to cause a severe disease [28-30]. It is also the most common mutation seen in other Asian populations [31-33]. In these populations, this mutant allele was associated with types 1,2 and 3 phenotypes in heterozygous or homozygous forms. In two of our patients who are siblings (Patients 1 and 2), L483P was seen in heterozygous combination with a rare allele P358A (previously P319A) of unknown severity [31]. Since these 2 patients presented with a chronic neuronopathic phenotype, P358A can be classified as severe. Patient 5 who had type 3 disease also had a severe allele G241R (previously G202R) in association with L483P. The parents' genotype showed that the father is a carrier for L483P/wild type while the mother has G241R/ wild type. The L483P and G241R mutations are known to occur in cis, and this is likely the case in this patient thereby accounting for the two G241R mutations as well as L483P. The "gene conversion" event between the active gene and pseudogene has probably been in the paternal side of the family for generations. G241R mutation destroys a Ncil restriction site in the GBA gene and has been reported in a patient who died of type 2 disease [28]. Patient 12 who had L483P as an heteroallele together with N409S (previously N307S) had a type 1 disease. N409S is known to be mild and even a severe or lethal mutation inherited with it results in type I disease [28,34]. F252I (previously F213I) is a severe mutation seen among Asians with types 2 or 3 disease. Patients homozygous or heterozygous for this mutant allele presented primarily with oculomotor apraxia and mental retardation

[26,32]. There were 2 homozygotes for F252I in this cohort. They are siblings (Patients 3 and 7) who are currently classified as having type 1 disease because of the absence of ocular or neurologic abnormalities. F252I is a rare allele that was previously reported in a non-Jewish English family whose 2 affected children presented with GD type 1 [35]. In the Japanese patients, F252I mutation was also seen among those who were originally classified as having type 1 disease but eventually transitioned to type 3 [26]. Thus, these siblings should be closely monitored for subtle signs of type 3 disease during their clinical course. The other patient (Patient 13) carrying the F252I mutation who had type 2 GD was a heterozygote for a splicing null mutation IVS 2 + 1 G > A that directs no functional enzyme activity [36]. The other patient with type 2 disease (Patient 14) had the severe mutation R159W (previously R120W) [29,32,33,35]. This patient also carried a novel mutation D257V (previously D218V) which is predicted to be probably damaging to protein function with a score of 0.99 (http://genetics.bwh. harvard.edu/pph2/bgi.shtml). Three patients in this cohort had another novel mutation N131S (previously N92S) which had only been reported among Filipinos and is likewise predicted to be probably damaging with a score of 1.00 (http://genetics.bwh.harvard.edu/pph2/bgi.shtml). This mutation was found in association with G416S (previously G377S) that was previously reported to be seen in type 3 disease [30,32] and with P358A. In the two patients who were compound heterozygotes for N131S and G416S, it is assumed that both gave rise to severe mutations leading to a neurologic phenotype.

In conclusion, our patients with Gaucher disease mostly had type 3 disease which is supported by the predominantly severe mutations noted in our population. Enzyme replacement therapy effectively improved the hematological impairments and hepatosplenomegaly, but due to the limited number of patients on ERT, deaths, and limitations on the use of recommended modalities for assessments, the results of the patterns of growth and skeletal manifestations may not accurately reflect the course of the disease. With the recent passage of the Rare Disease Act of the Philippines, it is hoped that physicians will be more aware of the presenting signs and symptoms of GD so that better access to diagnosis, and provision of a more comprehensive and sustainable treatment and monitoring will improve the neurologic and non-neurologic complications of the disease in this population.

## Funding

The research did not receive any grant from funding agencies in the public, commercial or not for profit sectors.

## Acknowledgements

We would like to thank Dr. Paul Hwu of the National Taiwan University Hospital for doing the  $\beta$  glucocerebrosidase enzyme assay of our patients and Dr. Ronald Scott of the University of Washington for performing the mutation analysis of our patients included in the Gaucher RegistryNXT. We would also like to extend our gratitude to Ms. Mikarla Lubat of the Department of Clinical Epidemiology, University of the Philippines-Manila for the assistance on the statistical analysis of the data. On behalf of the patients, we would like to express our gratefulness to Genzyme Sanofi for the compassionate enzyme replacement therapy given to our patients.

# References

- W.J. Huang, X. Zhang, W.W. Chen, Gaucher disease: a lysosomal neurodegenerative disorder, Eur. Rev. Med. Pharmacol. Sci. 19 (7) (2015) 1219–1226.
- [2] J. Stirnemann, N. Belmatoug, F. Camou, C. Serratrice, R. Froissart, C. Caillaud, T. Levade, L. Astudillo, J. Serratrice, A. Brassier, et al., A review of Gaucher disease pathophysiology, clinical presentation and treatments, Int. J. Mol. Sci. 18 (2) (2017).
- [3] E. Beutler, G.A. Grabowski, Gaucher disease, in: C.R. Scriver, A.L. Beaudet, W.S. Sly, D. Valle (Eds.), The Metabolic and Molecular Basis of Inherited Disease, McGraw-

Hill, New York, NY, 2001, pp. 3635–3668.

- [4] S. Linari, G. Castaman, Clinical manifestations and management of Gaucher disease, Clin. Cases Miner. Bone Metab. 12 (2) (2015) 157–164.
- [5] G.A. Grabowski, Phenotype, diagnosis, and treatment of Gaucher's disease, Lancet 372 (9645) (2008) 1263–1271.
- [6] P.K. Mistry, M.D. Cappellini, E. Lukina, H. Ozsan, S. Mach Pascual, H. Rosenbaum, M. Helena Solano, Z. Spigelman, J. Villarrubia, N.P. Watman, et al., A reappraisal of Gaucher disease-diagnosis and disease management algorithms, Am. J. Hematol. 86 (1) (2011) 110–115.
- [7] P.K. Mistry, E. Lukina, H. Ben Turkia, S.P. Shankar, H. Baris, M. Ghosn, A. Mehta, S. Packman, G. Pastores, M. Petakov, et al., Outcomes after 18 months of eliglustat therapy in treatment-naïve adults with Gaucher disease type 1: the phase 3 ENGAGE trial, Am. J. Hematol. 92 (11) (2017) 1170–1176.
- [8] A. Schlesinger, K. Edgar, L. Boxer, Volume of the spleen in children as measured on CT scans: normal standards as a function of body weight, AJR 160 (1993) 1107–1109.
- [9] E. Yetter, K. Acosta, M. Olson, K. Blundell, Estimating splenic volume: Sonographc measurements correlated with helical CT determination, AJR 181 (2003) 1615–1620.
- [10] R. Gajjar, E. Jalazo, Hematology, in: B. Engorn, J. Flerlage (Eds.), The Harriet Lane Handbook: A Manula for Pediatric House Officers, 20th ed., Mosby Elsevier, Philadelphia, 2015, p. 306.
- [11] G.A. Grabowski, A. Zimran, H. Ida, Gaucher disease types 1 and 3: phenotypic characterization of large populations from the ICGG Gaucher registry, Am. J. Hematol. 90 (Suppl. 1) (2015) S12–8.
- [12] J. Charrow, H.C. Andersson, P. Kaplan, E.H. Kolodny, P. Mistry, G. Pastores, B.E. Rosenbloom, C.R. Scott, R.S. Wappner, N.J. Weinreb, et al., The Gaucher registry: demographics and disease characteristics of 1698 patients with Gaucher disease, Arch. Intern. Med. 160 (18) (2000) 2835–2843.
- [13] P. Kaplan, H.C. Andersson, K.A. Kacena, J.D. Yee, The clinical and demographic characteristics of nonneuronopathic Gaucher disease in 887 children at diagnosis, Arch. Pediatr. Adolesc. Med. 160 (6) (2006) 603–608.
- [14] M. Andrade-Campos, P. Alfonso, P. Irun, J. Armstrong, C. Calvo, J. Dalmau, M.R. Domingo, J.L. Barbera, H. Cano, M.A. Fernandez-Galán, et al., Diagnosis features of pediatric Gaucher disease patients in the era of enzymatic therapy, a national-base study from the Spanish Registry of Gaucher disease, Orphanet J. Rare Dis. 12 (1) (2017) 84.
- [15] N. Gupta, I.M. Oppenheim, E.F. Kauvar, N. Tayebi, E. Sidransky, Type 2 Gaucher disease: phenotypic variation and genotypic heterogeneity, Blood Cells Mol. Dis. 46 (1) (2011) 75–84.
- [16] D.J. Barton, M.D. Ludman, K. Benkov, G.A. Grabowski, N.S. LeLeiko, Resting energy expenditure in Gaucher's disease type 1: effect of Gaucher's cell burden on energy requirements, Metabolism 38 (12) (1989) 1238–1243.
- [17] Pastores G, Hughes D. Gaucher disease. Genereviews https://www.ncbi.nlm.nih. gov/books/NBK1269 February 26, 2015. Accessed 28 August 2017.
- [18] A.P. Bussink, M. Verhoek, J. Vreede, K. Ghauharali-van der Vlugt, W.E. Donker-Koopman, R.R. Sprenger, C.E. Hollak, J.M. Aerts, R.G. Boot, Common G102S polymorphism in chitotriosidase differentially affects activity towards 4-methylumbelliferyl substrates, FEBS J. 276 (19) (2009) 5678–5688.
- [19] P. Lee, J. Waalen, K. Crain, A. Smargon, E. Beutler, Human chitotriosidase polymorphisms G354R and A442V associated with reduced enzyme activity, Blood Cells Mol. Dis. 39 (3) (2007) 353–360.
- [20] Y. Gielchensky, D. Elstein, I. Hadas-Halpern, A. Lahad, A. Abrahamov, A. Zimran, Is there a correlation between degree of splenomegaly, symptoms and hypersplenism? A study of 218 patients with Gaucher disease, Br. J. Haematol. 106 (3) (1999) 812–816.
- [21] N.J. Weinreb, Imiglucerase and its use for the treatment of Gaucher's disease, Expert. Opin. Pharmacother. 9 (11) (2008) 1987–2000.
- [22] G.M. Pastores, N.J. Weinreb, H. Aerts, G. Andria, T.M. Cox, M. Giralt, G.A. Grabowski, P.K. Mistry, A. Tylki-Szymańska, Therapeutic goals in the treatment of Gaucher disease, Semin. Hematol. 41 (4 Suppl. 5) (2004) 4–14.
- [23] N.J. Weinreb, J. Charrow, H.C. Andersson, P. Kaplan, E.H. Kolodny, P. Mistry, G. Pastores, B.E. Rosenbloom, C.R. Scott, R.S. Wappner, et al., Effectiveness of enzyme replacement therapy in 1028 patients with type 1 Gaucherdisease after 2 to 5 years of treatment: a report from the Gaucher registry, Am. J. Med. 113 (2) (2002) 112–119.
- [24] B.H. Yip, F.W. Yu, Z. Wang, V.W. Hung, T.P. Lam, B.K. Ng, F. Zhu, J.C. Cheng, Prognostic value of bone mineral density on curve progression: a longitudinal cohort study of 513 girls with adolescent idiopathic scoliosis, Sci. Rep. 6 (2016) 39220.
- [25] A. Sechi, L. Deroma, A. Dardis, G. Ciana, N. Bertin, D. Concolino, S. Linari, C. Perria, B. Bembi, Long term effects of enzyme replacement therapy in an Italian cohort of type 3 Gaucher patients, Mol. Genet. Metab. 113 (3) (2014) 213–218.
- [26] A. Tajima, T. Yokoi, M. Ariga, T. Ito, E. Kaneshiro, Y. Eto, H. Ida, Clinical and genetic study of Japanese patients with type 3 Gaucher disease, Mol. Genet. Metab. 97 (4) (2009) 272–277.
- [27] R.R. Lonser, S. Walbridge, G.J. Murray, M.R. Aizenberg, A.O. Vortmeyer, J.M. Aerts, R.O. Brady, E.H. Oldfield, Convection perfusion of glucocerebrosidase for neuronopathic Gaucher's disease, Ann. Neurol. 57 (4) (2005) 542–548.
- [28] E. Beutler, A. Demina, T. Gelbart, Glucocerebrosidase mutations in Gaucher disease, Mol. Med. 1 (1994) 82–92.
- [29] E. Beutler, T. Gelbart, C.R. Scott, Hematologically important mutations: Gaucher disease, Blood Cells Mol. Dis. 35 (2005) 355–364.
- [30] V. Koprivica, D.L. Stone, J.K. Park, M. Callahan, A. Frisch, I.J. Cohen, N. Tayebi, E. Sidransky, Analysis and classification of 304 mutant alleles in patients with type 1 and type 3 Gaucher disease, Am. J. Hum. Genet. 66 (6) (2000) 1777–1786.

- [31] H. Ida, O.M. Rennert, H. Kawame, K. Maekawa, Y. Eto, Mutation prevalence among 47 unrelated Japanese patients with Gaucher disease: identification of four novel mutations, J. Inherit. Metab. Dis. 20 (1997) 67–73.
- [32] S.Y. Jeong, S.J. Kim, J.A. Yang, J.H. Hong, S.J. Lee, H.J. Kim, Identification of a novel recombinant mutation in Korean patients with Gaucher disease using a longrange PCR approach, J. Hum. Genet. 56 (2011) 469–471.
- [33] L. Wan, C.M. Hsu, C.H. Tsai, C.C. Lee, W.L. Hwu, F.J. Tsai, Mutation analysis of Gaucher disease patients in Taiwan: high prevalence of the RecNciI and L444P mutations, Blood Cells Mol. Dis. 36 (2006) 422–425.
- [34] Y. Feng, Y. Huang, C. Tang, H. Hu, X. Zhao, H. Sheng, W. Zhang, M. Tan, T. Xie, J. Zheng, et al., Clinical and molecular characteristics of patients with Gaucher disease in Southern China, Blood Cells Mol. Dis. 16 (2016) 30150–30154.
- [35] G.S. He, M.E. Grace, G.A. Grabowski, Gaucher disease: four rare alleles encoding F213I, P289L, T323I, and R463C in type 1 variants, Hum. Mutat. 1 (5) (1992) 423–427.
- [36] G.S. He, G.A. Grabowski, Gaucher disease: a G+1 → A+1 IVS2 splice donor site mutation causing exon 2 skipping in the acid beta-glucosidase mRNA, Am. J. Hum. Genet. 51 (4) (1992) 810–820.