

REVIEW

Is pharmacological, H₂S-induced 'suspended animation' feasible in the ICU?

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Introduction

By definition, 'suspended animation' is a hypometabolic state characterized by the "the slowing of life processes by external means without termination" [1]. Various mammalian species are capable of nearly completely shutting down their vital functions in order to survive otherwise lethal environmental conditions, such as prolonged impairment of O₂ supply and/or extreme temperatures. First described and studied in patients as "hibernation artificielle" induced by the so-called "cocktail lytique" during the Indochina war in the early 1950 s, for obvious reasons the concept of inducing such a hypometabolic condition has attracted special interest in intensive care and emergency medicine. Originally, organ-protection, in particular for the central nervous system (CNS), was demonstrated when suspended animation was induced by rapidly cooling experimental animals to core body temperatures of about 10–15 °C using ice-cold infusions and/or cardiopulmonary bypass (CPB). Given the potential undesired adverse effects of hypothermia *per se*, e. g., metabolic acidosis, coagulopathy, prolonged inflammation, and impaired host defense, any pharmacological measure allowing for a therapeutic on-demand induction of suspended animation would be of particular interest. Moreover, more recently, it was suggested that the reduced visceral organ function present in critically ill patients and/or after overwhelming hyperinflammation could be referred to as an adaptive mechanism to maintain ATP-homeostasis due to reduced energy expenditure rather than to irreversible organ failure [2]. A landmark paper by Blackstone et al. produced much excitement among researchers in the field of shock and critical illness: These authors

demonstrated that mice inhaling hydrogen sulfide (H₂S) reversibly decreased their energy expenditure, which was associated with a fall in core temperature [3]. In the meantime, numerous pre-clinical studies have been published on the possible organ-protective effects of H₂S, the available data being equivocal depending on the model used and the type of shock investigated. In this context in particular, the impact of H₂S effects on energy metabolism remains a matter of debate. Therefore, the present chapter reviews the available data on H₂S-induced on-demand hypometabolism, and its relation (directly as well as *via* a possible consecutive drop in body temperature) to organ-protective properties of H₂S.

Rodent models

In their above-mentioned murine study, Blackstone et al. demonstrated, in awake, spontaneously breathing animals, that exposure to incremental, sub-toxic gaseous H₂S concentrations (20–80 ppm) dose-dependently decreased energy expenditure within a few minutes as assessed by calorimetric measurement of whole-body O₂ uptake and CO₂ production. This fall in metabolic activity was associated with bradypnea and consecutive hypothermia, with core temperature falling to levels close to ambient values [3]. After washout of H₂S, all these metabolic and cardiopulmonary effects were completely reversible, and animals showed no apparent sequelae. Subsequently, Volpato et al. reported that the reduced metabolic activity went along with bradycardia and, consequently, reduced cardiac output, whereas blood pressure and stroke volume remained unaffected [4]. Maintaining normothermia by external warming attenuated the metabolic depressor effect, but did not completely blunt the cardiovascular response [4]. Various other rodent models confirmed these observations: Inhaling gaseous H₂S [5]–[12] and infusing the soluble sulfide salts, NaSH or Na₂S [6], [13], [14], also induced a reversible reduction in energy expenditure with a

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subsequent fall in core temperature. Under stress conditions resulting from injurious mechanical ventilation [8], [13], ischemia/reperfusion [7], [9], [12], endotoxin challenge [11], or bacterial sepsis [14], this effect coincided with attenuation of lung [8], [12]–[14], liver [9], kidney [7] and heart [12] injury. Most importantly, survival was improved after otherwise lethal stress states, e. g., hemorrhagic shock [6] and exposure to hypoxic hypoxia (fraction of inspired O₂ [FiO₂] 5 %) [5]. In addition to anti-oxidant, anti-inflammatory, and anti-apoptotic properties, H₂S was associated with better maintenance of mitochondrial integrity and function [7], [15], [16]: Treatment with either gaseous H₂S treatment or injection of Na₂S prevented mitochondrial swelling, loss of crypts [7], [15], and, at least under hypothermic conditions, outer mitochondrial membrane rupture as documented by the lack of responsiveness of the mitochondrial respiratory chain to stimulation with exogenous cytochrome c [16].

It should be noted that most of the above-mentioned murine data originate from experiments in awake, spontaneously breathing animals. Consequently, the role of anesthesia for a putative H₂S-induced suspended animation remains unclear. Currently, scarce literature is available comparing the effects of anesthesia and H₂S *per se*. In spontaneously breathing mice, Li et al. demonstrated that H₂S (80 and 250 ppm) produced the same metabolic depression as 0.3 and 0.9 % of isoflurane, respectively, however, without any anesthesia-related muscle atonia. Strikingly, when combining these two interventions, H₂S even antagonized the isoflurane-induced metabolic depression [17]. Finally, in mechanically ventilated mice under continuous intravenous (i.v.) anesthesia, the metabolic depressor effect of H₂S was completely blunted when normothermia was maintained [16].

Large animal species and humans

Any metabolic depressant property of H₂S seems to be dependent on the animal size: In rats the H₂S-induced decrease in O₂ uptake was several-fold lower than in mice [18]. In larger species (swine, sheep), various authors failed to confirm any H₂S-related reduction in metabolic activity at all, regardless of whether inhalation of gaseous H₂S or injection of sulfide salts were studied [19]–[22]. Moreover, in sheep, Derwall et al. [23] demonstrated that during administration of gaseous H₂S via an extracorporeal, veno-arterial membrane oxygenator to avoid any airway mucosa damage related to the gas inhalation [24], [25], whole body O₂ uptake, CO₂ production, and cardiac output remained within the physiological range. At the highest doses administered (300 ppm), H₂S did not affect calorimetric energy expenditure either, but caused pulmonary vasoconstriction associated with arterial hypotension and metabolic acidosis [23]. Finally, in

human volunteers, inhalation of 10 ppm H₂S during exercise decreased O₂ uptake, and this effect was referred to a toxic reduction in maximal aerobic capacity rather than to a regulatory effect on mitochondrial respiration, as evidenced by a tendency for muscle lactate to increase and citrate synthase activity to decrease [26]. Consequently, it was questioned whether any therapeutic potential of the H₂S-induced “suspended animation”-like hypometabolism observed in mice and rats could be transferred to the clinical setting [27], [28]. On the other hand, when external measures to prevent hypothermia were withheld, Na₂S-related organ-protection after kidney ischemia/reperfusion-injury [29] or hemorrhage and resuscitation [30] coincided with a progressive decrease in core temperature (Figure 1). Moreover, in the latter experiments, immediate post-mortem liver tissue mitochondrial activity showed a tendency towards both reduced oxidative phosphorylation and maximal O₂ uptake in the uncoupled state, and, in particular, a significantly decreased “leak respiration”, i. e., the respiratory activity necessary to compensate for the proton leakage, slipping, and cation-exchange along the inner mitochondrial membrane (Figure 2). In other words, H₂S supplementation under these conditions provided protective reduction rather than toxic inhibition of cellular respiration.

How can these diverging findings be reconciled? Under stress conditions, e. g., in response to hypoxia or circulatory shock, small rodents can reduce their energy expenditure as a result of decreased ‘non-shivering thermogenesis’ [31], due to modulation of the uncoupling protein-1, mostly in the brown adipose tissue [32]. In these species, non-shivering thermogenesis represents a large proportion of total O₂ uptake, which can be rapidly decreased without affecting ATP formation [31]. This response is independent of any pharmacological intervention, and represents a unique protective adaptation present in numerous mammals [31] and even in humans, e. g., in neonates and during cold acclimatization [32]. However, due to the high area/volume ratio and, consequently, the higher heat dissipation, it is inversely related to body size [31], i. e., to the ratio of O₂ consumption and body weight. Two phenomena support this latter notion: i) No matter the species, newborns present with more pronounced hypoxia-induced hypometabolism than do adults [31]; ii) when the ratio of O₂ consumption and body weight *per se* is low (e. g., in adults of larger species), normoxic O₂ uptake (e. g., during exercise [31]) may be associated with hypoxia-induced hypometabolism. Hence, if possible at all, achieving a suspended animation-like status in larger animals and humans will be more difficult and require much more time because of the small surface area/mass ratio: In fact, in anesthetized and mechanically ventilated

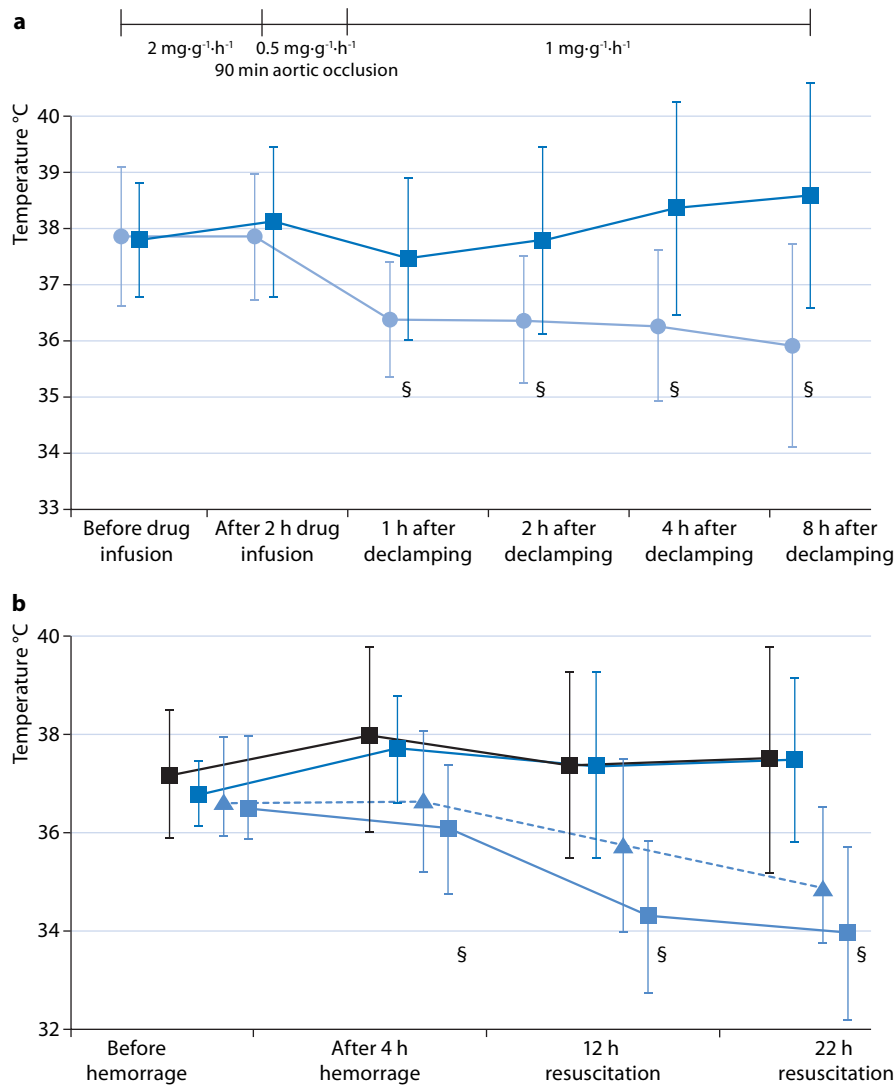
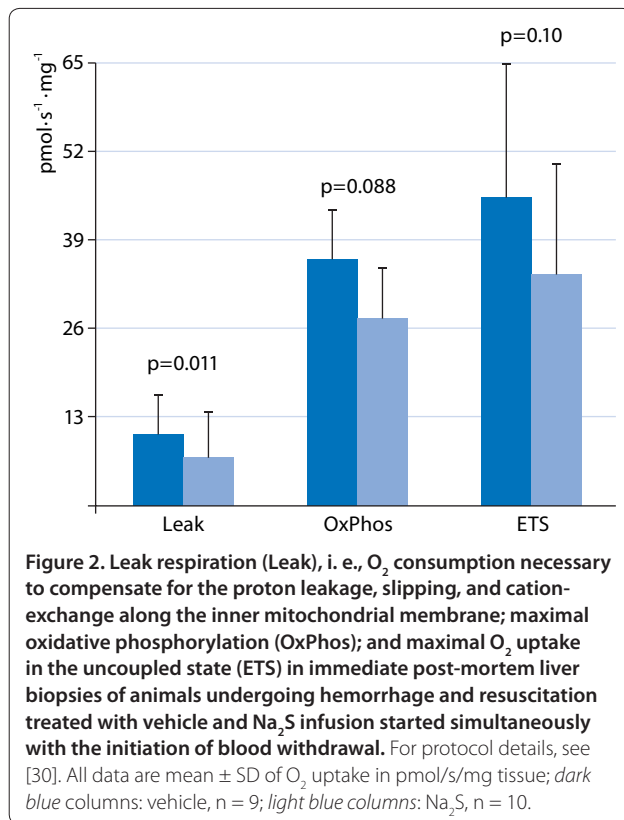


Figure 1. Time course of body core temperature in swine undergoing (a) 90 minutes intra-aortic balloon occlusion-induced kidney ischemia/reperfusion-injury (data are adapted from [29]: **Dark blue squares**, vehicle n = 10; **blue circles**, Na₂S n = 9; all data are mean ± SD, § designates p < 0.05 between groups); (b) hemorrhage and resuscitation (data are adapted from [30]: **black squares**, vehicle n = 14; **dark blue squares**, Na₂S started two hours before hemorrhage, n = 10; **light blue squares**, Na₂S started simultaneously with hemorrhage, n = 11; **blue triangles**, Na₂S started immediately after hemorrhage, n = 10; all data are mean ± SD, § designates p < 0.05 ‘simultaneous’ treatment vs. vehicle). Note that in both experimental series at least four hours of drug infusion were necessary to achieve a significant decrease in body temperature.

swine, after four hours of Na₂S infusion whole body O₂ uptake and CO₂ production started to decrease, subsequently resulting in a moderate decrease in core temperature at ten hours of drug infusion [23] (Fig. 1).

No matter the current debate on the feasibility of pharmacological induction of whole body suspended animation in larger animals, inducing hypometabolism to hibernate isolated organs and, thereby, prolong their tolerance against tissue ischemia or hypoxia remains an attractive option, in particular for organ transplantation.

Numerous studies in rodents have demonstrated that H₂S administration improved kidney, liver heart, and lung function and attenuated histological damage after orthotopic organ transplant. This beneficial effect of H₂S administration (NaSH 0.5 mmol/l over 10 minutes before and immediately after initiation of reperfusion) was confirmed in isolated porcine kidneys *ex vivo* undergoing normothermic reperfusion with autologous blood after 25 minutes of warm ischemia and subsequently 18 hours of storage at 4 °C [33].

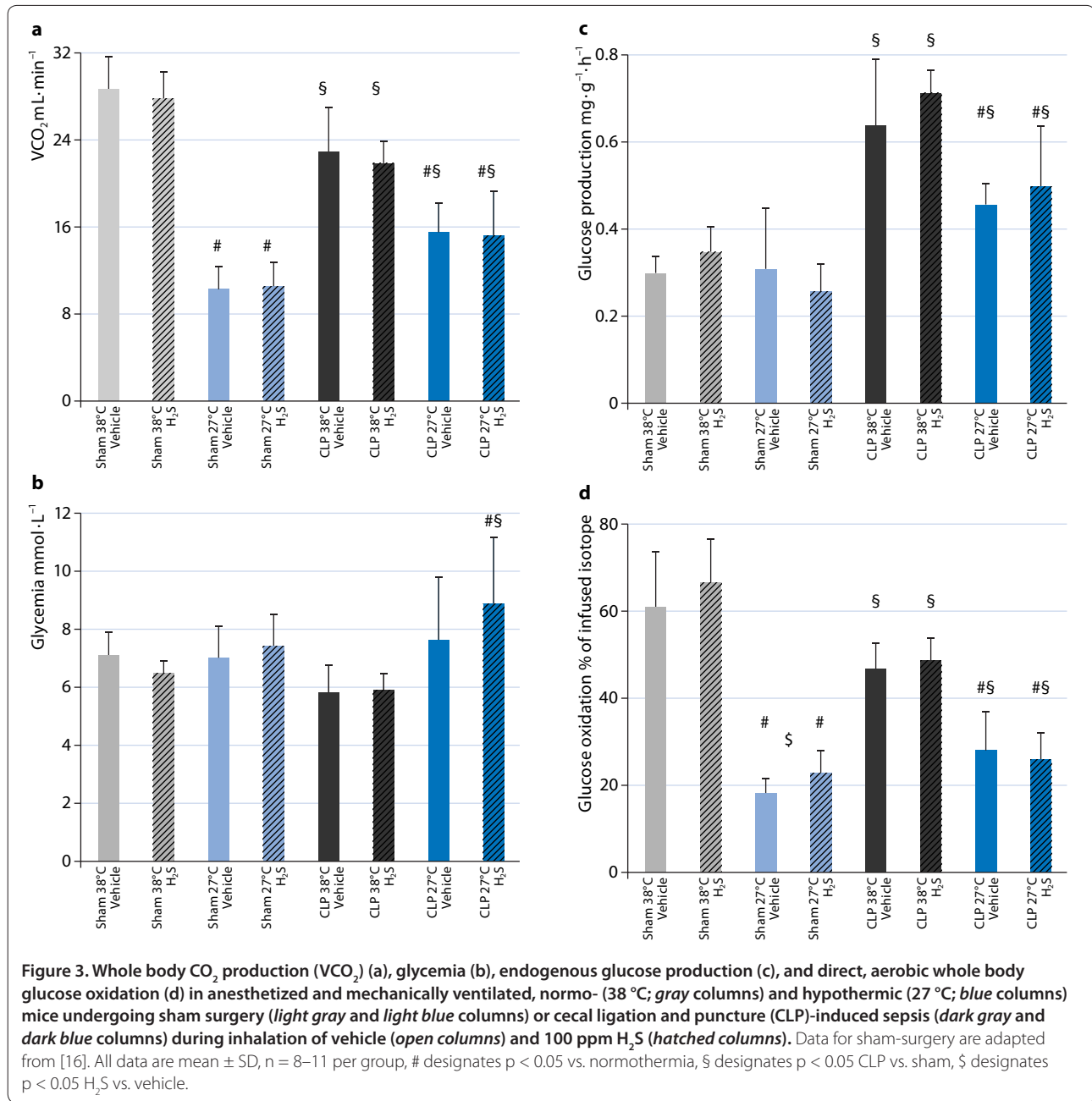


Hypothermia

Equivocal data are available whether hypothermia, caused by a possible H_2S -related fall in energy expenditure and/or due to external cooling measures, assumes importance for organ protection achieved during H_2S administration. Inhaling H_2S prior to myocardial ischemia at concentrations that had no metabolic depressant effect (10 ppm) attenuated organ damage, but to a lesser degree than concentrations that reduced energy expenditure (100 ppm) [12], suggesting that hypometabolism may indeed enhance the organ-protective properties of H_2S . Of note, in that study as well as in others demonstrating H_2S -related organ production coinciding with reduced metabolic activity, hypothermia was prevented [5], [7], [9], [14],[15] in order to elucidate the impact of a simultaneous drop in core temperature. Moreover, organ protection and improved survival were also shown to be in part [12], [13], [15], [34], [35] or even completely [8], [11], [36], [37] independent of any H_2S -induced metabolic depression at all. Finally, data obtained in large animal (swine or sheep) models of shock resulting from ischemia/reperfusion [29], [38]–[42], hemorrhage and resuscitation [30], or burn injury [36] also suggested that the beneficial effects of infusing Na_2S were at least in part independent of metabolic depression and/or a fall in core temperature. Hence, any moderate

hypothermia observed simultaneously with H_2S -induced organ-protection may also be due to attenuation of systemic inflammation rather than to reduced energy expenditure *per se*. In other words, such findings raise a ‘chicken and egg’ problem, which can be attributed to the so-called Q10 effect, i. e., the two to three fold reduction in all chemical reactions and thus metabolism associated with a 10 °C-reduction of body temperature [31]: As an example, during otherwise lethal porcine hemorrhage, therapeutic hypothermia was associated with reduced concentrations of pro-inflammatory cytokines [43]. The potential of H_2S acting as a metabolic depressant in larger species independent of any anti-inflammatory and antioxidant property still remains unsettled: In the above-mentioned swine study showing an H_2S -induced drop in O_2 uptake and CO_2 production as well as a consecutive moderate fall in core temperature, animals underwent a short period of aortic occlusion, which did not cause any increase in the blood levels of pro-inflammatory cytokines or markers of oxidative and nitrosative stress [23].

Irrespective of the question as to whether or not there is cause-effect relationship between H_2S -related organ protection and coinciding hypometabolism and/or hypothermia, hypothermia does assume importance for H_2S -induced effects on substrate utilization and mitochondrial function. It is well-established that H_2S toxicity is due to inhibition of mitochondrial respiration resulting from blockade of the complex IV of the respiratory chain, i. e., cytochrome c oxidase [44]. When compared to normothermia, hypothermia (27 °C) increased the Na_2S concentrations necessary to induce inhibition of mitochondrial respiratory activity (from $< 1 \mu\text{M}$ to 2–4 μM), and nearly doubled the Na_2S concentrations required for a 50 % reduction in mitochondrial respiratory activity [16], [45]. Hypothermia may also influence the effect of H_2S on substrate utilization and, thereby, may even improve the yield of the mitochondrial respiration: In anesthetized and ventilated mice, during normothermia, inhaling 100 ppm H_2S did not affect endogenous glucose production (as calculated from the rate of appearance of 1,2,3,4,5,6- $^{13}\text{C}_6$ -glucose during continuous i.v. isotope infusion), whole body CO_2 production, or direct, aerobic glucose oxidation rate (as derived from VCO_2 and the expiratory $^{13}\text{CO}_2/^{12}\text{CO}_2$ ratio) (Fig. 3). However, under hypothermic (core temperature 27 °C) conditions, the rate of direct, aerobic glucose oxidation increased, suggesting a shift toward preferential carbohydrate utilization [16] (Fig. 3). Such a switch in fuel utilization is associated with an improved yield of oxidative phosphorylation: The ATP synthesis/ O_2 consumption ratio is higher for glycolysis than for β -oxidation, because nicotinamide adenine dinucleotide (NADH) as an electron donor provides three coupling

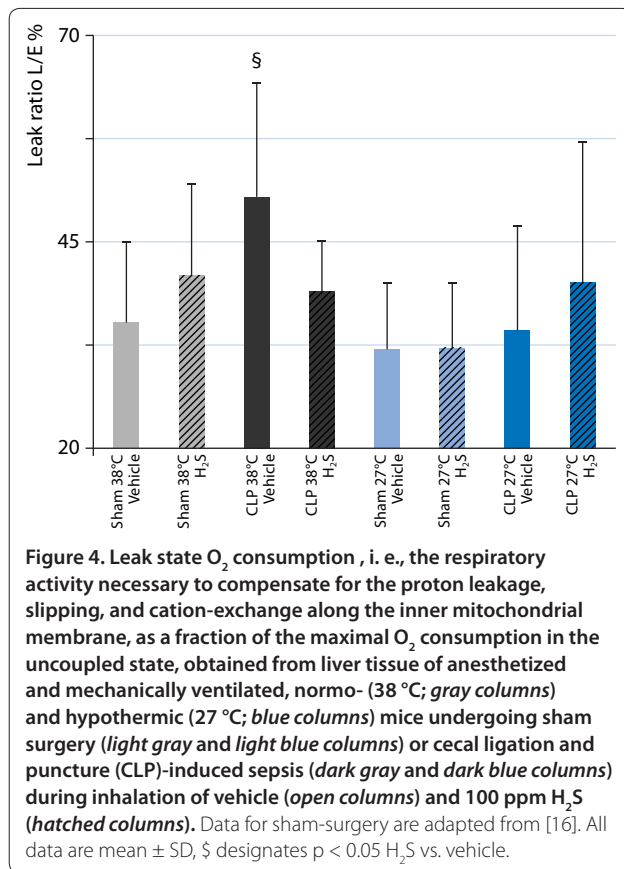


sites rather than just two from FADH₂ [46]. During cecal ligation and puncture-induced septic shock, the metabolic effects of inhaled H₂S partially disappeared: Inhaled H₂S affected neither the sepsis-induced metabolic acidosis [34] nor glucose utilization (Figure 3), nor the responsiveness to stimulation with exogenous cytochrome c oxidase. Nevertheless, H₂S did normalize the sepsis-related increase in “leak respiration” – which was less pronounced during hypothermia – thus allowing for better maintenance of mitochondrial function (Figure 4). It is unclear whether the lack of effect of H₂S on the

mitochondrial respiratory chain was due to the septic challenge *per se* and/or to the ongoing treatment: During sepsis, all mice needed continuous i.v. norepinephrine to achieve target hemodynamics characterized by a normotensive and hyperdynamic circulation. In turn, norepinephrine incubation was associated with impairment of tissue mitochondrial respiration.

Timing and dose

No matter the importance of hypometabolism for the organ-protective properties of H₂S administration *per se*,



the questions of timing and – due to the potential toxic inhibition of mitochondrial respiration – dosing of H₂S remain unsettled. Clearly, there are plenty of data available showing that inhalation of H₂S gas and/or the injection of NaSH or Na₂S can prevent organ damage when administered prior to or at least simultaneously with the initiation of shock. However, the very few studies comparing a pre- and post-treatment design in mice showed marked reduction [7] or even complete disappearance [37] of the protective potency. In swine undergoing long-term hemorrhage and resuscitation, the results were even more curious [47]: Primed-continuous Na₂S administration (initial bolus of 0.2 mg/kg, followed by 1 mg/kg/h over 12 hours of resuscitation) improved survival when compared to vehicle (survival: 71 %), regardless of whether the Na₂S infusion was started two hours before (pre-treatment: survival 100 %) or simultaneously with (survival 91 %) the initiation of blood withdrawal, or at the start of re-transfusion of shed blood (post-treatment: survival 90 %) [30]. However, a significant decrease in core temperature (Fig. 1b) and organ protection were only present in the group of animals treated simultaneously with the initiation of hemorrhage. Apparently, both the cumulative H₂S dose as well as the rate of its generation assume importance

for the effects on metabolism and organ protection, in particular under low flow conditions and/or circulatory shock: In swine undergoing cardiac arrest, primed-continuous Na₂S (0.3 mg/kg followed by 0.3 mg/kg/h over two hours) injected one minute after the start of cardiopulmonary resuscitation (CPR) reduced blood pressure and cardiac output during early resuscitation [21]. Increasing the Na₂S dose (1.0 mg/kg followed by 1.0 mg/kg/h) was associated with impaired neurological recovery. Even injection of comparable total amounts may have markedly different effects due to the different rate of H₂S generation: *In vitro* slow H₂S release from the H₂S donor GYY4137 exerted anti-inflammatory and -apoptotic effects, whereas short-term, high peak free sulfide levels resulting from incubation with NaSH induced the opposite response [48]. *In vivo*, this concept was confirmed in swine undergoing myocardial ischemia/reperfusion injury: A primed-continuous Na₂S infusion was superior to bolus injection [39].

Conclusions

The concept of “buying time in suspended animation” [49] has been discussed in the literature for more than a century. Originally induced by rapid external body cooling, any pharmacological measure allowing for a therapeutic, on demand induction of ‘suspended animation’ is of particular interest because of the undesired side effects of hypothermia *per se*. Therefore, the landmark paper demonstrating that inhaling H₂S could induce a reversible, suspended animation-like hypometabolism [3], produced much excitement among researchers in the field of shock and critical illness. Numerous pre-clinical studies are currently available on H₂S-related organ protection, but the effects on energy metabolism remain a matter of debate. In this context, the well-established toxic blockade of cytochrome c oxidase by H₂S may assume particular importance. Most studies so far suggest that the beneficial effects of H₂S are at least in part independent of an H₂S-induced metabolic depression and, in particular, any decrease in core temperature. However, other data suggest that H₂S-related hypometabolism may enhance the organ-protective properties. The mechanism behind H₂S-induced hypometabolism is still not fully understood, and, moreover, the feasibility of H₂S-induced suspended animation in larger animals has been questioned. Clearly, if possible at all, achieving a suspended animation-like status in larger animals and humans will be more difficult and require much more time because of the small surface area/mass ratio. Again the available data are equivocal, suggesting that at least hibernating isolated organs remains an option. Even in larger species, data on the effects of H₂S on mitochondrial function and morphology suggest that its supplementation during circulatory shock provides

protective reduction rather than toxic inhibition of cellular respiration. Finally, according to the currently available literature, neither inhalation of gaseous H₂S nor injection of the soluble sulfide salts, NaSH or Na₂S, is likely to become part of clinical practice because of damage to the airway mucosa and possibly toxic peak sulfide concentrations, respectively, but slow H₂S-releasing molecules may enable these limitations to be overcome. Hence, there is “nothing rotten about hydrogen sulfide’s medical promise” [50], and H₂S clearly remains a “hot molecule” [51] in the field of research for a possible pharmacological induction of suspended animation-like hypometabolism.

List of abbreviations used

CLP: cecal ligation and puncture; CNS: central nervous system; CPB: cardiopulmonary bypass; CPR: cardiopulmonary resuscitation; NADH: nicotinamide adenine dinucleotide.

Competing interests

The authors declare that they have no competing interests.

Declarations

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References

1. **Suspended animation.** [http://en.wikipedia.org/wiki/Suspended_animation] Accessed Nov 2013
2. Singer M, De Santis V, Vitale D, Jeffcoate W: **Multiorgan failure is an adaptive, endocrine-mediated, metabolic response to overwhelming systemic inflammation.** *Lancet* 2004, **364**:545–548.
3. Blackstone E, Morrison M, Roth MB: **H₂S induces a suspended animation-like state in mice.** *Science* 2005, **308**:518.
4. Volpato GP, Searles R, Yu B, Scherrer-Crosbie M, Bloch KD, Ichinose F, Zapol WM: **Inhaled hydrogen sulfide: a rapidly reversible inhibitor of cardiac and metabolic function in the mouse.** *Anesthesiology* 2008, **108**:659–668.
5. Blackstone E, Roth MB: **Suspended animation-like state protects mice from lethal hypoxia.** *Shock* 2007, **27**:370–372.
6. Morrison ML, Blackwood JE, Lockett SL, Iwata A, Winn RK, Roth MB: **Surviving blood loss using hydrogen sulfide.** *J Trauma* 2008, **65**:183–188.
7. Bos EM, Leuvenink HG, Snijder PM, Kloosterhuis NJ, Hillebrands JL, Leemans JC, Glorquin S, van Goor H: **Hydrogen sulfide-induced hypometabolism prevents renal ischemia/reperfusion injury.** *J Am Soc Nephrol* 2009, **20**:1901–1905.
8. Faller S, Ryter SW, Choi AM, Loop T, Schmidt R, Hoetzel A: **Inhaled hydrogen sulfide protects against ventilator-induced lung injury.** *Anesthesiology* 2010, **113**:104–115.
9. Bos EM, Snijder PM, Jekel H, Weij M, Leemans JC, van Dijk MC, Hillebrands JL, Lisman T, van Goor H, Leuvenink HG: **Beneficial effects of gaseous hydrogen sulfide in hepatic ischemia/reperfusion injury.** *Transpl Int* 2012, **25**:897–908.
10. Seitz DH, Fröba JS, Niesler U, Palmer A, Veltkamp HA, Braumüller ST, Wagner F, Wagner K, Bäder S, Wachter U, Calzia E, Radermacher P, Huber-Lang MS, Zhou S, Gebhard F, Knöferl MW: **Inhaled hydrogen sulfide induces suspended animation, but does not alter the inflammatory response after blunt chest trauma.** *Shock* 2012, **37**:197–204.
11. Tokuda K, Kida K, Marutani E, Crimi E, Bougaki M, Khatri A, Kimura H, Ichinose F: **Inhaled hydrogen sulfide prevents endotoxin-induced systemic inflammation and improves survival by altering sulfide metabolism in mice.** *Antioxid Redox Signal* 2012, **17**:11–21.
12. Snijder PM, de Boer RA, Bos EM, van den Born JC, Ruifrok WP, Vreeswijk-Baudoin I, van Dijk MC, Hillebrands JL, Leuvenink HG, van Goor H: **Gaseous hydrogen sulfide protects against myocardial ischemia-reperfusion injury in mice partially independent from hypometabolism.** *PLoS One* 2013, **8**:e63291.
13. Aslami H, Heinen A, Roelofs JJTH, Zuurbier CJ, Schultz MJ, Juffermans NP: **Suspended animation inducer hydrogen sulfide is protective in an in vivo model of ventilator-induced lung injury.** *Intensive Care Med* 2010, **36**:1946–1952.
14. Aslami H, Pulskens WP, Kuipers MT, Bos AP, van Küllenburg AB, Wander RJ, Roelofsens J, Roelofs JJ, Kerindongo RP, Beurskens CJ, Schultz MJ, Kulik W, Weber NC, Juffermans NP: **Hydrogen sulfide donor NaHS reduces organ injury in a rat model of pneumococcal pneumosepsis, associated with improved bio-energetic status.** *PLoS One* 2013, **8**:e63497.
15. Elrod JW, Calvert JW, Morrison J, Doeller JE, Kraus DW, Tao L, Jiao X, Scala R, Kiss L, Szabo C, Kimura H, Chow CW, Lefer DJ: **Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function.** *Proc Natl Acad Sci USA* 2007, **104**:15560–15565.
16. Baumgart K, Wagner F, Gröger M, Weber S, Barth E, Vogt JA, Wachter U, Huber-Lang M, Knöferl MW, Albuszies G, Georgieff M, Asfar P, Szabó C, Calzia E, Radermacher P, Simkova V: **Cardiac and metabolic effects of hypothermia and inhaled hydrogen sulfide in anesthetized and ventilated mice.** *Crit Care Med* 2010, **38**:588–595.
17. Li RQ, McKinstry AR, Moore JT, Caltagarone BM, Eckenhoff MF, Eckenhoff RG, Kelz MB: **Is hydrogen sulfide-induced suspended animation general anesthesia?** *J Pharmacol Exp Ther* 2012, **341**:735–742.
18. Haouzi P, Bell HJ, Notet V, Bihain B: **Comparison of the metabolic and ventilatory response to hypoxia and H₂S in unsedated mice and rats.** *Respir Physiol Neurobiol* 2009, **2**:316–322.
19. Li J, Zhang G, Cai S, Redington AN: **Effect of inhaled hydrogen sulfide on metabolic responses in anesthetized, paralyzed, and mechanically ventilated piglets.** *Pediatr Crit Care Med* 2008, **9**:110–112.
20. Haouzi P, Notet V, Chenuel B, Chalou B, Sponne I, Ogier V, Bihain B: **H₂S induced hypometabolism in mice is missing in sedated sheep.** *Respir Physiol Neurobiol* 2008, **160**:109–115.
21. Derwall M, Westerkamp M, Löwer C, Deike-Glindemann J, Schnorrenberger NK, Coburn M, Nolte KW, Gaisa N, Weis J, Siepmann k, Häusler M, Rossaint R, Fries M: **Hydrogen sulfide does not increase resuscitability in a porcine model of prolonged cardiac arrest.** *Shock* 2010, **34**:190–195.
22. Drabek T, Kochanek PM, Stezoski J, Wu X, Bayir H, Morhard RC, Stezoski RC, Tisherman SA: **Intravenous hydrogen sulfide does not induce hypothermia or improve survival from hemorrhagic shock in pigs.** *Shock* 2011, **35**:67–73.
23. Derwall M, Francis RC, Kida K, Bougaki M, Crimi E, Adrie C, Zapol WM, Ichinose F: **Administration of hydrogen sulfide via extracorporeal membrane lung perfusion in sheep with partial cardiopulmonary bypass perfusion: a proof of concept study on metabolic and vasomotor effects.** *Crit Care* 2011, **15**:R51.
24. Simon F, Giudici R, Duy CN, Schelzig H, Oter S, Gröger M, Wachter U, Vogt J, Speit G, Szabó C, Radermacher P, Calzia E: **Hemodynamic and metabolic effects of hydrogen sulfide during porcine ischemia/reperfusion injury.** *Shock* 2010, **33**:359–364.
25. Francis RC, Vaporidi K, Bloch KD, Ichinose F, Zapol WM: **Protective and detrimental effects of sodium sulfide and hydrogen sulfide in murine ventilator-induced lung injury.** *Anesthesiology* 2011, **115**:1012–1021.
26. Bhambhani Y, Burnham R, Snydmiller G, MacLean IJ: **Effects of 10-ppm hydrogen sulfide inhalation in exercising men and women. Cardiovascular, metabolic, and biochemical responses.** *J Occup Environ Med* 1997, **39**:122–129.
27. Tisherman SA, Drabek T: **Hydrogen sulfide: metabolic mediator or toxic gas?** *Pediatr Crit Care Med* 2008, **9**:129–130.
28. Haouzi P: **Murine models in critical care research.** *Crit Care Med* 2011, **39**:2290–2293.
29. Simon F, Scheuerle A, Gröger M, Stahl B, Wachter U, Vogt J, Speit G, Hauser B, Möller P, Calzi E, Szabó C, Schelzig H, Georgieff M, Radermacher P, Wagner F: **Effects of intravenous sulfide during porcine aortic occlusion-induced kidney ischemia/reperfusion injury.** *Shock* 2011, **35**:156–163.
30. Bracht H, Scheuerle A, Gröger M, Hauser B, Matallo J, McCook O, Seifritz A, Wachter U, Vogt JA, Asfar P, Matejovic M, Möller P, Calzia E, Szabó C, Stahl W,

- Hoppe K, Stahl B, Lampl L, Georgieff M, Wagner F, Radermacher P, Simon F: **Effects of intravenous sulfide during resuscitated porcine hemorrhagic shock.** *Crit Care Med* 2012, **40**:2157–2167.
31. Mortola JP: **Hypoxic hypometabolism in mammals.** *News Physiol Scie* 1993, **8**:79–82.
32. van Marken Lichtenbelt WD, Schrauwen P: **Implications of nonshivering thermogenesis for energy balance regulation in humans.** *Am J Physiol Regul Integr Comp Physiol* 2011, **301**:R285–R296.
33. Hosgood SA, Nicholson ML: **Hydrogen sulphide ameliorates ischaemia-reperfusion injury in an experimental model of non-heart-beating donor kidney transplantation.** *Br J Surg* 2010, **97**:202–209.
34. Wagner F, Wagner K, Weber S, Stahl B, Knöferl MW, Huber-Lang M, Seitz DH, Asfar P, Calzia E, Senfteleben U, Gebhard F, Georgieff M, Radermacher P, Hysa V: **Inflammatory effects of hypothermia and inhaled H₂S during resuscitated, hyperdynamic murine septic shock.** *Shock* 2011, **2**(35):396–402.
35. Wagner F, Scheuerle A, Weber S, Stahl B, McCook O, Knöferl MW, Huber-Lang M, Seitz DH, Thomas J, Asfar P, Szabó C, Möller P, Gebhard F, Georgieff M, Calzia E, Radermacher P, Wagner K: **Cardiopulmonary, histologic, and inflammatory effects of intravenous Na₂S after blunt chest trauma-induced lung contusion in mice.** *J Trauma* 2011, **71**:1659–1667.
36. Esehie A, Enkhbaatar P, Traber DL, Jonkam C, Lange M, Hamahata A, Djukom C, Whorton EB, Hawkins HK, Traber LD, Szabo C: **Beneficial effect of a hydrogen sulphide donor (sodium sulphide) in an ovine model of burn- and smoke-induced acute lung injury.** *Br J Pharmacol* 2009, **158**:1442–1453.
37. Minamishima S, Bougaki M, Sips PY, Yu JD, Minamishima YA, Elrod JW, Lefer DJ, Bloch KD, Ichinose F: **Hydrogen sulfide improves survival after cardiac arrest and cardiopulmonary resuscitation via a nitric oxide synthase 3-dependent mechanism in mice.** *Circulation* 2009, **120**:888–896.
38. Sodha NR, Clements RT, Feng J, Liu Y, Bianchi C, Horvath EM, Szabo C, Sellke FW: **The effects of therapeutic sulfide on myocardial apoptosis in response to ischemia-reperfusion injury.** *Eur J Cardiothorac Surg* 2008, **33**:906–913.
39. Osipov RM, Robich MP, Feng J, Liu Y, Clements RT, Glazer HP, Sodha NR, Szabo C, Bianchi C, Sellke FW: **Effect of hydrogen sulfide in a porcine model of myocardial ischemia-reperfusion: comparison of different administration regimens and characterization of the cellular mechanisms of protection.** *J Cardiovasc Pharmacol* 2009, **54**:287–297.
40. Sodha NR, Clements RT, Feng J, Liu Y, Bianchi C, Horvath EM, Szabo C, Stahl GL, Sellke FW: **Hydrogen sulfide therapy attenuates the inflammatory response in a porcine model of myocardial ischemia/reperfusion injury.** *J Thorac Cardiovasc Surg* 2009, **138**:977–984.
41. Osipov RM, Robich MP, Feng J, Chan V, Clements RT, Deyo RJ, Szabo C, Sellke FW: **Effect of hydrogen sulfide on myocardial protection in the setting of cardioplegia and cardiopulmonary bypass.** *Interact Cardiovasc Thorac Surg* 2010, **10**:506–512.
42. Hunter JP, Hosgood SA, Patel M, Rose R, Read K, Nicholson ML: **Effects of hydrogen sulphide in an experimental model of renal ischaemia-reperfusion injury.** *Br J Surg* 2012, **99**:1665–1671.
43. Chen Z, Chen H, Rhee P, Koustova E, Ayuste EC, Honma K, Nadel A, Alam HB: **Induction of profound hypothermia modulates the immune/inflammatory response in a swine model of lethal hemorrhage.** *Resuscitation* 2005, **66**:209–216.
44. Cooper CE, Brown GC: **The inhibition of mitochondrial cytochrome oxidase by the gases carbon monoxide, nitric oxide, hydrogen cyanide and hydrogen sulfide: chemical mechanism and physiological significance.** *J Bioenerg Biomembr* 2008, **40**:533–539.
45. Gröger M, Matallo J, McCook O, Wagner F, Wachter U, Bastian O, Gierer S, Reich V, Stahl B, Huber-Lang M, Szabó C, Radermacher P, Calzia E, Wagner K: **Temperature and cell-type dependency of sulfide effects on mitochondrial respiration.** *Shock* 2012, **38**:367–374.
46. Lerverve X, Batandier C, Fontaine E: **Choosing the right substrate.** *Novartis Found Symp* 2007, **280**:108–121.
47. Drabek T: **Hydrogen sulfide—curiouser and curiouser!** *Crit Care Med* 2012, **40**:2255–2256.
48. Whiteman M, Li L, Rose P, Tan C, Parkinson DB, Moore PK: **The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages.** *Antioxid Redox Signal* 2010, **12**:1147–1154.
49. Roth MB, Nystul T: **Buying time in suspended animation.** *Sci Am* 2005, **292**:48–55.
50. Leslie M: **Nothing rotten about hydrogen sulfide's medical promise.** *Science* 2008, **320**:1155–1157.
51. Aslami H, Schultz MJ, Juffermans NP: **Hydrogen sulfide: a hot molecule.** *Anesthesiology* 2011, **115**:921–922.

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