# Original

# **Concordance between Results of Medium-term Liver Carcinogenesis Bioassays and Long-term Findings for Carcinogenic 2-Nitropropane and Non-carcinogenic 1-Nitropropane in F344 Rats**

Yuko Doi<sup>1,3</sup>, Seiko Tamano<sup>1</sup>, Mayumi Kawabe<sup>1</sup>, Masashi Sano<sup>1</sup>, Norio Imai<sup>1,3</sup>, Hironao Nakashima<sup>1</sup>, Fumio Furukawa<sup>1</sup>, Akihiro Hagiwara<sup>1</sup>, Masanori Otsuka<sup>2, #</sup>, and Tomoyuki Shirai<sup>3</sup>

<sup>1</sup> DIMS Institute of Medical Science, Inc., 64 Goura, Nishiazai, Azai-cho, Ichinomiya 491-0113, Japan

<sup>2</sup>Chemicals Evaluation and Research Institute, Japan, 1-4-25 Kouraku, Bunkyou-ku, Tokyo 112-0004, Japan

<sup>3</sup> Department of Experimental Pathology and Tumor Biology, Nagoya City University Graduate School of Medical

Sciences, 1 Kawasumi, Mizuho-ku, Mizuho-cho, Nagoya 467-8601, Japan

#Present: Chemicals Evaluation and Research Institute, Japan, 3-2-7 Miyanojin, Kurume 839-0801, Japan

**Abstract:** This study was conducted to determine the concordance of results for a pair of structural isomers, 2-nitropropane (2-NP) and 1-nitropropane (1-NP), using the rat medium-term liver carcinogenesis bioassay (Ito test) and previously published long-term carcinogenicity tests. Male F344 rats were given a single intraperitoneal injection of DEN (200 mg/kg b.w.) to initiate hepatocarcinogenesis. After 2 weeks, they received per os 0, 0.8, 4 or 20 mg/kg/day of 2-NP or 1-NP six times a week and were subjected to two-thirds partial hepatectomy at week 3. Non-initiated groups receiving 0 or 20 mg/kg/day were also included. The animals were sacrificed for quantitative analysis of GST-P-positive foci at week 8. With the highest dose of 2-NP, significantly increased numbers and areas of GST-P-positive foci were demonstrated as compared with the respective control but were not noted with 1-NP. In the non-DEN-initiated groups, many small GST-P-positive foci of less than 0.2 mm in diameter were also induced in the rats treated with 2-NP at 20 mg/kg/ day but were lacking with 1-NP. These results strongly support that 2-NP is a complete hepatocarcinogen with a potent initiation activity, whereas 1-NP is not. (DOI: 10.1293/tox.24.207; J Toxicol Pathol 2011; 24: 207–213)

Key words: the rat medium-term liver carcinogenesis bioassay, Ito test, 1-nitropropane, 2-nitropropane

# Introduction

2-Nitropropane (2-NP) was once widely used as a chemical intermediate and as a solvent component of paints, inks and varnishes<sup>1,2</sup>. It has also been detected in cigarette smoke in significant quantities<sup>3</sup>. Results of *in vivo* long-term bioassays in rats given 2-NP via gavage or inhalation in fact showed the compound to be a potent liver carcinogen<sup>4,5</sup>, and the overall evaluation is category 2B in the IARC Monographs from positive animal data<sup>2</sup>. In contrast, 1-nitropro-

Received: 29 June 2011, Accepted: 22 August 2011

Mailing address: Seiko Tamano, DIMS Institute of Medical Science, Inc., 64 Goura, Nishiazai, Azai-cho, Ichinomiya 491-0113, Japan TEL: 81-586-51-1201 FAX: 81-586-51-5634

E-mail: tamano@dims.co.jp

©2011 The Japanese Society of Toxicologic Pathology

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives (by-ncnd) License <a href="http://creativecommons.org/licenses/by-nc-nd/3.0/">http://creativecommons.org/licenses/by-nc-nd/3.0/</a>>. pane (1-NP), a structural isomer utilized as a propellant fuel, gasoline additive and in chemical syntheses1, was not found to be a hepatocarcinogen in a series of experiments<sup>5-7</sup>. Furthermore, while 2-NP proved to be mutagenic in a variety of short-term mutagenesis assays, including the Ames/Salmonella<sup>8-10</sup>, in vitro sister chromatid exchange (SCE) and chromosome aberration<sup>11</sup>, V79/HGPRT forward mutation and in vitro and in vivo unscheduled DNA synthesis (UDS) assays<sup>12,13</sup>, positive data for 1-NP are limited to in vitro V79/ HGRPT cells<sup>12</sup>. Neither was found to be mutagenic in micronucleus tests with polychromatic erythrocytes14,15. 2-NP does not appear to resemble any of the known classes of chemical carcinogens. Regarding the mechanisms of its carcinogenicity, 2-NP causes oxidative DNA and RNA damage in the rat liver resulting from intracellular generation of reactive forms of oxygen and/or 8-hydroxyguanine and 8-hydroxy-2'-deoxyguanosine<sup>16,17</sup>. Sodum et al. proposed from in vivo and in vitro experimental evidence that activation to an aminating species by rat liver aryl sulfotransferase is involved<sup>18,19</sup>. In addition, it was suggested that 2-NP

and its nitronate, an NO species, may mediate or contribute to genotoxicity<sup>20</sup>. A positive association between increased cell proliferation as assessed by incorporation of bromodeoxyuridine and hepatocarcinogenesis has been reported for 2-NP, but not 1-NP<sup>21</sup>. Preneoplastic lesions,  $\gamma$ -glutamyl transpeptidase ( $\gamma$ -GT) and glutathione S-transferase placental form (GST-P)-positive foci, were also produced in Wistar rats injected intraperitoneally with 2-NP in the initiation stage<sup>22</sup>.

The structures of 1-NP and 2-NP differ by only a little, but they differ greatly in their carcinogenic activity in the liver. In the present experiment, we investigated whether the two isomers promote development of GST-P-positive foci, as end-point lesions, in a medium-term liver carcinogenesis bioassay<sup>23–25</sup> to determine their sensitivity and specificity for distinguishing carcinogenic from non-carcinogenic chemicals<sup>25–31</sup>.

## **Materials and Methods**

All experimental procedures were performed in accordance with the in-house guidelines for the Care and Use of Laboratory Animals at DIMS Institute of Medical Science, Inc.

#### Test chemicals and initiator

2-nitropropane (2-NP) and 1-nitropropane (1-NP) were purchased from Aldrich Chemical Co., Milwaukee, WI, USA. Diethylnitrosamine (DEN) was purchased from Tokyo Chemical Industry Co., Ltd., Tokyo, Japan and used as an initiator of liver carcinogenesis. Regarding the reason for the determined dose level of test chemicals, a daily oral dose of 20 mg/kg/day, which did not show hepatocellular injury and increase in cell proliferation in a 2-week short-term oral administration study of 2-NP<sup>21</sup>, was selected as the highest dose level in the present experiment. The lower levels were set at 4 and 0.8 mg/kg/day using a proportional factor of 5. The 1-NP dosage was the same as that for 2-NP to enable comparison.

#### Animals and maintenance

Male F344/DuCrj and F344/DuCrlCrlj rats, 5 weeks of age, were purchased from Charles River Laboratories Japan, Inc., Atsugi, Japan, and housed two or three to a polycarbonate cage with hardwood Beta chips (Northeastern Products Co., Warrensburg, NY, USA) for bedding. The animals were supplied with food (Oriental MF, Oriental Yeast Co., Ltd., Tokyo, Japan) and tap water *ad libitum*, and the room temperature and relative humidity were controlled at 20–24 °C and 48–59%, respectively. Fluorescent lighting provided a 12-hr light/dark cycle. After a one-week quarantine and acclimation period, during which body weight and health conditions were monitored, a total of 159 rats were used for two experiments at 6 weeks of age.

#### Experimental procedure and autopsy

In experiment 1, from a total 78 male F344/DuCrj rats, 72 were allocated to 6 groups (15 rats each for groups 1 to 4 and 6 rats each for groups 5 and 6) using a computerized stratified body weight technique. At 6 weeks of age, animals of groups 1 to 4 were initially given a single intraperitoneal injection of DEN at a dose of 200 mg/kg b.w. dissolved in saline to initiate hepatocarcinogenesis<sup>32</sup>. Rats of groups 5 and 6 received the saline vehicle instead. After 2 weeks, animals of groups 1 to 4 were given 0, 0.8, 4 or 20 mg/kg/ day per os of 2-NP, dissolved in corn oil, while non-DENinitiated groups 5 and 6 received 0 and 20 mg/kg/day of 2-NP, respectively, for 6 weeks 6 times per week, with no dosing on Sunday. All animals were subjected to two-thirds partial hepatectomy (PH) at the end of week 3. The animals were observed daily for abnormalities, and body weights and food consumption were measured once a week.

In experiment 2, from a total 97 male F344/DuCrlCrlj rats, 87 were allocated to 7 groups (15 rats each for groups 1 to 5 and 6 rats each for groups 6 and 7) using the same technique as detailed above. At 6 weeks of age, animals of groups 1 to 4 were used under the same experimental regime, given 0, 0.8, 4 or 20 mg/kg/day of 1-NP dissolved in corn oil in place of 2-NP as the test compound, with an additional group of 15 rats receiving 2-NP at 20 mg/kg/day as a positive control. Non-DEN-initiated groups 6 and 7 received 0 and 20 mg/kg/day of 1-NP, respectively. Two-thirds PH was also conducted at the end of week 3. The animals were observed daily for abnormalities, and body weights and food consumption were measured once a week. Surviving rats in each group were sacrificed at the end of experimental week 8. At sacrifice in both experiments, livers were immediately excised and weighed to allow calculation of the liver-to-body weight ratio. A total of three 4 to 5 mm-thick slices from the cranial and caudal parts of the right lateral lobes, and the cranial part of the caudate lobe were cut with a razor blade and fixed in 10% buffered formalin for immunohistochemical demonstration of glutathione S-transferase placental form (GST-P)-positive foci.

## *Immunohistochemical staining and measurement of GST-P-positive foci*

The avidin-biotin-peroxidase complex (ABC) method was used to stain GST-P-positive foci. After deparaffinization, liver sections were treated sequentially with normal goat serum, anti-rabbit GST-P antibody (Medical Biological Laboratories Co., Ltd., Nagoya, Japan; 1:2000) and biotinlabeled goat anti-rabbit IgG (1:200) for 1 hr using an Elite ABC kit (Vector Laboratories, Burlingame, CA, USA). The sites of peroxidase binding were visualized with diaminobenzidine tetrahydrochloride, and the nuclei were counterstained with hematoxylin. All GST-P-positive foci larger than 0.2 mm in diameter (the lowest limit for reliable evaluation) were measured using a color video image processor (IPAP-WIN, Sumika Technos, Osaka, Japan), and the numbers and areas of foci/square centimeter (cm<sup>2</sup>) of liver sections were calculated.

Group	Treatment		Effective	Final hady weight (g)	Liver weight <sup>a</sup>		
	DEN	2-NP (mg/kg/day)	No. of rats	Final body weight (g) <sup>a</sup>	Absolute (g)	Relative (g/100 g body weight)	
1	+	0	15	$240.5 \pm 9.8$	$5.63 \pm 0.35$	$2.34 \pm 0.07$	
2	+	0.8	13	$244.5 \pm 13.0$	$5.93 \pm 0.36*$	$2.42 \pm 0.07$ **	
3	+	4.0	15	$237.1 \pm 12.2$	$5.61 \pm 0.43$	$2.36 \pm 0.08$	
4	+	20.0	15	$239.7 \pm 8.5$	$6.31 \pm 0.36 **$	$2.63 \pm 0.10$ **	
5	-	0	6	$243.0 \pm 14.7$	$5.85 \pm 0.31$	$2.41 \pm 0.10$	
6	_	20.0	6	$253.0 \pm 11.0$	$6.68 \pm 0.34^{\#\#}$	$2.64 \pm 0.07^{\#\#}$	

 Table 1. Final Body and Liver Weights of Male F344/DuCrj Rats Initiated with or without DEN Followed by 2-Nitropropane (2-NP)

 Administration and Partial Hepatectomy in Experiment 1

\*,\*\* Significantly different from the control group (group 1) at P<0.05 and 0.01, respectively. ## Significantly different from the control group (group 5) at P<0.01. a Values indicate the mean  $\pm$  SD.

 Table 2. Final Body and Liver Weights of Male F344/DuCrlCrlj Rats Initiated with or without DEN Followed by 1-Nitropropane (1-NP) Administration and Partial Hepatectomy in Experiment 2

Group	Treatment		Effective	Final hadre maight (g)h	Liver weight <sup>b</sup>		
	DEN 1-NP (mg/kg/day)		No. of rats Final body weight (g) <sup>b</sup> -		Absolute (g)	Relative (g/100 g body weight)	
1	+	0	14	$233.2 \pm 12.9$	$5.58 \pm 0.44$	$2.39 \pm 0.09$	
2	+	0.8	14	$234.5 \pm 13.3$	$5.61 \pm 0.43$	$2.39 \pm 0.07$	
3	+	4.0	15	$233.5 \pm 8.8$	$5.68 \pm 0.32$	$2.43 \pm 0.11$	
4	+	20.0	14	$221.2 \pm 9.0$ **	$5.55 \pm 0.34$	$2.51 \pm 0.14$ **	
5ª	+	20.0	15	$231.7 \pm 12.6$	$6.31 \pm 0.48 **$	$2.72 \pm 0.10$ **	
6	_	0	6	$245.2 \pm 16.5$	$5.91 \pm 0.50$	$2.41 \pm 0.13$	
7	- 20.0		6	$243.0 \pm 13.8$	$6.15 \pm 0.61$	$2.52 \pm 0.12$	

\*\* Significantly different from the control group (group 1) at P<0.01. a 2-Nitropropane was used as a positive substance. b Values indicate the mean  $\pm$  SD.

Small GST-P-positive foci of less than 0.2 mm in diameter were found in the non-DEN-initiated rats. The sizes of these GST-P-positive foci were evaluated under a microscope as follows: single cell, 2 to 4 cells, 5 to 10 cells or 11 cells or more.

#### Statistical analysis

The significance of differences for each parameter (excluding general conditions and food consumption) was analyzed and evaluated at P<0.05 or 0.01. Statistical comparisons between group 1 and groups 2 to 4 for numerical data were conducted using the Bartlett's test (evaluated at P<0.05). If homogeneous, the data were analyzed with the Dunnett's multiple comparison test (one sided), and if not, they were analyzed with the Steel's test (one sided)<sup>33,34</sup>. Statistical comparisons between groups 5 and 6 (experiment 1) and between groups 1 and 5 and groups 6 and 7 (experiment 2) for numerical data were conducted using the *F* test. If homogeneous, the data were analyzed with the Student's *t*-test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test (two sided), and if not, they were analyzed with the Student's test.

# Results

Neither mortality nor clinical changes related to the test compound treatment were apparent in any of the groups. Average body weight values for animals exposed to DEN were lower than those of rats without DEN initiation at week 1 and thereafter continued to be depressed until termination at week 8. Throughout the period of test material treatment (weeks 3 to 8), the mean body weight values of rats given 2-NP or 1-NP were comparable to the corresponding control values, although the values for rats treated with 20 mg/kg of 1-NP with DEN initiation were significantly lower than the control values in experiment 2 (data not shown). Food consumption by 2-NP- or 1-NP-treated animals was comparable to the corresponding control values throughout the experiment (data not shown).

Final average body weights for rats given 20 mg/kg/day of 1-NP with DEN initiation were significantly lower than those of control group 1. Those of the other groups were comparable to control values (Tables 1 and 2). Absolute and relative liver weights of rats treated with 0.8 and 20 mg/kg/ day of 2-NP with DEN initiation were significantly higher than those of control group 1, and those of the rats treated with 20 mg/kg/day of 2-NP without DEN initiation were significantly higher as compared with control group 5 (Table 1). The relative liver weights of rats treated with 20 mg/ kg/day of 1-NP with DEN initiation were also significantly higher than those of control group 1 (Table 2). The absolute and relative liver weights of rats treated with 20 mg/kg/ day of 2-NP with DEN initiation, reference group 5, were significantly higher than those of control group 1 (Table 2). As macroscopic findings, pale color and several whitish/yellowish points in the liver were observed in rats treated with 20 mg/kg/day of 2-NP with DEN initiation (data not shown).

The numbers and areas of GST-P-positive foci for rats treated 20 mg/kg/day of 2-NP with DEN initiation were

**Table 3.** Numbers and Areas of GST-P-positive Foci (0.2 mm or More in Diameter) in Male F344/DuCrjRats Initiated with or without DEN Followed by 2-Nitropropane (2-NP) Administration andPartial Hepatectomy in Experiment 1

Group		Treatment	Effective	GST-P-positive foci <sup>a</sup>		
	DEN	2-NP (mg/kg/day)	No. of rats	Number (No./cm <sup>2</sup> )	Area (mm <sup>2</sup> /cm <sup>2</sup> )	
1	+	0	15	$3.39 \pm 2.20$	$0.25 \pm 0.17$	
2	+	0.8	13	$3.50 \pm 2.44$	$0.24 \pm 0.18$	
3	+	4.0	15	$4.60 \pm 1.94$	$0.33 \pm 0.18$	
4	+	20.0	15	$10.83 \pm 3.36 **$	$0.88 \pm 0.31 **$	
5	-	0	6	$0.00\pm0.00$	$0.00\pm0.00$	
6	-	20.0	6	$0.00\pm0.00$	$0.00\pm0.00$	

\*\* Significantly different from the control group (group 1) at P<0.01. a Values indicate the mean±SD.

 

 Table 4.
 Numbers and Areas of GST-P-positive Foci (0.2 mm or More in Diameter) in Male F344/Du-CrlCrlj Rats Initiated with or without DEN Followed by 1-Nitropropane (1-NP) Administration and Partial Hepatectomy in Experiment 2

Group		Treatment	Effective	GST-P-positive foci <sup>b</sup>			
	DEN	1-NP (mg/kg/day)	No. of rats	Number (No./cm <sup>2</sup> )	Area (mm <sup>2</sup> /cm <sup>2</sup> )		
1	+	0	14	$3.64 \pm 1.01$	$0.31 \pm 0.12$		
2	+	0.8	14	$4.95 \pm 1.99*$	$0.43 \pm 0.26$		
3	+	4.0	15	$3.91 \pm 1.78$	$0.33 \pm 0.18$		
4	+	20.0	14	$4.38 \pm 1.46$	$0.32 \pm 0.11$		
5ª	+	20.0	15	$11.32 \pm 1.54 **$	$0.97 \pm 0.17 **$		
6	-	0	6	$0.00 \pm 0.00$	$0.00 \pm 0.00$		
7	-	20.0	6	$0.00 \pm 0.00$	$0.00 \pm 0.00$		

\*.\*\* Significantly different from the control group (group 1) at *P*<0.05 and 0.01, respectively. a 2-nitropropane was used as a positive substance. b Values indicate the mean ± SD.

 Table 5. Quantitative Analysis of GST-P-positive Foci of Less than 0.2 mm in Diameter in Male F344/DuCrj Rats Initiated without DEN Followed by 2-Nitropropane (2-NP) Administration and Partial Hepatectomy in Experiment 1

Cassian		Treatment	Effective	Size distribution of GST-P-positive foci (No./cm <sup>2</sup> ) <sup>a</sup>				
Group	DEN	2-NP (mg/kg/day)	No. of rats	Single cell	2-4 Cells	5-10 Cells	>11 Cells	Total <sup>b</sup>
5	_	0	6	$0.00\pm0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.07 \pm 0.16$	$0.07 \pm 0.16$
6	-	20.0	6	$2.80 \pm 1.17$ **	$4.76 \pm 1.67 **$	$3.09 \pm 1.11$ **	$0.63\pm0.66*$	$8.48 \pm 2.45^{**}$

\*, \*\* Significantly different from the control group (group 5) at P<0.05 and 0.01, respectively. <sup>a</sup> Values indicate the mean ± SD. <sup>b</sup> Single GST-P-positive cells were excluded.

 Table 6. Quantitative Analysis of GST-P-positive Foci of Less than 0.2 mm in Diameter in Male F344/DuCrlCrlj Rats Initiated without DEN Followed by 1-Nitropropane (1-NP) Administration and Partial Hepatectomy in Experiment 2

Group	Treatment		Effective	Size distribution of GST-P-positive foci (No./cm <sup>2</sup> ) <sup>a</sup>				
	DEN	1-NP (mg/kg/day)	No. of rats	Single cell	2-4 Cells	5-10 Cells	>11 Cells	Total <sup>b</sup>
6	_	0	6	$0.42 \pm 0.37$	$0.07 \pm 0.17$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.07 \pm 0.17$
7	_	20.0	6	$0.00\pm0.00*$	$0.00\pm0.00$	$0.07\pm0.17$	$0.13\pm0.20$	$0.20\pm0.22$

\* Significantly different from the control group (group 6) at P<0.05. <sup>a</sup> Values indicate the mean  $\pm$  SD. <sup>b</sup> Single GST-P-positive cells were excluded.

significantly higher than those of control group 1 in experiment 1 (Table 3). On the other hand, 1-NP did not promote the development of GST-P-positive foci in experiment 2, although a slight significant increase in number was evident in rats given 0.8 mg/kg/day of 1-NP with DEN initiation, with no dose dependence (Table 4). In the non-DEN-initiated groups, a number of small GST-P-positive foci of less than 0.2 mm in diameter developed in the rats given 20 mg/ kg/day of 2-NP, and this was significantly higher than the control value (group 5) in experiment 1 (Table 5). However, no such findings were evident for 1-NP (group 6) in experiment 2 (Table 6).

## Discussion

In the present investigation, the liver carcinogen 2-NP gave an unequivocally positive result in the *in vivo* mediumterm liver carcinogenesis bioassay, whereas 1-NP proved negative. Preneoplastic GST-P-positive foci was significantly increased in the rats treated with 20 mg/kg/day of 2-NP with DEN initiation, but not 4 mg/kg/day of 2-NP or less (experiment 1), pointing to a dose threshold.

Most importantly, the results of the present carcinogenesis bioassay were in accordance with the earlier finding that 2-NP is a potent rat liver carcinogen according to longterm in vivo carcinogenicity tests with exposure by inhalation (207 ppm) and per os (1 mmol/kg)<sup>4,5</sup>. No toxicological effects were observed in the rats given 1-NP of 20 mg/kg/ day except for the retardation of body weight. In contrast, increased liver weights were shown in the rats treated with 2-NP of 20 mg/kg/day. Moreover, the number of small GST-P-positive foci of less than 0.2 mm in diameter was significantly increased in the rats treated with 2-NP of 20 mg/ kg/day without DEN initiation plus PH but not in the rats treated with 1-NP. Therefore, it was suggested that 2-NP is a complete hepatocarcinogen, displaying both promoting and initiating activities, in accordance with the results of a previous initiation bioassay focusing on induction of γ-GT-positive and GST-P-positive foci<sup>22</sup>. 2-NP, categorized as complete hepatocarcinogen, indicates increase of GST-P-positive foci in both with and without DEN initiation treatment. 1-NP, as non-hepatocarcinogen, demonstrates no increase in GST-P-positive foci even after DEN initiation treatment.

With regard to the mechanism of mutagenicity, oxidative damage to DNA and RNA was demonstrated for 2-NP, eliciting 8-hydroxyguanine and 8-hydroxy-2'-deoxyguanosine, products of hydroxyl radical attack that can cause DNA misreplication<sup>16,17</sup>. It is therefore very likely that a modification of DNA and/or RNA and hepatocellular proliferation are involved in 2-NP carcinogenicity<sup>21</sup>. Another mechanism might be related to NH<sub>2</sub><sup>+</sup>, an aminating species, activated by aryl sulfotransferase, resulting in 8-aminoguanine products. In contrast, the isomer 1-NP appears not to be a substrate for aryl sulfotransferase<sup>18,19</sup>.

With another pair of structural isomers, 2,4-diaminotoluene (2,4-DAT), but apparently not 2,6-diaminotoluene (2,6-DAT), proved to be hepatocarcinogenic<sup>35,36</sup>, although both were found to be genotoxic in the Ames/Salmonella assay, and the metabolism of both compounds is mediated in a similar manner<sup>37,38</sup>. Cunningham and Matthews have documented that the difference may depend upon induction of cell proliferation. In the toxicological profile of the liver, clinical chemistry does not appear to differ between 2-NP and 1-NP<sup>21</sup>. However, increased cell proliferation was limited to the case with 2-NP, which proved to be carcinogenic<sup>39</sup>. Hepatocellular proliferation was similarly displayed at extremely high levels in the rats treated with 2,4-DAT but was lacking with 2,6-DAT<sup>40</sup>. Cell proliferation, therefore, may be an essential correlate of carcinogenicity with both 2-NP and 2,4-DAT liver carcinogens. Thus, after initiated cells are generated in the liver in the two-step theory model with partial hepatectomy, whereby pluripotential cells in the intermediate cell population undergo replication, the probability that another genetic alteration mistake can occur that leads to the development of cancer is augmented. With repeated cell divisions, initiated populations colonally expand to produce identifiable specific intermediate lesions such as GST-P-positive foci, which can be quickly and easily assessed as evidence for carcinogenic or non-carcinogenic potential<sup>41,42</sup>.

In conclusion, the present medium-term liver bioassay clearly demonstrated 2-NP, but not 1-NP, to be a hepatocarcinogen, in accordance with published results for *in vivo* long-term carcinogenesis. Moreover, it was concluded that 2-NP was a complete hepatocarcinogen with genotoxic activity, showing that it displayed GST-P-positive activity both with and without DEN initiation treatment in this bioassay system.

Acknowledgements: This study was supported in part by the New Energy and Industrial Technology Development Organization (NEDO), Japan.

We also thank Dr. Malcolm A. Moore for critical reading of the manuscript.

#### References

- Baker PJ Jr, and Bollmeir AF Jr. Nitroparaffins. In: Kirk-Othmer Encyclopedia of Chemical Technology, 3rd ed. Vol. 15. John Wiley and Sons, Inc., New York. 969–987. 1978.
- IARC. Some Industrial Chemicals and Dyestuffs. IARC Monographs on the Evaluation of Carcinogenic Risk of Chemicals to Humans, Vol. 29. Lyon. 331–343. 1982.
- Hoffmann D, and Rathkamp G. Chemical studies on tabacco smoke. III. Primary and secondary nitroalkanes in cigarette smoke. Beitr Tabakforsch. 4: 124–134. 1968.
- Lewis TR, Ulrich CE, and Busey WM. Subchronic inhalation toxicity of nitromethane and 2-nitropropane. J Environ Pathol Toxicol. 2: 233–249. 1979. [Medline]
- Fiala ES, Czerniak R, Castonguay A, Conaway CC, and Rivenson A. Assay of 1-nitropropane, 2-nitropropane, 1-azoxypropane and 2-azoxypropane for carcinogenicity by gavage in Sprague-Dawley rats. Carcinogenesis. 8: 1947–1949. 1987. [Medline] [CrossRef]
- Hadidian Z, Fredrickson TN, Weisburger EK, Weisburger JH, Glass RM, and Mantel N. Tests for chemical carcinogens. Report on the activity of derivatives of aromatic amines, nitrosamines, quinolines, nitroalkanes, amides, epoxides, aziridines, and purine antimetabolites. J Natl Cancer Inst. 41: 985–1036. [Medline]
- Griffin TB, Stein AA, and Coulston F. Inhalation exposure of rats to vapors of 1-nitropropane at 100 ppm. Ecotoxicol Environ Saf. 6: 268–282. 1982. [Medline] [CrossRef]
- Hite M, and Skeggs H. Mutagenic evaluation of nitroparaffins in the *Salmonella typhimurium*/mammalian-microsome test and the micronucleus test. Environ Mutagen. 1: 383–389. 1979. [Medline] [CrossRef]

- Speck WT, Meyer LW, Zeiger E, and Rosenkranz HS. Mutagenicity and DNA-modifying activity of 2-nitropropane. Mutat Res. 104: 49–54. 1982. [Medline] [CrossRef]
- Löfroth G, Nilsson L, and Anderson JR. Structure–activity relationship of nitroalkane-induced mutagenicity in the Ames Salmonella assay. Prog Clin Biol Res. 209B: 149–155. 1986. [Medline]
- Bauchinger M, Kulka U, and Schmid E. Analysis of cytogenetic effect in human lymphocytes induced by metabolically activated 2-nitropropane. Mutat Res. 190: 217–219. 1987. [Medline] [CrossRef]
- Ziegler-Skylakakis K, Homfeldt H, and Andrae U. *In vitro*and *in vivo-*genotoxicity of 2-nitropropane and 1-nitropropane in mammalian cells. Naunyn-Schmiedeberg's Arch Pharmacol. **335**(Suppl.): R25. 1987.
- Andrae U, Homfeldt H, Vogl L, Lichtmannegger J, and Summer KH. 2-Nitropropane induces DNA repair synthesis in rat hepatocytes *in vitro* and *in vivo*. Carcinogenesis. 9: 811–815. 1988. [Medline] [CrossRef]
- Kliesch U, and Adler I-D. Micronucleus test in bone marrow of mice treated with 1-nitropropane, 2-nitropropane and cisplatin. Mutat Res. 192: 181–184. 1987. [Medline] [CrossRef]
- George E, Burlinson B, and Gatehouse D. Genotoxicity of 1- and 2-nitropropane in the rat. Carcinogenesis. 10: 2329– 2334. 1989. [Medline] [CrossRef]
- Fiala ES, Conaway CC, and Mathis JE. Oxidative DNA and RNA damage in the livers of Sprague-Dawley rats treated with the hepatocarcinogen 2-nitropropane. Cancer Res. 49: 5518–5522. 1989. [Medline]
- Conaway CC, Nie G, Hussain NS, and Fiala ES. Comparison of oxidative damage to rat liver DNA and RNA by primary nitroalkanes, secondary nitroalkanes, cyclopentanone oxime, and related compounds. Cancer Res. 51: 3143–3147. 1991. [Medline]
- Sodum RS, Nie G, and Fiala ES. 8-Aminoguanine: a base modification produced in rat liver nucleic acids by the hepatocarcinogen 2-nitropropane. Chem Res Toxicol. 6: 269– 276. 1993. [Medline] [CrossRef]
- Sodum RS, Sohn OS, Nie G, and Fiala ES. Activation of the liver carcinogen 2-nitropropane by aryl sulfotransferase. Chem Res Toxicol. 7: 344–351. 1994. [Medline] [CrossRef]
- Kohl C, Morgan P, and Gescher A. Metabolism of the genotoxicant 2-nitropropane to a nitric oxide species. Chem Biol Interact. 97: 175–184. 1995. [Medline] [CrossRef]
- Cunningham ML, and Matthews HB. Relationship of hepatocarcinogenicity and hepatocellular proliferation induced by mutagenic noncarcinogens vs carcinogens. II. 1- vs 2-Nitropropane. Toxicol Appl Pharmacol. 110: 505–513. 1991. [Medline] [CrossRef]
- Astorg P, Bergès R, and Suschetet M. Induction of γGT- and GST-P positive foci in the liver of rats treated with 2-nitropropane or propane 2-nitronate. Cancer Lett. **79**: 101–106. 1994. [Medline] [CrossRef]
- Pitot HC III, and Dragan YP. Chemical carcinogenesis. In: Casarett & Doull's Toxicology, The Basic Science of Poisons, 6<sup>th</sup> ed. CD Klaassen (ed). McGraw Hill, New York. 241–319. 2001.
- Ito N, Tamano S, and Shirai T. A medium-term rat liver bioassay for rapid *in vivo* detection of carcinogenic potential of chemicals. Cancer Sci. 94: 3–8. 2003. [Medline] [Cross-Ref]

- Ogiso T, Tatematsu M, Tamano S, Tsuda H, and Ito N. Comparative effects of carcinogens on the induction of placental glutathione S-transferase-positive liver nodules in a shortterm assay and of hepatocellular carcinomas in a long-term assay. Toxicol Pathol. 13: 257–265. 1985. [Medline] [Cross-Ref]
- 26. Ito N, Tsuda H, Tatematsu M, Inoue T, Tagawa Y, Aoki T, Uwagawa S, Kagawa M, Ogiso T, Masui T, Imaida K, Fukushima S, and Asamoto M. Enhancing effect of various hepatocarcinogens on induction of preneoplastic glutathione S-transferase placental form positive foci in rats — an approach for a new medium-term bioassay system. Carcinogenesis. 9: 387–394. 1988. [Medline] [CrossRef]
- 27. Tatematsu M, Mera Y, Inoue T, Satoh K, Sato K, and Ito N. Stable phenotypic expression of glutathione S-transferase placental type and unstable phenotypic expression of γ-glutamyltransferase in rat liver preneoplastic and neoplastic lesions. Carcinogenesis. 9: 215–220. 1988. [Medline] [CrossRef]
- Ogiso T, Tatematsu M, Tamano S, Hasegawa R, and Ito N. Correlation between medium-term liver bioassay system data and results of long-term testing in rats. Carcinogenesis. 11: 561–566. 1990. [Medline] [CrossRef]
- Hasegawa R, and Ito N. Liver medium-term bioassay in rats for screening of carcinogens and modifying factors in hepatocarcinogenesis. Food Chem Toxicol. 30: 979–992. 1992. [Medline] [CrossRef]
- Shirai T. A medium-term rat liver bioassay as a rapid *in vivo* test for carcinogenic potential: a historical review of model development and summary of results from 291 tests. Toxicol Pathol. 25: 453–460. 1997. [Medline] [CrossRef]
- 31. Shirai T, Hirose M, and Ito N. Medium-term bioassays in rats for rapid detection of the carcinogenic potential of chemicals. In: The Use of Short- and Medium-term Tests for Carcinogens and Data on Genetic Effects in Carcinogenic Hazard Evaluation. DB Mcgregor, JM Rice, and S Venitt (eds). IARC Scientific Publications No. 146. Lyon. 251–272. 1999.
- Shirai T, Hosoda K, Hirose K, Hirose M, and Ito N. Promoting effects of phenobarbital and 3'-methyl-4-dimethylaminoazobenzene on the appearance of γ-glutamyltranspeptidase positive foci in rat liver pretreated with varying doses of diethylnitrosamine. Cancer Lett. 28: 127–133. 1985. [Medline] [CrossRef]
- Dunnett CW. A multiple comparison procedure for comparing several treatments with a control. J Am Stat Assoc. 50: 1096–1121. 1955. [CrossRef]
- Steel RGD. A multiple comparison rank sum test: treatments versus control. Biometrics. 15: 560–572. 1959. [CrossRef]
- National Toxicology Program Bioassay of 2,4-diaminotoluene for possible carcinogenicity. Natl Cancer Inst Carcinog Tech Rep Ser. 162: 1–139. 1979. [Medline]
- National Toxicology Program Bioassay of 2,6-toluenediamine dihydrochloride for possible carcinogenicity (CAS No. 15481–70–6). Natl Toxicol Program Tech Rep Ser. 200: 1–123. 1980. [Medline]
- Cunningham ML, Burka LT, and Matthews HB. Metabolism, disposition, and mutagenicity of 2,6-diaminotoluene, a mutagenic noncarcinogen. Drug Metab Dispos. 17: 612– 617. 1989. [Medline]
- 38. Cunningham ML, Foley J, Maronpot RR, and Mat-

thews HB. Correlation of hepatocellullar proliferation with hepatocarcinogenicity induced by the mutagenic noncarcinogen:carcinogen pair—2,6- and 2,4-diaminotoluene. Toxicol Appl Pharmacol. **107**: 562–567. 1991. [Med-line] [CrossRef]

- Cunningham ML, and Matthews HB. Cell proliferation as a determining factor for the carcinogenicity of chemicals: studies with mutagenic carcingens and mutagenic noncarcingens. Toxicol Lett. 82/83: 9–14. 1995. [Medline] [CrossRef]
- 40. Toyoda-Hokaiwado N, Inoue T, Masumura K, Hayashi H,

Kawamura Y, Kurata Y, Takamune M, Yamada M, Sanada H, Umemura T, Nishikawa A, and Nohmi T. Integration of *in vivo* genotoxicity and short-term carcinogenicity assays using F344 *gpt* delta transgenic rats: *In vivo* mutagenicity of 2,4-diaminotoluene and 2,6-diaminotoluene structural isomers. Toxicol Sci. **114**: 71–78. 2010. [Medline] [CrossRef]

- Cohen SM, and Ellwein LB. Cell proliferation in carcinogenesis. Science. 249: 1007–1011. 1990. [Medline] [Cross-Ref]
- 42. Cohen SM, and Arnold LL. Cell proliferation and carcinogenesis. J Toxicol Pathol. **21**: 1–7. 2008. [CrossRef]