



⋒ OPEN ACCESS

Citation: Niang MN, Diop NS, Fall A, Kiori DE, Sarr FD, Sy S, et al. (2017) Respiratory viruses in patients with influenza-like illness in Senegal: Focus on human respiratory adenoviruses. PLoS ONE 12(3): e0174287. https://doi.org/10.1371/journal.pone.0174287

Editor: Oliver Schildgen, Kliniken der Stadt Köln qGmbH, GERMANY

Received: January 16, 2017

Accepted: March 6, 2017

Published: March 22, 2017

Copyright: © 2017 Niang et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: This work was supported by DHSS and Institut Pasteur de Dakar.

Competing interests: The authors have declared that no competing interests exist.

RESEARCH ARTICLE

Respiratory viruses in patients with influenzalike illness in Senegal: Focus on human respiratory adenoviruses

Mbayame Ndiaye Niang¹, Ndeye Sokhna Diop¹, Amary Fall¹, Davy E. Kiori¹, Fatoumata Diene Sarr², Sara Sy¹, Déborah Goudiaby¹, Mamadou Aliou Barry², Malick Fall³, Ndongo Dia¹*

- 1 Institut Pasteur de Dakar, Unité de Virologie Médicale, Dakar, Sénégal, 2 Institut Pasteur de Dakar, Unité d'Epidémiologie des maladies infectieuses, Dakar, Sénégal, 3 Département de Biologie Animale Faculté des Sciences et Techniques Université Cheikh Anta DIOP de Dakar, Dakar, Senegal
- * ndia@pasteur.sn

Abstract

Background

Human adenoviruses (HAdVs) are highly contagious pathogens that are associated with a wide spectrum of human illnesses involving the respiratory tract. In the present study, we investigate the epidemiologic and viral molecular features of HAdVs circulating in Senegal after 4 consecutive years of sentinel surveillance of influenza-like Illness cases.

Methodology and results

From January 2012 to December 2015 swabs were collected from consenting ILI outpatients. Adenoviral detection is performed by rRT-PCR with the Anyplex™ II RV16 Detection kit (Seegene) and molecular characterization was performed using a partial hexon gene sequence. 6381 samples were collected. More than half of patients (51.7%; 3297/6381) were children of ≤ 5 years. 1967 (30.8%) were positive for HAdV with 1561 (79.4%) found in co-infection with at least one another respiratory virus. The most common co-detections were with influenza viruses (53.1%; 1045/1967), rhinoviruses (30%; 591/1967), enteroviruses (18.5%; 364/1967) and RSV (13.5%; 266/1967). Children under 5 were the most infected group (62.2%; 1224/1967; p <0.05). We noted that HAdV was detected throughout the year at a high level with detection peaks of different amplitudes without any clear seasonality. Phylogenetic analysis revealed species HAdV-C in majority, species HAdV-B and one HAdV- 4 genome type. The 9 HAdV-B species like strains from Senegal grouped with genome types HAdV-7, HAdV-55 and HAdV-11 as shown by a phylogenetic branch with a high bootstrap value of (88%).

Conclusion

In conclusion, the results of the present study suggest strong year-round HAdV activity in Senegal, especially in children up to 5 years of age. Molecular studies revealed that the dominant species in circulation in patients with ILI appears to be HAdV-C and HAdV-B



species. The circulation of though HAdV-7 and HAdV-55 genome types is of note as these serotypes are recognized causes of more severe and even fatal acute respiratory infections.

Background

Human adenoviruses (HAdVs) are highly contagious pathogens that are associated with a wide spectrum of human illnesses involving the respiratory, ocular, gastrointestinal, and genitourinary systems [1]. They belong to the family Adenoviridae, genus Mastadenovirus with seven species (A-G), including each various types [2]. Ubiquitous in the environment, HAdVs are non-enveloped, double stranded DNA viruses that vary in size from 70 to 100 nm [3]. HAdVs are recognized as a common cause of respiratory infection in persons of all ages. The illnesses range from influenza-like fever and discomfort to pneumonia and death [4]. Indeed, HAdVs infections are usely mild but some groups such as very young children, elderly, immunocompromised persons, or persons with underlying pulmonary or cardiac disease, might be at higher risk degree for severe disease [5,6,7,8]. The most common HAdVs species that cause respiratory tract infections in children are B (HAdV-B3 and B7) and C (HAdV-C1, C2, and C5). Serotypes B3, B7, and B21 are the most frequent strains responsible for epidemics of acute febrile respiratory disease [9]. Circulating HAdVs can vary temporally and geographically with possibility of emergent genomic variants which can be associated with more severe illness [10,11].

In the present study, we investigate the epidemiologic and viral molecular features of HAdVs circulating in Senegal after 4 consecutive years of sentinel surveillance of influenza-like Illness cases.

Materiel and methods

Samples and data collection

From January 2012 to December 2015 we collected specimens (nasal-pharyngeal and oral-pharyngeal swabs) and surveillance data for influenza and other viral respiratory pathogens from outpatients presenting with influenza-like-illness (ILI) at different sentinel sites in Senegal. Once collected, swabs are placed in 2-mL cryovials with viral transport medium (Universal Transport Medium; COPAN Diagnostics Inc., Murrieta, CA), and transported at a controlled temperature of $2^{\circ}C$ — $8^{\circ}C$ to the laboratory. An ILI patient was defined as a person presenting with sudden onset of fever (>38 $^{\circ}C$) or history of sudden onset of fever in the recent past (\leq 3 days) and either cough or sore throat and/or rhinorrhea in the absence of other diagnosis, according to the CDC case definition. Each sample is accompanied by a case report form collecting demographic and clinical data. The questions included information on date of enrollment and symptom onset, sex, age, clinical symptoms, previous treatments, travelling history, vaccination status for influenza, and whether or not the patient was hospitalized. Upon arrival at the laboratory, the specimens were processed immediately for virus diagnosis. Aliquots of samples were also stored at $-80^{\circ}C$ for additional analysis (isolation and/or molecular characterization).

The data obtained daily were entered into an Epi Info database (Centers for Disease Control and Prevention, Atlanta, GA) and analyzed using Epi Info.



Nucleic acid extraction and viral detection

Total viral nucleic acid (DNA and RNA) was extracted from 140 µl of each clinical specimen using the PureLink™ Viral RNA/DNA Mini Kit (Invitrogen, Carlsbad CA, USA) according to the manufacturer's recommendation. DNA/RNA are eluted with 60 µl nuclease-free water and stored at −80°C until use.

A two-step multiplex real-time RT-PCR was performed with a Bio-Rad CFX-96 thermocycler (Bio-Rad Laboratories) and the Anyplex™ II RV16 Detection kit (Seegene) for a simultaneous testing of Influenza viruses (fluA and fluB), Human respiratory syncytial virus (RSVA and RSVB), Human adenoviruses (HAdV), Human metapneumovirus (HMPV), Human coronavirus (229E, NL63, OC43), Human parainfluenza virus (PIV1, -2, -3 and -4), Human rhinovirus (HRV), Human enterovirus (HEV) and Human bocavirus (HBoV), as previously described [12].

Molecular characterization of respiratory adenoviruses strains between 2012 and 2015 in Senegal

In consideration with low Ct-values, 80 HAdV positives samples (20 per year) were selected using a random number generator on MS Excel for further molecular characterization using classical PCR and sequencing. Viral DNA was extracted as previously described and eluted with 50 μ l water nuclease-free. DNAs were stored at -20° C until PCR reactions. For HAdV molecular characterization the last 300 base pairs (bp) of the hexon gene were amplified with the following specific primers: Adeno3 (5′-CCTTTGGCGCATCCCATTCT-3′) and Adeno4 (5′-TGGGCACCTATGACAAGCCC-3′) previously used by Garcia et al, [13]. The Phusion High-Fidelity PCR Master Mix with HF Buffer (New England Biolabs, Ipswich MA, USA) was used for amplifications. For each sample, PCR was carried out in a total reaction volume of 50 μ l consisting of 15 μ l H2O RNase free, 2.5 μ l of each primer (diluted at 10 μ M), 25 μ l of 2X Phusion Master Mix and 5 μ l of DNA template. Cycling conditions were as follows: denaturation step of 15 min at 95°C, 40 PCR cycles including 30 s at 95°C, 60 s at 55°C, 60 s at 72°C followed by an extension step of 10 min of 72°C.

Five microliters of the PCR product was then mixed with 1 μ l of 10X 5PRIME loading dye and loaded on to a 1% agarose gel along with an appropriated molecular weight markers (100 bp ladder, New England Biolabs), and gels were stained with ethidium bromide (0.5 μ g/ml) before visualization under UV.

For positive samples (380 bp size band), amplicons were cut and purified using the Gene-JET Gel Extraction Kit (Thermo Scientific). Purified products are then sent for sequencing to Beckman Coulter Services. Sequencing was performed in both directions with the same PCR primers (Adeno3 and Adeno4) on an ABI PRISM BigDye Terminator v3.1 Ready Reaction Cycle Sequencing kit (Applied Biosystems) on a 96-capillary ABI PRISM 3730-XL (Applied Biosystems). Data in FASTA format were then sent to the laboratory for analysis.

Sequence analysis and multiple sequence comparison

Sequences successfully obtained were aligned with representative GenBank sequences of previously published genotypes using the BioEdit Sequence alignment Editor [14]. The search for sequence similarities were carried out using the Basic Local Alignment Search Tool (Blastn) from NCBI BLAST web portal. Phylogenetic trees were performed in MEGA 6 software [15] using the neighbor-joining method, and the statistical significance of the tree topology tested by bootstrapping (1,000 replicates). The evolutionary distances were derived using the Tamura-Nei method. Bootstrap replicates with values \geq 70 are shown on the trees.



Statistical analysis. Regarding HAdV infection comparisons between age groups were performed using the Fisher's exact test. P value < 0.05 was considered statistically significant and the 0-5 year age group was used as reference group. HAdV mono-infections were also compared to HAdV co-infections. The R.3.0.1 tool was used to perform the analyses.

Ethical considerations

This study is a component of the 4S network syndromic surveillance [12]. The principles of the 4S network were approved by the Ministry of Health in its guidelines for influenza surveillance policy, finalized with the support of Pasteur Institute in Dakar and the Strengthening Influenza Sentinel Surveillance in Africa (SISA) project funded by the WHO. The protocol and oral consent were determined as routine surveillance activity, and therefore non-research by the Senegalese National Ethics committee and the steering committee for 4S network, an entity representing MoH, IPD, WHO and Clinicians in compliance with all applicable National regulations governing the protection of human subjects. Data were collected in an objective of surveillance and are anonymous. The information provided to participants was an informal description of the study. Respiratory specimens were collected, only after informed consent was granted, verbally, to local health care workers by the patients or parents in the case of minors. Oral consent was documented in the patient form with two questions about received information and about oral consent. Patients could refuse to participate, no specimen will be taken. For the surveillance activities, written consent is judged not necessary by the Senegalese national ethics committee, which has also previously approved the work of the National Influenza Center. Collections of non-sensitive data or an observation from normal care in which participants remain anonymous do not require ethics committee review. The patients included in this study were of all ages and consulted the sentinel sites due to influenza-like symptoms; the patients, or parents in the case of minors, accept the tests for respiratory viruses largely because they are free and safe.

Results

Patients' characteristics

Between January 2012 to December of 2015 a total of 6381 samples were collected from patients meeting case definition for ILI at the different sentinel sites, and analyzed: 1213 (19%) from 2012, 1519 (23.8%) from 2013, 1930 (30.2%) from 2014 and 1719 (26.9%) during 2015 (Table 1). Patient ages ranged from 1 month to 95 years. The mean age was 10 years 11 months and median age was 4 years. Approximately the male/female ratio was 0.99 (3163 [49.6%] males and 3185 [49.9%] females). For 33 (0.52%) patients the sex was not documented. More than half of patients (51.7%; 3297/6381) were children of \leq 5 years old followed by 5–10 years age group with 11.5% (731/6381) and 25-50 years age group with 10.9% (696/6381). Patients above 50 years old represented only 3.3% (210/6381) of enrolled patients and for 8.2% (526/6381) ages were not reported.

Patients and adenoviral infection

Of 6381 specimens tested, 1967 (30.8%) were positive for HAdV (Table 2). Detection rates over the study period are almost similar in the first 3 years (2012, 2013 and 2014) while in 2015 there is a marked decrease in adenoviral infections. The mean age of infected patients was 8 years 7 months and median age was 3 years.



Table 1. Demographical characteristics of patients and most prevalent symptoms.

Characteristics	Years	2012	2013	2014	2015	Total
	Total Number	(N = 1213)	(N = 1519)	(N = 1930)	(N = 1719)	(N = 6381)
	Gender no. (%)					
	Female	587(48.39)	767(50.49)	987(51.14)	844(49.10)	3185(49.9)
	Male	610(50.29)	744(48.98)	936(48.50)	873(50.79)	3163(49.6)
	Missing	16(1.32)	8(0.53)	7(0.36)	2(0.12)	33(0.52)
	Age no. (%)					
	0–5 yrs	749(61.75)	758(49.90)	941(48.78)	849(49.39)	3297(51.7)
	5–10 yrs	117(9.65)	163(10.73)	208(10.78)	243(14.14)	731(11.5)
	10–15 yrs	56(4.62)	72(4.74)	99(5.13)	109(6.34)	336(5.3)
	15–25 yrs	78(6.43)	125(8.23)	227(11.76)	155(9.02)	585(9.2)
	25–50 yrs	62(5.11)	124(8.16)	282(14.61)	228(13.26)	696(10.9)
	50+ yrs	21(1.73)	23(1.51)	91(4.72)	75(4.36)	210(3.3)
	Missing	130(10.72)	254(16.72)	82(4.25)	60(4.49)	526(8.2)
	Clinical signs no. (%)					
	Myalgia	125(10.31)	327(21.53)	306(15.85)	302(17.57)	1060(16.6)
	Fever	1129(93.08)	1350(88.87)	1864(96.58)	1652(96.10)	5995(93.9)
	Cough	824(67.93)	1099(72.35)	1554(80.52)	1373(79.87)	4850(76.0)
	Vomiting	124(10.22)	33(2.17)	94(4.87)	148(8.61)	399(6.2)
	Diarrhea	94(7.75)	30(1.97)	43(2.23)	39(2.27)	206(3.2)
	Headache	105(8.66)	182(11.98)	263(13.63)	305(17.74)	855(13.4)
	Dyspnea	20(1.65)	24(1.58)	87(4.51)	24(1.40)	155(2.4)
	Rhinitis	734(60.51)	959(63.13)	741(38.39)	522(30.37)	2956(46.3)
	Pharyngitis	109(8.99)	151(9.94)	409(21.19)	318(18.50)	987(15.5)

Among patients the most common respiratory symptoms were fever (94%; 5995/6381), cough (76%; 4850/6381) and rhinitis (46.3%; 2956/6381). Myalgia, pharyngitis, headache, dyspnea and diarrhea were also reported in smaller proportions.

https://doi.org/10.1371/journal.pone.0174287.t001

Table 2. Detection rates of human adenovirus infection in patients with ILI per year from 2012 to 2015 in Senegal and comparison of the distribution into the different age groups.

Year	2012	2013	2014	2015	Total	
Total Number	(N = 1213)	(N = 1519)	(N = 1930)	(N = 1719)	(N = 6381)	P-values
Positivity (per year)	34.71	33.2	33.2	23.3	30.8	
Sex male infection	224(53.2)	252(49.9)	313(48.8)	204(51.0)	993(50.5)	0.373
Age and infection						
0_5 years	324(80.0)	283(56.0)	372(58.0)	245(61.2)	1224(62.2)	< 2.2e-16
5_10 years	30(7.1)	47(9.3)	59(9.2)	54(13.5)	190(9.7)	
10_15 years	16(3.8)	20(4.0)	27(4.2)	15(3.7)	78(4.0)	
15_25 years	15(3.6)	32(6.3)	64(10.0)	22(5.5)	133(6.8)	
25_50 years	11(2.6)	35(6.9)	68(10.6)	38(9.5)	152(7.7)	
50+ years	5(1.2)	9(1.8)	26(4.1)	14(3.5)	54(2.7)	
Missing	20(4.7)	79(15.6)	25(3.9)	12(3.0)	136(6.9)	
Total	421(34.7)	505(33.2)	641(33.2)	400(23.3)	1967(30.8)	

https://doi.org/10.1371/journal.pone.0174287.t002



Year	2012	2013	2014	2015	Total	P-values
Total Number	(N = 1213)	(N = 1519)	(N = 1930)	(N = 1719)	(N = 6381)	
Positivity (per year)	34.71	33.25	33.21	23.27	30.83	
Clinical signs no. (%)						
Myalgia	29(6.9)	116(23.0)	78(12.2)	60(15.0)	283(14.4)	0.001437
Fever	396(94.1)	459(90.9)	613(95.6)	383(95.7)	1851(94.1)	0.9138
Cough	291(69.1)	386(76.4)	534(83.3)	331(82.7)	1542(78.4)	0.002881
Vomiting	55(13.1)	7(1.4)	30(4.7)	43(10.7)	135(6.9)	0.1789
Diarrhea	42(10.0)	14(2.8)	19(3.0)	14(3.5)	89(4.5)	9.194e-05
Headache	27(6.4)	67(13.3)	72(11.2)	68(17.0)	234(11.9)	0.01864
Dyspnea	5(1.2)	4(0.8)	29(4.5)	2(0.5)	40(2.0)	0.1707
Rhinitis	278(66.0)	344(68.1)	253(39.5)	106(26.5)	981(49.9)	0.0001482
Pharyngitis	29(6.9)	45(8.9)	118(18.4)	60(15.0)	252(12.8)	8.95e-05

Values in bold are considered statistically significant

https://doi.org/10.1371/journal.pone.0174287.t003

From the 1967 adenovirus positive cases, 1561 (79.4%) were found in co-infection with at least one respiratory virus. The most common were influenza viruses (53.1%; 1045/1967), rhinoviruses (30%; 591/1967), enteroviruses (18.5%; 364/1967) and RSV (13.5%; 266/1967).

Regarding the viral detection per age group, most of HAdV infected cases (62.2%; 1224/1967) were under 5 years patients, a statistically significant finding (p <0.05). However, the detection rates in the other groups including the elderly (above 50 years old) remain high. No significantly gender distribution of adenoviral infection was observed.

The comparison of symptoms prevalence between ILI patients with adenoviral infection and patients without adenoviral infection showed that cases of myalgia (P = 0.0014), cough (P = 0.0028), diarrhea (P < 0.001), rhinitis (P < 0.001) and headache (P = 0.01) are significantly higher in patients infected by adenoviruses (Table 3).

The Fig 1 shows the temporal distribution of HAdV positivity rate per month in Senegal from 2012 to 2015. We noted that HAdV was detected throughout the year at a high level with detection peaks of different amplitude. The highest peak, with 62% of detection rate, was recorded on December 2013. HAdV circulation pattern shows no seasonality even if results

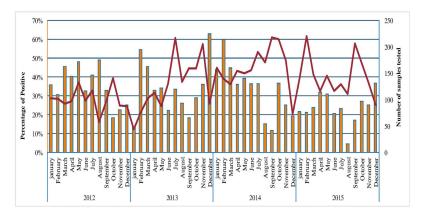


Fig 1. Distribution of adenovirus-positive cases among patients with influenza-like illness, by month and year. The curve represents the total number of influenza-like Illness cases tested for adenovirus. Bars represent the proportions (rates) of adenovirus-positive cases for each month.

https://doi.org/10.1371/journal.pone.0174287.g001



suggest a higher activity of these viruses during cold periods. It should be pointed out that the cold periods (between December and February) experience some instability in Senegal with possibilities of shifting.

Phylogenetic analysis and typing of HAdV

For phylogenetic analysis, we were able to obtain the partial hexon gene sequence from 54 HAdV-positive samples: 8 were from samples in 2012, 13 from 2013, 11 from 2014, 16 from 2015 and 6 from 2016. Unfortunately, some samples showed no amplification or poor-quality sequences. The low sensitivity of conventional PCR compared with real time PCR on samples with low viral load, and certainly non-specific amplifications could be the cause of these failures.

The nucleotide sequence alignment clustered the majority of Senegalese isolates into HAdV-C species (44/54). 9 isolates grouped with HAdV-B species and the remaining isolate, from 2012, seems close to the HAdV-4 genome type belonging to the HAdV-E species. In all cases bootstrap values are high (more than 85%). Within the HAdV-C species, 16 Senegalese isolates are grouped with the type HAdV-6 (36.4%); 2 isolates with HAdV-2 type (4.5%), 4 with HAdV-5 type (9%), and 22 isolates formed a subcluster with HAdV-1 and 57 types (Fig 2). The 9 HAdV-B like species from Senegal grouped with genome types HAdV-7, HAdV-55 and HAdV-11 as shown by a phylogenetic branch with a high bootstrap value of (88%). We also noted that this dominance of species C and B is confirmed over the years.

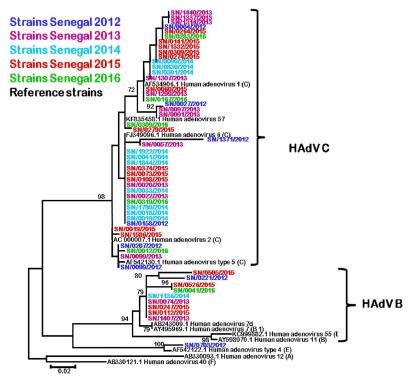


Fig 2. Molecular typing of Human Adenoviruses (HAdV) detected in patients with ILI in Senegal from 2012 to 2016. The last 300 nucleotides of the adenovirus hexon gene were amplified, sequenced, and compared to published sequences (in black) from GenBank using the neighbor-joining method with 1000 bootstrap replicates with MEGA 6 version. Senegalese strains are represented in different colors depending on the year of detection.

https://doi.org/10.1371/journal.pone.0174287.g002



Discussion

In this four-year retrospective study, we characterized HAdV isolates derived from an ILI surveillance program conducted as collaboration between Pasteur Institute of Dakar and the Senegalese Ministry of Health between 2012 and 2015. It is the first nationally molecular epidemiology investigation of HAdVs and even in West Africa.

Our study suggests that HAdV are strongly associated with ILI syndrome in Senegal with an overall detection rate of 30.8% among 6381 patients. This rate seems very higher in comparison with available data. Indeed, much lower rates are reported in similar other studies conducted in other countries. A study conducted in Kenya [16] on refugees from different countries (Somalia, Sudan, Ethiopia and Kenya) yielded a detection rate of 21.7%, in Gabon Douki et al [17] detected HAdV in 16.3% of outpatients with ILI. These detection rates are still lower in other geographical regions: South Korea with 10.1% or 0.6% [18,19], China with 2.7% or 6.3% [20,21], Philippines with 0.9% [22], Malaysia less than 2% [23], USA with 5.7% or 2.8% reported [24,25], Canada in the Ontario Provence with 0.9% [26], Peru with 6.2% [27], Venezuela with 1.6% [28], England with 6.6% [29]. The analysis of these data tends to confirm a higher prevalence of adenoviruses in the respiratory sphere in African populations. This trend was largely confirmed when we investigated the importance of HAdV in children with acute respiratory infections. Indeed we observed that proportions in Cameroon (27.3%) [30] and Senegal (29.2%) (Fall et al, on submission) were considerably higher than those found in other geographical areas: Nascimento-Carvalho et al., [31] in Brazil with 3%, Moe et al., [32] in Norway (1.7%), Wansaula et al., [33] in USA (1%) or Lu et al., [34] in China.

However, these discrepancies in HAdV detection rates can be also due to differences in technical approaches, virus burden geographical differences, the number of patients tested, the periods during which samples were collected and even the duration of the study. It should be also noted that adenoviral detection does not necessarily prove disease causation as coincidental upper airway infection, asymptomatic viral carrier state [35], or prolonged shedding [36] in a previous infection could explain adenoviral detection.

Regarding the group age, as expected, results showed that most patients with HAdV infection were younger than 5 years (62.2%), a statistically significant finding. These results are in concordance with those of other studies which findings concluded that most children are infected by adenovirus at an early age [25,37,38-41]. Indeed, it is well established that by 5 years of age, 70% to 80% of children demonstrate antibodies to at least one serotype [42]. Additionally more than 80% of diagnosed HAdV infections occur in children < 4 years old (due to lack of humoral immunity) [43]. Although most cases exhibit low to mild symptoms are and indistinguishable from other viral causes, acute respiratory infections caused by HAdV can be severe [44], or even fatal [7,45], and are associated with the highest risk of long term respiratory sequelae [46].

Consistent with the report from many other studies, results here showed that 79.4% of HAdV infected participants were co-infected with one or more other respiratory tract viruses. The most frequently co-detected viruses were influenza viruses (53.1%), rhinoviruses (30%), enteroviruses (18.5%) and RSV (13.5%). However, we noted no significant differences in clinical characteristics and laboratory findings between patients with single HAdV infection and those co-infected. The same observation was reported in studies conducted in diverse geographical contexts [27,41]. A previous study conducted in Chilean children stated that the clinical severity in patients with single HAdV infection and those with mixed infections was the same [47]. The overall finding is that the clinical value of such co-infections is not clear and still requires independent investigations in order to assess the association between co-infection and severe illness or symptoms.



Regarding the four years of surveillance, HAdV circulation pattern shows no clear seasonality even if results suggest a higher activity of these viruses during cold periods. This lack of seasonality of HAdV infection has been largely reported elsewhere [27,48,49]. However, seasonal peaks for HAdV infection were noted in summer in some China areas [50] or in spring in Northern China [51], Mexico [52] and Taiwan [53].

In our study, the last 300 bp region of the hexon gene were used for molecular studies of the different HAdV isolates. Phylogenetic analysis showed that among the 54 sequenced strains HAdV-C species were the most common HAdV detected (81.5%) in patients with ILI in Senegal from 2012 to 2016. Despite some divergences, the strains from Senegal were close to types 1, 2, 5, 6 and 57. This HAdV-C species predominance was reported in Malaysia [23], in Italy [54], in many Latina America countries [27,52,55] in contrast with studies done in the United States of America [56], United Kingdom [37], Korea [57], in Argentina [58] and China [59], where HAdV-B species were the most commonly isolated HAdV.

HAdV-B species were the second most common in Senegal with 9 strains, and only one type belonging to HAdV-E species was sequenced. HAdV-B species from Senegal clustered with genome types HAdV-7, HAdV-7d, HAdV-55 and HAdV-11 (88% bootstrap value). HAdV-7d serotype, firstly identified in 1980 in Beijing [59], is of particular concern as it was often associated with illnesses presenting with more severe and higher levels of morbidity than other respiratory HAdV pathogens, and also may result in higher levels of fatalities [60–62]. The HAdV-55 genome type, formerly known as HAdV-11a, is a genotype resulting from recombination between HAdV-11 and HAdV-14 [63]. The serotype has recently reemerged as a highly virulent pathogen, causing severe [64] and sometimes fatal pneumonia among immunocompetent adults, particularly in Asia [65–67]. So the circulation of such HAdV genome types in Senegal emphasizes the need to reinforce HAdV surveillance, especially in hospitalized patients, by including HAdV genome detection and genotyping in the documentation of severe respiratory infections.

The single HAdV-E species strain was typed as HAdV-4, the unique human type in this species, which is more commonly associated with high rates of febrile respiratory illness in US military recruits [68] though associated with viral conjunctivitis outbreak in Australia [69] for example.

We observed some limitations in our study. First, considering the vast number of HAdV positive samples, only a small number of HAdV were typed. So the sequencing results do not reflect the full spectrum of HAdV strains that may be circulating in ILI patients in Senegal, and even for selected samples it may have a bias toward samples with a high viral load. Another limitation concerned the molecular methods used for typing HAdVs in this study, a method which targeted a short hexon hypervariable region that has been shown to correlate closely with serotype. This method does not provide genomic detail and might miss recombination events located in other regions of the genome. Therefore, full-genome sequencing would be more informative on Senegalese strains, especially for HAdV-B7 and HAdV-B55 types. The results of this study should also be interpreted with caution especially for HAdV ILI causality (carriage in healthy or asymptomatic individuals).

Conclusion

In conclusion, the results of the present study suggest strong year-round HAdV activity in Senegal, especially in children up to 5 years of age. Molecular studies revealed that the dominant species in circulation in patients with ILI appears to be HAdV-C, HAdV-B species. The circulation of though HAdV-7d and HAdV-55 genome types is of note as these serotypes are recognized causes of more severe and even fatal acute respiratory infections. So in the interest of



global public health we strongly suggest molecular surveillance and genotyping of newly detected HAdV strains in Senegal and even by whole genome sequencing for some especial strains. Our study offers also an important perspective on the burden of adenovirus-associated respiratory illness in Senegal. Such a perspective, especially among children, should include asymptomatic controls, SARI cases, information on disease outcome, atypical clinical signs, duration of symptoms, and treatment. Data regarding viral load, shedding, and other possible etiologies (e.g., bacterial and other viruses) would also enable a more thorough assessment of the viral effective disease (or symptom) causality.

Acknowledgments

This study would not have been possible without the excellent support from all the health-care workers of the 4S network who contributes, every day, to the surveillance network. We convey our special thanks to Kathleen Victoir from the International Network of Pasteur Institutes for her unwavering support to the 4S network. We acknowledge the Senegalese Ministry of Health for their help in implementing the 4S Network.

Author Contributions

Conceptualization: ND MNN.

Data curation: ND AF. Formal analysis: ND AF.

Funding acquisition: ND MNN.

Investigation: AF NSD DEK DG SS.

Methodology: ND MNN.

Project administration: ND MNN.

Resources: MNN ND FDS MAB.

Supervision: ND MNN MF.

Validation: ND MNN.

Visualization: ND MNN.

Writing - original draft: ND MNN MF. Writing - review & editing: ND MNN.

References

- Lion T. (2014) Adenovirus Infections in Immunocompetent and Immunocompromised Patients. Clin. Microbiol. Rev. 27, 441-462. https://doi.org/10.1128/CMR.00116-13 PMID: 24982316
- 2. Harrach B, Benko M, Both GW (2011) Family Adenoviridae. In: King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (eds): Virus taxonomy. 9th Report of the International Committee on Taxonomy of Viruses. Elsevier, New York: 125-141.
- 3. San MC, Burnett RM (2003). Structural studies on adenoviruses. Curr Top Microbiol Immunol.; 272:57-94. PMID: 12747547
- 4. Schmitz H, Wigand R, Heinrich W (1983). Worldwide epidemiology of human adenovirus infections. Am J Epidemiol.; 117:455-466. PMID: 6301263
- 5. Ison MG (2006). Adenovirus infections in transplant recipients. Clin Infect Dis.; 43:331–9. http://dx.doi. org/10.1086/505498 https://doi.org/10.1086/505498 PMID: 16804849



- Lee J, Choi EH, Lee HJ (2010). Clinical severity of respiratory adenoviral infection by serotypes in Korean children over 17 consecutive years (1991–2007). J Clin Virol.; 49:115–20. http://dx.doi.org/10. 1016/j.jcv.2010.07.007 https://doi.org/10.1016/j.jcv.2010.07.007 PMID: 20692203
- 7. Moura PO, Roberto AF, Hein N, Baldacci E, Vieira SE, Ejzenberg B, et al. (2007). Molecular epidemiology of human adenovirus isolated from children hospitalized with acute respiratory infection in São Paulo, Brazil. J Med Virol.; 79:174-81. http://dx.doi.org/10.1002/jmv.20778 https://doi.org/10.1002/jmv. 20778 PMID: 17177301
- Kandel R, Srinivasan A, D'Agata EMC, Lu X, Erdman D, Jhung M (2010). Outbreak of adenovirus type 4 infection in a long-term care facility for the elderly. Infect Control Hosp Epidemiol.; 31:755-7. http:// dx.doi.org/10.1086/653612 https://doi.org/10.1086/653612 PMID: 20509762
- Kidd AH, Jonsson M, Garwicz D, Kajon AE, Wermenbol AG, Verweij MW, De Jong JC (1996). Rapid subgenus identification of human adenovirus isolates by a general PCR. J Clin Microbiol.; 34:622-627. PMID: 8904426
- Calder JA, Erdman DD, Ackelsberg J, Cato SW, Deutsch VJ, Lechich AJ, et al. (2004). Adenovirus type 7 genomic-type variant, New York City, 1999. Emerg Infect Dis.; 10:149-52. http://dx.doi.org/10.3201/ eid1001.020605 https://doi.org/10.3201/eid1001.020605 PMID: 15078614
- Lebeck MG, McCarthy TA, Capuano AW, Schnurr DP, Landry ML, Setterquist SF, et al. (2009). Emergent US adenovirus 3 strains associated with an epidemic and serious disease. J Clin Virol.; 46:331-6. http://dx.doi.org/10.1016/j.jcv.2009.09.023 https://doi.org/10.1016/j.jcv.2009.09.023 PMID: 19854101
- Dia N, Diene Sarr F, Thiam D, Faye Sarr T, Espié E, Ba O. et al (2014). Influenza-like illnesses in Senegal: not only focus on influenza viruses. PLoS One. Mar 27; 9(3):e93227. https://doi.org/10.1371/ journal.pone.0093227 PMID: 24675982
- Garcia J, Sovero M, Laguna-Torres VA, Gomez J, Chicaiza W, et al. (2009) Molecular characterization of adenovirus circulating in Central and South America during the 2006–2008 period. Influenza Other Respi Viruses 3: 327-330.
- Hall TA, (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95-98.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol. Dec; 30(12):2725-9. https://doi.org/10.1093/molbev/mst197 PMID: 24132122
- 16. Ahmed JA, Katz MA, Auko E, Njenga MK, Weinberg M, Kapella BK, et al. (2012). Epidemiology of respiratory viral infections in two long-term refugee camps in Kenya, 2007-2010. BMC Infect Dis. Jan 17; 12:7. https://doi.org/10.1186/1471-2334-12-7 PMID: 22251705
- Lekana-Douki SE, Nkoghe D, Drosten C, Ngoungou EB, Drexler JF, et al. (2014) Viral etiology and seasonality of influenza-like illness in Gabon, March 2010 to June 2011, BMC Infect Dis. Jul 7: 14:373. https://doi.org/10.1186/1471-2334-14-373 PMID: 25000832
- Kim JK, Jeon JS, Kim JW, Rheem I (2013). Epidemiology of respiratory viral infection using multiplex rt-PCR in Cheonan, Korea (2006–2010). J Microbiol Biotechnol. Feb; 23(2):267–73. PMID: 23412071
- Noh JY, Song JY, Cheong HJ, Choi WS, Lee J, et al. (2013) Laboratory Surveillance of Influenza-Like Illness in Seven Teaching Hospitals, South Korea: 2011-2012 Season. PLoS ONE 8(5): e64295. https://doi.org/10.1371/journal.pone.0064295 PMID: 23717587
- Fu Y, Pan L, Sun Q, Zhu W, Zhu L, Ye C, et al. (2015) The clinical and etiological characteristics of influ-20. enza-like illness (ILI) in outpatients in Shanghai, China, 2011 to 2013. PLoS One. Mar 30; 10(3): e0119513. https://doi.org/10.1371/journal.pone.0119513 PMID: 25822885
- Li H, Wei Q, Tan A, Wang L (2013). Epidemiological analysis of respiratory viral etiology for influenzalike illness during 2010 in Zhuhai, China. Virol J. May 7; 10:143. https://doi.org/10.1186/1743-422X-10-143 PMID: 23651577
- Otomaru H, Kamigaki T, Tamaki R, Opinion J, Santo A, et al. (2015) Influenza and Other Respiratory Viruses Detected by Influenza-Like Illness Surveillance in Leyte Island, the Philippines, 2010–2013. PLoS ONE 10(4): e0123755. https://doi.org/10.1371/journal.pone.0123755 PMID: 25893441
- Abd-Jamil J, Teoh BT, Hassan EH, Roslan N, Abubakar S (2010). Molecular identification of adenovirus causing respiratory tract infection in pediatric patients at the University of Malaya Medical Center. BMC Pediatr. Jul 2; 10:46. https://doi.org/10.1186/1471-2431-10-46 PMID: 20594359
- Fowlkes A, Giorgi A, Erdman D, Temte J, Goodin K, Di Lonardo et al (2014). Viruses associated with acute respiratory infections and influenza-like illness among outpatients from the Influenza Incidence Surveillance Project, 2010-2011. J Infect Dis. Jun 1; 209(11):1715-25. https://doi.org/10.1093/infdis/ jit806 PMID: 24338352
- Koren MA, Arnold JC, Fairchok MP, Lalani T, Danaher PJ, Schofield CM, et al. (2016) Type-specific clinical characteristics of adenovirus-associated influenza-like illness at five US military medical centers,



- 2009–2014. Influenza Other Respir Viruses. Sep; 10(5):414–20. https://doi.org/10.1111/irv.12392 PMID: 27062998
- Abbas KZ, Lombos E, Duvvuri VR, Olsha R, Higgins RR, Gubbay JB (2013). Temporal changes in respiratory adenovirus serotypes circulating in the greater Toronto area, Ontario, during December 2008 to April 2010. Virol J. Jan 7; 10:15. https://doi.org/10.1186/1743-422X-10-15 PMID: 23294909
- Ampuero JS, Ocana V, Gomez J, Gamero ME, Garcia J, Halsey ES, et al. (2012) Adenovirus respiratory tract infections in Peru. PLoS One; 7(10), e46898. https://doi.org/10.1371/journal.pone.0046898
 PMID: 23056519
- Comach G, Teneza-Mora N, Kochel TJ, Espino C, Sierra G, Camacho DE, et al. (2012) Sentinel surveillance of influenza-like illness in two hospitals in Maracay, Venezuela: 2006–2010. PLoS One.; 7(9): e44511. https://doi.org/10.1371/journal.pone.0044511 PMID: 22984519
- 29. Tanner H, Boxall E, Osman H (2012). Respiratory viral infections during the 2009–2010 winter season in Central England, UK: incidence and patterns of multiple virus co-infections. Eur J Clin Microbiol Infect Dis. Nov; 31(11):3001–6. https://doi.org/10.1007/s10096-012-1653-3 PMID: 22678349
- Kenmoe S, Tchendjou P, Vernet MA, Moyo-Tetang S, Mossus T, Njankouo-Ripa M, et al. (2016) Viral
 etiology of severe acute respiratory infections in hospitalized children in Cameroon, 2011–2013. Influenza Other Respir Viruses. Sep; 10(5):386–93. https://doi.org/10.1111/irv.12391 PMID: 27012372
- Nascimento-Carvalho CM, Ribeiro CT, Cardoso MR, Barral A, Araújo-Neto CA, Oliveira JR, et al. (2008) The role of respiratory viral infections among children hospitalized for community-acquired pneumonia in a developing country. Pediatr Infect Dis J; 27:939–941. https://doi.org/10.1097/INF.0b013e3181723751 PMID: 18756190
- Moe N, Pedersen B, Nordbø SA, Skanke LH, Krokstad S, Smyrnaios A, Døllner H (2016). Respiratory Virus Detection and Clinical Diagnosis in Children Attending Day Care. PLoS One Jul 19; 11(7): e0159196. https://doi.org/10.1371/journal.pone.0159196 PMID: 27433803
- Wansaula Z, Olsen SJ, Casal MG, Golenko C, Erhart LM, Kammerer P, et al. (2016) Surveillance for severe acute respiratory infections in Southern Arizona, 2010–2014. Influenza Other Respir Viruses. May; 10(3):161–9. https://doi.org/10.1111/irv.12360 PMID: 26590069
- Lu Y, Wang S, Zhang L, Xu C, Bian C, Wang Z, et al. (2013) Epidemiology of human respiratory viruses in children with acute respiratory tract infections in Jinan, China. 2013. Clin Dev Immunol.: 210490. https://doi.org/10.1155/2013/210490 PMID: 24363757
- Thavagnanam S, Christie SN, Doherty GM, Coyle PV, Shields MD, et al. (2010) Respiratory viral infection in lower airways of asymptomatic children. Acta Paediatr 99: 394–398. https://doi.org/10.1111/j. 1651-2227.2009.01627.x PMID: 20003105
- Madeley CRPM, McQuillin J (1996). Adenoviruses. In: Myint ST-RD, ed. Viral and Other Infections of the Human Respiratory Tract. London: Chapman & Hall: 169–190.
- Cooper RJ, Hallett R, Tullo AB, Klapper PE (2000) The epidemiology of adenovirus infections in Greater Manchester, UK 1982–96. Epidemiol Infect, 125:333–345. PMID: 11117957
- Williams BG, Gouws E, Boschi-Pinto C, Bryce J, Dye C (2002). Estimates of world-wide distribution of child deaths from acute respiratory infections. Lancet Infect Dis.; 2: 25–32. PMID: 11892493
- **39.** Pavia AT. Viral infections of the lower respiratory tract: old viruses, new viruses, and the role of diagnosis. Clin Infect Dis. 2011; Suppl 4:S284–S289.
- Demian PN, Horton KC, Kajon A, Siam R, Hasanin AM, Elgohary Sheta A, et al. (2014) Molecular identification of adenoviruses associated with respiratory infection in Egypt from 2003 to 2010. BMC Infect Dis. Jan 30; 14:50. https://doi.org/10.1186/1471-2334-14-50 PMID: 24479824
- Liu C, Xiao Y, Zhang J, Ren L, Li J, Xie Z, et al. (2015) Adenovirus infection in children with acute lower respiratory tract infections in Beijing, China, 2007 to 2012. BMC Infect Dis. Oct 1; 15:408. https://doi. org/10.1186/s12879-015-1126-2 PMID: 26429778
- Cherry JD (1998) Adenoviruses. In Textbook of Pediatric Infectious Diseases 4th edition. Edited by: Feigin RD, Cherry JD. Philadelphia: WB Saunders:1666–1684.
- Lynch JP 3rd, Kajon AE. (2016) Adenovirus: Epidemiology, Global Spread of Novel Serotypes, and Advances in Treatment and Prevention. Semin Respir Crit Care Med. Aug; 37(4):586–602. https://doi. org/10.1055/s-0036-1584923 PMID: 27486739
- 44. Chang SY, Lee CN, Lin PH, Huang HH, Chang LY, Ko W, et al. (2008) A community-derived outbreak of adenovirus type 3 in children in Taiwan between 2004 and 2005. J Med Virol; 80(1):102–112 https://doi.org/10.1002/jmv.21045 PMID: 18041026
- 45. Dudding BA, Wagner SC, Zeller JA, Gmelich JT, French GR, Top FH Jr (1972). Fatal pneumonia associated with adenovirus type 7 in three military trainees. N Engl J Med; 286(24):1289–1292. https://doi.org/10.1056/NEJM197206152862403 PMID: 4337012



- 46. Edmond K, Scott S, Korczak V, Ward C, Sanderson C, Theodoratou E, et al. (2012) Long term sequelae from childhood pneumonia; systematic review and meta-analysis. PLoS One; 7(2), e31239. https://doi.org/10.1371/journal.pone.0031239 PMID: 22384005
- **47.** Palomino MA, Larranaga C, Villagra E, Camacho J, Avendano LF (2004). Adenovirus and respiratory syncytial virus-adenovirus mixed acute lower respiratory infections in Chilean infants. Pediatr Infect Dis J.; 23(4):337–41. PMID: 15071289
- 48. Alonso WJ, Laranjeira BJ, Pereira SA, Florencio CM, Moreno EC, et al. (2012) Comparative dynamics, morbidity and mortality burden of pediatric viral respiratory infections in an equatorial city. Pediatr Infect Dis J 31: e9–14. https://doi.org/10.1097/INF.0b013e31823883be PMID: 22001966
- Jain S, Williams DJ, Arnold SR, Ampofo K, Bramley AM, Reed C, et al. (2015) Community-acquired pneumonia requiring hospitalization among U.S. children. N Engl J Med.; 372(9):835–45. https://doi. org/10.1056/NEJMoa1405870 PMID: 25714161
- Chen Y, Liu F, Wang C, Zhao M, Deng L, Zhong J, et al. (2016) Molecular Identification and Epidemiological Features of Human Adenoviruses Associated with Acute Respiratory Infections in Hospitalized Children in Southern China, 2012–2013. PLoS One. May 12; 11(5):e0155412. https://doi.org/10.1371/ journal.pone.0155412 PMID: 27171486
- Li Y, Zhou W, Zhao Y, Wang Y, Xie Z, Lou Y, et al. (2015) Molecular Typing and Epidemiology Profiles
 of Human Adenovirus Infection among Paediatric Patients with Severe Acute Respiratory Infection in
 China. Plos One; 10(4):e0123234. https://doi.org/10.1371/journal.pone.0123234 PMID: 25856575
- Rosete DP, Manjarrez ME, Barron BL (2008) Adenoviruses C in nonhospitalized Mexican children older than five years of age with acute respiratory infection. Mem Inst Oswaldo Cruz 103: 195–200. PMID: 18425273
- Cheng CC, Huang LM, Kao CL, Lee PI, Chen JM, Lu CY, et al. (2008) Molecular and clinical characteristics of adenoviral infections in Taiwanese children in 2004–2005. Eur J Pediatr.; 167(6):633–40. https://doi.org/10.1007/s00431-007-0562-4 PMID: 17876605
- 54. Esposito S, Zampiero A, Bianchini S, Mori A, Scala A, Tagliabue C, et al. (2016) Epidemiology and Clinical Characteristics of Respiratory Infections Due to Adenovirus in Children Living in Milan, Italy, during 2013 and 2014. PLoS One. Apr 5; 11(4):e0152375. https://doi.org/10.1371/journal.pone.0152375 PMID: 27045588
- Luiz LN, Leite JP, Yokosawa J, Carneiro BM, Pereira Filho E, et al. (2010) Molecular characterization of adenoviruses from children presenting with acute respiratory disease in Uberlandia, Minas Gerais, Brazil, and detection of an isolate genetically related to feline adenovirus. Mem Inst Oswaldo Cruz 105: 712–716. PMID: 20835622
- Fox JP, Hall CE, Cooney MK (1977). The Seattle virus watch. IV. Observations of adenovirus infections. Am J Epidemiol. 105:362–386. PMID: 192073
- Hong JY, Lee HJ, Piedra PA, Choi EH, Park KH, Koh YY, et al. (2001) Lower respiratory tract infections due to adenovirus in hospitalized Korean children: epidemiology, clinical features, and prognosis. *Clin Infect Dis*, 32:1423–1429. https://doi.org/10.1086/320146 PMID: 11317242
- Barrero PR, Valinotto LE, Tittarelli E, Mistchenko AS (2012). Molecular typing of adenoviruses in pediatric respiratory infections in Buenos Aires, Argentina (1999–2010). J Clin Virol 53: 145–150. https://doi.org/10.1016/j.jcv.2011.11.001 PMID: 22138300
- 59. Li Q. G., Zheng Q. J., Liu Y. H. & Wadell G (1996). Molecular epidemiology of adenovirus types 3 and 7 isolated from children with pneumonia in Beijing. Journal of medical virology 49, 170–177. https://doi.org/10.1002/(SICI)1096-9071(199607)49:3<170::AID-JMV3>3.0.CO;2-1 PMID: 8818961
- 60. Erdman DD, Xu W, Gerber SI, Gray GC, Schnurr D, Kajon AE. et al. (2002) Molecular Epidemiology of Adenovirus Type 7 in the United States, 1966–2000. Emerg. Infect. Dis. 8, 269–277. https://doi.org/10. 3201/eid0803.010190 PMID: 11927024
- 61. Zhao S, Wan C, Ke C, Seto J, Dehghan S, Zou L, et al. (2014) Re-emergent human adenovirus genome type 7d caused an acute respiratory disease outbreak in Southern China after a twenty-one year absence. Sci Rep. Dec 8; 4:7365. https://doi.org/10.1038/srep07365 PMID: 25482188
- 62. Yu Z, Zeng Z, Zhang J, Pan Y, Chen M, Guo Y, et al. (2016) Fatal Community-acquired Pneumonia in Children Caused by Re-emergent Human Adenovirus 7d Associated with Higher Severity of Illness and Fatality Rate. Sci Rep. Nov 16; 6:37216. https://doi.org/10.1038/srep37216 PMID: 27848998
- 63. Seto D, Jones MS, Dyer DW, Chodosh J (2013). Characterizing, typing, and naming human adenovirus type 55 in the era of whole genome data. J Clin Virol.; 58:741–2. https://dx.doi.org/10.1016/j.jcv.2013.09. 025 https://dx.doi.org/10.1016/j.jcv.2013.09. 025 https://
- 64. Lafolie J, Mirand A, Salmona M, Lautrette A, Archimbaud C, Brebion A, et al. (2016) Severe Pneumonia Associated with Adenovirus Type 55 Infection, France, 2014. Emerg Infect Dis. Nov; 22(11):2012–2014. https://doi.org/10.3201/eid2211.160728 PMID: 27767916



- 65. Kajon AE, Dickson LM, Metzgar D, Houng H-S, Lee V, Tan B-H (2010). Outbreak of febrile respiratory illness associated with adenovirus 11a infection in a Singapore military training camp. J Clin Microbiol.; 48:1438–41. http://dx.doi.org/10.1128/JCM.01928-09 https://doi.org/10.1128/JCM.01928-09 PMID: 20129957
- 66. Lu Q-B, Tong Y-G, Wo Y, Wang H-Y, Liu E-M, Gray GC, et al. (2014) Epidemiology of human adenovirus and molecular characterization of human adenovirus 55 in China, 2009–2012. Influenza Other Respir Viruses; 8:302–8. http://dx.doi.org/10.1111/irv.12232 https://doi.org/10.1111/irv.12232 PMID: 24467816
- **67.** Zhang S-Y, Luo Y-P, Huang D-D, Fan H, Lu Q-B, Wo Y, et al. (2016). Fatal pneumonia cases caused by human adenovirus 55 in immunocompetent adults. Infect Dis (Lond); 48:40–7.
- 68. Gray GC, Callahan JD, Hawksworth AW, Fisher CA, Gaydos JC (1999). Respiratory diseases among U.S. military personnel: countering emerging threats. Emerg Infect Dis; 5(3):379–85. https://doi.org/10.3201/eid0503.990308 PMID: 10341174
- Schepetiuk SK, Norton R, Kok T, Irving LG (1993). Outbreak of adenovirus type 4 conjunctivitis in South Australia. J Med Virol; 41(4):316–8 PMID: 8106866