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# Retino-protective effect of *Bucida buceras* against oxidative stress induced by H<sub>2</sub>O<sub>2</sub> in human retinal pigment epithelial cells line

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## Abstract

**Background:** Reactive Oxygen Species (ROS) impair the physiological functions of Retinal Pigment Epithelial (RPE) cells, which are known as one major cause of age-related macular degeneration and retinopathy diseases. The purpose of this study is to explore the cytoprotective effects of the antioxidant *Bucida buceras* extract in co-treatment with hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) delivery as a single addition or with continuous generation using glucose oxidase (GOx) in ARPE-19 cell cultures. The mechanism of *Bucida buceras* extract is believed to be associated with their antioxidant capacity to protect cells against oxidative stress.

**Methods:** A comparative oxidative stress H<sub>2</sub>O<sub>2</sub>-induced was performed by addition and enzymatic generation using glucose oxidase on human retinal pigment epithelial cells line. H<sub>2</sub>O<sub>2</sub>-induced injury was measured by toxic effects (cell death and apoptotic pathway) and intracellular redox status: glutathione (GSH), antioxidant enzymes (catalase and glutathione peroxidase) and reducing power (FRAP). The retino-protective effect of co-treatment with *Bucida buceras* extract on H<sub>2</sub>O<sub>2</sub>-induced human RPE cell injury was investigated by cell death (MTT assay) and oxidative stress biomarkers (H<sub>2</sub>O<sub>2</sub>, GSH, CAT, GPx and FRAP).

**Results:** *Bucida buceras* L. extract is believed to be associated with the ability to prevent cellular oxidative stress. When added as a pulse, H<sub>2</sub>O<sub>2</sub> is rapidly depleted and the cytotoxicity analyses show that cells can tolerate short exposure to high peroxide doses delivered as a pulse but are susceptible to lower chronic doses. Co-treatment with *Bucida buceras* was able to protect the cells against H<sub>2</sub>O<sub>2</sub>-induced injury. In addition to preventing cell death treatment with antioxidant plant could also reverse the significant decrease in GSH level, catalase activity and reducing power caused by H<sub>2</sub>O<sub>2</sub>.

**Conclusion:** These findings suggest that *Bucida buceras* could protect RPE against ocular pathogenesis associated with oxidative stress induced by H<sub>2</sub>O<sub>2</sub>-delivered by addition and enzymatic generation.

**Keywords:** *Bucida buceras* L, Retinal pigment epithelial cells, Oxidative stress, Hydrogen peroxide, Cellular redox status, Free radicals

## Background

Oxidative damage is involved in the pathogenesis of a variety of chronic degenerative and neurodegenerative diseases. Increasing evidence indicates that oxidative stress plays a major role in ocular pathologies including cataract, age-related macular degeneration (ARMD), glaucoma,

and diabetic retinopathy (DR). Under normal physiological states, ocular tissues possess several intrinsic antioxidant enzymes to cope with oxidative stress formed as a consequence of normal metabolism. During ocular injuries, overproduction of reactive oxygen species (ROS) and free radicals overwhelms the intrinsic antioxidant mechanisms resulting in oxidative stress and ultimately development of a pathological condition [1–4].

There is a clear difference between ROS required for basic cellular mechanisms like cellular signaling and excessive ROS production that might cause oxidative stress

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and contribute to the pathogenesis of major diseases, including diabetes, neuro-degeneration and cancer [5]. Among the various ROS, hydrogen peroxide ( $H_2O_2$ ) is perhaps the most ubiquitous of these species, which is found at measurable levels in all animal tissues.  $H_2O_2$  is most stable and can reach molecular targets distant from its site of generation. Because  $H_2O_2$  is a small, uncharged molecule, it easily crosses cell membranes and localizes in multiple subcellular compartment [6].

The effects of  $H_2O_2$  are concentration dependent and range from physiological signaling such as cell proliferation, migration, survival, differentiation, and gene expression [7–10] to overt cell death [11, 12]. At nanomolar levels,  $H_2O_2$  is a stimulant of cell growth and proliferation, whereas micromolar levels cause transient growth arrest and induce protective adaptive alterations in gene expression [13]. At millimolar levels, and above,  $H_2O_2$  is clearly a toxic oxidant species, causing a frank oxidative stress. The different sensitivities of the cells to  $H_2O_2$  are due to cell type, the species, and the differential antioxidant defense mechanisms to counteract the damaging effects of  $H_2O_2$  concentrations.

Hydrogen peroxide treatment of cultured cells is a commonly used model to test oxidative stress susceptibility or antioxidant efficiency in cell types that are at high risk for oxidative damage in vivo, such as cells of the retinal pigment epithelium (RPE). Although the retina is a complex multilayered structure, it can be functionally divide in two parts: the neuronal retina, composed by photoreceptors (cones and rods) and their neuronal connections, is responsible for photo transduction process; the RPE and its basal lamina known as Bruch's membrane maintain the integrity between retina and choroid. The RPE is composed of a polarized monolayer of pigmented hexagonal cells (melanin), and its integrity is essential for vision. Melanin in the RPE can act against ROS and protect the neural retina. Although the more popular mammalian RPE cell lines (e.g. ARPE19, D407, RPE-J) do not readily demonstrate melanogenesis, there have been numerous reports of repigmentation in ARPE19 and adult primary RPE cells [14–16].

The RPE is located adjacent to the outer retina, where it performs functions that are essential for the photoreceptor survival. Its main functions include nutrient, ion, and water transport; uptake of circulating vitamin A, its storage as an ester, its conversion to retinol, and then its transference to the photoreceptors; elimination of waste material accumulated by photoreceptors, diurnal phagocytosis and digestion of photoreceptor outer segment tips, light absorption, protection against photo-oxidation, and secretion of essential factors for maintaining the structural integrity of the retina [2, 3, 17, 18].

The retina is a part of the central nervous system (CNS), perceiving and processing visual information. But retinal photoreceptors are highly susceptible to oxidation

process because they are exposed to a range of light intensities [19]. The RPE is at high risk of oxidative stress because it resides in an environment of high oxygen tension and is exposed to phototoxic blue light [20]. Among the reactive oxygen species to which the cells are exposed is hydrogen peroxide. As in most cells,  $H_2O_2$  is generated during normal oxygen metabolism in mitochondria. In the RPE,  $H_2O_2$  is also produced during daily phagocytosis of shed photoreceptor outer segments and is generated as a consequence of light irradiation of the pigment melanin. The retinal-pigmented epithelium is armed with a robust antioxidant system and may contain cellular defense mechanisms against ROS elevation. Glutathione and its related enzymes are part of this antioxidant defense. Moreover increased macular pigment density by lutein and zeaxanthin may reduce the exposure to blue light and subsequently reduce the photo-mediated production of reactive oxygen species [1, 2, 21, 22].

Although  $H_2O_2$  addition to cell cultures is a common model of stress induction, its concentration in the medium over the period of cell treatment is usually not determined or controlled. The kinetics of  $H_2O_2$  decomposition over short time frames has been examined in cultures of several monolayer cell types including the RPE [22, 23]. The major focus of this investigation is to evaluate the protective effect of *Bucida buceras* against  $H_2O_2$ -delivered by two methods: as a single addition pulse or by continuous enzymatic generation on retinal pigment epithelial cells (ARPE-19) using cells death and oxidative stress indicators.

*Bucida buceras* L. (Combretaceae) is widely planted for shade and ornamental use in America. Very few studies have been done with *Bucida buceras*, but it contains functional components such as flavonoids and terpenes responsible of potent cytotoxicity activity against human tumor cell lines [24, 25]. Some authors have reported that extracts of *B. buceras* have antibacterial and antifungal activities (Adonizio et al., 2006; [26–28]). Our previous phytochemical screening of *Bucida buceras* give evidence of the presence of carotenes, triterpenes/steroids, lactonic groups, phenols/tannins, amines or amino acids, flavonoids/anthocyanins, saponins and reducing compounds in different extracts of *B. buceras*. These phytochemical compounds may be associated with the strong antioxidant potential found in this plant and it will contribute to protect against ocular pathogenesis associated with oxidative stress. The effect of *Bucida buceras*, in retinal health has not been studied.

## Methods

### Material

Hydrogen peroxide, catalase (from bovine liver), glucose oxidase (from *Aspergillus niger*), 3-(4,5-dimethylthiazo-2-yl)-2,5-diphenyl tetrazolium bromide (MTT), Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum

(FBS), penicillin, streptomycin, L-glutamine, trypsin-EDTA, iron (II) ammonium sulfate hexahydrate, xylenol orange disodium salt (3,3'-bis(N,N-di(carboxymethyl)-aminomethyl)-o-cresulfone-phatein), phosphate-buffered saline (PBS), 5,5'-dithiobis(2-nitrobenzoic acid) (DTNB), 2,2-difenil-1-picrilhidrazil (DPPH), and 2,4,6-tripyridyl-s-triazine (TPTZ), were purchased from Sigma (St. Louis, MO, USA). RIPA buffer from Thermo Scientific, caspase-3-substrate (Ac-DEVD-pNA) from Santa Cruz Biotechnology (Santa Cruz, CA, USA) and caspase-3 inhibitor I (DEVD-CHO) from Calbiochem (Merck, Millipore).

### Cell cultures

The hRPE cell line, ARPE-19 (ATCC, 2302) was kindly gifted from Dr. Horacio Rilo (Arizona, University, USA) and was used during all experiments. The cells were propagated in T 25 cm<sup>2</sup> culture flasks using twice-weekly feedings of DMEM containing 10 % fetal bovine serum and antibiotics (penicillin 100 U/ml and streptomycin 0.1 mg/ml) at 37 °C in a humidified 5 % CO<sub>2</sub> atmosphere. Attached and floating cells in the experimental dishes were collected after trypsinization with 1x trypsin-EDTA, centrifuged, re-suspended in fresh medium and counted using trypan blue exclusion assay.

### Plant collection and extract preparation

*Bucida buceras* L. was collected in April-May 2013 from Sonora, Mexico. The plant material was botanically identified by Dr. Jose Cosme Guerrero, department of Agriculture and Livestock, Sonora University (UNISON). A voucher specimen (RP-2014-10) was deposited at the Herbarium of UNISON.

The leaves were collected fresh and shade dried to obtain 500 g dry sample, then was later coarsely powdered in an electric chipper shredder (Chicago electric power tools) and used for solvent extraction. For sample preparation, 500 g of dried sample were extracted twice (1000 ml for each with absolute ethanol at 25 °C for 1 week and concentrated using a rotary evaporator (Buchi R-210, Switzerland) at 37 °C and preserved at 4 °C. Dried sample of ethanol extract (200 mg) was dissolved 1 ml of dimethyl sulfoxide as a stock solution and stored at -20 °C.

### Determination of cell viability via trypan blue method

Viability and cell numbers were determined by staining cells with trypan blue dye. Cells were harvested, collected and re-suspended in 3 ml medium. Then the cells were stained with a final concentration of 0.04 % for 5 min at room temperature. Stained and unstained cells were counted using a hemacytometer in optic microscope. Unstained cells represent the viable cells, which were not permeable to trypan blue dye.

### H<sub>2</sub>O<sub>2</sub> exposure on ARPE-19 cells

In H<sub>2</sub>O<sub>2</sub> treatment experiments, the cells were plated at 5 × 10<sup>4</sup> cell/well in 96-well plates to adhere overnight before exposure to oxidant. Two treatment protocols were used: pulse delivery of a range of micro-molar and millimolar concentrations of H<sub>2</sub>O<sub>2</sub> and addition of glucose oxidase to initiate continuous enzymatic generation of the oxidant.

For pulse delivery, the culture medium was first removed and cells were fed with DMEM in presence or absence of serum (10 %) and exposed to different ranges of concentrations of H<sub>2</sub>O<sub>2</sub>, freshly prepared in medium: treatment 1-(12.5, 25, 50, 100, 200, 400, 800, 1200, 1600 μM) during 24 h with intervals of up to 30 min during the first 2 h and treatment-2 (0.01, 0.05, 0.1, 0.5, 1, 5, 10, 25 mM) in 30, 60 and 120 min. Control cells were cultured in H<sub>2</sub>O<sub>2</sub>-free medium and H<sub>2</sub>O<sub>2</sub> was also added to medium in culture wells lacking cells to determine H<sub>2</sub>O<sub>2</sub> depletion in the absence of culture monolayers.

For continuous enzymatic generation of H<sub>2</sub>O<sub>2</sub>, the cultures were first refed with fresh DMEM containing 10 % FBS (D-10). Glucose oxidase (GOx) was then added to the medium to initiate the generation of H<sub>2</sub>O<sub>2</sub> by oxidation of the glucose contained in DMEM (4.5 mg/ml D-glucose). Stock solutions of GOx were prepared by solubilizing the enzyme in 50 mM sodium acetate buffer, pH 5.1, at a concentration of 10 KU/ml and storing aliquots at -20 °C. Just before use, stock solutions were thawed, diluted, and added to the culture medium to produce final concentrations of 3–100 mU/ml. GOx was added as well to medium in culture wells lacking cells to determine H<sub>2</sub>O<sub>2</sub> depletion in the absence of culture monolayers.

After addition of H<sub>2</sub>O<sub>2</sub> (pulse delivery) or GOx (to initiate continuous H<sub>2</sub>O<sub>2</sub> generation), aliquots of culture medium were retrieved at intervals to determine H<sub>2</sub>O<sub>2</sub> levels, and cells were harvested after 24 h (treatment-1 and GOx) or 2 h (treatment-2) to perform cytotoxicity assays by the methods described below.

### Hydrogen peroxide determination

Hydrogen peroxide concentration in culture medium was determined by a modified ferrous oxidation-xylenol orange (FOX) assay reported by Gil et al. [29]. Complete FOX reagent consisted in a mix of two solutions: contained 25 mM ammonium ferrous sulfate in 3.5 M H<sub>2</sub>SO<sub>4</sub> (solution A) and 0.125 mM xylenol orange disodium salt (solution B). Just before use 1 volume of solution A was added to 100 volumes of solution B to produce complete FOX reagent. For the assay, an aliquot of medium retrieved from cultures (50 μl DMEM or D10 in the presence and absence of ARPE-19 cells) were mixed with 500 μl of complete FOX reagent.

The oxidation of Fe<sup>2+</sup> in the presence of hydroperoxides forms Fe<sup>3+</sup>, which reacts with xylenol orange to

produce a chromosphere. Samples were incubated for 30 min at room temperature. Absorbance of the supernatant was read spectrophotometrically at 560 nm against PBS as background control. 50  $\mu$ l of known concentrations of H<sub>2</sub>O<sub>2</sub> (3.125 to 100  $\mu$ M) were used as standard calibration curve and a best-fit line for the data was plotted using linear regression. Experimental H<sub>2</sub>O<sub>2</sub> concentrations were then extrapolated substituting the unknown values into the equation derived for the standard curve.

#### Assay of mitochondrial viability (MTT assay)

In the MTT assay, cell respiration, and thus cell viability, was assessed based on the mitochondrial-dependent reduction of MTT to form the colored product (purple) reported by Mossman [30]. Cytotoxicity was determined in different times after treatment of ARPE-19 cultures with H<sub>2</sub>O<sub>2</sub>, GOx or co-treated with the indicated concentrations of *Bucida buceras* ethanol extract. Afterward the medium was replaced by MTT (0.5 mg/ml in medium), and the conversion of MTT into an insoluble formazan crystals for 4 h at 37 °C were solubilized in acidified isopropanol. The absorbance was monitored at 570 nm and 655 nm in a microplate reader (Thermo Scientific Multiskan Spectrum). Cell viability was expressed as a percentage of the control value (untreated cells), which was set to 100 %. Control cells were cultured in medium supplemented with 0.1 % dimethyl sulfoxide, which was used as the solvent for the extract.

#### Preparation of cell lysates from ARPE-19 cells and protein quantification

One million cells were plated in 12-well plates and allowed to adhere overnight before hydrogen peroxide-stress. Cultures were treated with H<sub>2</sub>O<sub>2</sub> (800, 1000, 1600  $\mu$ M) in addition or GOx (12.5, 25, 50 mU/ml) alone or cells treated with H<sub>2</sub>O<sub>2</sub> followed by co-treatment with *Bucida buceras* extract (1600  $\mu$ g/ml) for 24 h. Control cells were cultured in medium in absence of H<sub>2</sub>O<sub>2</sub>. After incubation time the medium was carefully removed and washed twice with cold PBS buffer, trypsinized and centrifuged at 1500 rpm for 5 min. For making whole cell lysates, the treated and untreated cells were lysed in phosphate-buffered saline (PBS; 136.75 mM NaCl, 2.68 mM KCl, 1.47 mM KH<sub>2</sub>PO<sub>4</sub>, 8.10 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 7.4) by three freeze-thaw cycles (-77 °C/25 °C) and then disrupted by sonication. The homogenate was centrifuged at 14,000 g at 4 °C for 15 min. Protein concentration determination was performed by the Bradford method using bovine serum albumin as the standard at 595 nm [31].

#### Measurement of apoptosis/caspase-3 assay

After treatment the medium was aspirated and cells were washed once in cold PBS and afterward counted

using the trypan blue exclusion method. An amount of 1x10<sup>6</sup> cell/ml was transferred to lysis RIPA buffer. The activity of caspase-3-like protease in the lysate was measured using colorimetric caspase-3 assay kit according to the reported by Bai et al. [32]. In brief, cytosolic protein (100  $\mu$ g) was mixed with caspase-3-specific substrate acetyl-Asp-Glu-Val-Asp-p-nitronilide (final concentration, 200  $\mu$ M) and incubated at 37 °C for 90 min. The absorbance was read at 405 nm. Apoptotic cell lysates containing active caspase-3 produce a considerable activity compared to non-apoptotic cell lysates.

To confirm that substrate cleavage was due to caspase-3 activity, extracts were incubated in the presence of caspase-3-specific inhibitor acetyl-DEVD-CHO (final concentration, 20  $\mu$ M) at 37 °C, before the addition of substrate. The value (in arbitrary absorbance units) of the absorbance signal of the inhibited sample was subtracted from the no inhibited sample.

#### Antioxidant activities

##### Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging activity assay

Hydrogen peroxide scavenging activity of *Bucida buceras* ethanol extract was measured by the FOX assay [29]. H<sub>2</sub>O<sub>2</sub> (100  $\mu$ L of 1600  $\mu$ M or GOx 50 mU/ml) and 100  $\mu$ L of various concentrations (400, 800 and 1600  $\mu$ g/ml) of the extract were mixed. After 1 h, 50  $\mu$ l were mixed with 500  $\mu$ l of complete FOX reactive and incubated for 30 min at room temperature.

##### Total antioxidant capacity

Total antioxidant capacity in the lysates cells and extract were assayed using the FRAP assay of Benzie and Strain [33] with some modifications according to Iloki et al. [34]. The FRAP reactive was prepared in acetate buffer (pH 3.6), 10 mmol 2,4,6-tripyridyl-s-triazine (TPTZ) solution in 40 mmol HCL and 20 mmol iron (III) chloride solution in proportions of 10:1:1 (v/v), respectively. The FRAP reactive was prepared fresh on a daily basis and warmed to 37 °C in water bath prior to use. 5  $\mu$ l of sample diluted with 15  $\mu$ l of PBS were added to 150  $\mu$ l of FRAP reagent. The absorbance of the mixture was measured using a microplate spectrophotometer reader Thermo Scientific Multiskan Spectrum at 595 nm after 5 min. The standard curve was prepared by ascorbic acid (AA) solution, and the results were expressed as  $\mu$ MAA/mg extract.

##### DPPH-scavenging capacity

*Bucida buceras* ethanol extract was evaluated for its activity to scavenge the stable DPPH radical (0.1 mM in ethanol) according to a previously described method [35] with some modifications according to Lewis et al. [36]. The affinity of the test material to quench the DPPH free radical was evaluated according to the

equation scavenging % =  $(A_c - A_s) / A_c * 100$  %.  $A_s$  and  $A_c$  are the absorbance at 517 nm of the reaction mixture with sample and control respectively.

### Redox homeostasis

#### Cellular thiol quantification

Total intracellular thiol levels were quantified spectrophotometrically using 5,5'-dithiobis (2-nitrobenzoic acid (DTNB)), as described previously by Gil et al. [29] with glutathione reduced (GSH) as standard. Briefly, this method is based on the reaction of the glutathione present in the lysates with DTNB to generate oxidized GSH (GSSG) and 2-nitro-5-thiobenzoic acid, a yellow compound that absorbs at 412 nm.

#### Activity of antioxidant enzymes

Catalase (CAT) activity was measured by using a protocol adapted from [29]. Catalase activity was assayed adding 50  $\mu$ l of sample to 450  $\mu$ l of 50 mM phosphate buffer, pH 7.0, and 250  $\mu$ l of 50 mM  $H_2O_2$  in a final volume of 750  $\mu$ L. Absorbance decrease was measured at 240 nm in a Cary spectrophotometer for 1 min and calculations were performed using an extinction coefficient of  $0.043 \text{ mM}^{-1} \text{ cm}^{-1}$ . Catalase activity is expressed as  $\mu\text{mol of } H_2O_2 / \text{min} * \text{ml}$ . All experiments were carried out using three biological replicates and three technical replicates for each sample.

#### Assay of GSH-peroxidase activity

GSH (200  $\mu$ M) and  $H_2O_2$  (200  $\mu$ M) were incubated at 37 °C for 1 h in PBS containing various lysates of ARPE-19. The volume of the enzymatic reaction was 120  $\mu$ l. Residual peroxide was measured spectrophotometrically at 560 nm by a modified ferrous oxidation-xylenol orange (FOX) assay.

#### Microscopic imaging

Samples from ARPE-19 cells treated or untreated with  $H_2O_2$  and *Bucida buceras* were evaluated using an inverted microscope (Faga-Lab-Labomed, TCM 400) to detect structural changes. A series of structural images were collected with a bottom-mount digital camera for the analysis of treatments.

#### Statistical analysis

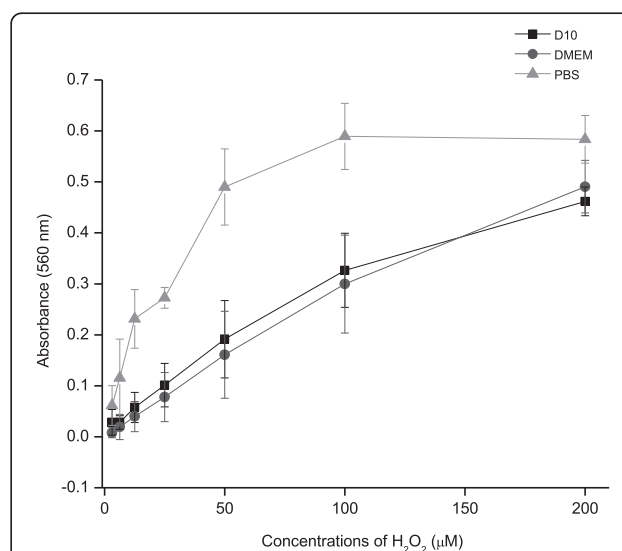
Data is expressed as the mean  $\pm$  standard deviation (SD). Significance between experimental groups was determined by GLM-ANOVA followed by post hoc Tukey test using NCSS 2007 software. Values of  $p \leq 0.05$  were considered statistically significant. Figures were performed using Origin 8.0 software and the error bars represent the SD.

## Results

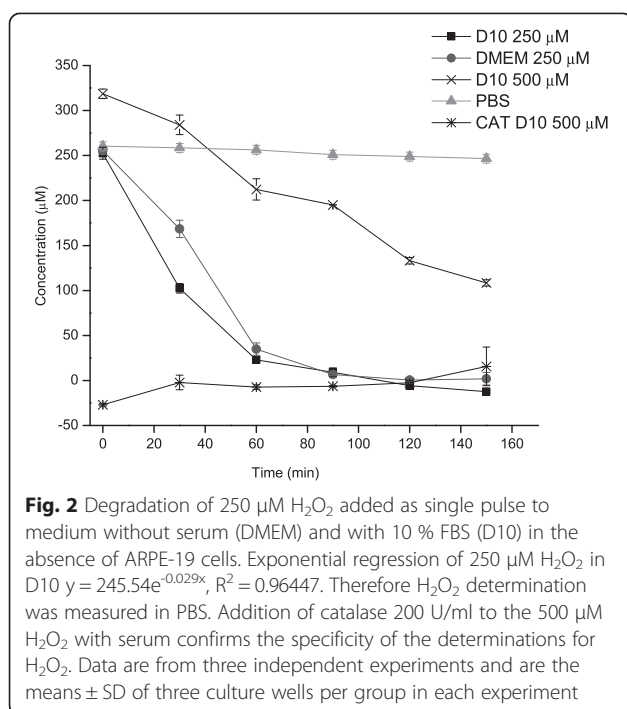
### Pulse delivery of $H_2O_2$

Preliminary experiments were conducted to confirm that the presence of serum did not affect  $H_2O_2$  measurement. As shown (Fig. 1), hydrogen peroxide determinations did not differ for a range of concentrations of  $H_2O_2$  (10–200  $\mu$ M) added to DMEM either lacking serum or containing 10 % FBS. However previous  $H_2O_2$  concentrations in PBS are nearly at an order of magnitude higher than that in medium. It is not clear which component(s) of DMEM is/are responsible for the slow but measurable decomposition of  $H_2O_2$  in the medium.

Exogenously added of  $H_2O_2$  has a short half-life as a result of its rapid degradation in culture medium. For monitoring the decay of the  $H_2O_2$  bolus in our experiments and to know how long and to what extent  $H_2O_2$  levels remain elevated after a bolus treatment, the kinetics of  $H_2O_2$  degradation was determined in the presence and absence of cells. In the absence of cells, the initial concentration is not sustained in the medium (Fig. 2) and therefore the concentration of the agent and the time of exposure are not well controlled. As illustrated for an initial nominal concentration of 250  $\mu$ M, the depletion rate is almost similar over 2 h at 37 °C in the absence and presence of serum. Although, in the presence of serum (D10)  $H_2O_2$  concentration undergoes a more rapid exponential depletion, the half time of depletion is 30 min with 10 % FBS while in DMEM (without serum) it is 45 min. The rapid degradation of  $H_2O_2$  in medium



**Fig. 1** Calibration curves for  $H_2O_2$  levels in the range 10–200  $\mu$ M using ferrous-xylenol orange with 10 % FBS (D10; linear regression  $y = 0.0022x + 0.0149$ ,  $R^2 = 0.98616$  and without FBS (DMEM linear regression:  $y = 0.0025x + 0.0157$ ,  $R^2 = 0.98643$ ). Therefore  $H_2O_2$ -determination was measured in PBS. Data are from three independent experiments and are the means  $\pm$  SD of three culture wells per group in each experiment



with serum dependent on its own serum antioxidant joined to its amino acids present in the medium.  $\text{H}_2\text{O}_2$  concentration in PBS declines slowly with time.

The rapid elimination of  $\text{H}_2\text{O}_2$  in culture medium is dependent of initial nominal concentrations. While 250  $\mu\text{M}$ , was totally diminished in the medium in the first our; an increases in  $\text{H}_2\text{O}_2$  concentration (500  $\mu\text{M}$ ) delay the rapid degradation but during the time was eliminated. Because of oxidation of ferrous to ferric ions in biological samples could also result from the interaction of ferrous ions with organic hydroperoxides,  $\text{H}_2\text{O}_2$  specificity was confirmed by measuring concentrations of 500  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in D10 without and with catalase at a final concentration of 200 U/ml in the assay mix (Fig. 2). Catalase (CAT) broke down  $\text{H}_2\text{O}_2$  into  $\text{H}_2\text{O}$  and  $\text{O}_2$ . Catalase is likely to be of particular relevance when cells are exposed to high  $\text{H}_2\text{O}_2$  concentrations, on account of its essentially non-saturable first-order kinetics.

The relationship between  $\text{H}_2\text{O}_2$  concentration and incubation time shows an exponential decline (Fig. 3). Exponential decay was more rapid in medium with serum in presence of ARPE-19-cells ( $k_d = -0.065 \text{ min}^{-1}$ ), similar decay rate constants ( $k_d$ ) were found in medium with 10 % of serum and DMEM without serum in presence of ARPE-19 cells ( $-0.041$  and  $-0.037 \text{ min}^{-1}$  respectively) (Fig. 3a). The near perfect exponential decay (shown for 400  $\mu\text{M}$  bolus) in Fig. 3a demonstrates that  $\text{H}_2\text{O}_2$  degradation closely follows first-order kinetics with dependence on substrate concentration. Fitting of the measured concentrations to the function.

$$[\text{H}_2\text{O}_2] = [\text{H}_2\text{O}_2]_{\text{ini}} \times e^{-kt}$$

Where  $[\text{H}_2\text{O}_2]_{\text{ini}}$  is the initial concentration and  $k$  ( $\text{min}^{-1}$ ) the first order rate constant. The rate constant is used to calculate elimination half-life ( $t_{1/2}$ ) and the time of 99 %  $\text{H}_2\text{O}_2$  disappears from the supernatant ( $t$ ).

$$t_{1/2} = \ln 2/k, t_{1/2} = \ln 2/0.065, t_{1/2} = 0.693/0.065 \quad t_{1/2} = 10.67 \text{ min}$$

$$t = \ln(0.01)/-k; t = \ln(0.01)/-0.065; t = 70.85 \text{ min}$$

For ARPE-19 cells grown in DMEM with 10 % FBS using the specific elimination rate constant,  $k_d = -0.065 \text{ min}^{-1}$  derived from the relationship between peroxide concentration and the time (Fig. 3), it can be estimated that the half-life elimination ( $t_{1/2}$ ) was 10.67 min and the elimination total time was 70.85 min. By this time the concentration of  $\text{H}_2\text{O}_2$  in culture medium had diminished more than 10-fold.

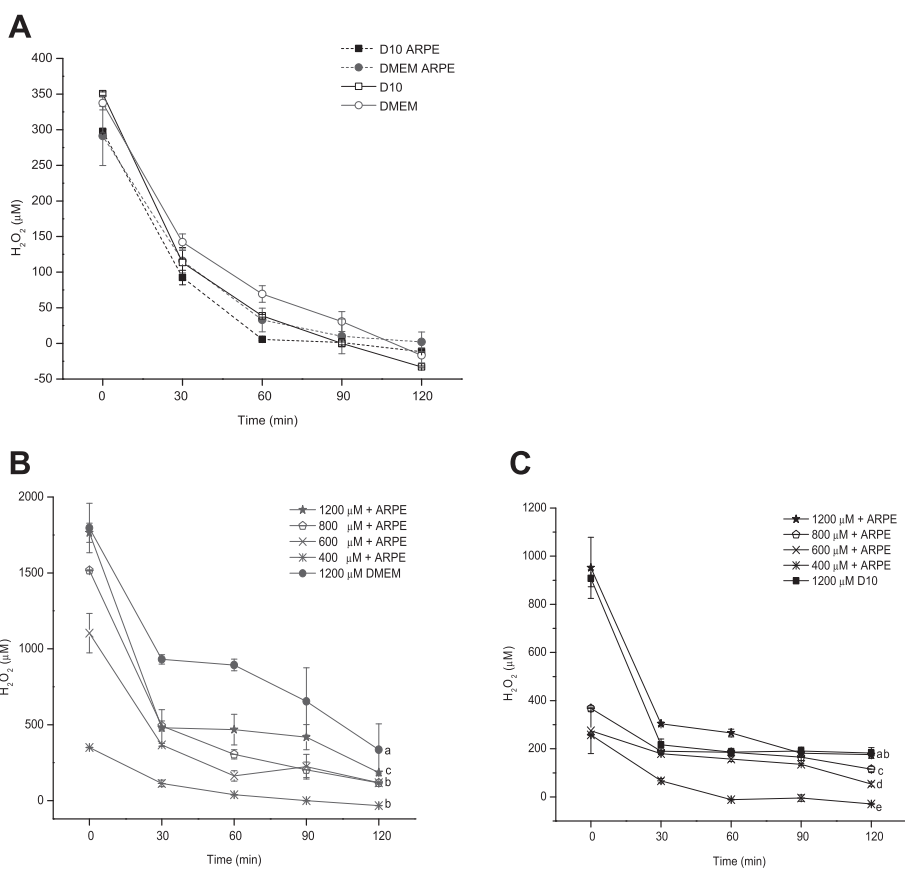
ARPE-19 cells were exposed to a variety of  $\text{H}_2\text{O}_2$  nominal concentrations (400–1200  $\mu\text{M}$ ), by delivering oxidant to culture as a single addition (pulse) with serum-free culture medium (Fig. 3b) and with 10 % of serum (Fig. 3c). The concentration-effect relationship shows that the response of cultured cells is probably determined by both the concentration of the agent and the time of exposure (Fig. 3).

A rapid exponential depletion of  $\text{H}_2\text{O}_2$  from serum-free medium exposed to RPE culture was obtained (Fig. 3b), although elimination of  $\text{H}_2\text{O}_2$  in ARPE-19 cell culture is much more rapid in the presence of serum (Fig. 3c). The concentration of  $\text{H}_2\text{O}_2$  in the medium of ARPE-19 cell cultures with serum starts to decrease immediately after administration of the peroxide. At least for the first 30 min of incubation  $\text{H}_2\text{O}_2$  disappears from the culture. By 2 h the concentration of  $\text{H}_2\text{O}_2$  in culture medium had diminished dramatically, but residual high  $\text{H}_2\text{O}_2$  concentrations remain in serum-free media.

An initial exponential decline of the  $\text{H}_2\text{O}_2$  concentration in the culture medium was observed with  $\text{H}_2\text{O}_2$  concentrations between 100 and 1600  $\mu\text{M}$ . In general, concentrations  $\leq 100 \mu\text{M}$   $\text{H}_2\text{O}_2$  were completely eliminated during the first 30 min after administration. At initial  $\text{H}_2\text{O}_2$  concentrations of  $\geq 200 \mu\text{M}$  after the initial rapid phase, the elimination became slower and the concentration of  $\text{H}_2\text{O}_2$  leveled off. Regardless of the kinetics, from the pragmatic point of view, the significant observation is that  $\text{H}_2\text{O}_2$  is not sustained at the added concentration when the agent is delivered as a pulse.

#### Influence of cell concentration on the elimination $\text{H}_2\text{O}_2$ concentrations

ARPE-19 cultures with different cell number (2, 4,  $8 \times 10^5$  cells/well) were exposed to various nominal  $\text{H}_2\text{O}_2$  concentrations (400, 800, 1600  $\mu\text{M}$ ) in 1 ml /well to evaluate the



**Fig. 3** Dissipation of 400 μM nominal concentration of H<sub>2</sub>O<sub>2</sub> (a), different nominal concentrations 400, 600, 800 and 1200 μM with serum-free medium (b) and with 10 % FBS (c) from the culture medium in presence or absence of ARPE-19 cells (50, 000 cell/well) with or without serum. Decay rate constants were  $k_d = -0.065, -0.041, -0.037, -0.026 \text{ min}^{-1}$  for D10 + ARPE-19 cells, D10, DMEM + ARPE-19 cells and DMEM respectively. Data represent means of triplicate measurements of 3 independent experiments. 1200 μM of H<sub>2</sub>O<sub>2</sub> in absence of ARPE-19 cells was determined in parallel culture

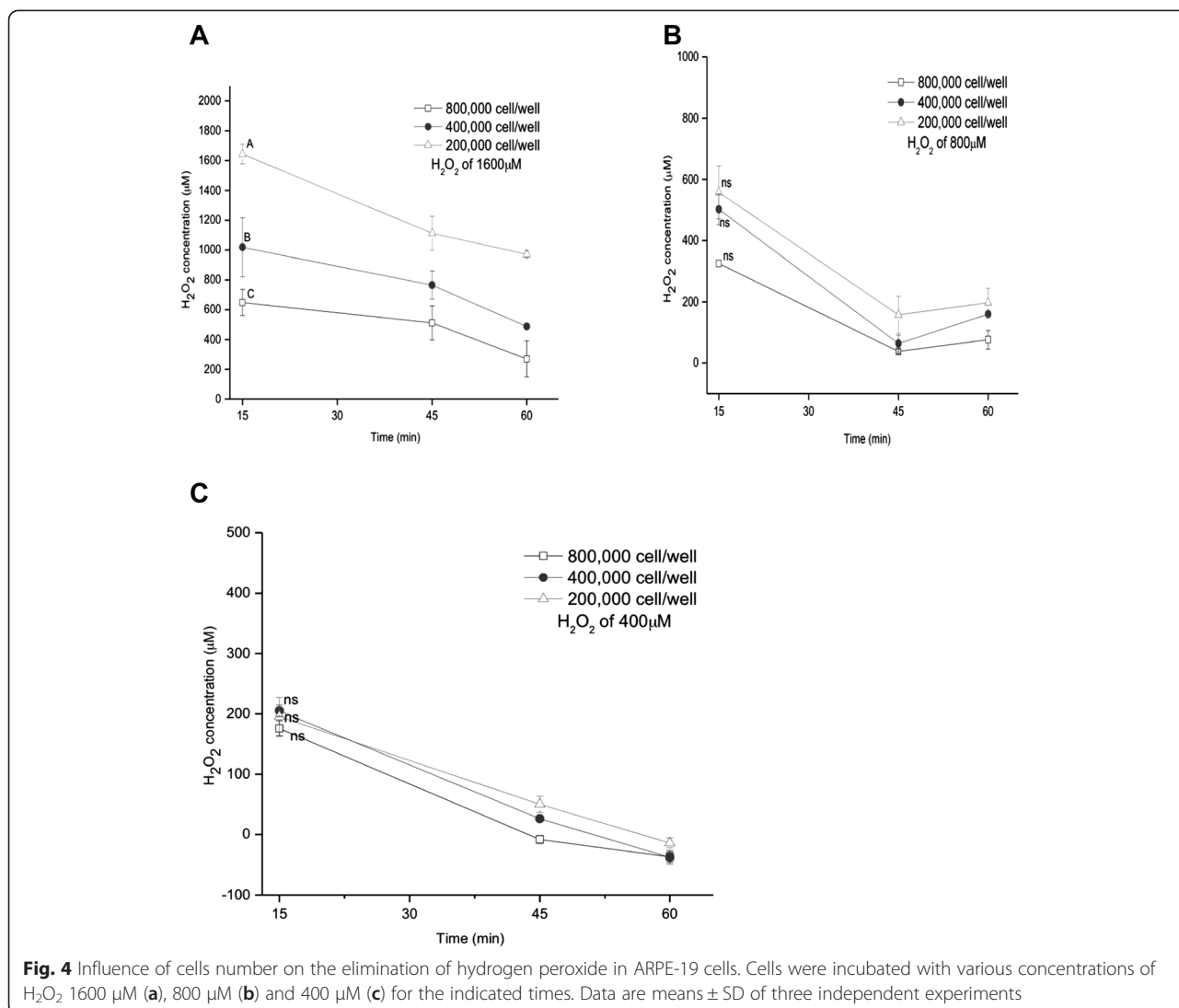
elimination of H<sub>2</sub>O<sub>2</sub> from culture medium and the cytotoxic action. Figure 4 shows that increasing the cell concentration accelerate the peroxide depletion. Raising the cell concentration by increasing the cell number resulted in a shift of the nominal concentration-effect curves for the elimination of H<sub>2</sub>O<sub>2</sub> toward higher concentrations (Fig. 4a, b). The elimination of H<sub>2</sub>O<sub>2</sub> changes with variation of cell number, although this effect is almost independent of the cell number when the dose of H<sub>2</sub>O<sub>2</sub> is low (Fig. 4c).

**Cytotoxicity in addition of H<sub>2</sub>O<sub>2</sub>**

It is well established that H<sub>2</sub>O<sub>2</sub> added to the medium of ARPE-19 cells as a single pulse produces an oxidant-dependent cytotoxicity that can be detected by MTT assay 24 h after treatment (Fig. 5). Because H<sub>2</sub>O<sub>2</sub> is largely depleted from culture medium within 2 h (Fig. 3), one would expect similar cytotoxicity whether the medium was replaced with fresh medium at 4 h after oxidant addition or remained unchanged until the time of assay.

The figure reveals that H<sub>2</sub>O<sub>2</sub> concentrations ≤100 μM are not cytotoxic at 24 h when H<sub>2</sub>O<sub>2</sub> is delivered as a pulse, since the oxidant that is depleted fairly rapidly in these concentrations in the culture medium. Additionally ARPE-19 cell is not very sensitive to oxidative stress. Although 200 μM had little effect, but cytotoxicity was substantial at higher concentrations. The minimal concentration necessary to elicit 50 % of the cytotoxic effect (IC<sub>50</sub>) was calculated to be 948 μM at 24 h similar to 964 μM whether the medium was replaced with fresh medium at 4 h after oxidant addition. As shown, similar outcomes were in fact obtained (Fig. 5). The manifestation of the incipient cytotoxicity of H<sub>2</sub>O<sub>2</sub> at 4 h means that cytotoxicity of H<sub>2</sub>O<sub>2</sub> takes several hours longer than the exposure to H<sub>2</sub>O<sub>2</sub>

In the case of H<sub>2</sub>O<sub>2</sub>, incubation time is not equivalent to exposure time when oxidant is delivery as single addition. The clearance measurements reveal that H<sub>2</sub>O<sub>2</sub> that is rapidly eliminated obviously the cell death is not immediate and the window between initial exposure and



cell death is relevant because it is in this interval that antioxidants could function to increase the fraction of surviving cells.

Additionally the ARPE-19 cells were exposed to a short exposure time (30, 60 and 120 min) with H<sub>2</sub>O<sub>2</sub> with or without serum to find the cytotoxic effect (Fig. 6). The amount of H<sub>2</sub>O<sub>2</sub> that has to be added to ARPE-19 cell cultures to produce 50 % cytotoxicity increased about 10-fold by shortening the incubation time. Exposure to H<sub>2</sub>O<sub>2</sub> concentrations ≤ 0.5 mM showed little effect on both culture cells, however, 5 mM H<sub>2</sub>O<sub>2</sub> exposure by 60 and 120 min resulted in a significant loss of cell viability, with a 50 % reduction in serum-free culture compared to 10 mM for the ARPE-19 cells containing serum in the same short periods. Increasing exposure to 25 mM resulted in an 80 % loss of viability in ARPE-19 cells in both cultures types at 120 min. Short-term exposure to high H<sub>2</sub>O<sub>2</sub> levels induced marked toxicity, suggesting that

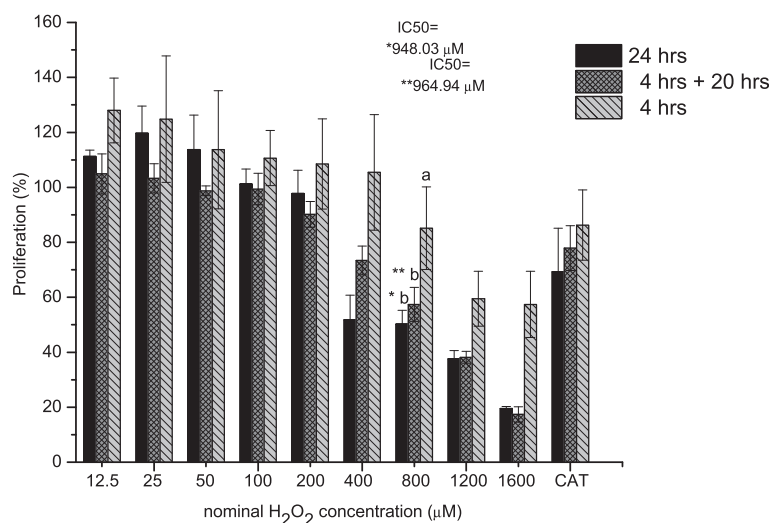
under these experimental conditions necrosis was the predominant mode of cell death.

The IC<sub>50</sub> value revealed that the cytotoxic potency in serum-free culture medium was increased nearly two-fold over the ARPE-19 culture grown in 10 % FBS (Fig. 6). This high sensitivity to oxidation was attributed to serum starvation-induced apoptosis owing to the lack of serum antioxidants. The duration of incubation with H<sub>2</sub>O<sub>2</sub> is important for the determination of H<sub>2</sub>O<sub>2</sub> concentration required to induce cellular death.

#### Continuous enzymatic generation of H<sub>2</sub>O<sub>2</sub>

Because H<sub>2</sub>O<sub>2</sub> is labile in culture medium, sustained exposure to a given concentration is difficult to achieve when the agent is delivered in a single pulse. An alternative method for sustained treatment of cultures is to continuously generate the product from glucose medium using GOx. Glucose oxidase catalyzes the direct two-





**Fig. 5** Cytotoxic action of hydrogen peroxide in ARPE-19 cells. A range of concentrations of H<sub>2</sub>O<sub>2</sub> from 12.5 to 1600 μM was added to serum-containing DMEM. Cytotoxicity was estimated 24 h after addition by the MTT assay in cultures in which the medium was not replaced (solid bars) or was replaced at 4 h with fresh H<sub>2</sub>O<sub>2</sub>-free DMEM (hatched bars) or was measured at 4 h (gray-hatched bars). Addition of catalase 200 U/ml to the 1600 μM H<sub>2</sub>O<sub>2</sub> as CAT control cells. \* IC<sub>50</sub> in 24 h \*\* IC<sub>50</sub> 4 h with replace (4 h + 20 h). Data expressed as a percentage of the untreated control, are from three independent experiments and are the means ± SD

electron reduction of oxygen to H<sub>2</sub>O<sub>2</sub>, using reducing equivalents from the oxidation of glucose. This approach has been used for short-term treatment of cultured cells including, ARPE-19 cells [30].

The addition of GOx to culture medium (DMEM) produces a linear rate of accumulation of H<sub>2</sub>O<sub>2</sub> as a function of enzyme concentration during the first hour of incubation in both, with the absence and the presence of ARPE-19 cells (Fig. 7), although the rate of accumulation in the absence of cells is approximately twice as great as when cells are present. The generation of H<sub>2</sub>O<sub>2</sub> by GOx can be assuming pseudo-zero order kinetics (i.e., assuming an unlimited supply of Glc and O<sub>2</sub>).

In the absence of cells, H<sub>2</sub>O<sub>2</sub> concentration continues to increase over 48 h at a higher rate than during the first hours, perhaps owing to the partial H<sub>2</sub>O<sub>2</sub> depletion by medium that overcame by hydrogen peroxide accumulation for glucose oxidase (Fig. 8a). In the presence of cells the concentration of H<sub>2</sub>O<sub>2</sub> in the culture medium over time after GOx addition is strikingly different in the absence of cells, and the pattern differs with enzyme concentration (Fig. 8b).

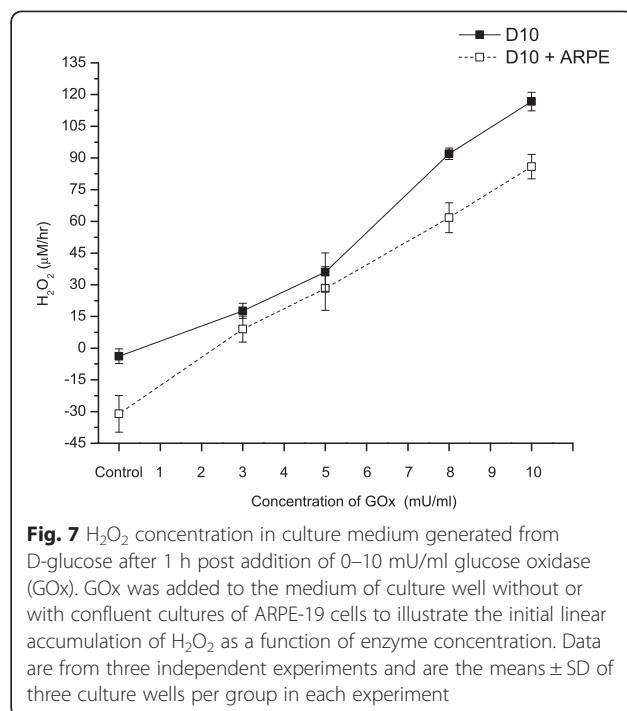
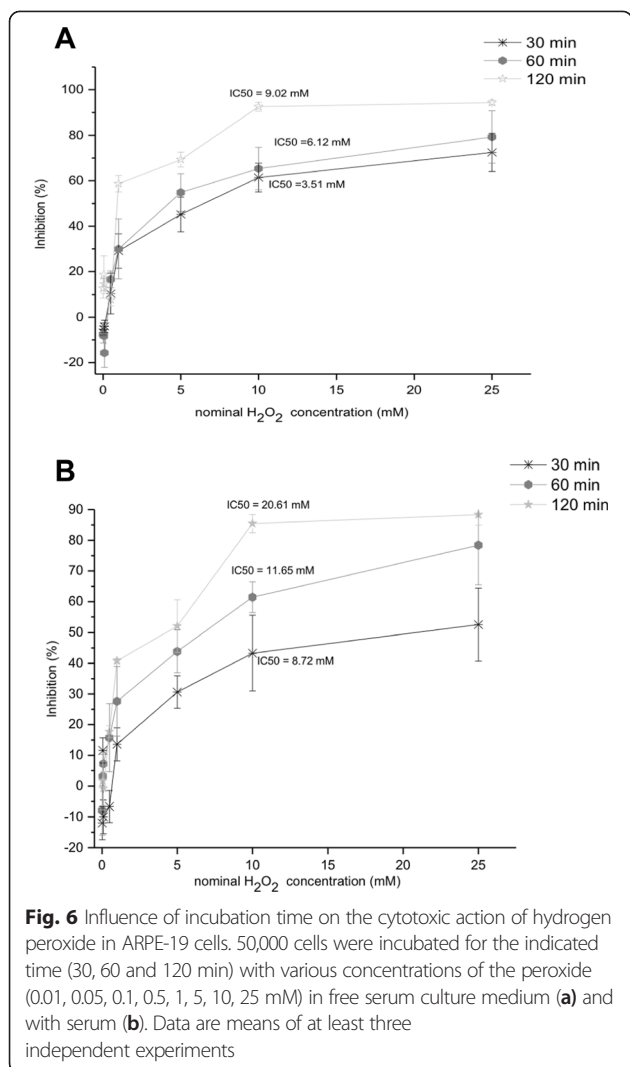
With a lower amount of GOx (5 mU/ml) H<sub>2</sub>O<sub>2</sub> concentration in the medium is nearly stationary in the presence of cells during 48 h. The production rates and decomposition of H<sub>2</sub>O<sub>2</sub> are approximately equal. At middle GOx amounts (8 or 10 mU/ml), H<sub>2</sub>O<sub>2</sub> concentration exhibits a complex dynamic. The concentration continues to rise after the first hour of enzyme addition, peaks at approximately 6 h, and then decreases thereafter. Using 10 mU/ml GOx delivery to the ARPE-19

cultures a maximum concentration of 60 μM is observed at 6 h and this declined to 0 μM by 48 h. With higher concentration (25 mU/ml) the rate of H<sub>2</sub>O<sub>2</sub> production continues to increase over 48 h, reaching a maximum value of 280 μM much lower than the seen in the absence of cells (Fig. 8b). These changes in medium content of H<sub>2</sub>O<sub>2</sub> are related to the cytotoxic response of ARPE-19 cells.

#### Cytotoxicity in enzymatic generation of H<sub>2</sub>O<sub>2</sub>

As for delivery of H<sub>2</sub>O<sub>2</sub> in a single pulse (Fig. 5), GOx addition to culture medium also produces a dose-dependent cytotoxic response in confluent ARPE-19 cultures quantified by the MTT assay at different times (Fig. 9). Low amounts of added enzyme (3 or 5 mU/ml) had little effect, but cytotoxicity was substantial at higher concentrations. This observation is noteworthy because the peak concentrations of H<sub>2</sub>O<sub>2</sub>, achieved at 6 h after addition of middle concentrations enzyme (Fig. 8b), were relatively low compared to concentrations that produced cytotoxicity when H<sub>2</sub>O<sub>2</sub> was added in a single pulse (Fig. 5).

Using 19.5 mU/ml GOx, the concentration of 228.6 μM of H<sub>2</sub>O<sub>2</sub> produced by this amount of enzyme reduced MTT in 50 % at 48 h, while 31.1 mU/ml achieved 486.04 μM of H<sub>2</sub>O<sub>2</sub> with almost 50 % of inhibition at 24 h (Fig. 9), yet 900–1000 μM H<sub>2</sub>O<sub>2</sub> was required to produce comparable MTT reductions when the oxidant was delivered as a pulse (Fig. 5). Clearly, however, the dynamics of oxidant exposure to cells differ under both conditions. When delivered as a pulse, H<sub>2</sub>O<sub>2</sub> is depleted fairly rapidly,



whereas the GOx-treated cells were exposed to a more sustained moderate concentration over the full 48-h incubation time.

**Oxidative stress index**

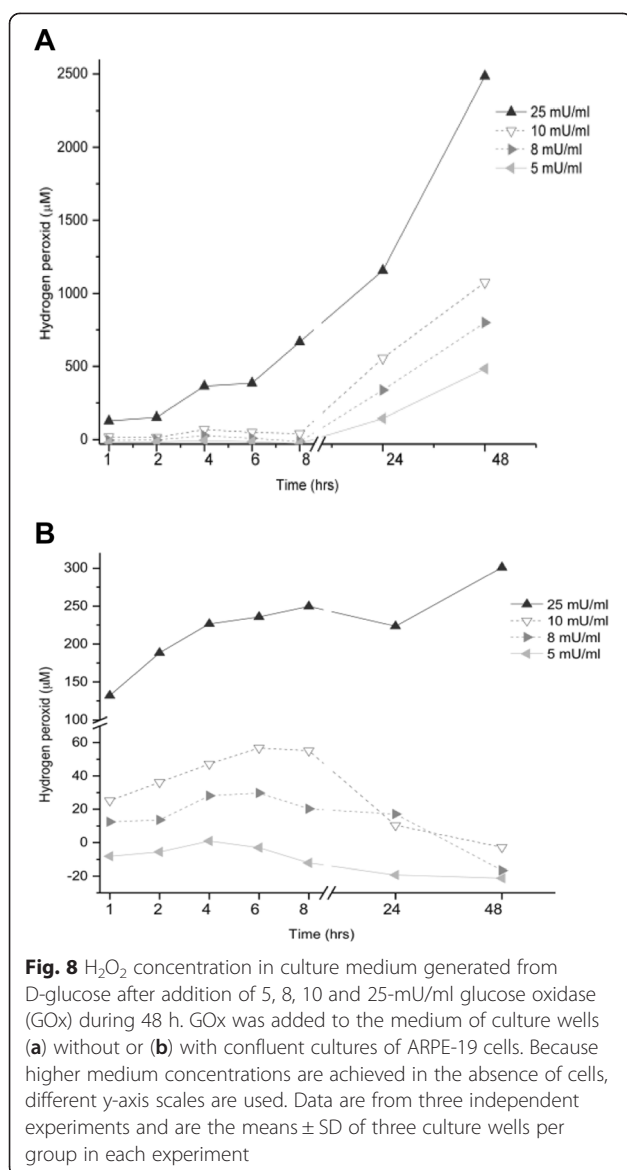
**Antioxidant activity of *Bucida buceras* extract**

The levels of oxidant and antioxidant capacity were simultaneously measured to assess oxidative stress and determine the antioxidant power in the control cells and cells exposed to H<sub>2</sub>O<sub>2</sub> with or without *Bucida buceras* ethanol extract. The hydrogen peroxide scavenging activity of extract was evaluated previously. The concentration of *Bucida buceras* achieving a 50 % reduction in 1600 µM of H<sub>2</sub>O<sub>2</sub> (pulse) and GOx 50 mU/ml (enzymatic) over 1 h is shown in Fig. 10a–b. From the results, ethanol extract showed concentration dependent activity and the H<sub>2</sub>O<sub>2</sub> scavenging effect was higher in system of pulse delivery (IC<sub>50</sub> = 1356.29 µg/ml) compared to a continuous generation (IC<sub>50</sub> = 2050.09 µg/ml).

To evaluate the antioxidant activity, we started by investigating its DPPH-scavenging action. The DPPH stable free radical method is an easy, rapid, and sensitive way to survey the antioxidant activity of compounds or extracts. Figure 10c demonstrates that DPPH-scavenging potentials increased as the concentrations of *Bucida buceras* extract increased. The IC<sub>50</sub> value 26.14 µg/ml was obtained through extrapolation from linear regression analysis and denoted the concentration of sample required to scavenge 50 % of DPPH radical. This result indicates the considerable antioxidant effect of *Bucida buceras* extract and can be related with phenolic compounds and carotenes present in this plant. Standards such as ascorbic acid, catechin and rutin had IC<sub>50</sub> values of 7.88, 7.24 y 13.6 µg/ml respectively.

The ferric ion reducing antioxidant power (FRAP) of *Bucida buceras* extract was estimated from its ability to reduce TPTZ-Fe (III) to TPTZ-Fe (II). The intensity of blue color is a significant indicator of potential antioxidant activity. The reducing power of extract correlated well with increased concentrations (Fig. 10d). The value of reducing power was 1327.56 µM AA/mg, which suggests that the plant extract has a significant antioxidant effect. Also it may be useful for reducing free radicals, which can retard the free radical chain reaction in the propagation of the oxidation mechanism.

It is well established that the detoxification of H<sub>2</sub>O<sub>2</sub> can be achieved through non-enzymatic (glutathione) and enzymatic ways; two different enzymatic systems, namely, catalase (CAT) and glutathione peroxidase



(GPx). In this study glutathione, CAT and GPx were measured.

#### Evaluation of the intracellular glutathione levels

To determine whether exposure to oxidant agents (single high dose or enzymatic generation) influenced intracellular levels of reduced glutathione, and the impact of *Bucida buceras* extract in this regard, the glutathione levels were monitored. GSH, a thiol-containing tripeptide, is the major intracellular antioxidant but it can also be the major substrate of peroxidases reducing hydroperoxides. The sulfhydryl group (SH) serves as an electron donor and reacts with ROS (H<sub>2</sub>O<sub>2</sub>, OH, O<sub>2</sub>) maintaining a reducing cellular environment.

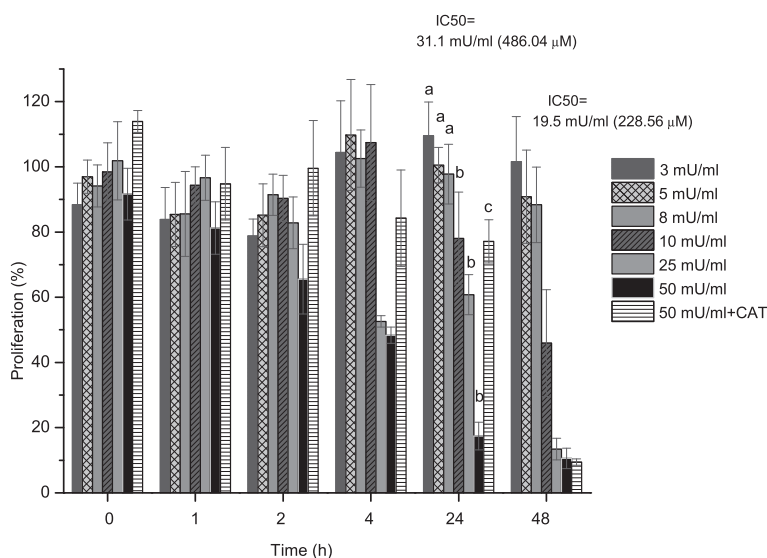
The results revealed that cells treated with GOx had the lowest amount of GSH, i.e., 40 to 80 µg/ml of SH

content, while the cells exposed to H<sub>2</sub>O<sub>2</sub> addition significantly increased the level (60–100 µg/ml) compared to GOx cells (Fig. 11a). As can be seen, following H<sub>2</sub>O<sub>2</sub> treatment, the glutathione levels decreased in a dose-dependent manner in ARPE-19 cells, reflecting the antioxidant protection by GSH against H<sub>2</sub>O<sub>2</sub>-induced oxidative stress. The results showed that moderate but sustained H<sub>2</sub>O<sub>2</sub> concentrations (GOx) and pulse (1600 µM H<sub>2</sub>O<sub>2</sub>) deplete the intracellular reduced glutathione content almost entirely ( $p < 0.05$ ). However, lower concentrations of added H<sub>2</sub>O<sub>2</sub> (800 and 1000 µM) achieved only a minor decrease in reduced glutathione.

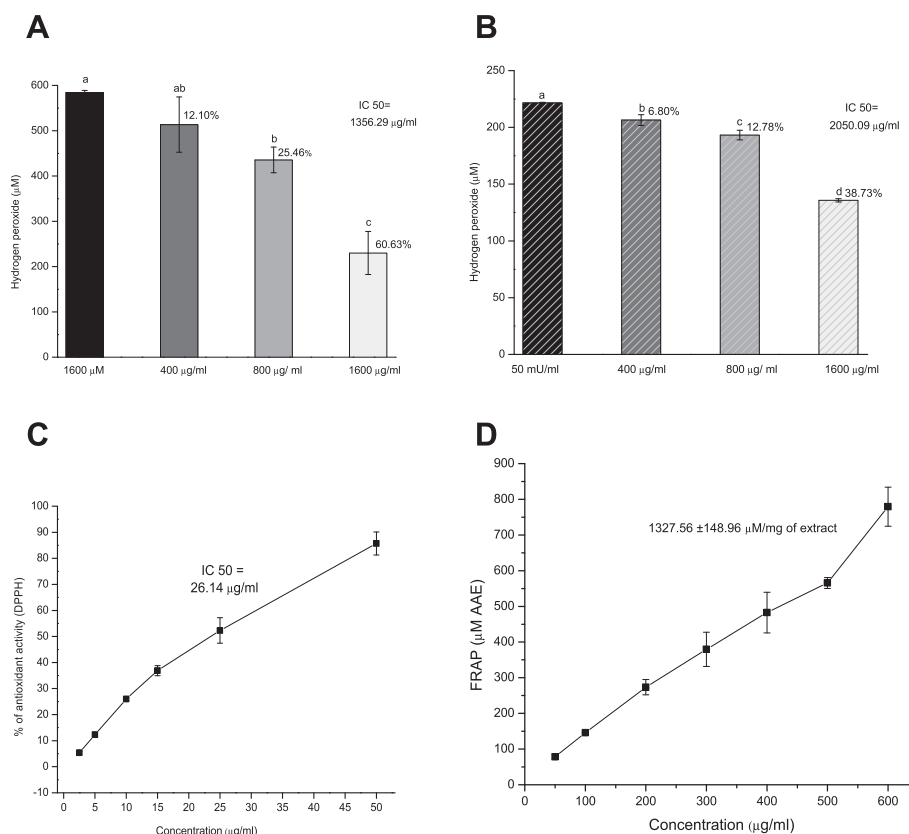
GSH levels were significantly higher ( $p < 0.05$ ) in the ARPE-19 cells exposed to H<sub>2</sub>O<sub>2</sub> with *Bucida buceras* extract and showed a 1.5 to 2-fold increased levels of GSH over the treated H<sub>2</sub>O<sub>2</sub> cells (Fig. 11a). The increase of glutathione in cells treated with H<sub>2</sub>O<sub>2</sub> in pulse in presence of antioxidant plant was comparable to that in the untreated control cells, indicating the possible involvement of GSH in the cytoprotection of *Bucida buceras*. Catalase inhibited the release of H<sub>2</sub>O<sub>2</sub> from ARPE-19 cells (GOx-50 mU/ml) and the GSH increased 1.5-fold compared to cells treated with GOx (mU/ml). The lower amount of endogenous reduced glutathione indicates the increased consumption of antioxidant to counteract the elevated level of H<sub>2</sub>O<sub>2</sub> (Fig. 11b). This behavior results mainly by the accumulation of H<sub>2</sub>O<sub>2</sub> as a function of enzymatic concentration, whereas the delivery of H<sub>2</sub>O<sub>2</sub> in a single pulse is not sustained.

The GSH redox cycle represents the most important H<sub>2</sub>O<sub>2</sub> elimination pathway in the RPE cells. In our study, we found that GOx (12.5-50 mU/ml) or addition of 1600 µM H<sub>2</sub>O<sub>2</sub> caused a dose-dependent decrease in reduced glutathione of up to 60 %, exceeding the 40 % depletion levels that render cells vulnerability to oxidative injury. *Bucida buceras* can prevent the H<sub>2</sub>O<sub>2</sub>-induced depletion of GSH, indicating that antioxidant-co-treated cells were less oxidized during oxidative stress. The recovery of GSH level may be explained by the H<sub>2</sub>O<sub>2</sub>-scavenging activity of antioxidant upon H<sub>2</sub>O<sub>2</sub>-treatment. *Bucida buceras* can increase GSH availability and as a result promote the elimination of H<sub>2</sub>O<sub>2</sub> by increasing the catalase activity.

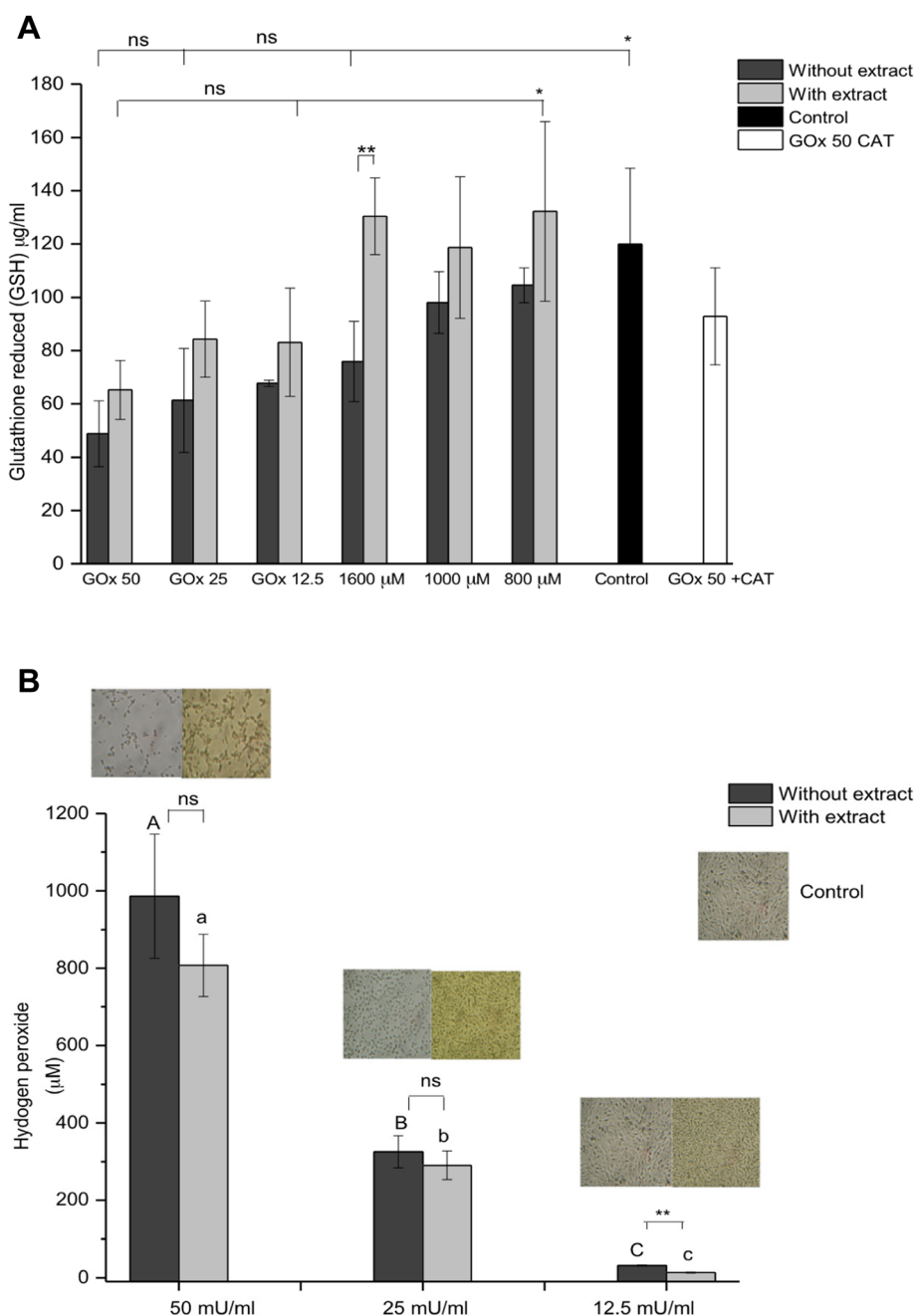
GSH serves as an antioxidant by scavenging ROS, which oxidize the cysteine moiety. Oxidation of GSH drives the formation of glutathione disulfide (GSSG); which can then be directly recycled to GSH through the enzyme glutathione disulfide reductase, a reaction requiring NADPH. The ratio of GSH to GSSG is often used as an indicator of intracellular redox status; more highly oxidized redox state is associated with differentiation and apoptosis. GSH can also act as a cofactor for GSH-utilizing antioxidant enzymes, such as GSH peroxidase, glutaredoxin and glutathione S-transferases.



**Fig. 9** Influence of glucose oxidase concentrations on the cytotoxic action of hydrogen peroxide in confluent cultures of ARPE-19 cells. The viability was determined as a percentage of the untreated controls using the MTT assay for the indicated times. IC<sub>50</sub> values have 31.1 mU/ml and 19.5 mU/ml at 24 h y 48 h respectively. Data are from at least three independent experiments and are the means ± SD of three culture wells per group in each experiment



**Fig. 10** Hydrogen peroxide scavenging of ethanol *Bucida buceras* extract (400–1600 μg/ml) at 1 h in (a) 1600 μM H<sub>2</sub>O<sub>2</sub> pulse  $y = 0.0415x - 3.2327$ ,  $R^2 = 0.99336$  (IC<sub>50</sub> = 1356.29 μg/ml) and (b) 50 mU/ml of GOx  $y = 0.0277x - 3.8591$ ,  $R^2 = 0.98872$  (IC<sub>50</sub> = 2050.09 μg/ml). (c) DPPH-scavenging activity of *Bucida buceras* (5 a 50 μg/ml),  $y = 1.6486x + 6.8934$ ,  $R^2 = 0.976$  IC<sub>50</sub> = 26.14 μg/ml. (d) The ferric ion reducing antioxidant power (FRAP) in concentrations of 100 to 800 μg/ml. Values above the bars indicate % of inhibition. Each value is expressed as mean ± SD (n = 3)

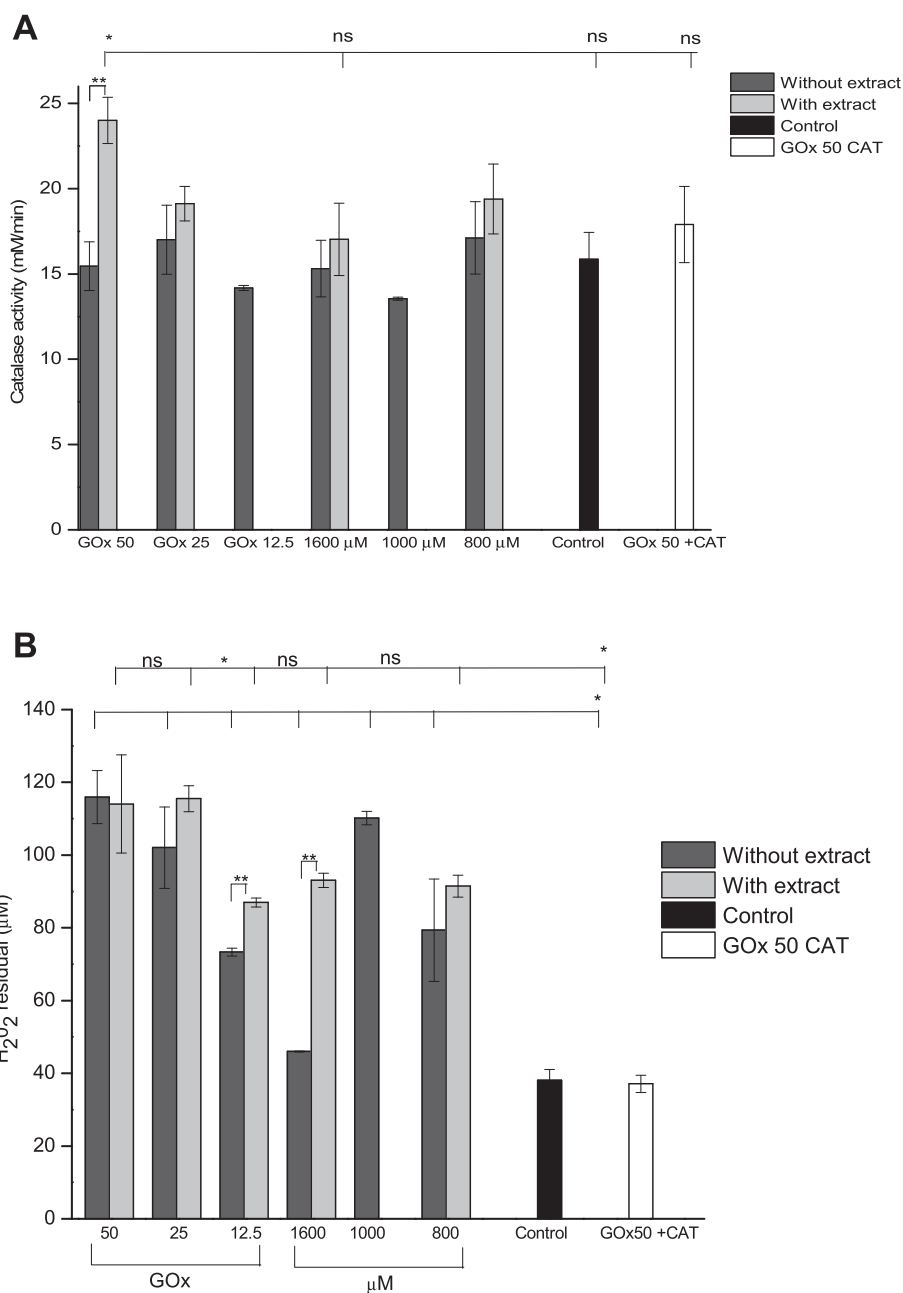


**Fig. 11** Effects of GOx and H<sub>2</sub>O<sub>2</sub> in addition on glutathione reduced (GSH) at 24 h (a). ARPE-19 cells treated with 12.5, 25 and 50 mU/ml of GOx and pulse of H<sub>2</sub>O<sub>2</sub> 800, 1000, 1600 µM and ARPE-19 cells treated with 1600 µg/ml of ethanolic extract of *Bucida buceras*, control cells (untreated with H<sub>2</sub>O<sub>2</sub>) or GOx (50 mU/ml) + 200 U/ml of catalase as CAT control; (b) The levels of H<sub>2</sub>O<sub>2</sub> in GOx at 24 h in 12.5, 25 and 50 mU/ml, the medium was analyzed using FOX reagent. Values were expressed in means ± SD in µM (n = 3 independent experiments). \*p < 0.05 significantly different from value of cells vs. H<sub>2</sub>O<sub>2</sub>-treated \*\*p < 0.05 significantly different from value of antioxidant treatment vs. H<sub>2</sub>O<sub>2</sub>-treated, n.s not significant differences

**Evaluation of catalase (CAT) and glutathione peroxidase (GPx) activities**

It is well established that most of the H<sub>2</sub>O<sub>2</sub> in cells is eliminated by the intracellular antioxidant enzymes, catalase and GPx. The effect of different treatments on

the activity of catalase and glutathione peroxidase is shown in Fig. 12. The CAT activity was slightly lower in the H<sub>2</sub>O<sub>2</sub> cells than in the control cells (p > 0.05). It was found that treatment with antioxidant extract raises the activity of catalase compared to that of the untreated



**Fig. 12** The effect of hydrogen peroxide by enzymatic generation (GOx 50, 25 and 12.5 mU/ml) and pulse (1600, 1000, 800 μM) in presence or absence of co-treatment with *Bucida buceras* ethanol extract (1600 μg/ml) on catalase activity (a) and GSH-peroxidase activity assay (b) H<sub>2</sub>O<sub>2</sub> (initially, 200 μM in PBS was incubated for 1 h, at 37 °C with different cell lysates, control GOx (50 mU/ml + catalase 200 U/ml) or control cells (untreated with H<sub>2</sub>O<sub>2</sub>). Results are expressed as the means ± SD of three independent studies (each analyzed in triplicate). \**p* < 0.05 significantly different from value of cells control (untreated) vs. H<sub>2</sub>O<sub>2</sub>-treated \*\**p* < 0.05 significantly different from value of antioxidant treatment vs. H<sub>2</sub>O<sub>2</sub>-treated, n.s not significant differences

control and H<sub>2</sub>O<sub>2</sub> treated . In cells treated with 50 mU/ml in co-treatment with *Bucida buceras* CAT activity increased (*p* < 0.05) (Fig. 12a). *Bucida buceras* elevates the activity of catalase indicating its preventive nature. Catalase is likely to be of particular relevance when cells are exposed to high H<sub>2</sub>O<sub>2</sub> concentrations, due to its essentially

non-saturable first-order kinetics. Catalase exerts a “filter” function that very efficiently prevents accumulation of cytosolic H<sub>2</sub>O<sub>2</sub> levels by decomposition into H<sub>2</sub>O and O<sub>2</sub>.

GSH-peroxidase activity was evidenced by measuring the residual H<sub>2</sub>O<sub>2</sub> during the incubation of 200 μM H<sub>2</sub>O<sub>2</sub> with different treatments or in the presence of

500  $\mu\text{M}$  GSH. We evaluated whether ARPE-19 cultures with  $\text{H}_2\text{O}_2$  or GOx had different depleting effect (Fig. 12b). The untreated cells control, GOx (50 mU/ml + catalase 200 U/ml) and GSH (500  $\mu\text{M}$ ) had a depleting effect higher than oxidant treatments. The depletion induced by GSH is because of a passive nonenzymatic scavenging effect. In enzymatic generation the residual peroxide was diminished concentration-dependent. At the highest concentration tested (50 mU/ml), there was a decrease in the GSH-peroxidase activity as indicated by the higher  $\text{H}_2\text{O}_2$  residual. In addition, the *Bucida buceras* extract had little effect on peroxidase activity. In contrast, 1600  $\mu\text{M}$  of  $\text{H}_2\text{O}_2$  added to the medium of ARPE-19 cells as a single pulse produces the maximum activity.

The higher CAT activity and the higher-level concentrations of glutathione appear to act as the first line of defense against  $\text{H}_2\text{O}_2$  formation and oxidative damage in ARPE-19 cells co-treated with *Bucida buceras*. However GPx activity is quite low between the cells and cells with extract, suggesting that GPx has a minor role in detoxifying  $\text{H}_2\text{O}_2$ . An explanation for the relative importance of catalase and not of GPx in detoxifying  $\text{H}_2\text{O}_2$  in ARPE-19 cells relates to the intracellular location and kinetic of these enzymes. Catalase enzyme is located in peroxisomes and its access to cytosolic is limited. However GPx, unlike catalase, is mainly present in cytosol and can be first saturated by  $\text{H}_2\text{O}_2$ .

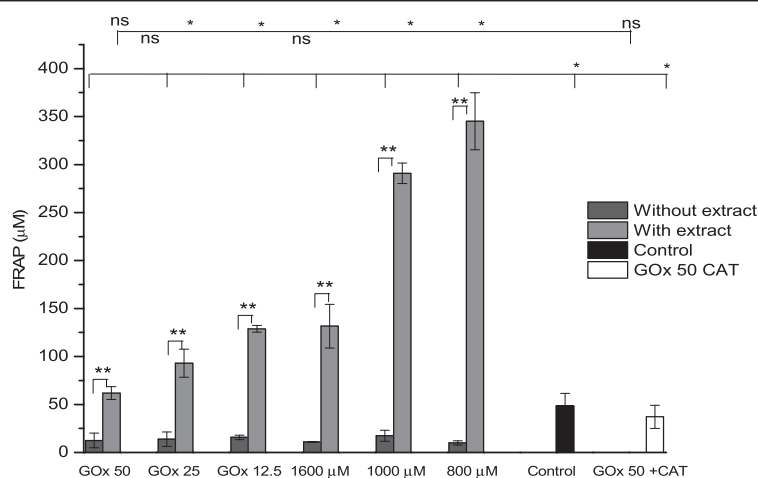
Antioxidant capacity by measuring the ferric reducing antioxidant power decreased after treatment with  $\text{H}_2\text{O}_2$  addition or with  $\text{H}_2\text{O}_2$ -generating GOx compared to culture control. As *Bucida buceras* has proposed to protect against oxidative damage, we first analyzed the effect of the antioxidant plus treatment  $\text{H}_2\text{O}_2$  on power

antioxidant reducing ferric (FRAP). Treatment with  $\text{H}_2\text{O}_2$  or GOx for 24 h significantly decreased ( $p < 0.05$ ) around threefold the antioxidant power in cells in the absence of extract compared to control cells (Fig. 13). Antioxidant extract treatment significantly increased ( $p < 0.05$ ) the basal antioxidant capacity from 48.5 to 345.15  $\mu\text{M}$ . Our results showed that RPE cells treated with extract exhibited a significant increase in oxidative protection in both  $\text{H}_2\text{O}_2$  treatments (enzymatic and pulse). Similarly, there was an increment in antioxidant capacity of GOx-50 mU/ml plus catalase comparable to untreated cells but lower than *Bucida buceras* treatments.

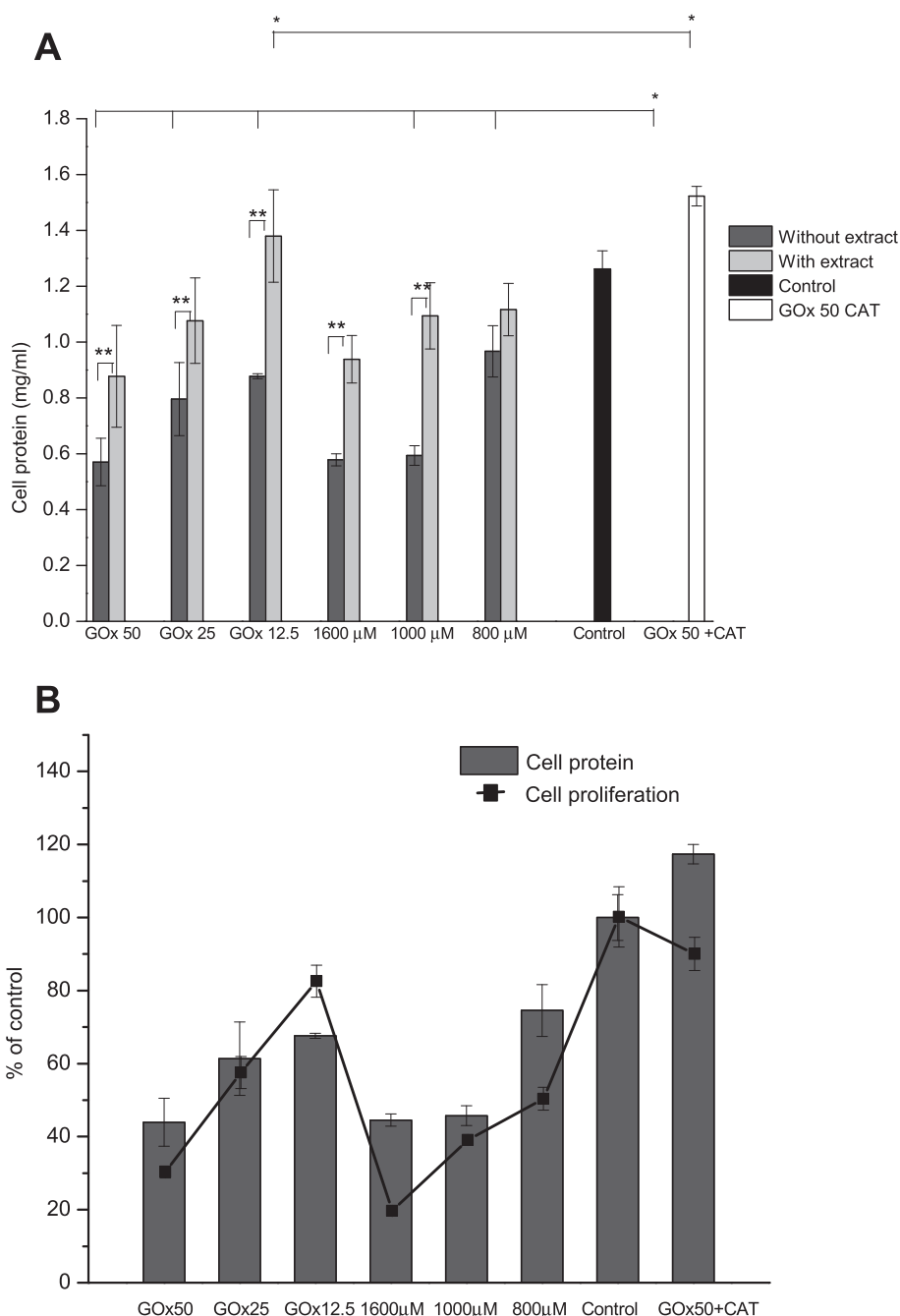
### Cell protein

The significant change in cellular morphology and the increase in  $\text{H}_2\text{O}_2$  of levels in ARPE-19 cells were associated with a dose-dependent decrease in protein concentrations, but antioxidant treatment prevented this decrease in cell protein (Fig. 14a). From these results it is apparent that *Bucida buceras* treatment exerted a beneficial effect on oxidative stress tolerance. The loss of protein from the cell layer compared to the untreated control cultures was related with cytotoxic action of  $\text{H}_2\text{O}_2$ . Because the decrease in protein concentrations could represent a cytostatic and cytotoxic effect; MTT assays were performed in control (untreated cells) and  $\text{H}_2\text{O}_2$  treated cells. Whereas (100  $\mu\text{M}$  of  $\text{H}_2\text{O}_2$  or 5 mU/ml GOx) treatment was nontoxic at 24 h, the surviving fraction decreased approximately 50 % in 800  $\mu\text{M}$  of  $\text{H}_2\text{O}_2$  addition or 32.5 mU/ml GOx and 80 % in 1600  $\mu\text{M}$  or 50 mU/ml GOx treated cells (Fig. 14b).

In Fig. 14b it is clearly visible that hydrogen peroxide-treated cells are losing their dividing ability in a concen



**Fig. 13** Effect of co-treatment with *Bucida buceras* (1600  $\mu\text{g}/\text{ml}$ ) in glucose oxidase (GOx) or  $\text{H}_2\text{O}_2$  addition treatment on antioxidant capacity measured by FRAP assay. Control GOx + CAT (50 mU/ml + catalase 200 U/ml) or control cells (untreated with  $\text{H}_2\text{O}_2$ ). \* $p < 0.05$  significantly different from value of cells control vs.  $\text{H}_2\text{O}_2$ -treated \*\* $p < 0.05$  significantly different from value of antioxidant treatment vs.  $\text{H}_2\text{O}_2$ -treated, n.s not significant differences



**Fig. 14** Effects of *Bucida buceras* on cell protein concentration in ARPE-19 cells H<sub>2</sub>O<sub>2</sub>-induced oxidative damage. H<sub>2</sub>O<sub>2</sub> generated by GOx (50, 25, 12.5 mU/ml) or in addition (1600, 1000 y 800 μM) (a). Relationship between values for cell proliferation (line) and the protein concentration (bars) (b). Values are presented as means ± SD of at least 3 independent experiments. Control GOx + CAT (50 mU/ml + catalase 200 U/ml) or control cells (untreated with H<sub>2</sub>O<sub>2</sub>). \**p* < 0.05 significantly different from value of cells control vs. H<sub>2</sub>O<sub>2</sub>-treated \*\**p* < 0.05 significantly different from value of antioxidant treatment vs. H<sub>2</sub>O<sub>2</sub>-treated, n.s not significant differences

tration-dependent manner. The proliferating ability had not recovered after 24 h after the start of oxidative challenge. This permanent inhibition in proliferation might be attributed to the persistent presence of protein aggregates in ARPE-19 cells.

The oxidative attack leads first to a slight protein oxidation, causing misfolding in the native protein state; if the stress persists, further oxidation aims at the hydrophobic residues that are normally buried inside proteins and these become heavily oxidized, which favors the formation of



protein aggregates. Accumulation of oxidized proteins is normally degraded by the proteasome, however this process may be impaired when the rate of oxidized protein formation exceeds proteasomal capacity or when the proteasome itself has decreased its activity. The formed aggregates interact with the proteasome, inhibit its activity, and lead to a loss of functionality, e.g. proliferation.

**Effect of *Bucida buceras* extract on cell survival after exposure to H<sub>2</sub>O<sub>2</sub>**

**MTT cytotoxicity assay**

Cellular viability was determined through the MTT assay in ARPE-19 cells in a range of concentration 12.5 to 1600 µg/ml of extract. The present study confirmed the low toxicity of *Bucida buceras* up to the highest concentration analyzed (Fig. 15 inset). It was further studied whether *Bucida buceras* could attenuate H<sub>2</sub>O<sub>2</sub>-stimulated ARPE-19 cell death. The Figs 5 and 9 demonstrate that exposure of ARPE-19 cells to GOx 25 or 50 mU/ml and 800–1600 µM H<sub>2</sub>O<sub>2</sub> in addition increased the cell death.

Treatment of ARPE-19 cells with antioxidant decreased the cytotoxic effect of different oxidant treatments (Fig. 15). The co-treatment with 1600 µg/ml of extract enhanced the cell viability by ≈ 20 % compared to H<sub>2</sub>O<sub>2</sub>-treated cells alone, mainly in 1200 and 1600 µM. Exposure to 1600 µg/ml was found to exert a minor cytotoxic effect on ARPE-19 cells. Thedata indicated that *Bucida buceras* exerted a cytoprotective effect against H<sub>2</sub>O<sub>2</sub> induced oxidative stress. This

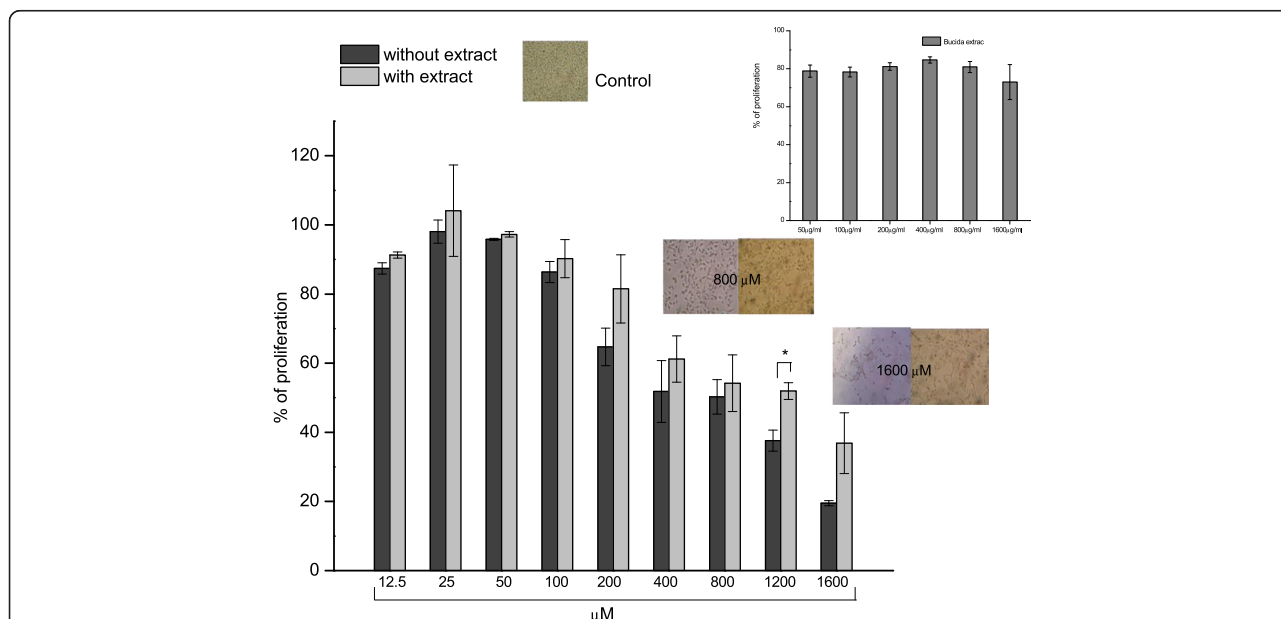
protection against H<sub>2</sub>O<sub>2</sub>-induced cell injury can be through direct H<sub>2</sub>O<sub>2</sub> scavenging activity and its potent anti-oxidative activity.

This study demonstrates for the first time that *Bucida buceras* can prevent partially H<sub>2</sub>O<sub>2</sub>-induced cell death. It was demonstrated that treatment with extract could improve the GSH and catalase activity. This demonstrated the anti-oxidative effects of the antioxidant extract in the retinal pigmented epithelium and supports the notion that the consumption of *Bucida buceras* may provide retinal-protective effects against oxidative stress by modulating the intracellular redox status.

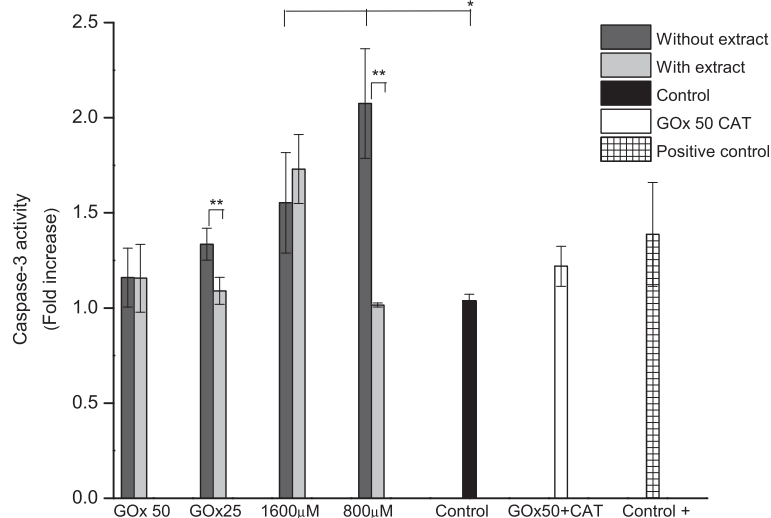
**Caspase-3 activity**

It is well known that caspase-3 has a primordial role in triggering the cascade of events that lead to apoptosis [2, 32, 37]. Cell lysates from the experimental cultures were tested for functional caspase activity using the caspase-3-specific substrate Ac-DEVD, this peptide-PNA conjugate is nonchromophore until PNA is cleaved from the peptide by an active caspase, and consequently, increased absorbance is indicative of caspase activity.

Treatment with H<sub>2</sub>O<sub>2</sub> by addition or by enzymatic generation significantly increased the activity of caspase-3 relative to ARPE-19 control cells (Fig. 16). The control cells (untreated) showed a very minor caspase-3 activity (≈1); however, once the cells were treated with 800–1600 µM



**Fig. 15** Protective effects of *Bucida buceras* on H<sub>2</sub>O<sub>2</sub>-treated ARPE-19 cells. Cells were co-treated with 1600 µg/ml in addition with various concentrations of H<sub>2</sub>O<sub>2</sub>-treatment by 24 h. Cell viability was measured by MTT assay: values are expressed as the percentage of cell survival relative to the medium control cells. Values are presented as means ± SD of 5–7 independent experiments. The insets show the low cytotoxicity of *Bucida buceras* (50 to 1600 µg/ml) on ARPE-19 cells. Cell morphology is also presented (original magnification: 200x). \*p < 0.05 significantly different from value of antioxidant treatment vs. H<sub>2</sub>O<sub>2</sub>-treated alone



**Fig. 16** Caspase-3 activity induced by  $H_2O_2$  addition (800 or 1600  $\mu M$ ) and GOx (25 or 50 mU/ml) in ARPE-19 cells for 24 h. Control,  $C^+$  cells were untreated with  $H_2O_2$ ,  $C^+$  (the specific caspase-3 control). Cells were co-treated with 1600  $\mu g/ml$  of *Bucida buceras* extract. Control GOx + CAT (50 mU/ml + catalase 200 U/ml) or control cells (untreated with  $H_2O_2$ ). Values are presented as means  $\pm$  SD of at least 3 independent experiments. \* $p < 0.05$  significantly different from value of control vs. Control GOx + CAT (50 mU/ml + catalase 200 U/ml) or control cells (untreated with  $H_2O_2$ ) *Bucida buceras*, or  $H_2O_2$ -treated only; \*\* $p < 0.05$  significantly different from value of antioxidant treatment vs.  $H_2O_2$ -treated alone

$H_2O_2$ , in addition the caspase-3 activity was increased by 1.5 and 2 fold ( $p < 0.05$ ). Although  $H_2O_2$  addition caused greater increase in the activation of caspase-3 compared to the enzymatic method. Indeed, severe oxidative stress conditions resulted in a drastic drop in the apoptosis (i.e. GOx 50 mU/ml and 1600  $\mu M$ ), consistent with the notion that pro-oxidant conditions can inactivate caspases by oxidative modification of key cysteine residues, thereby impairing the apoptotic program and promoting a switch to necrosis [5].

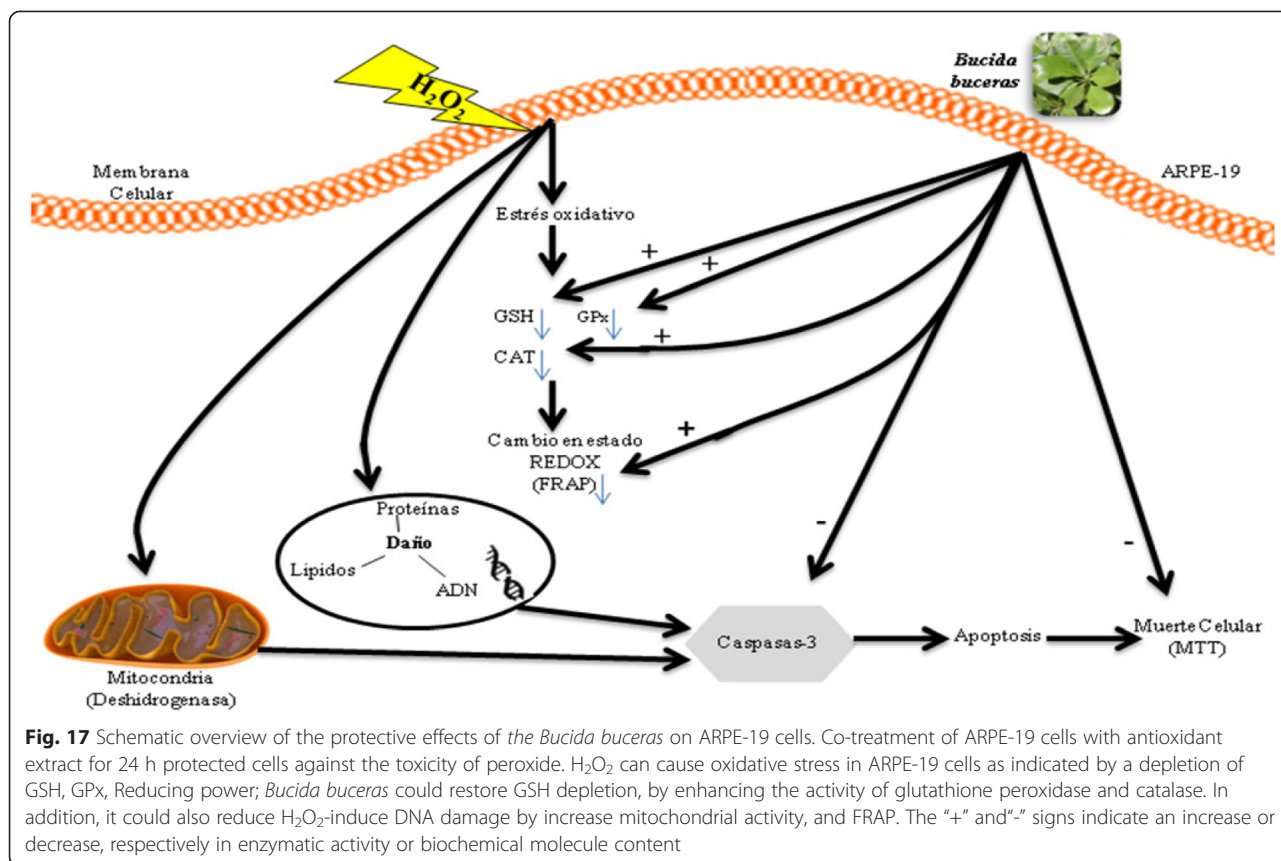
Exposure to high  $H_2O_2$  concentrations (1600  $\mu M$  or 50 mU/ml) exerted extensive damage leads to massive cell death. Necrotic-like cell death was reported, rather than apoptosis to high concentrations. At a moderate  $H_2O_2$  cytotoxic dose (800  $\mu M$  or 25 mU/ml), there was a significant increase in caspase-3 activity. Our current data further confirmed that caspase-3 might be a partial target involved in  $H_2O_2$ -induced apoptosis in the retinal-pigmented epithelium. On the other hand, the low correlation between caspase-3 activity and the level of cell death indicates that concentrations of  $H_2O_2$  influence the mode of cell death, i.e., apoptosis vs. necrosis. The higher  $H_2O_2$  concentrations induced severe toxicity, with over 90 % cell death, suggesting that under these experimental conditions necrosis was the predominant mode of cell death.

Since *Bucida buceras* treatment inhibited the cytotoxic effect of  $H_2O_2$ -generated oxidative stress, it was studied the direct effect on caspase-3 activity. Figure 16 shows that *Bucida buceras* exerted significant decrease on caspase-3 activity, mainly when these treatments were

studied at moderate concentrations (addition system-800  $\mu M$  or enzymatic-25 mU/ml). *Bucida* extract was able to prevent  $H_2O_2$ -induced increase in caspase-3 activity, with reductions of 16 and 38 % in 1600  $\mu M$  and 800  $\mu M$  respectively. GOx 25 mU/ml + antioxidant treatment exerted similar inhibitory effect (30 % decrease) but at GOx 50 mU/ml + extract treatment did not inhibited this activity.

#### Effect of *Bucida buceras* on death cell and redox status

Aging of the retina and RPE is associated with increased level of oxidative damage. It is generally assumed that oxidative stress also plays a role in the development or progression of age-related macular degeneration (AMD). The loss of photoreceptor function is as a consequence of oxidative damage to the RPE [1, 2]. This study employed several methods to determine whether co-treatment with extract may alter the response of ARPE-19 cells to  $H_2O_2$  on cell death and redox homeostasis. The data presented showed that *Bucida buceras* extract can partially protect ARPE-19 cells against  $H_2O_2$ -induced cell death. The change in the intracellular redox state can increase the ability of cells to handle oxidative stress. On the other hand, treatment with antioxidant extract was able to prevent the  $H_2O_2$ -induced damage due to an increase in the reducing power, glutathione level, and catalase activity (Fig. 17). These findings, lead to the conclusion that regulation of cellular redox state may be the major action of *Bucida buceras*.



### Discussion

A variety of continuous RPE cell lines provide the possibility to analyze in detail their morphology, functions, and molecular and genomic properties under normal and pathological conditions, which is hardly possible *in vivo*. A major advantage of such line is that they can be subcultured over more than hundreds of passages maintaining their characteristics over a number of passages and it have longer survival times, compared to primary cultures. Another important feature is that they have a uniform cell composition, although this may be evidence that these lines have lost certain properties essential to the initial cell material. On the other hand, cell culture, as any artificial system, obviously has certain disadvantages. Unlike cells *in vivo*, cultured cells are devoid of their native 3D microenvironment.

Additional limitations on the use of RPE cell cultures arise because of the genetic instability of continuous cell lines, which results from their unstable aneuploidy chromosome constitution, and heterogeneity of short-term cultures in terms of growth rate and capacity for intrapopulation differentiation, with consequent variation in their properties between passages. Despite all these circumstances, however, cultured cells retain many specialized functions, and cell lines have become an important tool in studies on RPE. ARPE-19 cells have been widely

used in studies on oxidative stress, retinal pathogenesis, and signaling pathways and also in research related to drugs and toxicity testing [12, 38–42].

Numerous studies have analyzed the response of cultured cells to H<sub>2</sub>O<sub>2</sub> when added to the culture medium as a single or repeated bolus. Usually, the purpose of these experiments is to investigate the cellular response to “oxidative stress”, as it may occur naturally in retinal pigment epithelium that are at high risk for oxidative damage *in vivo*. Most of these studies had been conducted by adding diluted H<sub>2</sub>O<sub>2</sub> to the cell culture medium (H<sub>2</sub>O<sub>2</sub> bolus treatment) assuming that this at least partially mimics conditions and elicits responses that may also occur *in vivo*. H<sub>2</sub>O<sub>2</sub> was utilized to generate oxidative stress due to the fact that do multiple oxidative pathways produce a common intermediate and it is involved in cellular redox signaling.

Nominal concentrations of H<sub>2</sub>O<sub>2</sub> in cell cultures are in fact initial concentrations that are eliminated rapidly from the culture medium [5, 22, 43–47]. Exogenously added H<sub>2</sub>O<sub>2</sub> has a short half-life cause by its rapid degradation by cellular catalase and peroxidases. Therefore amino acids present in the medium, such as methionine, cysteine and tryptophan, interact with H<sub>2</sub>O<sub>2</sub> leading to its consumption [22, 48]. On the other hand, different supplementation of culture medium with antioxidants and

precursors, as well as different contents of peroxide-consuming additives in the culture medium such as pyruvate and serum albumin is a known  $H_2O_2$  scavenger [46, 49, 50]. A rapid exponential depletion of  $H_2O_2$  is consistent with results reported for other cell types [46, 51] and for RPE [22]. Different supplementation of culture medium exerts variability in the cytotoxic potency of peroxides.

In our study we investigated the minimal concentration necessary to elicit 50 % of the cytotoxic effect ( $IC_{50}$ ). This value was calculated to be  $\approx 900 \mu M$  in contrast to the found by Zhang et al. [37], who reported that ARPE-19 cells needed 24 h to develop half cytotoxicity effect at 300  $\mu M$  nominal concentration but the cells were grown in serum starvation in DMEM/F-12 Su et al. [52] reported similar behavior when neuronal cells were cultured in serum-free medium for 48 h they experienced about 40 % cell death. This cell population displayed decrease in S phase and an emergence of sub- $G_0/G_1$  phase compared with those cultured in normal serum. Serum starvation successfully blocked the progression of cells from  $G_0/G_1$  phase into S phase and drove cells toward apoptosis.

When  $H_2O_2$  is delivered or generated outside cells, a gradient is established across the cell membrane that depends not only on the extracellular concentration of the oxidant, but also on plasma membrane permeability to  $H_2O_2$  and, on the amount and availability of enzymes that decompose  $H_2O_2$  [5, 44, 45].  $H_2O_2$  was found to be distributed in various subcellular compartments, i.e., the cytosol, the mitochondria, and the nucleus [5].

The low cytotoxicity of  $H_2O_2$  at 4 h in ARPE-19 cells means that cytotoxicity of  $H_2O_2$  takes several hours longer than the exposure to  $H_2O_2$ . Gulden et al. [46] revealed that the cytotoxic potency of  $H_2O_2$  in C6 astrogloma cell increased about 17 fold by prolonging the incubation time. Antunes and Cadenas [44] observed that Jurkat cells needed 12 h after 1-h exposure to a steady-state concentration 25  $\mu M$   $H_2O_2$  to develop maximum cytotoxicity. It has to be noted that cell lines vary in their sensitivity to  $H_2O_2$ . ARPE-19 cells are not very sensitive to low concentrations of  $H_2O_2$  (i.e. 12.5 to 200  $\mu M$ ), presumably because their antioxidant defense mechanisms are sufficient to counteract the damaging effects of the low  $H_2O_2$  concentrations applied. However at high  $H_2O_2$  concentrations (i.e. 400–1600  $\mu M$ ), the antioxidant defense mechanism in the ARPE-19 cell line appears to be insufficient to cope with such high oxidant insult, resulting in a high percentage of cell death.

The retinal pigmented epithelium is armed with a robust antioxidant system because of the high ambient oxygen tension required to maintain its high metabolism that is necessary to maintain the health and function of the overlying photoreceptors, and the unique, constant exposure to photo-oxidative stress [53]. Despite a potent

antioxidant system, the RPE cell became progressively dysfunctional and finally died by apoptosis. Part of this deterioration has been attributed to inadequately neutralized oxidative stress. Oxidative damaged in biomolecules have been identified in the RPE from AMD samples [53, 54].

GSH had an essential role in preserving nuclear function from deleterious oxidative modifications. A minimal GSH concentration is essential to protect the nucleus against oxidative damage and the maintaining of nuclear function during mild  $H_2O_2$  treatment is the key parameter of cell survival. When GSH concentration is high ( $>1$  mM), all cell functions are preserved from oxidative inhibition. As a result, enzymes of the detoxification machinery are induced and intracellular  $H_2O_2$  is rapidly degraded. At a lower concentration ( $\approx 0.1$  mM), GSH loses its cytosolic protective effect but it remains efficient for the protection of the nucleus against oxidative damage. Despite the transcriptional induction of stress-responsive genes, the detoxification machinery remains at a low level, because of the inhibition of translation. Therefore, is a slowly degraded and oxidized protein accumulating in the cytosol. But upon stress release, the functional nucleus can support growth resumption. When GSH concentration drops under 0.03 mM, both nuclear and cytosolic components become highly sensitive to oxidative damage, leading to cell death [55].

The protective effect of *Bucida buceras* in the cell viability of ARPE-19 is comparable to results reported in human colorectal cancer cells. Where cells exposed to 0.5 mM  $H_2O_2$  treatment with 8 mM GSH increased cell viability by  $\approx 15$  % and reduced oxidative microsatellite mutations by  $\approx 75$  %. However N-acetylcysteine (NAC) added at 10 mM did not further affect cell viability that had already been reduced by 0.5 mM  $H_2O_2$  although NAC suppressed oxidative microsatellite mutations [56].

Progression of death stimuli to apoptosis and necrosis depended on the mitochondrial-mediated damage and on ATP levels. The presence of ATP is essential for the activation of apoptosis protease and subsequent activation of caspases that induce apoptosis. The data obtained from this work indicate that moderate oxidative stress induced by  $H_2O_2$  is capable of inducing cell death by apoptosis dependent of caspase-3 in ARPE-19. Yu et al. [57] found that cell injury with 200  $\mu M$   $H_2O_2$ , in human lens epithelial B3 (HLE B3) is mediated primarily by caspase-associated apoptosis. HLE B3 cells were grown in serum deprivation MEM before exposure to a bolus of  $H_2O_2$ . While Zhang et al. [37] found that the aqueous extract of the root of *Salvia miltiorrhiza* (Salvianolic acid A) inhibited  $H_2O_2$ -induced caspase-3 cleavage in RPE cells. The cytoprotective effect of Sal A on RPE cells was also due to the activation of the transcription factor Nrf2 and its dependent antioxidant-responsive element (ARE)- genes.

In the RPE, oxidative stress is a powerful risk factor for age-related macular degeneration (AMD) and retinopathy diseases. The susceptibility of RPE cells to oxidative stress progressively increases with age, and cumulative oxidative damage contributes to RPE dysfunction and apoptosis [58]. Thurman et al. [59] found that H<sub>2</sub>O<sub>2</sub> depleted nuclear factor erythroid-derived 2-related factors 2 (Nrf2). Nrf2 regulates the inducible expression of antioxidant and cytoprotective enzymes via the cis-acting antioxidant response element. The *Bucida buceras* extract may affect genomic expression, which in turn regulates the expression of certain cytoprotective enzymes (i.e., catalase or GPx). We speculate therefore that long-term increment in antioxidant status will probably reduce RPE oxidative damage in the human eye and may delay onset of age-related visual impairment.

## Conclusion

These findings suggest that *Bucida buceras* could protect RPE against ocular pathogenesis associated with oxidative stress induced by H<sub>2</sub>O<sub>2</sub>-delivered by addition and enzymatic generation. The change in the intracellular redox state can increase the ability of cells to handle oxidative stress. On the other hand, treatment with antioxidant extract was able to prevent the H<sub>2</sub>O<sub>2</sub>-induced damage due to an increase in the reducing power, glutathione level, and catalase activity. The regulation of cellular redox state may be the major action of *Bucida buceras*.

## Competing interests

The authors declare that they have no competing interest.

## Authors' contributions

L.L.M. and I.A.S.B. proposed the ideas, planning the experimental methods, analyzed the data and wrote the manuscript; F.A.D. collected the samples and carried out the chemical analyses; G.S.A.A., L.E.C.L. and R.P.J.L. statistical analysis of data, reviewing and editing the manuscript. All authors read and approved the final manuscript.

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## Highlights

► *Bucida buceras* could protect RPE against oxidative stress. ► Cytotoxicity analyses induced by H<sub>2</sub>O<sub>2</sub> depend of addition or enzymatic generation methods. ► H<sub>2</sub>O<sub>2</sub>-induced injury decrease metabolic indicators: cellular proliferation, glutathione, antioxidant enzymes and reducing power.

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