



Article

The Unexpected Holiday Souvenir: The Public Health Risk to UK Travellers from Ticks Acquired Overseas

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Abstract: Overseas travel to regions where ticks are found can increase travellers' exposure to ticks and pathogens that may be unfamiliar to medical professionals in their home countries. Previous studies have detailed non-native tick species removed from recently returned travellers, occasionally leading to travel-associated human cases of exotic tick-borne disease. There are 20 species of tick endemic to the UK yetUK travellers can be exposed to many other non-native species whilst overseas. Here wereport ticks received by Public Health England's Tick Surveillance Scheme from humans with recent travel history between January 2006 and December 2018. Altogether, 16 tick species were received from people who had recently travelled overseas. Confirmed imports (acquired outside of the UK) were received from people who recently travelled to 22 countries. Possible imports (acquired abroad or within the UK) were received from people who had recently travelled to eight European countries. Species-specific literature reviews highlighted nine of the sixteen tick species are known to vector at least one tick-borne pathogen to humans in the country of acquisition, suggesting travellers exposed to ticks may be at risk of being bitten by a species that is a known vector withimplications for novel tick-borne disease transmission to travellers.

Keywords: tick-borne pathogens; *Ixodes ricinus*; *Amblyomma americanum*; *Dermacentor*; *Hyalomma*; *Rhipicephalus*; *Ixodes*; *Amblyomma*

1. Introduction

Since the 1950s, there has been year-on-year increases in worldwide international tourist arrivals [1]. In 2017 alone, there were 1,323 million worldwide tourist arrivals, increasing by 84 million international arrivals than 2016 [1]. Similar tourism increases have been seen in the United Kingdom (UK): overseas resident arrivals increased by 4% in 2017 compared with 2016 and3% more overseas visits were made by UK residents in the same time period [2]. Such frequent movement could increase exposure of travellers to ticks and their pathogens. As ticks are the second most common vectors of disease-causing pathogens in humans [3] such exposure could present unique public health challenges asticks from one region may transmit pathogens that are unfamiliar to medical professionals in other parts of the world [4].

To date, several reports have detailed tick detection and removal on recently returned travellers and ticks acquired in Africa, Asia, Australia, North and South America have been removed from residents in Asia, Europe NewZealand and the USA [5–13]. In New Zealand, 50% of all tick importations that were intercepted at the border were associated with human travel [11]. There have also been cases of illness caused by tick-borne pathogens in recently returned travellers. More than

350 travel-associated cases of African tick bite fever (caused by *Rickettsia africae*) have been reported in Europe, USA, Australia, Argentina and Japan [14] and patients may not recall a tick bite, despite suffering from a tick-borne illness. For example twoUSA residents developed skin lesions and flu-like symptoms within eight days of returning from Swaziland and were diagnosed with African tick bite fever yetneither patient reported tick bites during the trips [15].

Whilst 20 tick species are considered endemic in the UK [16,17] UKresidents can be exposed to non-native tick species whilst travelling abroad. As tick bites acquired overseas can present different health risks to those acquired in the UK itis vital that existing public health guidance promotes the risks to both public health professionals and travellers. The following paper summarises imported ticks received by Public Health England's passive Tick Surveillance Scheme (TSS) from humans with recent travel history and investigates the potential public health risk to UK residents bitten by ticks when travelling abroad.

2. Materials and Methods

Samples submitted to the TSS (see [18]) consisting of ticks likely acquired from outside of the UK between January 2006 and December 2018 were received from medical staff and members of the public. Upon arrival, specimens suspected to have been acquired overseas were frozen at -80 °C for 48 hours. Records were classed as imported if the recorder clearly stated that the tick had been acquired outside of the UK thespecies was not endemic to the UK orthe tick could not have been acquired in the UK based on the level of engorgement and supplied travel information. All submissions of endemic species with a recent travel history that could not be confirmed as definitely acquired outside of the UK were classed as possible importation events aslocal acquisition of the ticks could not be ruled out.

To identify species keysfor European ticks were initially used [19–21] withadditional keys consulted for non-European species (e.g., [22]). Tick experts in the country of origin were contacted to verify specimens where necessary. The identification result was relayed to the person submitting the tick aswell as signposting to information about possible tick-borne diseases that are known to be transmitted by the tick species in the country of origin andwhere necessary, follow-up questions were used to obtain more information about the tick encounter and the health of the person who had been bitten.

Following identification areview of published literature was conducted using PubMed to further understand the ecology and potential public health risk following a bite from each tick species. First, searches were carried out for each tick species in the country of origin forexample 'Ixodes ricinus AND France' andthen pathogens forexample 'Ixodes ricinus AND France AND Borrelia burgdorferi'. The literature review of pathogens detected in ticks focused on articles that were published in English between January 2010 and March 2019 aswell as the references cited therein. The relevance of articles identified by the database search (>900) were assessed first by their titles and abstracts, followed by an in-depth review of tick ecology and pathogen information.

3. Results

3.1. Tick Surveillance Scheme Imported Ticks

Between January 2006 and December 2018, 59 records were received from people with a recent travel history (Table 1). Records were comprised of 66 individual ticks belonging to 16 species plustwo damaged specimens that could only be identified at the genus level. In total, 76% of records (n = 45) were confirmed as imported and 24% (n = 14) were possible imports, where it was not possible to determine whether an endemic species received from someone with recent travel history was acquired abroad or in the UK. Considering only confirmed imported tick records, records were received in 2006 (n = 1), 2008 (n = 1), 2012 (n = 1), 2013 (n = 2), 2014 (n = 3), 2015 (n = 9), 2016 (n = 10) and 2017 (n = 10) and 2018. Possible imported records were received during 2013 (n = 2), 2015 (n = 2), 2016 (n = 2), 2017 (n = 3) and 2018 (n = 5). Across the years, ticks were removed from people with a history of travel during every month apart from March; the highest numbers were removed

in June (n = 11), followed by July (n = 10), April (n = 9) and May (n = 8), totalling 63% of all records (see Figure 1 for a breakdown of confirmed and possible imported tick records). In addition, 20% of records (n = 12) were removed from travellers between November and February; half of these records originated from Southern Hemisphere countries (Figure 1).

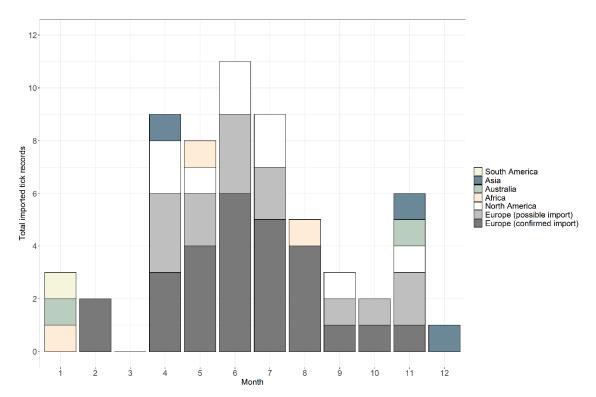


Figure 1. Seasonality of tick records received by the Tick Surveillance Scheme from people with a history of travel that were acquired in South America (yellow) Asia(blue-grey), Australia (green), Africa (beige) and North America (white) andpossible imports from Europe (light grey) and confirmed imports from Europe (dark grey).

Ticks were received from people who had recently travelled to six continents: 70% of records were associated with travel in Europe, 15% from North America, 5% from both Africa and Asia, 3% from Australia and 2% from South America (Table 1; Figure 2). Confirmed imported ticks were removed from people that had recently travelled to 22 countries (Figure 2, Table 1) andpossible imported ticks were removed from people with recent travel history to eight European countries (Figure 2, Table 1). There were two instances where the country of origin could not be determined as travel through multiple countries was reported (Table 1). One record, *Dermacentor marginatus*, was likely acquired either in Holland, Germany or France (although *D. marginatus* is not present in Holland (see [23])), whilst the second record, *Dermacentor andersoni* camefrom either the Canadian Rockies or Washington State in north-western USA; despite the country of origin being unknown bothrecords were confirmed importation events as neither species is endemic to the UK. Exact locations where ticks were likely acquired were provided for 54% of records (n = 32), whilst regional/country information was provided for 46% (n = 27) of records.

Int. J. Environ. Res. Public Health 2020, 17, 7957

Table 1. Ticks submitted to Public Health England's Tick Surveillance Scheme from people with recent travel history outside of the UK between 2006 and 2018. The tick species, history of travel, number of records associated with each tick species (including confirmed (C) and possible (P) imported ticks) and number of each life stage identified are shown (no larvae were submitted).

Tick Species	Continent	History of Travel	No. Records	No. Confirmed/Possible Imported Records	Males	Females	Nymphs	Total Ticks
Amblyomma americanum	North America	USA	5	5/0	-	3	3	6
Amblyomma cajennense s.l.	North America	Mexico	1	1/0	-	1	-	1
Amblyomma species *	South America	Peru	1	1/0	-	-	1	1
Dermacentor andersoni	North America	USA/Canada	1	1/0	1	-	-	1
Dermacentor auratus	Asia	Nepal	1	1/0	-	-	1	1
Demucentor untutus		Sri Lanka	1	1/0	-	-	1	1
Dermacentor marginatus	Europe	Holland/Germany/France	1	1/0	-	1	-	1
Dermineenter mangimune		Italy	2	2/0	2	-	-	2
Dermacentor variablis	North America	USA	1	1/0	1	-	-	1
Haemaphysalis hystricis	Asia	Java, Indonesia	1	1/0	-	1	-	1
Hyalomma lusitanicum	Europe	Spain	1	1/0	1	-	-	1
Hyalomma truncatum	Africa	Botswana	1	1/0	1	-	-	1
Ixodes hexagonus	Europe	Madeira	1	0/1	-	1	-	1
ixoues hexagonus		Portugal	1	1/0	-	1	-	1
Ixodes holocyclus	Australia	Australia	2	2/0	-	1	1	2
Ixodes pacificus	North America	USA	1	1/0	-	-	1	1

Int. J. Environ. Res. Public Health 2020, 17, 7957

 Table 1. Cont.

Tick Species	Continent	History of Travel	No. Records	No. Confirmed/Possible Imported Records	Males	Females	Nymphs	Total Ticks
		Czechia	1	1/0	-	-	1	1
	Europe	Denmark	1	0/1	-	1	-	1
		France	10	6/4	-	3	10	13
Toro dos misiros		Germany	4	3/1	-	1	4	5
Ixodes ricinus		Ireland	3	1/2	-	-	4	4
		Italy	4	2/2	-	1	3	4
		Norway	2	2/0	3	1	1	5
		Poland	3	1/2	-	2	1	3
		Slovenia	1	1/0	-	-	1	1
		Spain	1	1/0	-	1	-	1
		Sweden	2	2/0	-	1	1	2
Ixodes species *	Europe	Belgium	1	0/1	-	1	-	1
Rhipicephalus appendiculatus	Africa	Eswatini	1	1/0	-	-	1	1
Rhipicephalus gertrudae	Africa	South Africa	1	1/0	1	-	-	1
Rhipicephalus sanguineus s.l.	Europe	Croatia	1	1/0	1	-	-	1
		Italy	1	1/0	-	-	1	1
	Total		59	45/14	11	21	36	68

^{*} indicates that the tick was damaged and could only be identified at the species level. Total numbers of records and ticks are shown in bold.



Figure 2. Map showing recent travel history of people submitting confirmed and possible imported tick records to Public Health England's Tick Surveillance Scheme between January 2006 and December 2018. The numbers of records received from each country are indicated by the size of the circle. Two records are not shown: one record that was acquired in either Holland, Germany or France; another record that was acquired in either the Canadian Rockies or north-western USA (see Table 1). Maps have been reproduced with permission from Ordnance Survey on behalf of Her Majesty's Stationery Office, © Crown Copyright and database right. 2020. All rights reserved. Ordnance Survey License number 100016969/100022432.

In total, 16 tick species belonging to six genera were identified. Two species endemic to the UK were received: *Ixodes hexagonus* and *Ixodes ricinus*. In addition, fourteen non-native species were identified: *Amblyomma americanum*, *Amblyomma cajennense*, *D. andersoni*, *Dermacentor auratus*, *D. marginatus*, *Dermacentor variablis*, *Haemaphysalis hystricis*, *Hyalomma lusitanicum*, *Hyalomma truncatum*, *Ixodes holocyclus*, *Ixodes pacificus*, *Rhipicephalus appendiculatus*, *Rhipicephalus gertrudae* and *Rhipicephalus sanguineus* s.l. The most common tick species received was *I. ricinus*; there were 30 confirmed records and 12 possible records, representing 54% of all records, followed by *A. americanum*, which represented 8% (n = 5) of records. Nymphs were the most common (53% of all ticks) life stage received, followed by females (31%) and males (16%); no larvae were received. The ecology of each tick species, including distribution, habitat, hosts and seasonality ispresented in Table 2. Whilst human biting by the majority of the received species is commonly reported, human infestation of *Hy. lusitanicum*, *Hy. truncatum* and *H. hystricis* is considered rare or occasional (Table 2).

All records involving the importation of species endemic to the UK (n = 35) were acquired in Europe (Figure 2): 21 records (60%) were definitively confirmed as imported, whilst 14 records (40%) were possible imports (Table 1). There was one confirmed import of *I. hexagonus* from Portugal and a possible import of *I. hexagonus* from Madeira. Six confirmed *I. ricinus* imports originated from France, three from Germany twoeach from Italy, Norway and Sweden and one each from Czechia, Ireland, Poland, Slovenia and Spain. In addition fourpossible imported records were received from France twoeach from Ireland, Italy and Poland and one from Germany.

Int. J. Environ. Res. Public Health 2020, 17, 7957

Table 2. Review of distribution, habitat and climate requirements, hosts, seasonality and commonality of human biting of each confirmed and possible imported tick species, along with the prevalence of the most common human pathogens detected in the focal tick species in the country where the tick was confirmed or suspected to have been acquired. References are mostly focused on countries listed in the history of travel butliterature searches were extended if there was no information available for the tick species in the country.

Tick Species	Distribution	Habitat and Climate Requirements	Hosts	Seasonality in Countries of Interest	Commonality of Human Biting
Ixodes ricinus (sheep or castor bean tick)	Most widespread tick across Europe alsopresent in parts of North Africa and Asia [24]	Deciduous and mixed forests, scrub, parks	Larvae: small mammals Nymphs: woodland and game birds Adults: large mammals, particularly deer [25–31]	Active year-round; peaks in larvae during June. Nymphs and females peak in May-June withsmaller peak in September [32]	Most common tick species found biting humans in Europe
Dermacentor marginatus (ornate sheep tick)	Found south of the Alps, stretching from Portugal across Southern Europe intonorthern Africa and mountain region of central Asia [23,33]	Dry habitats with sparse vegetation, e.g., alpine and forest steppes and semi-desert regions; tolerant to a wide range of environmental conditions [34–36]	Larvae: small mammals Nymphs: small mammals Adults: larger mammals including horses, cattle, dogs, deer, sheep [34]	Larvae: June and July Nymphs: July and August Adults: March–May, recommences in September & October [37,38]	Adults are known to bite humans [23,39]
Rhipicephalus sanguineus s.l. (brown dog tick)	Most widely distributed tick species in the world; more commonly associated with Mediterranean areas in Europe [40]	Capable of surviving in temperate regions. Can survive inside buildings, particularly those housing dogs but also residential homes	Dogs are the primary host of all stages, although immature stages can feed on rodents and small mammals	Active in temperate regions between spring and autumn [40]. Can be active year-round inside properties	Low probability of biting humans butcan be a problem in infested residential homes [39,41–44]
Hyalomma lusitanicum	Limited to a narrow strip from Sicily to Portugal and North Africa [45,46]	Adapted to meso-Mediterranean climate summer conditions [47]	Larvae and nymphs: Mediterranean rabbits (Oryctolagus cuniculus huxleyi) Adults: ungulates, insectivores, carnivores [45,48,49]	Adult questing begins in March, peaks May–July then declines witha small increase in September–October [47,50,51]	Human biting considered rare and accidental [49]

Table 2. Cont.

Tick Species	Distribution	Habitat and Climate Requirements	Hosts	Seasonality in Countries of Interest	Commonality of Human Biting
Amblyomma americanum (lone star tick)	Found in 39 states plus the District of Columbia in USA, stretches across eastern, south-eastern, mid-western and north-eastern USA [52,53]	Wooded areas with thick underbrush aswell as scrub and meadows, where the primary host (white-tailed deer, <i>Odocoileus virginianus</i>) resides	Larvae and nymphs feed on small- and medium-sized hosts, adults feed on medium- and large-sized mammals. Host range is vast butwhite-tailed deer are the most important host and can feed all life stages—recent expansions in lone star tick populations due to increase in white-tailed deer populations [52]	Larval activity is highest during the summer, nymph and adult activity peaks April–June and then declines [52]	One of the most aggressive human-biting tick species, often the most abundant human biting species in many studies [52–56]
Dermacentor variabilis (American dog tick or wood tick)	Found in USA from Gulf of Mexico to New England and through the mid-western states to the east of the Rocky Mountains withan independent population also found in California. Also present in parts of Mexico	Found in areas with limited tree cover likebrushy field habitat and scrubland [57]	Larvae and nymphs: small mammals Adults: dogs, cervids, ruminants	Two peaks in adult questing: mid-April to late May; July [58]	Adults are the only stage known to bite humans [59]
Dermacentor andersoni (Rocky Mountain wood tick)	Found throughout the western United States and is established in at least 14 states [60]	Found in semiarid and mountainous areas with woodland, scrub and grassy areas [60–62]	Larvae and nymphs: small mammals, e.g., rats, squirrels, chipmunks Adults: larger hosts such as cattle, deer, horses and elk [60].	Larvae and nymphs occur between March and October: larval peaks occur from June to July whilst nymphs peak from May to June [60]. Adults are active between February and November withgreatest abundance recorded from March to April [60]	Larvae and nymphs rarely bite humans

Table 2. Cont.

Tick Species	Distribution	Habitat and Climate	Hosts	Seasonality in Countries of Interest	Commonality of Human Biting
Ixodes pacificus (western blacklegged tick)	Found in western parts of the United States, particularly Washington, Oregon, California, Arizona, Nevada and Utah [63]	Requirements More nymphs found in warmer and drier hardwood-dominated woodlands compared with cooler and more humid woodlands dominated by redwoods [64]. Survival is limited by summer drought conditions; larvae that hatch in the early summer remain in behavioral diapause until the following spring to avoid desiccation [64,65]	Larvae and nymphs: birds (migratory and non-migratory), rodents and lizards Adults: birds and lizards aswell as larger mammals such as cats dogsand ruminants [66–68]	Larval and nymphal infestation of birds and lizards peaks in April and May [66,69,70]. Questing nymphs active March–September, peaking in May and June, whilst adults are active October–June with peaks in November–January and March [71,72]	Primary human-biting species in California [71]
Ixodes holocyclus (paralysis/scrub typhus tick)	Found along the East Coast of Australia fromnorthern Queensland down to Victoria [73]	The limited distribution is thought to be driven by climatic conditions, particularly humidity andit is often found in wet forested (scrub) areas [73]	Generalist feeder hasbeen recorded infesting mammals and birds [74]. In south-eastern Queensland, northern brown bandicoots (Isoodon macrourus) and long-nosed bandicoots (Perameles nasuta) are required for I. holocyclus populations to persist from one season to the next [74]	Nymphs most abundant between April and September; female adults peak between October and December [74]	Known to frequently bite humans [75]
Ixodes hexagonus (hedgehog tick)	Widespread distribution across temperate Europe and North Africa [34]	Nidicolous, found in dark humid places e.g., nests/burrows [76,77]; distribution is not limited by or dependent upon ambient climatic conditions	Hedgehogs are the most common hosts butalso found on a range of mammals [34]	Infestations occur throughout the year [78–81]	Human biting has been reported [17,82–86]
Amblyomma cajennense s.l.	Range spans from parts of southern USA, across Central America theCaribbean, extending into South America to parts of northern Argentina [87]	Found in a wide range of habitats including dry grasslands, tropical forests and highlands of the Peruvian Andes [87]	Giant anteaters wildpigs, tapir, water buffalo, dogs, horses, capybara and small mammals are common hosts [88–93]	Larvae most active between August and February peaknymph activity between December and March, adult activity increases when daily mean temperature reaches 20 °C, peaking between February and September [89,90,94–97]	All life stages bite humans andin South America it bites humans almost twice as frequently as all other tick species combined [98,99]

Table 2. Cont.

Tick Species	Distribution	Habitat and Climate Requirements	Hosts	Seasonality in Countries of Interest	Commonality of Human Biting
Hyalomma truncatum	Widely distributed throughout sub-Saharan Africa	Unknown	Parasitises a wide range of mammals, including rodents, lagomorphs, canids, ungulates and carnivores [9,100,101]	Unknown	Human infestation is rare [102] butthere are at least two reports of this tick species found attached to returned travellers [7,9]
Rhipicephalus gertrudae	Found extensively across South Africa withits distribution reaching the southern province of Western Cape aswell as northwards into Namibia and eastwards to the Free State province [103]	Unknown	Larvae and nymphs: small mammals [104,105] Adults: cattle, goats, sheep, horses, zebra, antelopes, primates, dogs cats[22,106–110]	Highest numbers of biting ticks occur between September and February [103,107,111]	Adults most commonly bite humans [111]
Rhipicephalus appendiculatus (brown ear tick)	Widely distributed between southern Sudan and the east coast of South Africa witha noticeable absence in the Horn of Africa [112]	Unable to survive on open plains andpredominantly found in woodland and savannah with good vegetation cover butcan disappear following overgrazing [111]	Larvae and nymphs: small antelopes and hares Adults: large ungulates including cattle, buffalo, wildebeest [113]	Larvae active late summer to winter (April-August) Nymphs active in winter and early spring (July to October) Adults active in rainy season (December to March) [113]	All stages are known to bite humans [111]
Haemaphysalis hystricis	Distributed from the eastern Himalayas to India, through the coasts of Vietnam and China to Taiwan and Japan andfurther south including Thailand, Malaysia and Indonesia [114–119]	Unknown	Commonly infests many hosts including cats, dogs, Japanese marten wildboar, wildcats [115,117,120,121]	Unknown	Occasionally bite humans
Dermacentor auratus	Widespread distribution across much of Asia, including Borneo, China, India, Java, Laos, Malaysia, Myanmar, Nepal SriLanka, Sumatra, Thailand and Vietnam [39,122–132]	It is a common tick species found in forests below 400 m altitude, although there are data of tick bites occurring at higher altitudes [131]	Larvae infest rodents; adults found on wild pigs, cattle, buffalo, deer dogsand birds [125,126,129,131–133]	Unknown	Intra-aural infestation in particular has been reported [122,125,126,128,129,131,134]

3.2. Literature Review of Pathogens Detected in Ticks in the Native Country

Literature reviews for pathogens infecting tick species in the country of origin highlighted nine of the received species (*Ixodes ricinus*, *D. marginatus*, *R. sanguineus*, *Hy. lusitanicum*, *A. americanum*, *D. variabilis*, *D. andersoni*, *I. pacificus*, *I. holocyclus*), comprising 81% of all records areknown to transmit at least one pathogenic organism to humans; the most important pathogens in terms of human health in the country where the imported tick was likely acquired are shown in Table 2.

3.2.1. Europe—Ixodes Ricinus

The most common pathogen detected in *I. ricinus* in Europe is the spirochete *Borrelia burgdorferi* sensu lato (s.l.) thecausative agent of Lyme borreliosis (prevalence 0.06–46.6% [135]). Despite only being prevalent in the Northern Hemisphere Lymeborreliosis is the most common tick-borne disease in the world [24]. It is transmitted by ticks belonging to the *I. ricinus* complex across Europe and North America aswell as in parts of northern Africa and Asia. The true incidence of infection in Europe is difficult to quantify, however asLyme borreliosis is not a notifiable disease. In Europe theprimary vectors of Lyme borreliosis are *I. ricinus* and *I. persulcatus* butlaboratory evidence suggests that *I. hexagonus* is also an efficient vector of *B. burgdorferi* s.l. [136,137].

There are multiple genospecies of *B. burgdorferi* s.l. circulating in Europe: *Borrelia afzelii, Borrelia* bissettiae, Borrelia bavariensis (formerly B. garinii OspA serotype 4), Borrelia burgdorferi sensu stricto (s.s.), Borrelia carolensis, Borrelia finlandensis, Borrelia garinii, Borrelia lusitaniae, Borrelia spielmanii, Borrelia turdi and Borrelia valaisiana [135,138]. Throughout Europe theprevalence of genospecies varies geographically [138], which is likely driven by genospecies associated with different reservoir hosts. For example, whilst B. afzelii is associated with small mammals [139–141], ground-foraging birds are competent reservoirs of B. garinii and B. valaisiana butare not able to maintain B. afzelii [142–144]. Not all genospecies are pathogenic and in Europe at least five genospecies are well recognised as being pathogenic to humans—B. afzelii, B. bavariensis, B. burgdorferi s.s., B. garinii and B. spielmanii [145–147]. A meta-analysis investigating the prevalence of genospecies in questing *I. ricinus* across Europe found that B. afzelii (46.6%) and B. garinii (23.8%) were the most common genospecies, followed by B. valaisiana (11.4%), B. burgdorferi s.s. (10.2%), B. lusitaniae (7.0%), B. bavariensis (2.0%), B. spielmanii (1.7%), B. finlandensis (0.2%) and B. bissettiae (0.06%) [135]. Focusing on I. ricinus in the countries where recorders had a recent history of travel and subsequent tick bite themost common genospecies reported in ticks, B. afzelii, B. burgdorferi s.s. and B. garinii havebeen recorded from all focal European countries. In addition, other pathogenic species have been found: B. bavariensis in I. ricinus from Czechia and Germany and B. spielmanii in Czechia, Denmark, France, Germany, Poland and Sweden.

Tick-borne encephalitis (TBE) is a severe neurological disease caused by a flavivirus transmitted by ticks in parts of Europe and Asia [148], although infection can be acquired to a lesser extent through consumption of raw milk from infected animals [149]. Tick-borne encephalitis virus (TBEV) is the most important tick-borne arboviral disease in Europe; it is endemic in 27 European countries and is expanding northwards and to higher altitude as a result of numerous factors including warming temperatures [150-152]. Adults were traditionally considered most at risk of TBE infection as the prognosis worsens with age, whereas children suffer milder symptoms [153–157]. A recent review of data from the European Centre for Disease Prevention and Control (ECDC), however, found that there is a greater risk of long-term cognitive impairment in children following TBE infection [158]. There are three subtypes of TBEV: European, Siberian and Far Eastern. The European subtype is transmitted by I. ricinus, whilst the other two are transmitted by I. persulcatus [155] (see Lindquist & Vapalahti, 2008). The virus is transmitted to a susceptible tick when it feeds adjacent to an infected tick aprocess known as co-feeding [159] andkey wildlife hosts are required for TBEV to persist in a population [160,161]. In southern Germany and Slovakia mostTBE cases occurred during June Julyand August [162,163], which is likely linked to increased human activity during the summer period, despite reduced tick numbers [164]. Between 2012 and 2016, 12,500 cases of TBE were reported from 23 European countries and of these, Czechia and Lithuania accounted for 38.6% of all reported

cases [165]. Prevalence < 0.1–5% [166]. In the focal countries for the current study TBEVhas been found in *I. ricinus* in all countries apart from Ireland and Spain (Table 2) and the highest number of cases of TBEV have been reported from Czechia [165]. To date fiveimported cases of TBE have been reported in the UK, although the country where the infection was acquired is unknown [165].

3.2.2. Europe: *Dermacentor marginatus* in France, Germany and Italy

Tick-borne lymphadenopathy (TIBOLA) or *Dermacentor*-borne necrosis erythema lymphadenopathy (DEBONEL) is caused by three species of *Rickettsia*: *R. slovaca*, *R. raoultii* and *R. rioja* [167–172]. Cases of infection have been reported from Bulgaria, France, Hungary, Italy, Portugal and Spain [168,173–179]. Compared to Lyme borreliosis, TIBOLA/DEBONEL is a relatively rare and mild infection butawareness of symptoms is still important [180]. In Europe, *D. marginatus* is considered to be the main vector of TIBOLA/DEBONEL and the peak in diagnosed cases coincides with the peak in adult activity [175]. The prevalence of *R. slovaca* infecting *D. marginatus* is between 15.7% and 50% [33,173,181–185], whilst the prevalence of *R. raoultii* is lower at 8.3–8.4% [33,184]. Of the focal countries in the current study, infection with *R. slovaca* has been reported in *D. marginatus* from France, Germany and Italy and *R. raoultii* has been detected in *D. marginatus* from Italy.

3.2.3. Europe: Rhipicephalus sanguineus s.l. in Croatia and Italy

The most frequent rickettsiosis in Europe is Mediterranean spotted fever (MSF alsonamed fièvre boutonneuse), caused by Rickettsia conorii and cases have been reported from much of Western and Southern Europe aswell as parts of Eastern Europe [175]. Mortality can occur and fatality rate of 32.3% of hospitalized patients with severe morbidity in southern Portugal has been reported [186]. As Rh. sanguineus is thought to act as both the vector and reservoir of the rickettsia somemammals present in the Mediterranean may also be reservoir hosts [175]. Although Rh. sanguineus has a low probability of biting humans mostcases of infection are diagnosed in the spring and summer when Rh. sanguineus activity peaks [175]. In an MSF-endemic region in Spain, however onlyone out of 4049 ticks was infected [187]. Very low infection prevalence (<1%) has been reported [188,189] butcan be higher (see [190]) As other Rickettsia species were more prevalent theauthors suggested that many of the cases that had been attributed to MSF may have actually been caused by other Rickettsia species [187]. For example, *Rickettsia conorii israelensis*, which has also been detected in European *Rh*. sanguineus populations, results in a sudden fever, accompanied by headache, rash andgastrointestinal symptoms [191]. Mortality has been reported in less than a third of patients [191] butis higher than that reported for MSF, suggesting R. conorii israelensis is more virulent than R. conorii [192]. Another Spotted Fever group rickettsia which has been detected in multiple species belonging to the Rhipicephalus genus [193], including Rh. sanguineus, is R. massiliae. Symptoms include fever, headache, muscular pain rashand eschar [190,194–196] and Italy and France, R. massiliae is considered the causative agent of a MSF-like illness in patients [190,194–196]. During the current study twoimported Rh. sanguineus were received; one from Italy and one from Croatia. Whilst to date, there is no information on pathogen prevalence in Rh. sanguineus in Croatia, R. conorii, R. conorii israelensis and R. massiliae have all been detected in *Rh. sanguineus* from Italy.

3.2.4. Europe: *Hyalomma lusitanicum* in Spain

Crimean Congo haemorrhagic fever virus (CCHFV) is caused by a member of the Nairovirus genus and is transmitted via bites from *Hyalomma* species (particularly *Hy. marginatum marginatum* but also *Hy. lusitanicum*) orfollowing contact with infected body fluids or tissues [197]. To date twostudies have detected CCHFV in *Hy. lusitanicum* at a prevalence of 3.2% [45,198] feeding from red deer (*Cervus elaphus*) in Spain between 2010 and 2015, suggesting that CCHFV is circulating in southwestern Europe [45,198] butwhether *Hy. lusitanicum* is a vector of the virus remains unclear. During 2016 twoautochthonous cases of CCHFV were reported in Spain [199], followed by a subsequent fatal case reported in August 2018 [200]. Until the end of August 2020, there have been three autonomous

cases of CCHF in Spain and Bulgaria, including a fatality [201–203]. To date, there have been three CCHFV cases reported in the UK. First in1997 assuspected case of CCHFV was diagnosed in a UK resident recently returned from Zimbabwe [204]. In 2012, there was a fatal CCHFV case in a UK national returning from Afghanistan [205]. Finally inJune 2014, there was a case in a UK national that had received a tick bite whilst in Bulgaria [206].

3.2.5. North America: Amblyomma americanum in USA

In the USA Lymeborreliosis is caused by the genospecies *B. burgdorferi s.s.*, and in the north-eastern USA where Lyme borreliosis is most prevalent, *I. scapularis* is the primary vector, whilst in the western states thebacteria are vectored by *I. pacificus*. Infection of *A. americanum* with *B. burgdorferi* s.s. has been reported from a number of states [207–210] yetit is not considered a competent vector [211–213]. Furthermore Lymeborreliosis-like symptoms have been associated with bites from *A. americanum* in some southern states when no trace of antibodies to *B. burgdorferi* s.l. have been detected in patients [214, 215]. The spirochete responsible has been identified and termed *'Borrelia lonestari'* [216] andhas been detected at low prevalence (0.35–9.1% [216–227]) in questing *A. americanum* from multiple locations [216–227]). The resulting illness is named Master's disease or southern-tick-associated rash-illness (STARI) andis clinically indistinguishable from the early stages of Lyme borreliosis [228]; however thepublic health significance of *B. lonestari* remains unclear.

Ehrlichia chaffeensis and E. ewingii are gram-negative bacteria that are the causative agents of human ehrlichiosis [229,230]. Whilst ehrlichiosis caused by E. chaffeensis is considered to be underreported and probably as prevalent as Rocky Mountain spotted fever (see below) lessthan 20 cases of *E. ewingii* have been documented and are mostly reported in immunosuppressed patients [52,231]. The prevalence of E. chaffeensis in ticks is higher (0.6–19%) [217,220–222,232–239] than E. ewingii (0.24–7.1%) [217,220–222,233–236,238]. It is thought that *E. chaffeensis* and *E. ewingii* are maintained in cycles involving a wide variety of competent vertebrate reservoir hosts such as white-tailed deer and A. americanum as the primary vector [52,235,240]. Laboratory experiments have demonstrated transmission of E. ewingii from A. americanum to dogs, highlighting the importance of A. americanum in maintaining and transmitting ehrlichiae to hosts [241]. In 2006, Panola Mountain Ehrlichia (PME), caused by a disease agent similar to Ehrlichia ruminantium wasdiscovered in a goat from Panola Mountain State Park in Georgia [242] and was later associated with a report of human illness following a bite from an Amblyomma nymph also acquired at the State Park [243]. Furthermore, A. americanum infected with PME have been collected from ten states on the east coast of the United States between 1998 and 2006, suggesting the infection has an extensive distribution throughout the A. americanum range and has not been recently introduced [244].

3.2.6. North America: Dermacentor variabilis in USA

Dermacentor variabilis is a primary vector USA [245,246]. Caused by the bacteria Rickettsia rickettsii RMSFis the most common tick-borne rickettsial disease in the USA [247]. Severe illness can develop if symptoms remain untreated andfatalities are possible [247]. Low prevalence rates of less than 2% in D. variabilis populations [248,249] have led some authors to question whether other tick species may play a more important vectoral role than D. variabilis [249]. For example, Rh. sanguineus was the cause of an outbreak of RMSF in eastern Arizona [250]. Furthermore, whilst A. americanum historically has not been considered to be a major vector of RMSF [251] muchof the distribution overlaps with D. variabilis [252] andas both species feed on similar hosts, there is potential for A. americanum larvae to become infected with R. rickettsii either whilst feeding on an infected host or cofeeding next to an infected D. variabilis [252]. Under laboratory conditions, A. americanum has acquired the infection from guinea pigs, maintained infection throughout moulting and transmitted the bacteria to susceptible hosts during subsequent feeding, suggesting that under laboratory conditions, A. americanum is capable of acquiring, maintaining and transmitting R. rickettsii [252]. To date, however, there is no evidence to suggest that A. americanum is a competent vector of R. rickettsii in the USA. Rickettsia

montanensis is another spotted fever group Rickettsia that has been detected in *D. variabilis*. Studies in the USA and Canada have found a mean infection prevalence of *D. variabilis* with *R. montanensis* of 1.2–10.5% [56,59,248,253–259]. In addition achild from Georgia USAdeveloped an afebrile rash illness after being bitten by an individual *D. variabilis* infected with *R. montanensis*, suggesting the possibility that *R. montanensis* may be a human pathogen [260].

3.2.7. North America: Ixodes pacificus in USA

Ixodes pacificus is the primary vector of Lyme disease in western USA [110] and the highest regional incidence occurs in north-western California, where dense oak woodlands support high densities of *I. pacificus* [261]. In California, cases of Lyme borreliosis peak between May and July, following the nymph peak between April and June, suggesting nymphs are responsible for most cases in California [72].

Borrelia miyamotoi belongs to the relapsing fever group of Borrelia andis distinctly related to B. burgdorferi s.l. It is transmitted by the same tick species as B. burgdorferi s.l., namely I. ricinus and I. persulcatus in Europe and I. scapularis and I. pacificus in the USA. Three distinct genotypes of B. miyamotoi have been identified in the USA, Europe and Japan [262–264] and all three groups include strains that are pathogenic to humans [265]. Unlike the spirochetes that cause Lyme borreliosis, B. miyamotoi spirochetes are able to be maintained in ticks via both transstadial and transovarial transmission [266,267]. The first human cases of infection with B. miyamotoi were reported in Russia in 2011 [268] and further cases have been reported from elsewhere in Europe (Netherlands and France) and the USA [269–272]. A study of people bitten by ticks which tested positive for B. miyamotoi who went on to develop clinical disease estimated transmission efficiency in humans to be 8.3% (2/24 patients), compared with 4.4% (3/68) of humans bitten by ticks infected with B. burgdorferi s.l. who went on to develop symptoms [273,274]. In the USA, white-footed mice (*P. leucopus*) are important hosts of the bacteria [266] and a high prevalence of infection (58%) has been detected in wild turkeys (Maleagris gallopavo) [275]. Prevalence rates of infection in ticks are significantly lower (0.4–5% [276–282]) than B. burgdorferi s.l. (0.2–24.7% [64,261,276,277,279,281,283–295]), although a comparable prevalence of B. miyamotoi and B. burgdorferi in adult I. pacificus in California suggests that there is a similar risk of exposure to both species [276]. As well as I. pacificus from USA, B. miyamotoi infection has been reported in I. ricinus from Czechia, France, Germany, Norway, Poland, Spain and Sweden (Table 2). In Europe bankvoles (Myodes glareolus) and yellow-necked mice (Apodemus flavicollis) are reservoirs for B. miyamotoi [263,296,297] and whilst bacteria have been detected in engorged I. ricinus feeding on wild boar (Sus scrofa) roedeer (Capreolus capreolus) and a blackbird (Turdus merula) itis unknown whether these species are reservoir hosts of *B. miyamotoi* [298].

Anaplasma phagocytophilum is an obligate intracellular bacterium that is responsible for causing human granulocytic anaplasmosis (HGA) amoderate flu-like illness in humans. In the USA HGAis the second most important tick-borne disease after Lyme borreliosis andis a notifiable infection, although many infections may result in minimal or no clinical manifestations [299]. Small mammals, which feed immature tick stages arethe primary reservoirs for *A. phagocytophilum* [300–302]. In eastern USA themain vector of HGA is *I. scapularis*, whilst *I. pacificus* is considered the main vector in western parts of the USA [303,304]. Further theprevalence of *A. phagocytophilum* in *I. pacificus* is significantly lower (0–11%) than the prevalence in *I. scapularis* (0-51%) [277,283,285,288,295,300,305–307].

3.2.8. North America: Dermacentor andersoni in USA/Canada

Along with being a primary vector of RMSF [308], *D. andersoni* is also the principal vector of Colorado tick fever virus, which is found in western USA. The disease is considered under-reported asnon-specific symptoms may be misdiagnosed as other infections including RMSF [309]. Whilst producing a febrile illness in humans thevirus does not cause illness in its natural reservoir hosts. It was first detected in Montana, where adult *D. andersoni* were found to be infected with the virus andinfection was also present in ground squirrels thepreferred host for immature stages of *D. andersoni* [310,311]. A prevalence of 6.6%

to 21% in *D. andersoni* has been reported [312,313]. Further, clinical cases of Colorado tick fever peak between May and July, concurrently with the peak of ticks [314].

Dermacentor andersoni has been associated with tick paralysis withrapidly ascending paralysis occurring five to seven days after tick attachment. The tick often attaches to the head andso can be hidden by hair; removal of the tick leads to an immediate improvement in symptoms [315]. In a review of 33 cases of tick paralysis that were reported to the Washington State Department of Health between 1946 and 1996, *D. andersoni* was identified as the culpable species [316].

3.2.9. North America: Amblyomma cajennense Sensu Lato in Mexico

There are no studies published to date reporting the association of *A. cajennense* with human disease in Mexico. In South America, however, *A. cajennense* is considered to be the most important vector of *R. rickettsii* (the causative bacteria of RMSF alsocalled Brazilian spotted fever (BSF) in Brazil [99]). Considered to be the deadliest of the spotted fevers in the world BSFhas a fatality rate of 31.5%, compared with 5–10% fatality for RMSF in the USA [317,318]. A laboratory experiment found that it was possible for transovarial transmission from a female *A. cajennense* to her offspring butonly some of the eggs became infected, suggesting that *R. rickettsii* is not able to be maintained by transovarial transmission only [319]. In addition, *A. cajennense* larvae that fed on guinea pigs experimentally infected with *R. rickettsii* were less susceptible to infection with *R. rickettsii* than other tick species [320].

3.2.10. Australia: Ixodes holocyclus

Ixodes holocyclus is responsible for the majority of (though not all) cases of tick paralysis in Australia [316]. Tick paralysis is a rare but potentially fatal condition. Between 1904 and 1945, there were 20 fatalities reported in Australia due to tick paralysis with70% of all deaths being children butsince 1945, there have been no further fatalities [321]. Ticks do not secrete detectable levels of the paralysis-inducing toxin until three days of feeding on a host [73]. As such, clinical signs of tick paralysis do not become apparent until three days following a tick bite [322,323]. Prompt tick removal is an important part of patient treatment [324], although, following removal of *I. holocyclus*, there can be a deterioration in the health of the patient [321]. Whilst nymphs may cause a mild paralysis, females are responsible for the most severe symptoms and a bite from a single infected female tick is sufficient to cause paralysis [74,322,325]. Cases of paralysis occur year-round butare most common in spring and summer [323]. It appears that children up to 5 years old are most affected [74]. Furthermore, *I. holocyclus* is usually found on the scalp and paralysis is less likely to occur if ticks are attached to other parts of the body [74].

3.2.11. Africa: Hyalomma truncatum in Botswana

Whilst there have been no published studies looking at the pathogen prevalence in *H. truncatum* collected in Botswana to date, horizontal transfer of CCHFV from larval *H. truncatum* to laboratory mice has been reported [326] andtransovarial and sexual transmission of CCHFV in *H. truncatum* has been seen in laboratory studies [327,328]. In addition, whilst Rift Valley fever virus is predominantly associated with mosquito transmission or contact with infected blood/body fluids ithas been suggested that *H. truncatum* is a possible vector of the virus [329,330]. Horizontal and transstadial transmission has been demonstrated [329] andthe geographical distribution of *H. truncatum* correlates with the incidence of Rift Valley fever virus [330]. *Rickettsia aeschlimannii* and *R. africae* have been detected in *H. truncatum* [331–335], although there does not appear to be confirmed evidence of transmission from laboratory experiments. The prevalence of *R. aeschlimannii* in *H. truncatum* was lower compared to other *Hyalomma* species, suggesting that *H. truncatum* is not the principal vector of the bacteria [331].

3.2.12. Africa: Rhipicephalus gertrudae in South Africa

To date, there is no information on whether *Rh. gertrudae* is involved in the transmission of any pathogens to humans in South Africa or further afield.

3.2.13. Africa: *Rhipicephalus appendiculatus* in Eswatini

Although *Rh. appendiculatus* is a known vector of diseases affecting cattle in Southern Africa forexample EastCoast fever [336], there have been no published studies to date to suggest it to vector any diseases which can be transmitted to humans in Eswatini or further afield.

3.2.14. Asia: Haemaphysalis hystricis in Java, Indonesia

Whilst to date there have been no studies published looking at the pathogen prevalence in H. hystricis collected specifically in Java or across Indonesia thefindings of studies that have been conducted in other Asian countries suggest that H. hystricis could act as a vector in Java. Haemaphysalis hystricis is considered to be one of the most important vectors of Rickettsia japonica thebacteria responsible for Japanese spotted fever [120] and infected ticks have been found in Japan and China [337,338] (reviewed in Mahara, 1997; Lu et al., 2017). Following the diagnosis of a patient with Japanese spotted fever in Fukuoka Prefecture in Japan aninvestigation of ticks found in the area where it was thought that the patient most likely acquired the tick bite found larvae were infected with R. japonica [339]. Similarly in the Hubei province of China, R. japonica was detected in a single H. hystricis [338]. Focusing on other infections, Coxiella species have been detected in H. hystricis in Thailand, although infection prevalence in *H. hystricis* is thought to be lower than in other *Haemaphysalis* species also present in Thailand [340]. Following on from this astudy of *H. hystricis* from Malaysia suggested the possibility of infection with C. burnetii based on the phylogenetic clustering of the bacteria [341]. Finally, whilst Borrelia burgdorferi s.l. has not been detected in H. hystricis from China [115], Borrelia sp. closely related to the relapsing fever group (e.g. Borrelia miyamotoi) have been detected in H. hystricis from Malaysia [116], although it should be noted that the bacteria was only detected in a single tick.

3.2.15. Asia: Dermacentor auratus in Nepal and Sri Lanka

To date, there has been no investigation into pathogens found in D. auratus in either Nepal or Sri Lanka. Kyasanur forest disease and Lanjan virus have been isolated in questing D. auratus in India and Malaya, respectively [131,342], although it is unclear whether D. auratus also transmits the pathogens. A study of ticks removed from wild boar in Thailand found D. auratus to be infected with Rickettsia raoultii (3/11 = 27%) and Francisella-like endosymbionts (2/11 = 18%), whilst Coxiella bacteria were also investigated but not detected [132].

4. Discussion

Between January 2006 and December 2018, Public Health England's Tick Surveillance Scheme (TSS) received 59 records from humans with recent travel history, comprised of 66 individual ticks belonging to 16 tick species that were associated with travel outside of the UK. Records were submitted by people with recent travel history to 25 different countries with70% of confirmed and possible imported records being associated with travel to Europe. Although less than five percent of total records received by the TSS are imported ticks [18] thefindings suggest that UK travellers are exposed to a variety of tick species when abroad. Whilst it was not possible to test the ticks for pathogens in the current study, literature reviews suggest that just over half (9/16) of the received tick species are known to vector at least one tick-borne pathogen in the country of acquisition, suggesting that travellers exposed to ticks may be at risk of being bitten by a species that is a known vector, which could have implications for the transmission of novel tick-borne diseases to UK travellers.

Whilst there are several records of non-native ticks entering the UK previously on animal hosts, including dogs [44,343–352], horses [353], migratory birds [354,355] and reptiles [356–358] thecurrent study is an extensive report of tick exposure to UK travellers submitted to the TSS. Reports of two records included in the current study have been published previously [6,359]. First afemale *I. holocyclus* was detected on a traveller recently returned from Australia who presented with swelling at the bite site and was prescribed antibiotics as an infected mole or wart was suspected [6]. In the second

case afemale *D. marginatus* was received from a patient with no history of travel experiencing swelling, swollen glands and flu-like symptoms [359]. As *D. marginatus* is not endemic in the UK, further investigations revealed that the patient had had contact with several European visitors in the month preceding the bite anda family member had driven her car through Holland, Germany and France the week before soit was suggested that the tick was imported by one of the travellers [359]. Such an event highlights the importance for clinicians to obtain substantial information from patients presenting with tick bites. Investigations should include questions regarding extended travel history, including several weeks prior to tick removal asseveral non-native tick records were received where the date of removal was several weeks after the person had returned from abroad. In addition, questions on contact with visitors who have recently been overseas should also be considered.

Some of the imported species received by the TSS have been reported as imported in the UK previously. Two days after returning from a two-month trip to Missouri an84-year-old male presented to his general practitioner in Northern Ireland andan almost fully fed female A. americanum was detected [8]. Another study described 16 tick species removed from UK domestic and international travellers; six non-native species detected in the paper are reported in the current study (A. americanum, A. cajennense, D. variabilis, D. marginatus, I. scapularis and Rh. sanguineus), whilst six additional non-native species were also described (A. maculatum, A. hebraeum, A. variegatum, H. concinna, Hy. marginatum marginatum and Hy. marginatum rufipes) aswell as four species acquired within the UK (D. reticulatus, H. punctata, I. ricinus and I. hexagonus) [13]. Whilst I. ricinus was the most common species detected, 25 of 28 records were acquired within the UK, whereas of tick records acquired from outside of the UK themost common species detected was D. marginatus (four records), acquired in France, Italy, Greece and Romania [13]. Other published studies have described non-native tick species detected on imported dogs in the UK, including Hy. lusitanicum [346], D. variabilis [351], I. holocyclus [343] and Rh. sanguineus [351]. Four of the species reported in the current study, however havebeen detected on UK travellers for the first time: H. hystricis, Hy. truncatum, Rh. appendiculatus and Rh. gertrudae. As imported tick species can transmit a range of pathogens UKtravellers may be exposed to novel pathogens which are unfamiliar to public health professionals in the UK. As such, issuing appropriate public health guidance on tick awareness and avoidance prior to travel could reduce the risk of acquiring tick bites and subsequent tick-borne infections whilst overseas.

Confirmed and possible imported ticks were removed from people with recent travel history in all months apart from March. Sixty-three percent of records detailed ticks removed between April and July, which may reflect travelling activity asmost travel abroad by UK residents occurs during these months [360]. Seasonality of adult and nymph *I. ricinus* in the UK and Europe is highest between April and June [361–363], however soit is also possible that the peak in imported ticks between April and July may be a consequence of 54% of all records being *I. ricinus*. It should be noted, however that 20% of imported records were received between November and February when *I. ricinus* activity is at its lowest in the UK [18]. The results from the current study suggest that the risk of being bitten by a tick whilst travelling occurs throughout the year butis likely to vary depending on travel destination soadvice on tick awareness, removal and tick-borne infections should be provided to travellers year-round with special attention paid during late spring and early summer when tick bite risk is highest in the most common destinations for UK travellers.

Seventy percent of ticks received by the TSS from people with a recent travel history were associated with recent travel to Europe andas Europe is a common destination for UK travellers therisk of ticks and tick-borne diseases being imported from Europe is higher. The predominant species received from people recently returned from Europe was *I. ricinus*, which is also the most common tick species found throughout the UK [18]. In Europe, *I. ricinus* is the predominant vector of *B. burgdorferi* s.l. thecausative agent of Lyme borreliosis, which is the most common tick-borne infection in Europe with approximately 65,000 cases reported annually between 1987–2006 [364–366]. Overall thehighest incidence of Lyme borreliosis has been reported in eastern and central Europe anddecreases from east to west, although local incidence can vary [135,367–371]. Lyme borreliosis should be considered in

patients returning from Europe with a history of tick bites yetwhether there is an increased or reduced risk of UK travellers being bitten by *B. burgdorferi*-infected *I. ricinus* in European countries compared with the UK will be dependent upon the country of travel. As the incidence of Lyme borreliosis is higher in eastern and central Europe [135,367–371], travellers to these regions in particular should be made aware of the increased risk and symptoms of Lyme borreliosis.

Along with Lyme borreliosis TBEshould also be considered in patients presenting with symptoms and a history of tick bites following travel to Europe. It is the second most important tick-borne disease transmitted by I. ricinus in Europe andto date, infected I. ricinus have been detected from nine of the eleven European countries where recorders had a recent history of travel and subsequent tick bite. Several cases of travellers contracting TBE whilst abroad have been previously reported. Five imported cases of TBE were reported in the USA between 2000 and 2009 from patients who had travelled to Czechia, Sweden, Russia and China and four out of five of the patients described having multiple tick bites whilst abroad [372]. Between 2012 and 2016 fiveimported cases of TBE were described in the UK, although the country of origin was unknown [165]. In 2011 twoDutch travellers returning from Austria were diagnosed with TBE; and similarly to the previous study bothpatients reported tick bites [373]. In parts of Austria TBEVis endemic and the risk of a tourist acquiring the infection after spending four weeks in an endemic region is estimated as 1 in 10,000 and following this itis estimated that 60 travel-associated (i.e., exported) cases of TBE were likely to occur over the whole summer period [374]. As the popularity of outdoor activities such as hiking and biking increases, people may be more exposed to tick bites than in the past [375]. The risk of acquiring TBEV infection when abroad, therefore, should be made clear to travellers from non-endemic regions [375]. Furthermore theincidence of TBE is expanding northwards to higher latitudes and altitudes [150–152] and could result in increased human exposure to infected ticks in the future.

Whilst Lyme borreliosis is the most important tick-borne infection in Europe and North America itis important to remember this is not the case in other countries sopatients with travel history to other continents may present with different symptoms. It is also important to consider that the same organism may have a different pathogenicity in a different country. An example of this is the causative agent of human granulocytic anaplasmosis (HGA), *Anaplasma phagocytophilum*. In the USA HGAis a notifiable infection andthe second most important tick-borne disease (after Lyme borreliosis) withincreasing infection incidence from 1.4 cases per million persons in 2000 to 17.9 cases per million persons in 2017 [376]. In comparison only70 cases have ever been reported in Europe and are sporadic [377,378]. It is unclear whether the small number of confirmed cases in Europe is due to under-reporting, under-diagnosis or low pathogenicity of *A. phagocytophilum* strains circulating in Europe [299].

5. Conclusions

Since 2006, 16 tick species removed from humans with a recent travel history were submitted to the TSS. Furthermore, literature reviews suggested that nine of the received species are known to vector at least one organism that is pathogenic to humans in the country of travel. Such findings suggest that travellers exposed to ticks may be at risk of being bitten by species known to transmit at least one pathogen. As 70% of received ticks were confirmed or suspected to have been acquired in Europe andEuropean countries are common destinations for UK travellers therisk of imported ticks and tick-borne diseases from Europe is higher andUK travellers should be made aware of the risk preceding and following European travel.

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References

- 1. World Tourism Organization. UNWTO Annual Report 2017. UNWTO Madr. 2018, 6, 1–28.
- 2. Office for National Statistics. Travel Trends: 2017. Available online: https://www.ons.gov.uk/peoplepopulationandcommunity/leisureandtourism/articles/traveltrends/2017 (accessed on 13 June 2019).
- 3. Parola, P.; Raoult, D. Ticks and Tickborne Bacterial Diseases in Humans: An Emerging Infectious Threat. *Clin. Infect. Dis.* **2001**, *32*, 897–928. [CrossRef] [PubMed]
- 4. Parola, P.; Paddock, C.D. Travel and tick-borne diseases: Lyme disease and beyond. *Travel Med. Infect. Dis.* **2018**, *26*, 1–2. [CrossRef] [PubMed]
- 5. Pek, C.H.; Cheong, C.S.J.; Yap, Y.L.; Doggett, S.; Lim, T.C.; Ong, W.C.; Lim, J. Rare Cause of Facial Palsy: Case Report of Tick Paralysis by *Ixodes holocyclus* Imported by a Patient Travelling into Singapore from Australia. *J. Emerg. Med.* **2016**, *51*, e109–e114. [CrossRef]
- 6. Pietzsch, M.E.; Hansford, K.; Medlock, J.M.; Doggett, S.L. Australian paralysis tick imported on a traveller returning to the UK. *Travel Med. Infect. Dis.* **2014**, *12*, 196–197. [CrossRef]
- 7. Mathison, B.A.; Gerth, W.J.; Pritt, B.S.; Baugh, S. Introduction of the exotic tick *Hyalomma truncatum* on a human with travel to Ethiopia: A case report. *Ticks Tick-Borne Dis.* **2015**, *6*, 152–154. [CrossRef]
- 8. Alderdice, J.M.; Burgess, I.F. The travels of a lone star tick. J. Clin. Pathol. 1998, 51, 403. [CrossRef]
- 9. Molaei, G.; Andreadis, T.G.; Anderson, J.F.; Iii, K.C.S. An Exotic Hitchhiker: A Case Report of Importation into Connecticut from Africa of the Human Parasitizing Tick, Hyalomma truncatum(Acari: Ixodidae). *J. Parasitol.* **2018**, 104, 302–305. [CrossRef]
- 10. Burridge, M.J.; Simmons, L.A.; Simbi, B.H.; Mahan, S.M.; Fournier, P.-E.; Raoult, D. Introduction of the Exotic Tick *Amblyomma hebraeum* into Florida on a Human Host. *J. Parasitol.* **2002**, *88*, 800. [CrossRef]
- 11. Heath, A.; Hardwick, S. The role of humans in the importation of ticks to New Zealand: A threat to public health and biosecurity. *N. Z. Med. J.* **2011**, *124*, 67–82.
- 12. Anderson, J.F.; Magnarelli, L.A.; Burgdorfer, W.; Casper, E.A.; Philip, R.N. Importation Into the United States from Africa of *Rhipicephalus simus* on a Boutonneuse Fever Patient. *Am. J. Trop. Med. Hyg.* **1981**, *30*, 897–899. [CrossRef] [PubMed]
- 13. McGarry, J.W. Travel and disease vector ticks. *Travel Med. Infect. Dis.* **2011**, *9*, 49–59. [CrossRef] [PubMed]
- 14. Chmielewski, T.; Szymanek-Pasternak, A.; Mączka, I.; Fiecek, B.; Simon, K.; Tylewska-Wierzbanowska, S. Case report of African tick-bite fever from Poland. *Advances in Dermatology and Allergology/Postępy Dermatol. Alergol.* **2013**, *30*, 396–398. [CrossRef] [PubMed]
- 15. Neal, S.P. African tick- bite fever among international travelers—Oregon, 1998. *Morb. Mortal. Wkly. Rep.* **1998**, 47, 950–952.
- 16. Martyn, K.P. *Provisional Atlas of the Ticks (Ixodoidea) of the British Isles*; Biological Records Centre Institute of Terrestrial Ecology: Huntingdon, UK, 1988.
- 17. Jameson, L.J.; Medlock, J.M. Tick Surveillance in Great Britain. *Vector-Borne Zoonotic Dis.* **2011**, 11, 403–412. [CrossRef] [PubMed]
- 18. Cull, B.; Pietzsch, M.E.; Hansford, K.M.; Gillingham, E.L.; Medlock, J.M. Surveillance of British ticks: An overview of species records hostassociations andnew records of Ixodes ricinus distribution. *Ticks Tick-Borne Dis.* **2018**, *9*, 605–614. [CrossRef]
- 19. Clifford, C.M.; Arthur, D.R. British Ticks. J. Parasitol. 1964, 50, 285. [CrossRef]
- 20. Hillyard, P.D. *Ticks of North-West Europe (Synopses of the British Fauna)*; The Linnean Society of London: London, UK, 1996.

- 21. Estrada-Peña, A.; Nava, S.; Petney, T. Description of all the stages of *Ixodes inopinatus* n. sp. (Acari: Ixodidae). *Ticks Tick-Borne Dis.* **2014**, *5*, 734–743. [CrossRef]
- 22. Walker, A.; Keirans, J.E.; Horak, I.G. *The Genus Rhipicephalus (Acari: Ixodidade): A Guide to the Brown Ticks of the World*; Cambridge University Press: Cambridge, UK, 2005.
- 23. Rubel, F.; Brugger, K.; Pfeffer, M.; Chitimia-Dobler, L.; Didyk, Y.M.; Leverenz, S.; Dautel, H.; Kahl, O. Geographical distribution of *Dermacentor marginatus* and *Dermacentor reticulatus* in Europe. *Ticks Tick-Borne Dis.* **2016**, *7*, 224–233. [CrossRef]
- 24. Rizzoli, A.; Silaghi, C.; Obiegala, A.; Rudolf, I.; Hubálek, Z.; Földvári, G. *Ixodes ricinus* and Its Transmitted Pathogens in Urban and Peri-Urban Areas in Europe: New Hazards and Relevance for Public Health. *Front. Public Health* **2014**, *2*, 251. [CrossRef]
- 25. Pato, F.J.; Panadero, R.; Vázquez, L.; López, C.M.; Díaz, P.; Vázquez, E.; Díez-Baños, P.; Morrondo, P.; Díaz, P. Seroprevalence of *Borrelia burgdorferi* sensu lato in roe deer (*Capreolus capreolus*) from northwestern Spain. *J. Zoo Wildl. Med.* **2013**, 44, 660–665. [CrossRef] [PubMed]
- 26. Hoby, S.; Mathis, A.; Doherr, M.G.; Robert, N.; Ryser-Degiorgis, M.-P. *Babesia capreoli* infections in Alpine chamois (*Rupicapra r. rupicapra*) roedeer (*Capreolus c. capreolus*) and red deer (*Cervus elaphus*) from Switzerland. *J. Wildl. Dis.* **2009**, 45, 748–753. [CrossRef] [PubMed]
- 27. Vázquez, L.; Panadero, R.; DaCal, V.; Pato, F.J.; López, C.; Díaz, P.; Arias, M.S.; Fernández, G.; Díez-Baños, P.; Morrondo, P.; et al. Tick infestation (Acari: Ixodidae) in roe deer (*Capreolus capreolus*) from northwestern Spain: Population dynamics and risk stratification. *Exp. Appl. Acarol.* 2010, 53, 399–409. [CrossRef] [PubMed]
- 28. Pacilly, F.; Benning, M.; Jacobs, F.; Leidekker, J.; Sprong, H.; Van Wieren, S.; Takken, W. Blood feeding on large grazers affects the transmission of *Borrelia burgdorferi* sensu lato by *Ixodes ricinus*. *Ticks Tick-Borne Dis*. **2014**, *5*, 810–817. [CrossRef] [PubMed]
- 29. Cull, B.; Vaux, A.G.C.; Ottowell, L.J.; Gillingham, E.L.; Medlock, J.M. Tick infestation of small mammals in an English woodland. *J. Vector Ecol.* **2017**, *42*, 74–83. [CrossRef] [PubMed]
- 30. Hoodless, A.N.; Kurtenbach, K.; Peacey, M. The role of pheasants as hosts for ticks and Lyme disease spirochaetes in southern England. *Game Wildl.* **1998**, *15*, 477–490.
- 31. Zdrazilova-Dubska, L.; Literak, I.; Kocianova, E.; Taragelova, V.; Sverakova, V.; Sychra, O.; Hromadko, M. Synanthropic Birds Influence the Distribution of *Borrelia Species*: Analysis of *Ixodes ricinus* Ticks Feeding on Passerine Birds. *Appl. Environ. Microbiol.* **2010**, 77, 1115–1117. [CrossRef]
- 32. Cayol, C.; Koskela, E.; Mappes, T.; Siukkola, A.; Kallio, E.R. Temporal dynamics of the tick *Ixodes ricinus* in northern Europe: Epidemiological implications. *Parasites Vectors* **2017**, *10*, 166. [CrossRef]
- 33. Selmi, M.; Ballardini, M.; Salvato, L.; Ricci, E. Rickettsia spp. in *Dermacentor marginatus* ticks: Analysis of the host-vector-pathogen interactions in a northern Mediterranean area. *Exp. Appl. Acarol.* **2017**, 72, 79–91. [CrossRef]
- 34. Petney, T.N.; Pfäffle, M.; Skuballa, J.D. An annotated checklist of the ticks (Acari: Ixodida) of Germany. *Syst. Appl. Acarol.* **2012**, *17*, 115–170. [CrossRef]
- 35. Walter, M.; Brugger, K.; Rubel, F. The ecological niche of *Dermacentor marginatus* in Germany. *Parasitol. Res.* **2016**, 115, 2165–2174. [CrossRef] [PubMed]
- 36. Selmi, M.; Tomassone, L.; Ceballos, L.A.; Crisci, A.; Ragagli, C.; Pintore, M.D.; Mignone, W.; Pautasso, A.; Ballardini, M.; Casalone, C.; et al. Analysis of the environmental and host-related factors affecting the distribution of the tick *Dermacentor marginatus*. *Exp. Appl. Acarol.* **2018**, 75, 209–225. [CrossRef] [PubMed]
- 37. Nosek, J. The ecology and public health importance of *Dermacentor marginatus* and *D. reticulatus* in Central Europe. *Folia Parasitol.* **1972**, 19, 93–102.
- 38. Hornok, S.; Farkas, R. Influence of biotope on the distribution and peak activity of questing ixodid ticks in Hungary. *Med. Vet. Entomol.* **2009**, 23, 41–46. [CrossRef]
- 39. Estrada-Peña, A.; Jongejan, F. Ticks Feeding on Humans: A Review of Records on Human-Biting Ixodoidea with Special Reference to Pathogen Transmission. *Exp. Appl. Acarol.* **1999**, 23, 685–715. [CrossRef] [PubMed]
- 40. Keirans, J.E.; Durden, L.A. Invasion: Exotic ticks (Acari: Argasidae, Ixodidae) imported into the United States. A review and new records. *J. Med. Entomol.* **2001**, *38*, 850–861. [CrossRef] [PubMed]
- 41. Dantas-Torres, F.; Figueredo, L.A.; Brandão-Filho, S.P. *Rhipicephalus sanguineus* (Acari: Ixodidae) thebrown dog tick, parasitizing humans in Brazil. *Rev. Soc. Bras. Med. Trop.* **2006**, *39*, 64–67. [CrossRef]

- 42. Dantas-Torres, F. The brown dog tick, *Rhipicephalus sanguineus* (Latreille, 1806) (Acari: Ixodidae): From taxonomy to control. *Vet. Parasitol.* **2008**, *152*, 173–185. [CrossRef]
- 43. Renvoisé, A.; Delaunay, P.; Blanchouin, E.; Cannavo, I.; Cua, E.; Socolovschi, C.; Parola, P.; Raoult, D. Urban family cluster of spotted fever rickettsiosis linked to *Rhipicephalus sanguineus* infected with *Rickettsia conorii subsp. caspia* and *Rickettsia massiliae*. *Ticks Tick-Borne Dis.* **2012**, *3*, 389–392. [CrossRef]
- 44. Hansford, K.M.; Phipps, L.P.; Cull, B.; Pietzsch, M.E.; Medlock, J.M. *Rhipicephalus sanguineus* importation into the UK: Surveillance, risk, public health awareness and One Health response. *Vet. Rec.* **2016**, *180*, 119. [CrossRef]
- 45. Estrada-Peña, A.; Palomar, A.M.; Santibáñez, P.; Sánchez, N.; Habela, M.A.; Portillo, A.; Romero, L.; Oteo, J.A. Crimean-Congo Hemorrhagic Fever Virus in Ticks, Southwestern Europe, 2010. *Emerg. Infect. Dis.* **2012**, *18*, 179–180. [CrossRef]
- 46. Apanaskevich, D.A.; Santos-Silva, M.M.; Horak, I.G. The genus *Hyalomma* Koch, 1844. IV. Redescription of all parasitic stages of *H.* (*Euhyalomma*) *lusitanicum* Koch, 1844 and the adults of *H.* (*E.*) *franchinii* Tonelli Rondelli, 1932 (Acari: Ixodidae) with a first description of its immature stages. *Folia Parasitol.* **2008**, *55*, 61–74. [CrossRef]
- 47. Valcárcel, F.; González, J.; Sánchez, J.L.P.; Jaime, J.M.T.; Olmeda, A.S. Long-Term Ecological Study of Host-Seeking Adults of *Hyalomma lusitanicum* (Acari: Ixodidae) in a Meso-Mediterranean Climate. *J. Med. Entomol.* **2015**, *53*, 221–224. [CrossRef]
- 48. Ruiz-Fons, F.; Fernández-De-Mera, I.G.; Acevedo, P.; Höfle, U.; Vicente, J.; De La Fuente, J.; Gortazár, C. Ixodid ticks parasitizing Iberian red deer (*Cervus elaphus hispanicus*) and European wild boar (*Sus scrofa*) from Spain: Geographical and temporal distribution. *Vet. Parasitol.* **2006**, 140, 133–142. [CrossRef] [PubMed]
- 49. Santos-Silva, M.M.; Beati, L.; Santos-Silva, M.M.; De Sousa, R.; Núncio, M.S.; Melo, P.; Santos-Reis, M.; Fonseca, C.M.M.S.; Formosinho, P.; Vilela, C.; et al. The hard-tick fauna of mainland Portugal (Acari: Ixodidae): An update on geographical distribution and known associations with hosts and pathogens. *Exp. Appl. Acarol.* **2011**, *55*, 85–121. [CrossRef] [PubMed]
- 50. Barandika, J.F.; Olmeda, S.A.; Casado-Nistal, M.A.; Hurtado, A.; Juste, R.A.; Valcárcel, F.; Anda, P.; Garcia-Perez, A.L. Differences in Questing Tick Species Distribution Between Atlantic and Continental Climate Regions in Spain. *J. Med. Entomol.* **2011**, *48*, 13–19. [CrossRef] [PubMed]
- 51. Requena-García, F.; Cabrero-Sanudo, F.J.; Olmeda-García, S.; González, J.; Valcárcel, F. Influence of environmental temperature and humidity on questing ticks in central Spain. *Exp. Appl. Acarol.* **2017**, 71, 277–290. [CrossRef]
- 52. Childs, J.E.; Paddock, C.D. The ascendancy of *Amblyomma americanum* as a vector of pathogens affecting humans in the United States. *Annu. Rev. Entomol.* **2003**, *48*, 307–337. [CrossRef] [PubMed]
- 53. Springer, Y.P.; Eisen, L.; Beati, L.; James, A.M.; Eisen, R.J. Spatial Distribution of Counties in the Continental United States With Records of Occurrence of *Amblyomma americanum* (Ixodida: Ixodidae). *J. Med. Entomol.* **2014**, *51*, 342–351. [CrossRef]
- 54. A Merten, H.; Durden, L. A state-by-state survey of ticks recorded from humans in the United States. *J. Vector Ecol.* **2000**, *25*, 102–113.
- 55. Stromdahl, E.Y.; Hickling, G.J. Beyond Lyme: Aetiology of Tick-borne Human Diseases with Emphasis on the South-Eastern United States. *Zoonoses Public Health* **2012**, *59*, 48–64. [CrossRef]
- 56. Nadolny, R.M.; Wright, C.L.; Sonenshine, D.E.; Hynes, W.L.; Gaff, H.D. Ticks and spotted fever group rickettsiae of southeastern Virginia. *Ticks Tick-Borne Dis.* **2014**, *5*, 53–57. [CrossRef]
- 57. Minigan, J.N.; Hager, H.A.; Peregrine, A.S.; Newman, J.A. Current and potential future distribution of the American dog tick (*Dermacentor variabilis*, Say) in North America. *Ticks Tick-Borne Dis.* **2018**, *9*, 354–362. [CrossRef]
- 58. Burg, J.G. Seasonal activity and spatial distribution of host-seeking adults of the tick *Dermacentor variabilis*. *Med. Vet. Entomol.* **2001**, *15*, 413–421. [CrossRef]
- 59. Stromdahl, E.Y.; Jiang, J.; Vince, M.; Richards, A.L. Infrequency of *Rickettsia rickettsii* in *Dermacentor variabilis* removed from humans withcomments on the role of other human-biting ticks associated with spotted fever group rickettsiae in the United States. *Vector-Borne Zoonotic Dis.* **2011**, *11*, 969–977. [CrossRef]
- 60. James, A.M. Distribution, seasonality andhosts of the Rocky Mountain wood tick in the United States. *J. Med. Entomol.* **2006**, *43*, 17–24. [CrossRef]

- 61. Lane, R.S. Ecology of tick-borne agents in California. II. Further observations on rickettsiae. In *Rickettsiae and Rickettsial Diseases*; Burgdorfer, W., Ed.; Academic Press: Cambridge, MA, USA, 1981; pp. 575–584.
- 62. Eads, R.B.; Smith, G.C. Seasonal activity and Colorado tick fever virus infection rates in Rocky Mountain wood ticks, *Dermacentor andersoni* (Acari: Ixodidae) innorth-central Colorado, USA. *J. Med. Entomol.* 1983, 20, 49–55. [CrossRef]
- 63. Eisen, R.J.; Eisen, L.; Beard, C.B. County-Scale Distribution of *Ixodes scapularis* and *Ixodes pacificus* (Acari: Ixodidae) in the Continental United States. *J. Med. Entomol.* **2016**, *53*, 349–386. [CrossRef]
- 64. Eisen, R.J.; Eisen, L.; Castro, M.B.; Lane, R.S. Environmentally Related Variability in Risk of Exposure to Lyme Disease Spirochetes in Northern California: Effect of Climatic Conditions and Habitat Type. *Environ. Entomol.* **2003**, *32*, 1010–1018. [CrossRef]
- 65. Padgett, K.; Lane, R.S. Life Cycle of *Ixodes pacificus* (Acari: Ixodidae): Timing of Developmental Processes Under Field and Laboratory Conditions. *J. Med. Entomol.* **2001**, *38*, 684–693. [CrossRef]
- 66. Dingler, R.J.; Wright, S.A.; Donohue, A.M.; Macedo, P.A.; Foley, J. Surveillance for *Ixodes pacificus* and the tick-borne pathogens *Anaplasma phagocytophilum* and *Borrelia burgdorferi* in birds from California's Inner Coast Range. *Ticks Tick-Borne Dis.* **2014**, *5*, 436–445. [CrossRef]
- 67. Castro, M.B.; Wright, S.A. Vertebrate hosts of *Ixodes pacificus* (Acari: Ixodidae) in California. *J. Vector Ecol.* **2007**, 32, 140–149. [CrossRef]
- 68. Eisen, L.; Eisen, R.J.; Lane, R.S. The roles of birds, lizards androdents as hosts for the western black-legged tick Ixodes pacificus. *J. Vector Ecol.* **2004**, *29*, 295–308.
- 69. Eisen, R.J. Prevalence and abundance of *Ixodes pacificus* immatures (Acari: Ixodidae) infesting western fence lizards (*Sceloporus occidentalis*) in northern California: Temporal trends and environmental correlates. *J. Parasitol.* **2001**, *87*, 1301–1307. [CrossRef]
- 70. Eisen, L.; Eisen, R.J.; Lane, R.S. Seasonal activity patterns of *Ixodes pacificus* nymphs in relation to climatic conditions. *Med. Vet. Entomol.* **2002**, *16*, 235–244. [CrossRef]
- 71. Clover, J.R.; Lane, R.S. Evidence Implicating Nymphal *Ixodes pacificus* (Acari: Ixodidae) in the Epidemiology of Lyme Disease in California. *Am. J. Trop. Med. Hyg.* **1995**, 53, 237–240. [CrossRef]
- 72. Salkeld, D.J.; Castro, M.B.; Bonilla, D.; Kjemtrup, A.; Kramer, V.L.; Lane, R.S.; Padgett, K.A. Seasonal activity patterns of the western black-legged tick, *Ixodes pacificus* inrelation to onset of human Lyme disease in northwestern California. *Ticks Tick-Borne Dis.* **2014**, *5*, 790–796. [CrossRef]
- 73. Masina, S.; Broady, K. Tick paralysis: Development of a vaccine. Int. J. Parasitol. 1999, 29, 535–541. [CrossRef]
- 74. Barker, S.C.; Walker, A.R. Ticks of domestic animals and humans in Australia. *Zootaxa* **2014**, *3816*, 1–144. [CrossRef]
- 75. Hall-Mendelin, S.; Craig, S.B.; Hall, R.; O'Donoghue, P.; Tulsiani, S.M.; Graham, G.C. Tick paralysis in Australia caused by *Ixodes holocyclus* Neumann. *Ann. Trop. Med. Parasitol.* **2011**, *105*, 95–106. [CrossRef]
- 76. Jahfari, S.; Ruyts, S.C.; Frazer-Mendelewska, E.; Jaarsma, R.I.; Verheyen, K.; Sprong, H. Melting pot of tick-borne zoonoses: The European hedgehog contributes to the maintenance of various tick-borne diseases in natural cycles urban and suburban areas. *Parasites Vectors* **2017**, *10*, 134. [CrossRef]
- 77. Sándor, A.D.; D'Amico, G.; Gherman, C.M.; Dumitrache, M.O.; Domṣa, C.; Mihalca, A.D. Mesocarnivores and macroparasites: Altitude and land use predict the ticks occurring on red foxes (*Vulpes vulpes*). *Parasites Vectors* **2017**, *10*, 173. [CrossRef]
- 78. Estrada-Peña, A.; Roura, X.; Sainz, A.; Miró, G.; Solano-Gallego, L. Species of ticks and carried pathogens in owned dogs in Spain: Results of a one-year national survey. *Ticks Tick-Borne Dis.* **2017**, *8*, 443–452. [CrossRef]
- 79. Pfäffle, M.; Petney, T.; Skuballa, J.; Taraschewski, H. Comparative population dynamics of a generalist (*Ixodes ricinus*) and specialist tick (*I. hexagonus*) species from European hedgehogs. *Exp. Appl. Acarol.* **2011**, *54*, 151–164. [CrossRef]
- 80. Beichel, E.; Petney, T.N.; Hassler, D.; Brückner, M.; Maiwald, M. Tick infestation patterns and prevalence of *Borrelia burgdorferi* in ticks collected at a veterinary clinic in Germany. *Vet. Parasitol.* **1996**, *65*, 147–155. [CrossRef]
- 81. Meyer-Kayser, E.; Hoffmann, L.; Silaghi, C.; Pfister, K.; Mahling, M.; Passos, L.M.F. Dynamics of tick infestations in foxes in Thuringia, Germany. *Ticks Tick-Borne Dis.* **2012**, *3*, 232–239. [CrossRef]
- 82. Faulde, M.K.; Rutenfranz, M.; Hepke, J.; Rogge, M.; Görner, A.; Keth, A. Human tick infestation pattern, tick-bite rate and associated *Borrelia burgdorferi* s.l. infection risk during occupational tick exposure at the Seedorf military training area, northwestern Germany. *Ticks Tick-Borne Dis.* **2014**, *5*, 594–599. [CrossRef]

- 83. Nijhof, A.M.; Bodaan, C.; Postigo, M.; Nieuwenhuijs, H.; Opsteegh, M.; Franssen, L.; Jebbink, F.; Jongejan, F. Ticks and Associated Pathogens Collected from Domestic Animals in the Netherlands. *Vector-Borne Zoonotic Dis.* **2007**, *7*, 585–596. [CrossRef]
- 84. Sanogo, Y.O.; Parola, P.; Shpynov, S.; Camicas, J.L.; Brouqui, P.; Caruso, G.; Raoult, D. Genetic Diversity of Bacterial Agents Detected in Ticks Removed from Asymptomatic Patients in Northeastern Italy. *Ann. N. Y. Acad. Sci.* **2003**, *990*, 182–190. [CrossRef]
- 85. Hubbard, M.J.; Baker, A.S.; Cann, K.J. Distribution of *Borrelia burgdorferi* s.l. spirochaete DNA in British ticks (Argasidae and Ixodidae) since the 19th Century, assessed by PCR. *Med. Vet. Entomol.* **1998**, 12, 89–97. [CrossRef]
- 86. Carter, W.I. A case of human parasitization by *Ixodes hexagonus*, Leach (hedgehog tick). *Br. Med. J.* **1955**, 22, 1012. [CrossRef]
- 87. Estrada-Peña, A.; Guglielmone, A.; Mangold, A.J. The distribution and ecological 'preferences' of the tick *Amblyomma cajennense* (Acari: Ixodidae) anectoparasite of humans and other mammals in the Americas. *Ann. Trop. Med. Parasitol.* **2004**, *98*, 283–292. [CrossRef]
- 88. Pires, M.S.; Dos Santos, T.M.; Santos, H.A.; Vilela, J.A.R.; Peixoto, M.P.; Roier, E.C.R.; Da Silva, C.B.; Barreira, J.D.; De Lemos, E.R.S.; Massard, C.L. *Amblyomma cajennense* infestation on horses in two microregions of the state of Rio de Janeiro, Brazil. *Rev. Bras. Parasitol. Vet.* **2013**, 22, 235–242. [CrossRef]
- 89. Oliveira, P.R.; Borges, L.M.F.; Leite, R.C.; Freitas, C.M.V. Seasonal dynamics of the Cayenne tick, *Amblyomma cajennense* on horses in Brazil. *Med. Vet. Entomol.* **2003**, 17, 412–416. [CrossRef] [PubMed]
- 90. Labruna, M.B.; Kasai, N.; Ferreira, F.; Faccini, J.L.; Gennari, S.M. Seasonal dynamics of ticks (Acari: Ixodidae) on horses in the state of São Paulo, Brazil. *Vet. Parasitol.* **2002**, *105*, 65–77. [CrossRef]
- 91. Ramos, V.D.N.; Piovezan, U.; Franco, A.H.A.; Osava, C.F.; Herrera, G.P.; Szabó, M.P.J. Feral pigs as hosts for *Amblyomma sculptum* (Acari: Ixodidae) populations in the Pantanal MatoGrosso do Sul, Brazil. *Exp. Appl. Acarol.* **2014**, *64*, 393–406. [CrossRef]
- 92. Martins, T.F.; Barbieri, A.R.M.; Costa, F.B.; Terassini, F.A.; Camargo, L.M.A.; Peterka, C.R.L.; Pacheco, R.D.C.; Dias, R.A.; Nunes, P.H.; Marcili, A.; et al. Geographical distribution of Amblyomma cajennense (sensu lato) ticks (Parasitiformes: Ixodidae) in Brazil withdescription of the nymph of A. cajennense (sensu stricto). *Parasites Vectors* 2016, *9*, 1–14. [CrossRef]
- 93. Saraiva, D.G.; Fournier, G.F.S.R.; Martins, T.F.; Leal, K.P.G.; Vieira, F.N.; Câmara, E.M.V.C.; Costa, C.G.; Onofrio, V.C.; Barros-Battesti, D.M.; Guglielmone, A.A.; et al. Ticks (Acari: Ixodidae) associated with small terrestrial mammals in the state of Minas Gerais, southeastern Brazil. *Exp. Appl. Acarol.* **2012**, *58*, 159–166. [CrossRef] [PubMed]
- 94. Labruna, M.B.; Terassini, F.A.; Camargo, L.M.A. Notes on Population Dynamics of *Amblyomma* Ticks (Acari: Ixodidae) in Brazil. *J. Parasitol.* **2009**, 95, 1016–1018. [CrossRef] [PubMed]
- 95. Oliveira, P.R.; Borges, L.; Lopes, C.; Leite, R.C. Population dynamics of the free-living stages of *Amblyomma cajennense* (Fabricius, 1787) (Acari: Ixodidae) on pastures of Pedro Leopoldo, Minas Gerais State, Brazil. *Vet. Parasitol.* **2000**, *92*, 295–301. [CrossRef]
- 96. Brites-Neto, J.; Nieri-Bastos, F.A.; Brasil, J.; Duarte, K.M.R.; Martins, T.F.; Veríssimo, C.J.; Barbieri, A.R.M.; Labruna, M.B. Environmental infestation and rickettsial infection in ticks in an area endemic for Brazilian spotted fever. *Rev. Bras. Parasitol. Vet.* **2013**, 22, 367–372. [CrossRef]
- 97. Beck, D.L.; Zavala, J.; Montalvo, E.O.; Quintana, F.G. Meteorological indicators for *Amblyomma cajennense* and population dynamics in the Tamaulipan Biotic Province in Texas. *J. Vector Ecol.* **2011**, *36*, 135–146. [CrossRef]
- 98. Guglielmone, A.A.; Beati, L.; Barros-Battesti, D.M.; Labruna, M.B.; Nava, S.; Venzal, J.M.; Mangold, A.J.; Szabó, M.P.J.; Martins, J.R.; González-Acuña, D.; et al. Ticks (Ixodidae) on humans in South America. *Exp. Appl. Acarol.* **2006**, *40*, 83–100. [CrossRef] [PubMed]
- 99. Labruna, M.B. Ecology of Rickettsia in South America. Ann. N. Y. Acad. Sci. 2009, 1166, 156–166. [CrossRef]
- 100. Apanaskevich, D.A.; Horak, I.G. The genus *Hyalomma*. VI. Systematics of *H. (Euhyalomma) truncatum* and the closely related species, *H. (E.) albiparmatum* and *H. (E.) nitidum* (Acari: Ixodidae). *Exp. Appl. Acarol.* **2008**, 44, 115–136. [CrossRef]
- 101. Magano, S.R.; Els, D.; Chown, S. Feeding patterns of immature stages of *Hyalomma truncatum* and *Hyalomma marginatum rufipes* on different hosts. *Exp. Appl. Acarol.* **2000**, 24, 301–313. [CrossRef]

- 102. Apanaskevich, D.A.; Schuster, A.L.; Horak, I.G. The genus Hyalomma: VII. Redescription of all parasitic stages of *Hy.* (*Euhyalomma*) *dromedarii* and *H.* (*E.*) *schulzei* (Acari: Ixodidae). *Morpho. System. Evo.* **2008**, 45, 817–831. [CrossRef]
- 103. Dreyer, K.; Fourie, L.; Kok, D.J. Tick diversity, abundance and seasonal dynamics in a resource-poor urban environment in the Free State Province. *Onderstepoort J. Vet. Res.* **1998**, *65*, 305–316.
- 104. Horak, I.G.; Fourie, L.; Braack, L. Small mammals as hosts of immature ixodid ticks. *Onderstepoort J. Vet. Res.* **2005**, 72, 255–261. [CrossRef] [PubMed]
- 105. Horak, I.; Golezardy, H.; Uys, A. Ticks associated with the three largest wild ruminant species in Southern Africa. *Onderstepoort J. Vet. Res.* **2007**, *74*, 231–242. [CrossRef]
- 106. Walker, J.B. A review of the Ixodid ticks (Acari, Ixodidae) occurring in southern Africa. *Onderstepoort J. Vet. Res.* **1991**, *58*, 81–105.
- 107. Fourie, L.; Kok, D.J.; Heyne, H. Adult ixodid ticks on two cattle breeds in the south-western Free State and their seasonal dynamics. *Onderstepoort J. Vet. Res.* **1996**, *63*, 19–23.
- 108. Horak, I.G.; Matthee, S. Parasites of domestic and wild animals in South Africa. XLIII. Ixodid ticks of domestic dogs and cats in the Western Cape Province. *Onderstepoort J. Vet. Res.* **2003**, *70*, 187–195.
- 109. Brain, C.; Bohrmann, R. Tick infestation of baboons (*Papio ursinus*) in the Namib Desert. *J. Wildl. Dis.* **1992**, 28, 188–191. [CrossRef]
- 110. Horak, I.; Fourie, L. Parasites of domestic and wild animals in South Africa. XXXI. Adult ixodid ticks on sheep in the Cape Province and in the Orange Free State. *Onderstepoort J. Vet. Res.* **1992**, *59*, 275–283.
- 111. Horak, I.; Fourie, L.; Heyne, H.; Walker, J.B.; Needham, G. Ixodid ticks feeding on humans in South Africa: With notes on preferred hosts, geographic distribution, seasonal occurrence and transmission of pathogens. *Exp. Appl. Acarol.* **2002**, 27, 113–136. [CrossRef]
- 112. Lessard, P.; L'Eplattenier, R.; Norval, R.; Kundert, K.; Dolan, T.T.; Croze, H.; Walker, J.B.; Irvin, A.D.; Perry, B.D. Geographical information systems for studying the epidemiology of cattle diseases caused by *Theileria parva*. *Vet. Rec.* **1990**, *126*, 255–262.
- 113. Walker, A.R. *Ticks of Domestic Animals in Africa: A Guide to Identification of Species;* Bioscience Reports: Edinburgh, UK, 2003.
- 114. Hoogstraal, H. Studies on southeast Asian Haemaphysalis ticks (Ixodoidea, Ixodidae). The identity, distribution andhosts of H. (Kaiseriana) hystricis Supino. *J. Parasitol.* **1965**, *51*, 467–480. [CrossRef]
- 115. Hou, J.; Ling, F.; Chai, C.; Lu, Y.; Yu, X.; Lin, J.; Sun, J.; Chang, Y.; Ye, X.; Gu, S.; et al. Prevalence of *Borrelia burgdorferi* sensu lato in ticks from eastern China. *Am. J. Trop. Med. Hyg.* **2014**, 92, 262–266. [CrossRef]
- 116. Khoo, J.J.; Lim, F.S.; Tan, K.-K.; Chen, F.S.; Phoon, W.H.; Khor, C.S.; Pike, B.L.; Chang, L.Y.; Abubakar, S. Detection in Malaysia of a *Borrelia* sp. From *Haemaphysalis hystricis* (Ixodida: Ixodidae). *J. Med. Èntomol.* **2017**, *54*, 1444–1448. [CrossRef]
- 117. Shimada, Y.; Beppu, T.; Inokuma, H.; Okuda, M.; Onishi, T. Ixodid tick species recovered from domestic dogs in Japan. *Med. Vet. Entomol.* **2003**, *17*, 38–45. [CrossRef]
- 118. Durden, L.A.; Merker, S.; Beati, L. The tick fauna of Sulawesi, Indonesia (Acari: Ixodoidea: Argasidae and Ixodidae). *Exp. Appl. Acarol.* **2008**, *45*, 85–110. [CrossRef]
- 119. Grassman, L.I.; Sarataphan, N.; Tewes, M.E.; Silvy, N.J.; Nakanakrat, T. Ticks (Acari: Ixodidae) Parasitizing Wild Carnivores in Phu Khieo Wildlife Sanctuary, Thailand. *J. Parasitol.* **2004**, *90*, 657–659. [CrossRef]
- 120. Yamauchi, T.; Yano, S.; Yamamoto, T.; Yamamoto, E.; Miyamoto, T. Ticks (Acari: Ixodidae) from medium-sized to large mammals in Ehime Prefecture, Japan. *Exp. Appl. Acarol.* **2012**, *60*, 263–270. [CrossRef]
- 121. Tateno, M.; Sunahara, A.; Nakanishi, N.; Izawa, M.; Matsuo, T.; Setoguchi, A.; Endo, Y. Molecular survey of arthropod-borne pathogens in ticks obtained from Japanese wildcats. *Ticks Tick-Borne Dis.* **2015**, *6*, 281–289. [CrossRef]
- 122. Ajithkumar, K.; Ravindran, R.; Ghosh, S. *Dermacentor auratus* Supino, 1897 (Acarina, Ixodidae) reported from Wayanad, Kerala. *Indian J. Med. Res.* **2012**, *135*, 435–436.
- 123. Mariana, A.; Zuraidawati, Z.; Ho, T.M.; Kulaimi, B.M.; Saleh, I.; Shukor, M.N.; Shahrul-Anuar, M.S. Ticks (Ixodidae) and other ectoparasites in Ulu Muda Forest Reserve, Kedah, Malaysia. *Southeast Asian J. Trop. Med. Public Health* 2008, 39, 496–506.
- 124. Vongphayloth, K.; Hertz, J.C.; Lakeomany, K.; Apanaskevich, D.; Robbins, R.G.; Sutherland, I.W.; Brey, P.T. The Genus *Dermacentor* (Acari: Ixodidae) in Laos: A Review and Update of Species Records. *J. Med. Entomol.* **2018**, *55*, 1047–1050. [CrossRef]

- 125. Parola, P.; Cornet, J.-P.; Sanogo, Y.O.; Miller, R.S.; Van Thien, H.; Gonzalez, J.-P.; Raoult, D.; Telford, S.R., III; Wongsrichanalai, C. Detection of *Ehrlichia* spp., *Anaplasma* spp., *Rickettsia* spp. andOther Eubacteria in Ticks from the Thai-Myanmar Border and Vietnam. *J. Clin. Microbiol.* **2003**, *41*, 1600–1608. [CrossRef]
- 126. Ariyarathne, S.; Dilrukshi, P.R.M.P.; Amarasinghe, P.H.; Rajakaruna, R.S. Intra-aural ecdysis of *Dermacentor auratus* Supino, 1897 ina human host. *Ceylon Med. J.* **2011**, *56*, 133. [CrossRef]
- 127. Sumrandee, C.; Baimai, V.; Trinachartvanit, W.; Ahantarig, A. Hepatozoon and *Theileria* species detected in ticks collected from mammals and snakes in Thailand. *Ticks Tick-Borne Dis.* **2015**, *6*, 309–315. [CrossRef]
- 128. Kirwan, E.O. A tick on the upper eye-lid. Br. J. Ophthalmol. 1935, 19, 659-661. [CrossRef]
- 129. Liyanaarachchi, D.; Rajakaruna, R.; Dikkumbura, A.; Rajapakse, R. Ticks infesting wild and domestic animals and humans of Sri Lanka with new host records. *Acta Trop.* **2015**, *142*, 64–70. [CrossRef]
- 130. Ariyarathne, S.; Apanaskevich, D.A.; Amarasinghe, P.H.; Rajakaruna, R.S. Diversity and distribution of tick species (Acari: Ixodidae) associated with human otoacariasis and socio-ecological risk factors of tick infestations in Sri Lanka. *Exp. Appl. Acarol.* **2016**, *70*, 99–123. [CrossRef]
- 131. Hoogstraal, H.; Wassef, H.Y. *Dermacentor (Indocentor) auratus* (Acari: Ixodoidea: Ixodidae): Hosts, Distribution and Medical Importance in Tropical Asia. *J. Med. Entomol.* **1985**, 22, 170–177. [CrossRef]
- 132. Sumrandee, C.; Baimai, V.; Trinachartvanit, W.; Ahantarig, A. Molecular detection of *Rickettsia, Anaplasma, Coxiella* and *Francisella* bacteria in ticks collected from Artiodactyla in Thailand. *Ticks Tick-Borne Dis.* **2016**, 7, 678–689. [CrossRef]
- 133. Ghosh, S.; Bansal, G.C.; Gupta, S.C.; Ray, D.; Khan, M.Q.; Irshad, H.; Shahiduzzaman, M.; Seitzer, U.; Ahmed, J.S. Status of tick distribution in Bangladesh, India and Pakistan. *Parasitol. Res.* **2007**, *101*, 207–216. [CrossRef]
- 134. Edussuriya, B.D.; Weilgama, D. Case reports: Intra-aural tick infestations in humans in Sri Lanka. *Trans. R. Soc. Trop. Med. Hyg.* **2004**, 97, 412–413. [CrossRef]
- 135. Strnad, M.; Hönig, V.; Růžek, D.; Grubhoffer, L.; Rego, R.O.M. Europe-Wide Meta-Analysis of *Borrelia burgdorferi* Sensu Lato Prevalence in Questing *Ixodes ricinus* Ticks. *Appl. Environ. Microbiol.* **2017**, *83*, e00609–e00617. [CrossRef]
- 136. Gern, L.; Toutoungi, L.N.; Hu, C.M.; Aeschlimann, A. *Ixodes (Pholeoixodes) hexagonus* anefficient vector of *Borrelia burgdorferi* in the laboratory. *Med. Vet. Entomol.* **1991**, *5*, 431–435. [CrossRef]
- 137. Toutoungi, L.N.; Gern, L. Ability of transovarially and subsequent transstadially infected *Ixodes hexagonus* ticks to maintain and transmit Borrelia burgdorferi in the laboratory. *Exp. Appl. Acarol.* **1993**, 17, 581–586. [CrossRef]
- 138. Franke, J.; Hildebrandt, A.; Dorn, W. Exploring gaps in our knowledge on Lyme borreliosis spirochaetes Updates on complex heterogeneity, ecology and pathogenicity. *Ticks Tick-Borne Dis.* **2013**, *4*, 11–25. [CrossRef]
- 139. Humair, P.-F.; Rais, O.; Gern, L. Transmission of *Borrelia afzelii* from *Apodemus* mice and *Clethrionomys* voles to Ixodes ricinus ticks: Differential transmission pattern and overwintering maintenance. *Parasitology* **1999**, 118, 33–42. [CrossRef]
- 140. Kurtenbach, K.; Peacey, M.; Rijpkema, S.G.T.; Hoodless, A.N.; Nuttall, P.A.; Randolph, S.E. Differential Transmission of the Genospecies of *Borrelia burgdorferi* Sensu Lato by Game Birds and Small Rodents in England. *Appl. Environ. Microbiol.* **1998**, *64*, 1169–1174. [CrossRef]
- 141. Humair, P.-F.; Peter, O.; Wallich, R.; Gern, L. Strain Variation of Lyme Disease Spirochetes Isolated from *Ixodes ricinus* Ticks and Rodents Collected in Two Endemic Areas in Switzerland. *J. Med. Entomol.* **1995**, 32, 433–438. [CrossRef]
- 142. Kurtenbach, K.; Carey, D.; Hoodless, A.N.; Nuttall, P.A.; Randolph, S.E. Competence of Pheasants as Reservoirs for Lyme Disease Spirochetes. *J. Med. Entomol.* **1998**, *35*, 77–81. [CrossRef]
- 143. Humair, P.-F.; Postic, D.; Wallich, R.; Gern, L. An Avian Reservoir (*Turdus merula*) of the Lyme Borreliosis Spirochetes. *Zentralblatt Fur Bakteriol.* **1998**, 287, 521–538. [CrossRef]
- 144. Kurtenbach, K.; De Michelis, S.; Etti, S.; Schäfer, S.M.; Sewell, H.-S.; Brade, V.; Kraiczy, P. Host association of *Borrelia burgdorferi* sensu lato—The key role of host complement. *Trends Microbiol.* **2002**, *10*, 74–79. [CrossRef]
- 145. Baranton, G.; Postic, D.; Girons, I.S.; Boerlin, P.; Piffaretti, J.-C.; Assous, M.; Grimont, P.A.D. Delineation of *Borrelia burgdorferi* Sensu Stricto, *Borrelia garinii* sp. nov. and Group VS461 Associated with Lyme borreliosis. *Int. J. Syst. Bacteriol.* **1992**, 42, 378–383. [CrossRef]
- 146. Richter, D.; Schlee, D.B.; Matuschka, F.-R. Relationships of a Novel Lyme Disease Spirochete, *Borrelia spielmani* sp. nov. withIts Hosts in Central Europe. *Appl. Environ. Microbiol.* **2004**, *70*, 6414–6419. [CrossRef]

- 147. Stanek, G.; Reiter, M. The expanding Lyme Borrelia complex—Clinical significance of genomic species? *Clin. Microbiol. Infect.* **2011**, 17, 487–493. [CrossRef]
- 148. Gritsun, T.S.; Lashkevich, V.A.; Gould, E.A. Tick-borne encephalitis. *Antivir. Res.* **2003**, *57*, 129–146. [CrossRef]
- 149. Holzmann, H.; Aberle, S.W.; Stiasny, K.; Werner, P.; Mischak, A.; Zainer, B.; Netzer, M.; Koppi, S.; Bechter, E.; Heinz, F.X. Tick-borne Encephalitis from Eating Goat Cheese in a Mountain Region of Austria. *Emerg. Infect. Dis.* **2009**, *15*, 1671–1673. [CrossRef]
- 150. Dobler, G.; Gniel, D.; Petermann, R.; Pfeffer, M. Epidemiology and distribution of tick-borne encephalitis. *Wien. Med. Wochenschr.* **2012**, *162*, 230–238. [CrossRef]
- 151. Amicizia, D.; Domnich, A.; Panatto, D.; Lai, P.L.; Cristina, M.; Avio, U.; Gasparini, R. Epidemiology of tick-borne encephalitis (TBE) in Europe and its prevention by available vaccines. *Hum. Vaccines Immunother.* **2013**, *9*, 1163–1171. [CrossRef]
- 152. Danielová, V.; Daniel, M.; Schwarzová, L.; Materna, J.; Rudenko, N.; Golovchenko, M.; Holubová, J.; Grubhoffer, L.; Kilian, P. Integration of a Tick-Borne Encephalitis Virus and *Borrelia burgdorferi* sensu lato into Mountain Ecosystems, Following a Shift in the Altitudinal Limit of Distribution of Their Vector, *Ixodes ricinus* (Krkonoše Mountains, Czech Republic). *Vector-Borne Zoonotic Dis.* **2010**, *10*, 223–230. [CrossRef]
- 153. Arnez, M.; Avšič-Županc, T. Tick-borne encephalitis in children: An update on epidemiology and diagnosis. *Expert Rev. Anti Infect. Ther.* **2009**, *7*, 1251–1260. [CrossRef]
- 154. Haglund, M.; Günther, G. Tick-borne encephalitis—pathogenesis, clinical course and long-term follow-up. *Vaccine* **2003**, *21*, S11–S18. [CrossRef]
- 155. Lindquist, L.; Vapalahti, O. Tick-borne encephalitis. Lancet 2008, 371, 1861–1871. [CrossRef]
- 156. Logar, M.; Bogovič, P.; Cerar, D.; Avšič-Županc, T.; Strle, F. Tick-borne encephalitis in Slovenia from 2000 to 2004: Comparison of the course in adult and elderly patients. *Wien. Klin. Wochenschr.* **2006**, *118*, 702–707. [CrossRef]
- 157. Logar, M.; Arnez, M.; Kolbl, J.; Avšič-Županc, T.; Strle, F. Comparison of the epidemiological and clinical features of tick-borne encephalitis in children and adults. *Infection* **2000**, *28*, 74–77. [CrossRef]
- 158. Steffen, R. Tick-borne encephalitis (TBE) in children in Europe: Epidemiology, clinical outcome and comparison of vaccination recommendations. *Ticks Tick-Borne Dis.* **2018**, *10*, 100–110. [CrossRef]
- 159. Labuda, M.; Jones, L.D.; Williams, T.; Danielová, V.; Nuttall, P.A. Efficient Transmission of Tick-Borne Encephalitis Virus Between Cofeeding Ticks. *J. Med. Entomol.* 1993, 30, 295–299. [CrossRef]
- 160. Labuda, M.; Kozuch, O.; Zuffová, E.; Elecková, E.; Hails, R.S.; Nuttall, P.A. Tick-Borne Encephalitis Virus Transmission between Ticks Cofeeding on Specific Immune Natural Rodent Hosts. *Virology* **1997**, 235, 138–143. [CrossRef]
- 161. Perkins, S.E.; Cattadori, I.M.; Tagliapietra, V.; Rizzoli, A.; Hudson, P.J. Empirical evidence for key hosts in persistence of a tick-borne disease. *Int. J. Parasitol.* **2003**, *33*, 909–917. [CrossRef]
- 162. Kaiser, R. The clinical and epidemiological profile of tick-borne encephalitis in southern Germany 1994-98. A prospective study of 656 patients. *Brain* 1999, 122, 2067–2078. [CrossRef]
- 163. Lesnicar, G.; Poljak, M.; Seme, K.; Lesnicar, J. Pediatric tick-borne encephalitis in 371 cases from an endemic region in Slovenia, 1959 to 2000. *Pediatr. Infect. Dis. J.* 2003, 22, 612–617. [CrossRef]
- 164. Randolph, S.; Asokliene, L.; Avsic-Zupanc, T.; Bormane, A.; Burri, C.; Gern, L.; Golovljova, I.; Hubalek, Z.; Knap, N.; Kondrusik, M.; et al. Variable spikes in tick-borne encephalitis incidence in 2006 independent of variable tick abundance but related to weather. *Parasites Vectors* **2008**, *1*, 44. [CrossRef]
- 165. Beauté, J.; Spiteri, G.; Warns-Petit, E.; Zeller, H. Tick-borne encephalitis in Europe, 2012 to 2016. *Eurosurveillance* 2018, 23, 1800201. [CrossRef]
- 166. Süss, J. Epidemiology and ecology of TBE relevant to the production of effective vaccines. *Vaccine* **2003**, 21, S19–S35. [CrossRef]
- 167. Raoult, D.; Berbis, P.; Roux, V.; Xu, W.; Maurin, M. A new tick-transmitted disease due to *Rickettsia slovaca*. *Lancet* **1997**, 350, 112–113. [CrossRef]
- 168. Oteo, J.; Ibarra, V.; Blanco, J.; De Artola, V.M.; Márquez, F.J.; Portillo, A.; Raoult, D.; Anda, P. *Dermacentor*-borne necrosis erythema and lymphadenopathy: Clinical and epidemiological features of a new tick-borne disease. *Clin. Microbiol. Infect.* **2004**, *10*, 327–331. [CrossRef] [PubMed]

- 169. Ibarra, V.; Portillo, A.; Santibáñez, S.; Blanco, J.; Pérez-Martínez, L.; Márquez, F.J.; Oteo, J.A. DEBONEL/TIBOLA: Is *Rickettsia slovaca* the Only Etiological Agent? *Ann. N. Y. Acad. Sci.* **2005**, *1063*, 346–348. [CrossRef] [PubMed]
- 170. Portillo, A.; Ibarra, V.; Santibáñez, S.; Pérez-Martínez, L.; Blanco, J.; Oteo, J.A. Genetic characterisation of ompA ompBand gltA genes from Candidatus *Rickettsia rioja*. *Clin. Microbiol. Infect.* **2009**, *15*, 307–308. [CrossRef] [PubMed]
- 171. Mediannikov, O.; Matsumoto, K.; Samoylenko, I.; Drancourt, M.; Roux, V.; Rydkina, E.; Davoust, B.; Tarasevich, I.; Brouqui, P.; Fournier, P.-E. *Rickettsia raoultii* sp. nov. aspotted fever group rickettsia associated with *Dermacentor* ticks in Europe and Russia. *Int. J. Syst. Evol. Microbiol.* **2008**, *58*, 1635–1639. [CrossRef] [PubMed]
- 172. Pérez-Pérez, L.; Portillo, A.; Allegue, F.; Zulaica, A.; Oteo, J.; Caeiro, J.; Fabeiro, J. *Dermacentor*-borne Necrosis Erythema and Lymphadenopathy (DEBONEL): A Case Associated with *Rickettsia rioja*. *Acta Derm. Venereol.* **2010**, *90*, 214–215. [CrossRef]
- 173. Selmi, M.; Bertolotti, L.; Tomassone, L.; Mannelli, A. Rickettsia slovaca in *Dermacentor marginatus* and Tick-borne Lymphadenopathy, Tuscany, Italy. *Emerg. Infect. Dis.* **2008**, *14*, 817–820. [CrossRef]
- 174. Lakos, A. Tick-borne lymphadenopathy–a new rickettsial disease? Lancet 1997, 350, 1006. [CrossRef]
- 175. Oteo, J.A.; Portillo, A. Tick-borne rickettsioses in Europe. Ticks Tick-Borne Dis. 2012, 3, 271–278. [CrossRef]
- 176. Komitova, R.; Lakos, A.; Aleksandrov, A.; Christova, I.; Murdjeva, M. A case of tick-transmitted lymphadenopathy in Bulgaria associated with *Rickettsia slovaca*. *Scand. J. Infect. Dis.* **2003**, 35, 213. [CrossRef]
- 177. Raoult, D.; Lakos, A.; Fenollar, F.; Beytout, J.; Brouqui, P.; Fournier, P.-E. Spotless Rickettsiosis Caused by *Rickettsia slovaca* and Associated with *Dermacentor* Ticks. *Clin. Infect. Dis.* **2002**, *34*, 1331–1336. [CrossRef] [PubMed]
- 178. Parola, P.; Rovery, C.; Rolain, J.M.; Brouqui, P.; Davoust, B.; Raoult, D. *Rickettsia slovaca* and *R. raoultii* in Tick-borne Rickettsioses. *Emerg. Infect. Dis.* **2009**, *15*, 1105–1108. [CrossRef]
- 179. Ibarra, V.; Oteo, J.A.; Portillo, A.; Santibáñez, S.; Blanco, J.; Metola, L.; Eiros, J.; Pérez-Martínez, L.; Sanz, M. *Rickettsia slovaca* Infection: DEBONEL/TIBOLA. *Ann. N. Y. Acad. Sci.* **2006**, *1078*, 206–214. [CrossRef] [PubMed]
- 180. Földvári, G.; Rigó, K.; Lakos, A. Transmission of *Rickettsia slovaca* and *Rickettsia raoultii* by male *Dermacentor marginatus* and *Dermacentor reticulatus* ticks to humans. *Diagn. Microbiol. Infect. Dis.* **2013**, 76, 387–389. [CrossRef] [PubMed]
- 181. Sanogo, Y.; Davoust, B.; Parola, P.; Camicas, J.L.; Brouqui, P.; Raoult, D. Prevalence of *Rickettsia* spp. in *Dermacentor marginatus* ticks removed from game pigs (*Sus scrofa*) in southern France. *Ann. N. Y. Acad. Sci.* **2003**, 990, 191–195. [CrossRef]
- 182. Márquez, F.J.; Rojas, A.; Ibarra, V.; Cantero, A.; Rojas, J.; Oteo, J.A.; Muniain, M. Prevalence Data of Rickettsia slovaca and Other SFG Rickettsiae Species in *Dermacentor marginatus* in the Southeastern Iberian Peninsula. *Ann. N. Y. Acad. Sci.* **2006**, *1078*, 328–330. [CrossRef]
- 183. Masala, G.; Chisu, V.; Satta, G.; Socolovschi, C.; Raoult, D.; Parola, P. Rickettsia slovaca from *Dermacentor marginatus* ticks in Sardinia, Italy. *Ticks Tick-Borne Dis.* **2012**, *3*, 393–395. [CrossRef]
- 184. Špitalská, E.; Štefanidesová, K.; Kocianová, E.; Boldiš, V. *Rickettsia slovaca* and *Rickettsia raoultii* in *Dermacentor marginatus* and *Dermacentor reticulatus* ticks from Slovak Republic. *Exp. Appl. Acarol.* **2012**, *57*, 189–197. [CrossRef]
- 185. Maioli, G.; Pistone, D.; Bonilauri, P.; Pajoro, M.; Barbieri, I.; Patrizia, M.; Vicari, N.; Dottori, M. Ethiological agents of rickettsiosis and anaplasmosis in ticks collected in Emilia-Romagna region (Italy) during 2008 and 2009. *Exp. Appl. Acarol.* **2012**, *57*, 199–208. [CrossRef]
- 186. De Sousa, R.; Nóbrega, S.D.; Bacellar, F.; Torgal, J. Mediterranean spotted fever in Portugal: Risk factors for fatal outcome in 105 hospitalized patients. *Ann. N. Y. Acad. Sci.* **2003**, *990*, 285–294. [CrossRef]
- 187. Fernández-Soto, P.; Pérez-Sánchez, R.; Álamo-Sanz, R.; Encinas-Grandes, A. Spotted Fever Group Rickettsiae in Ticks Feeding on Humans in Northwestern Spain. *Ann. N. Y. Acad. Sci.* **2006**, *1078*, 331–333. [CrossRef]
- 188. Parola, P.; Paddock, C.D.; Raoult, D. Tick-Borne Rickettsioses around the World: Emerging Diseases Challenging Old Concepts. *Clin. Microbiol. Rev.* **2005**, *18*, 719–756. [CrossRef] [PubMed]

- 189. Márquez, F.J.; Rodríguez-Liébana, J.J.; Soriguer, R.C.; Muniáin, M.A.; Bernabeu-Wittel, M.; Caruz, A.; Contreras-Chova, F. Spotted fever group *Rickettsia* in brown dog ticks *Rhipicephalus sanguineus* in southwestern Spain. *Parasitol. Res.* **2008**, *103*, 119–122. [CrossRef] [PubMed]
- 190. Parola, P.; Socolovschi, C.; Jeanjean, L.; Bitam, I.; Fournier, P.-E.; Sotto, A.; Labauge, P.; Raoult, D. Warmer Weather Linked to Tick Attack and Emergence of Severe Rickettsioses. *PLoS Negl. Trop. Dis.* **2008**, 2, e338. [CrossRef] [PubMed]
- 191. Colomba, C.; Trizzino, M.; Giammanco, A.; Bonura, C.; Di Bona, D.; Tolomeo, M.; Cascio, A. Israeli Spotted Fever in Sicily. Description of two cases and minireview. *Int. J. Infect. Dis.* **2017**, *61*, 7–12. [CrossRef]
- 192. de Sousa, R. Host and microbial risk factors and pathophysiology of fatal *Rickettsia conorii* infection in Portuguese patients. *J. Infect. Dis.* **2008**, *198*, 576–585. [CrossRef]
- 193. Parola, P.; Paddock, C.D.; Socolovschi, C.; Labruna, M.B.; Mediannikov, O.; Kernif, T.; Abdad, M.Y.; Stenos, J.; Bitam, I.; Fournier, P.-E.; et al. Update on Tick-Borne Rickettsioses around the World: A Geographic Approach. *Clin. Microbiol. Rev.* **2013**, *26*, 657–702. [CrossRef]
- 194. Vitale, G.; Mansueto, S.; Rolain, J.-M.; Raoult, D. *Rickettsia massiliae* Human Isolation. *Emerg. Infect. Dis.* **2006**, 12, 174–175. [CrossRef]
- 195. García-García, J.C. Case report: A patient from Argentina infected with *Rickettsia massiliae*. *Am. J. Trop. Med. Hyg.* **2010**, *82*, 691–692. [CrossRef]
- 196. Cascio, A.; Torina, A.; Valenzise, M.; Blanda, V.; Camarda, N.; Bombaci, S.; Iaria, C.; De Luca, F.; Wasniewska, M. Scalp Eschar and Neck Lymphadenopathy Caused by *Rickettsia massiliae*. *Emerg. Infect. Dis.* **2013**, *19*, 836–837. [CrossRef]
- 197. Önder, E. Crimean-Congo haemorrhagic fever. Lancet Infect. Dis. 2006, 6, 203-214. [CrossRef]
- 198. Negredo, A.; Habela, M.Á.; De Arellano, E.R.; Diez, F.; LaSala, F.; López, P.; Sarriá, A.; Labiod, N.; Calero-Bernal, R.; Arenas, M.; et al. Survey of Crimean-Congo Hemorrhagic Fever Enzootic Focus, Spain, 2011-2015. *Emerg. Infect. Dis.* **2019**, 25, 1177–1184. [CrossRef] [PubMed]
- 199. *Rapid Risk Assessment: Crimean-Congo Haemorrhagic Fever in Spain*; European Centre for Disease Prevention and Control: Solna stad, Sweden, 9 September 2016.
- 200. *Communicable Disease Threats Report Week33*; European Centre for Disease Prevention and Control: Solna stad, Sweden, 12–18 August 2018.
- 201. *Communicable Disease Threats Report Week33*; European Centre for Disease Prevention and Control: Solna stad, Sweden, 9–15 August 2020.
- 202. Communicable Disease Threats Report Week33; European Centre for Disease Prevention and Control: Solna stad, Sweden, 28 June–4 July 2020.
- 203. *Communicable Disease Threats Report Week*33; European Centre for Disease Prevention and Control: Solna stad, Sweden, 7–13 June 2020.
- 204. Stuart, J. Suspected case of Crimean-Congo haemorrhagic fever in British traveller returning from Zimbabwe. *Eurosurveillance* **1998**, *2*, 1256. [CrossRef]
- 205. Atkinson, B.; Latham, J.; Chamberlain, J.; Logue, C.; O'Donoghue, L.; Osborne, J.; Carson, G.; Brooks, T.; Carroll, M.; Jacobs, M.; et al. Sequencing and phylogenetic characterisation of a fatal Crimean-Congo haemorrhagic fever case imported into the United Kingdom, October 2012. *Eurosurveillance* 2012, 17, 20327. [PubMed]
- 206. Lumley, S.; Atkinson, B.; Dowall, S.D.; Pitman, J.K.; Staplehurst, S.; Busuttil, J.; Simpson, A.J.; Aarons, E.J.; Petridou, C.; Nijjar, M.; et al. Non-fatal case of Crimean-Congo haemorrhagic fever imported into the United Kingdom (ex Bulgaria) June2014. *Eurosurveillance* 2014, 19, 20864. [CrossRef] [PubMed]
- 207. Teltow, G.J.; Rawlings, J.A.; Fournier, P.V. Isolation of *Borrelia burgdorferi* from arthropods Collected in Texas. *Am. J. Trop. Med. Hyg.* **1991**, 44, 469–474. [CrossRef]
- 208. Feir, D.; Masters, E.; Weil, G.; Santanello, C.R.; Xie, C.-S.; Li, B.-W.; Marconi, R. Evidence Supporting the Presence of *Borrelia burgdorferi* in Missouri. *Am. J. Trop. Med. Hyg.* **1994**, *51*, 475–482. [CrossRef]
- 209. Schulze, T.L.; Bowen, G.S.; Bosler, E.M.; Lakat, M.F.; E Parkin, W.; Altman, R.; Ormiston, B.G.; Shisler, J.K. *Amblyomma americanum*: A potential vector of Lyme disease in New Jersey. *Science* **1984**, 224, 601–603. [CrossRef]
- 210. Rudenko, N.; Golovchenko, M.; Clark, K.; Oliver, J.H.; Grubhoffer, L. Detection of *Borrelia burgdorferi* sensu stricto in *Amblyomma americanum* ticks in the southeastern United States: The case of selective compatibility. *Emerg. Microbes Infect.* **2016**, *5*, e48–e53. [CrossRef]

- 211. Oliver, J.H.; Chandler, F.W.; Luttrell, M.P.; James, A.M.; Stallknecht, D.E.; McGuire, B.S.; Hutcheson, H.J.; Cummins, G.A.; Lane, R.S. Isolation and transmission of the Lyme disease spirochete from the southeastern United States. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 7371–7375. [CrossRef]
- 212. Piesman, J.; Sinsky, R.J. Ability of *Ixodes scapularis*, *Dermacentor variabilis* and *Amblyomma americanum* (Acari: Ixodidae) to Acquire, Maintain and Transmit Lyme Disease Spirochetes (*Borrelia burgdorferi*). *J. Med. Entomol.* 1988, 25, 336–339. [CrossRef] [PubMed]
- 213. Sanders, F.H.; Oliver, H. Evaluation of *Ixodes scapularis*, *Amblyomma americanum* and *Dermacentor variabilis* (Acari: Ixodidae) from Georgia as vectors of a Florida strain of the Lyme disease spirochete, Borrelia burgdorferi. *J. Med. Entomol.* **1995**, 32, 402–406. [CrossRef] [PubMed]
- 214. Armstrong, P.M.; Brunet, L.R.; Spielman, A.; Telford, S.R. Risk of Lyme disease: Perceptions of residents of a Lone Star tick-infested community. *Bull. World Health Organ.* **2001**, *79*, 916–925.
- 215. Kirkland, K.B. Erythema migrans-like rash illness at a camp in North Carolina: A new tick-borne disease? *Arch. Intern. Med.* **1997**, *157*, 2635–2641. [CrossRef] [PubMed]
- 216. Barbour, A.G.; Maupin, G.O.; Teltow, G.J.; Carter, C.J.; Piesman, J. Identification of an Uncultivable *Borrelia* Species in the Hard Tick *Amblyomma americanum*: Possible Agent of a Lyme Disease-like Illness. *J. Infect. Dis.* 1996, 173, 403–409. [CrossRef] [PubMed]
- 217. Killmaster, L.F.; Loftis, A.D.; Zemtsova, G.E.; Levin, M.L. Detection of Bacterial Agents in *Amblyomma americanum* (Acari: Ixodidae) From Georgia, USA and the Use of a Multiplex Assay to Differentiate *Ehrlichia chaffeensis* and *Ehrlichia ewingii*. *J. Med. Entomol.* **2014**, *51*, 868–872. [CrossRef] [PubMed]
- 218. Mixson, T.R. Prevalence of *Ehrlichia*, *Borrelia* and Rickettsial agents in *Amblyomma americanum* (Acari: Ixodidae) collected from nine states. *J. Med. Entomol.* **2006**, *43*, 1261–1268. [CrossRef]
- 219. Stegall-Faulk, T.; Clark, D.C.; Wright, S.M. Detection of *Borrelia lonestari* in *Amblyomma americanum* (Acari: Ixodidae) from Tennessee. *J. Med. Entomol.* 2003, 40, 100–102. [CrossRef]
- 220. Hudman, D.; Sargentini, N. Detection of *Borrelia*, *Ehrlichia* and *Rickettsia* spp. in ticks in northeast Missouri. *Ticks Tick-Borne Dis.* **2016**, 7, 915–921. [CrossRef]
- 221. A Hudman, D.; Sargentini, N.J. Prevalence of Tick-Borne Pathogens in Northeast Missouri. *Mo. Med.* **2018**, 115, 162–168.
- 222. Fritzen, C.M.; Huang, J.; Dunlap, B.; Moncayo, A.C.; Westby, K.; Dunn, J.R.; Yabsley, M.J.; Jones, T.F.; Schardein, M.; Freye, J.D. Infection Prevalences of Common Tick-borne Pathogens in Adult Lone Star Ticks (*Amblyomma americanum*) and American Dog Ticks (*Dermacentor variabilis*) in Kentucky. *Am. J. Trop. Med. Hyg.* 2011, 85, 718–723. [CrossRef] [PubMed]
- 223. Schulze, T.L.; Jordan, R.A.; Schulze, C.J.; Mixson, T.; Papero, M. Relative Encounter Frequencies and Prevalence of Selected *Borrelia*, *Ehrlichia* and *Anaplasma* Infections in *Amblyomma americanum* and *Ixodes scapularis* (Acari: Ixodidae) Ticks from Central New Jersey. *J. Med. Entomol.* 2005, 42, 450–456. [CrossRef]
- 224. Schulze, T.L.; Jordan, R.A.; Healy, S.P.; Roegner, V.E.; Meddis, M.; Jahn, M.B.; Guthrie, D.L. Relative abundance and prevalence of selected *Borrelia* infections in *Ixodes scapularis* and *Amblyomma americanum* (Acari: Ixodidae) from publicly owned lands in Monmouth County NewJersey. *J. Med. Entomol.* 2006, 43, 1269–1275. [CrossRef]
- 225. Williamson, P.C.; Billingsley, P.M.; Teltow, G.J.; Seals, J.P.; Turnbough, M.A.; Atkinson, S.F. Borrelia, Ehrlichia andRickettsia spp. in Ticks Removed from Persons, Texas, USA. *Emerg. Infect. Dis.* **2010**, *16*, 441–446. [CrossRef] [PubMed]
- 226. Mitchell, E.A.; Williamson, P.C.; Billingsley, P.M.; Seals, J.P.; Ferguson, E.E.; Allen, M.S. Frequency and Distribution of Rickettsiae, Borreliae and Ehrlichiae Detected in Human-Parasitizing Ticks, Texas, USA. *Emerg. Infect. Dis.* 2016, 22, 312–315. [CrossRef]
- 227. A Sayler, K.; Loftis, A.; Beatty, S.S.K.; Boyce, C.L.; Garrison, E.; Clemons, B.; Cunningham, M.; Alleman, A.R.; Barbet, A.F. Prevalence of Tick-Borne Pathogens in Host-Seeking *Amblyomma americanum* (Acari: Ixodidae) and *Odocoileus virginianus* (Artiodactyla: Cervidae) in Florida. *J. Med. Entomol.* 2016, 53, 949–956. [CrossRef]
- 228. James, A.M.; Liveris, D.; Wormser, G.P.; Schwartz, I.; Montecalvo, M.A.; Johnson, B.J.B. *Borrelia lonestari* Infection after a Bite by an *Amblyomma americanum* Tick. *J. Infect. Dis.* **2001**, *183*, 1810–1814. [CrossRef] [PubMed]
- 229. Anderson, B.; E Dawson, J.; Jones, D.C.; Wilson, K.H. *Ehrlichia chaffeensis* anew species associated with human ehrlichiosis. *J. Clin. Microbiol.* **1991**, 29, 2838–2842. [CrossRef] [PubMed]

- 230. Buller, R.S. *Ehrlichia ewingii* anewly recognised agent of human ehrlichiosis. *N. Engl. J. Med.* **1999**, 341, 148–155. [CrossRef]
- 231. Ismail, N.; Bloch, K.C.; McBride, J.W. Human Ehrlichiosis and Anaplasmosis. *Clin. Lab. Med.* **2010**, 30, 261–292. [CrossRef]
- 232. Little, S.E.; Barrett, A.W.; Nagamori, Y.; Herrin, B.H.; Normile, D.; Heaney, K.; Armstrong, R. Ticks from cats in the United States: Patterns of infestation and infection with pathogens. *Vet. Parasitol.* **2018**, 257, 15–20. [CrossRef] [PubMed]
- 233. Lee, S.; Kakumanu, M.L.; Ponnusamy, L.; Vaughn, M.F.; Funkhouser, S.W.; Thornton, H.; Meshnick, S.R.; Apperson, C.S. Prevalence of Rickettsiales in ticks removed from the skin of outdoor workers in North Carolina. *Parasites Vectors* **2014**, *7*, 1–14. [CrossRef]
- 234. Maegli, A.; Loy, J.D.; Cortinas, R. Note on *Ehrlichia chaffeensis*, *Ehrlichia ewingii* and *"Borrelia lonestari"* infection in lone star ticks (Acari: Ixodidae), Nebraska, USA. *Ticks Tick-Borne Dis.* **2016**, *7*, 154–158. [CrossRef] [PubMed]
- 235. Wright, C.L.; Gaff, H.D.; Hynes, W.L. Prevalence of *Ehrlichia chaffeensis* and *Ehrlichia ewingii* in *Amblyomma americanum* and *Dermacentor variabilis* collected from southeastern Virginia, 2010–2011. *Ticks Tick-Borne Dis.* **2014**, *5*, 978–982. [CrossRef] [PubMed]
- 236. Harmon, J.R.; Scott, M.C.; Baker, E.M.; Jones, C.J.; Hickling, G.J. Molecular identification of *Ehrlichia* species and host bloodmeal source in *Amblyomma americanum* L. from two locations in Tennessee, United States. *Ticks Tick-Borne Dis.* **2015**, *6*, 246–252. [CrossRef] [PubMed]
- 237. Ijdo, J.W.; Wu, C.; Magnarelli, L.A.; Stafford, K.C.; Anderson, J.F.; Fikrig, E. Detection of *Ehrlichia chaffeensis* DNA in *Amblyomma americanum* Ticks in Connecticut and Rhode Island. *J. Clin. Microbiol.* **2000**, *38*, 4655–4656. [CrossRef] [PubMed]
- 238. Steiert, J.G.; Gilfoy, F. Infection rates of Amblyomma americanum and Dermacentor variabilis by Ehrlichia chaffeensis and Ehrlichia ewingii in southwest Missouri. *Vector-Borne Zoonotic Dis.* **2002**, *2*, 53–60. [CrossRef]
- 239. Simpson, D.T.; Teague, M.; Weeks, J.K.; Lewis, A.D.; D'Addio, P.M.; Moore, J.D.; Thompson, J.; Harris, A.C.; Cannella, R.T.; Kaup, B.Z.; et al. Broad, Multi-Year Sampling Effort Highlights Complex Dynamics of the Tick-Borne Pathogen *Ehrlichia chaffeensis* (Rickettsiales: Anaplasmatacae). *J. Med. Entomol.* **2018**, *56*, 162–168. [CrossRef]
- 240. Yabsley, M.J. Natural History of *Ehrlichia chaffeensis*: Vertebrate hosts and tick vectors from the United States and evidence for endemic transmission in other countries. *Vet. Parasitol.* **2010**, *167*, 136–148. [CrossRef]
- 241. Anziani, O.S.; A Ewing, S.; Barker, R.W. Experimental transmission of a granulocytic form of the tribe Ehrlichieae by *Dermacentor variabilis* and *Amblyomma americanum* to dogs. *Am. J. Vet. Res.* **1990**, 51, 929–931.
- 242. Loftis, A.D.; Reeves, W.K.; Spurlock, J.P.; Mahan, S.M.; Troughton, D.R.; A Dasch, G.; Levin, M.L. Infection of a goat with a tick-transmitted *Ehrlichia* from Georgia, U.S.A. thatis closely related to *Ehrlichia ruminantium*. *J. Vector Ecol.* 2006, *31*, 213–223. [CrossRef]
- 243. Reeves, W.K.; Loftis, A.; Nicholson, W.L.; Czarkowski, A.G. The first report of human illness associated with the Panola Mountain *Ehrlichia* species: A case report. *J. Med. Case Rep.* **2008**, *2*, 139–143. [CrossRef] [PubMed]
- 244. Loftis, A.; Mixson, T.R.; Stromdahl, E.Y.; Yabsley, M.J.; E Garrison, L.; Williamson, P.C.; Fitak, R.R.; Fuerst, P.; Kelly, D.J.; Blount, K.W. Geographic distribution and genetic diversity of the *Ehrlichia* sp. from Panola Mountain in *Amblyomma americanum*. *BMC Infect. Dis.* **2008**, *8*, 54. [CrossRef] [PubMed]
- 245. Burgdorfer, W. Review Article: A Review of Rocky Mountain Spotted Fever (Tick-Borne Typhus) itsAgent and its Tick Vectors in the United States. *J. Med. Entomol.* **1975**, 12, 269–278. [CrossRef]
- 246. McDade, J.E.; Newhouse, M.J. Natural history of *Rickettsia rickettsii*. *Annu. Rev. Microbiol.* **1986**, 40, 287–309. [CrossRef]
- 247. Dantas-Torres, F. Rocky Mountain spotted fever. Lancet Infect. Dis. 2007, 7, 724–732. [CrossRef]
- 248. Kakumanu, M.L.; Ponnusamy, L.; Sutton, H.; Meshnick, S.R.; Nicholson, W.L.; Apperson, C.S. Prevalence of *Rickettsia* Species (Rickettsiales: Rickettsiaceae) in *Dermacentor variabilis* Ticks (Acari: Ixodidae) in North Carolina. *J. Med. Entomol.* **2018**, *55*, 1284–1291. [CrossRef]
- 249. Hecht, J.A.; Allerdice, M.E.; Dykstra, E.A.; Mastel, L.; Eisen, R.J.; Johnson, T.L.; Gaff, H.D.; Varela-Stokes, A.S.; Goddard, J.; Pagac, B.B.; et al. Multistate Survey of American Dog Ticks (*Dermacentor variabilis*) for Rickettsia Species. *Vector-Borne Zoonotic Dis.* **2019**, *19*, 652–657. [CrossRef]

- 250. Demma, L.J.; Traeger, M.S.; Nicholson, W.L.; Paddock, C.D.; Blau, D.M.; Eremeeva, M.E.; Dasch, G.A.; Levin, M.L.; Singleton, J.; Zaki, S.R.; et al. Rocky Mountain Spotted Fever from an Unexpected Tick Vector in Arizona. *N. Engl. J. Med.* 2005, 353, 587–594. [CrossRef]
- 251. Goddard, J.; Norment, B.R. Spotted Fever Group Rickettsiae in the Lone Star Tick, *Amblyomma americanum* (Acari: Ixodidae). *J. Med. Entomol.* **1986**, 23, 465–472. [CrossRef]
- 252. Levin, M.L.; Zemtsova, G.E.; Killmaster, L.F.; Snellgrove, A.; Schumacher, L.B. Vector competence of *Amblyomma americanum* (Acari: Ixodidae) for *Rickettsia rickettsii*. *Ticks Tick-Borne Dis.* **2017**, *8*, 615–622. [CrossRef] [PubMed]
- 253. Wood, H.; Dillon, L.; Patel, S.N.; Ralevski, F. Prevalence of *Rickettsia* species in *Dermacentor variabilis* ticks from Ontario, Canada. *Ticks Tick-Borne Dis.* **2016**, 7, 1044–1046. [CrossRef]
- 254. Dergousoff, S.J.; Gajadhar, A.J.A.; Chilton, N.B. Prevalence of *Rickettsia Species* in Canadian Populations of *Dermacentor andersoni* and *D. variabilis. Appl. Environ. Microbiol.* **2009**, 75, 1786–1789. [CrossRef] [PubMed]
- 255. Yunik, M.E.; Galloway, T.D.; Lindsay, L.R. Assessment of Prevalence and Distribution of Spotted Fever Group Rickettsiae in Manitoba, Canada in the American Dog Tick, *Dermacentor variabilis* (Acari: Ixodidae). *Vector-Borne Zoonotic Dis.* **2015**, *15*, 103–108. [CrossRef] [PubMed]
- 256. Ammerman, N.C. Spotted-fever group *Rickettsia* in *Dermacentor variabilis*, Maryland. *Emerg. Infect. Dis.* **2004**, 10, 1478–1481. [CrossRef] [PubMed]
- 257. Pagac, B.B.; Miller, M.K.; Mazzei, M.C.; Nielsen, D.H.; Jiang, J.; Richards, A.L. *Rickettsia parkeri* and *Rickettsia montanensis*, Kentucky and Tennessee, USA. *Emerg. Infect. Dis.* **2014**, 20, 1750–1752. [CrossRef]
- 258. Smith, M.P.; Ponnusamy, L.; Jiang, J.; Abu Ayyash, L.; Richards, A.L.; Apperson, C.S. Bacterial Pathogens in Ixodid Ticks from a Piedmont County in North Carolina: Prevalence of Rickettsial Organisms. *Vector-Borne Zoonotic Dis.* 2010, 10, 939–952. [CrossRef]
- 259. Fryxell, R.T.T.; Hendricks, B.M.; Pompo, K.; Mays, S.E.; Paulsen, D.J.; Operario, D.J.; Houston, A.E. Investigating the Adult Ixodid Tick Populations and Their Associated *Anaplasma*, *Ehrlichia* and *Rickettsia* Bacteria at a Rocky Mountain Spotted Fever Hotspot in Western Tennessee. *Vector-Borne Zoonotic Dis.* 2017, 17, 527–538. [CrossRef]
- 260. McQuiston, J.H.; Zemtsova, G.; Perniciaro, J.; Hutson, M.; Singleton, J.; Nicholson, W.L.; Levin, M.L. Afebrile Spotted Fever Group *Rickettsia* Infection After a Bite from a *Dermacentor variabilis* Tick Infected with *Rickettsia montanensis*. *Vector-Borne Zoonotic Dis.* **2012**, 12, 1059–1061. [CrossRef]
- 261. Macdonald, A.J.; Hyon, D.W.; Brewington, J.B.; O'Connor, K.E.; Swei, A.; Briggs, C.J. Lyme disease risk in southern California: Abiotic and environmental drivers of *Ixodes pacificus* (Acari: Ixodidae) density and infection prevalence with *Borrelia burgdorferi*. *Parasites Vectors* 2017, 10, 7. [CrossRef]
- 262. Crowder, C.D.; Carolan, H.E.; Rounds, M.A.; Hönig, V.; Mothes, B.; Haag, H.; Nolte, O.; Luft, B.J.; Grubhoffer, L.; Ecker, D.J.; et al. Prevalence of *Borrelia miyamotoi* in Ixodes Ticks in Europe and the United States. *Emerg. Infect. Dis.* **2014**, 20, 1678–1682. [CrossRef] [PubMed]
- 263. Cosson, J.-F.; Michelet, L.; Chotte, J.; Le Naour, E.; Cote, M.; Devillers, E.; Poulle, M.-L.; Huet, D.; Galan, M.; Geller, J.; et al. Genetic characterization of the human relapsing fever spirochete *Borrelia miyamotoi* in vectors and animal reservoirs of Lyme disease spirochetes in France. *Parasites Vectors* **2014**, *7*, 233. [CrossRef] [PubMed]
- 264. Mukhacheva, T.; Salikhova, I.I.; Kovalev, S.Y. Multilocus spacer analysis revealed highly homogeneous genetic background of Asian type of *Borrelia miyamotoi*. *Infect. Genet. Evol.* **2015**, *31*, 257–262. [CrossRef] [PubMed]
- 265. Siński, E.; Welc-Faleciak, R.; Zajkowska, J. Borrelia miyamotoi: A human tick-borne relapsing fever spirochete in Europe and its potential impact on public health. *Adv. Med. Sci.* **2016**, *61*, 255–260. [CrossRef]
- 266. Scoles, G.A.; Papero, M.; Beati, L.; Fish, D. A Relapsing Fever Group Spirochete Transmitted by *Ixodes scapularis* Ticks. *Vector-Borne Zoonotic Dis.* **2001**, *1*, 21–34. [CrossRef]
- 267. Barbour, A.G.; Fish, D.; Hoen, A.G.; Tsao, J.I.; Diuk-Wasser, M.A.; Bunikis, J.; Travinsky, B. Niche Partitioning of *Borrelia burgdorferi* and *Borrelia miyamotoi* in the Same Tick Vector and Mammalian Reservoir Species. *Am. J. Trop. Med. Hyg.* **2009**, *81*, 1120–1131. [CrossRef]
- 268. Platonov, A.E.; Karan, L.S.; Kolyasnikova, N.M.; Makhneva, N.A.; Toporkova, M.G.; Maleev, V.V.; Fish, D.; Krause, P.J. Humans Infected with Relapsing Fever Spirochete *Borrelia miyamotoi*, Russia. *Emerg. Infect. Dis.* **2011**, *17*, 1816–1823. [CrossRef]

- 269. Krause, P.J.; Narasimhan, S.; Wormser, G.P.; Rollend, L.; Fikrig, E.; Lepore, T.; Barbour, A.; Fish, D. Human *Borrelia miyamotoi* Infection in the United States. *N. Engl. J. Med.* **2013**, *368*, 291–293. [CrossRef]
- 270. Gugliotta, J.L.; Goethert, H.K.; Berardi, V.P.; Telford, S.R. Meningoencephalitis from *Borrelia miyamotoi* in an Immunocompromised Patient. *N. Engl. J. Med.* 2013, 368, 240–245. [CrossRef]
- 271. Hovius, J.W.R. A case of meningoencephalitis by the relapsing fever spirochaete *Borrelia miyamotoi* in Europe. *Lancet* **2013**, *382*, 658. [CrossRef]
- 272. Aubry, C.; Socolovschi, C.; Raoult, D.; Parola, P. Bacterial agents in 248 ticks removed from people from 2002 to 2013. *Ticks Tick-Borne Dis.* **2016**, *7*, 475–481. [CrossRef] [PubMed]
- 273. Sarksyan, D.S.; Platonov, A.E.; Karan, L.S.; Shipulin, G.; Sprong, H.; Hovius, J.W. Probability of Spirochete *Borrelia miyamotoi* Transmission from Ticks to Humans. *Emerg. Infect. Dis.* **2015**, 21, 2273–2274. [CrossRef]
- 274. Hofhuis, A.; Herremans, T.; Notermans, D.W.; Sprong, H.; Fonville, M.; van der Giessen, J.W.B.; Van Pelt, W. A Prospective Study among Patients Presenting at the General Practitioner with a Tick Bite or Erythema Migrans in the Netherlands. *PLoS ONE* **2013**, *8*, e64361. [CrossRef] [PubMed]
- 275. Scott, M.C.; Rosen, M.E.; Hamer, S.A.; Baker, E.; Edwards, H.; Crowder, C.; Tsao, J.I.; Hickling, G.J. High-Prevalence *Borrelia miyamotoi* Infection Among Wild Turkeys (*Meleagris gallopavo*) in Tennessee. *J. Med. Entomol.* 2010, 47, 1238–1242. [CrossRef] [PubMed]
- 276. Padgett, K.; Bonilla, D.; Kjemtrup, A.; Vilcins, I.-M.; Yoshimizu, M.H.; Hui, L.; Sola, M.; Quintana, M.; Kramer, V. Large Scale Spatial Risk and Comparative Prevalence of *Borrelia miyamotoi* and *Borrelia burgdorferi* Sensu Lato in *Ixodes pacificus*. *PLoS ONE* **2014**, *9*, e110853. [CrossRef]
- 277. Eshoo, M.W.; Carolan, H.E.; Massire, C.; Chou, D.M.; Crowder, C.D.; Rounds, M.A.; Phillipson, C.A.; Schutzer, S.E.; Ecker, D.J. Survey of *Ixodes pacificus* Ticks in California Reveals a Diversity of Microorganisms and a Novel and Widespread Anaplasmataceae Species. *PLoS ONE* **2015**, *10*, e0135828. [CrossRef]
- 278. Lynn, G.E.; Graham, C.B.; Horiuchi, K.; Eisen, L.; Johnson, T.L.; Lane, R.S.; Eisen, R.J. Prevalence and Geographic Distribution of *Borrelia miyamotoi* in Host-Seeking *Ixodes pacificus* (Acari: Ixodidae) Nymphs in Mendocino County, California. *J. Med. Entomol.* 2018, 55, 711–716. [CrossRef]
- 279. Salkeld, D.J.; Cinkovich, S.; Nieto, N.C. Tick-borne Pathogens in Northwestern California, USA. *Emerg. Infect. Dis.* **2014**, 20, 493–494. [CrossRef]
- 280. Mun, J.; Eisen, R.J.; Eisen, L.; Lane, R.S. Detection of a *Borrelia miyamotoi* Sensu Lato Relapsing-Fever Group Spirochete from *Ixodes pacificus* in California. *J. Med. Entomol.* **2006**, *43*, 120–123. [CrossRef]
- 281. Fedorova, N.; Kleinjan, J.E.; James, D.; Hui, L.T.; Peeters, H.; Lane, R.S. Remarkable diversity of tick or mammalian-associated Borreliae in the metropolitan San Francisco Bay Area, California. *Ticks Tick-Borne Dis.* **2014**, *5*, 951–961. [CrossRef]
- 282. Cutler, S.J.; Vayssier-Taussat, M.; Estrada-Peña, A.; Potkonjak, A.; Mihalca, A.D.; Zeller, H. A new *Borrelia* on the block: *Borrelia miyamotoi*—A human health risk? *Eurosurveillance* 2019, 24, 1800170. [CrossRef] [PubMed]
- 283. Lane, R.S.; Steinlein, D.B.; Mun, J. Human Behaviors Elevating Exposure *toIxodes pacificus* (Acari: Ixodidae) Nymphs and Their Associated Bacterial Zoonotic Agents in a Hardwood Forest. *J. Med. Entomol.* **2004**, 41, 239–248. [CrossRef] [PubMed]
- 284. Newman, E.A.; Eisen, L.; Eisen, R.J.; Fedorova, N.; Hasty, J.M.; Vaughn, C.; Lane, R.S. *Borrelia burgdorferi* Sensu Lato Spirochetes in Wild Birds in Northwestern California: Associations with Ecological Factors BirdBehavior and Tick Infestation. *PLoS ONE* **2015**, *10*, e0118146. [CrossRef] [PubMed]
- 285. Holden, K.; Boothby, J.T.; Kasten, R.W.; Chomel, B.B. Co-detection of *Bartonella henselae*, *Borrelia burgdorferi* and *Anaplasma phagocytophilum* in *Ixodes pacificus* Ticks from California, USA. *Vector-Borne Zoonotic Dis.* 2006, 6, 99–102. [CrossRef] [PubMed]
- 286. Rose, I.; Yoshimizu, M.H.; Bonilla, D.L.; Fedorova, N.; Lane, R.S.; Padgett, K. Phylogeography of *Borrelia* spirochetes in *Ixodes pacificus* and *Ixodes spinipalpis* ticks highlights differential acarological risk of tick-borne disease transmission in northern versus southern California. *PLoS ONE* **2019**, *14*, e0214726. [CrossRef]
- 287. Eisen, R.J.; Mun, J.; Eisen, L.; Lane, R.S. Life Stage-Related Differences in Density of Questing Ticks and Infection with *Borrelia burgdorferi* sensu lato Within a Single Cohort of *Ixodes pacificus*(Acari: Ixodidae). *J. Med. Entomol.* **2004**, *41*, 768–773. [CrossRef]
- 288. Lane, R.S. Acarologic risk of exposure to emerging tick-borne bacterial pathogens in a semi-rural community in northern California. *Vector-Borne Zoonotic Dis.* **2001**, *1*, 197–210. [CrossRef]

- 289. Wright, S.A.; Thompson, M.A.; Miller, M.J.; Knerl, K.M.; Elms, S.L.; Karpowicz, J.C.; Young, J.F.; Kramer, V.L. Ecology ofBorrelia burgdorferiin Ticks (Acari: Ixodidae), Rodents andBirds in the Sierra Nevada Foothills, Placer County, California. *J. Med. Entomol.* **2000**, *37*, 909–918. [CrossRef]
- 290. Lane, R.S.; Fedorova, N.; Kleinjan, J.E.; Maxwell, M. Eco-epidemiological factors contributing to the low risk of human exposure to ixodid tick-borne borreliae in southern California, USA. *Ticks Tick-Borne Dis.* **2013**, *4*, 377–385. [CrossRef]
- 291. Salkeld, D.J.; Nieto, N.C.; Carbajales-Dale, P.; Dale, M.; Cinkovich, S.S.; Lambin, E.F. Disease Risk & Landscape Attributes of Tick-Borne Borrelia Pathogens in the San Francisco Bay Area, California. *PLoS ONE* **2015**, *10*, e0134812. [CrossRef]
- 292. Davis, R.S.; Ramirez, R.A.; Anderson, J.L.; Bernhardt, S. Distribution and Habitat of *Ixodes pacificus* (Acari: Ixodidae) and Prevalence of *Borrelia burgdorferi* in Utah. *J. Med. Entomol.* **2015**, *52*, 1361–1367. [CrossRef] [PubMed]
- 293. Eisen, R.J.; Eisen, L.; Girard, Y.A.; Fedorova, N.; Mun, J.; Slikas, B.; Leonhard, S.; Kitron, U.; Lane, R.S. A spatially-explicit model of acarological risk of exposure to *Borrelia burgdorferi*-infected *Ixodes pacificus* nymphs in northwestern California based on woodland type, temperature andwater vapor. *Ticks Tick-Borne Dis.* 2010, 1, 35–43. [CrossRef]
- 294. Swei, A.; Meentemeyer, R.; Briggs, C.J. Influence of abiotic and environmental factors on the density and infection prevalence of *Ixodes pacificus* (Acari:Ixodidae) with Borrelia burgdorferi. *J. Med. Entomol.* **2011**, *48*, 20–28. [CrossRef] [PubMed]
- 295. Xu, G.; Pearson, P.; Dykstra, E.; Andrews, E.S.; Rich, S.M. Human-Biting Ixodes Ticks and Pathogen Prevalence from California, Oregon and Washington. *Vector-Borne Zoonotic Dis.* **2019**, *19*, 106–114. [CrossRef] [PubMed]
- 296. Burri, C.; Schumann, O.; Gern, L.; Schümann, C. Are Apodemus spp. mice and Myodes glareolus reservoirs for Borrelia miyamotoi, Candidatus Neoehrlichia mikurensis, Rickettsia helvetica, R. monacensis and Anaplasma phagocytophilum? *Ticks Tick-Borne Dis.* **2014**, *5*, 245–251. [CrossRef] [PubMed]
- 297. Szekeres, S.; Coipan, E.C.; Rigó, K.; Majoros, G.; Jahfari, S.; Sprong, H.; Földvári, G. Eco-epidemiology of *Borrelia miyamotoi* and Lyme borreliosis spirochetes in a popular hunting and recreational forest area in Hungary. *Parasites Vectors* **2015**, *8*, 309. [CrossRef] [PubMed]
- 298. Wodecka, B.; Rymaszewska, A.; Skotarczak, B. Host and pathogen DNA identification in blood meals of nymphal *Ixodes ricinus* ticks from forest parks and rural forests of Poland. *Exp. Appl. Acarol.* **2013**, *62*, 543–555. [CrossRef]
- 299. Stuen, S.; Granquist, E.G.; Silaghi, C. *Anaplasma phagocytophilum*—A widespread multi-host pathogen with highly adaptive strategies. *Front. Cell. Infect. Microbiol.* **2013**, *3*, 31. [CrossRef]
- 300. Foley, J.; Piovia-Scott, J. Vector biodiversity did not associate with tick-borne pathogen prevalence in small mammal communities in northern and central California. *Ticks Tick-Borne Dis.* **2014**, *5*, 299–304. [CrossRef]
- 301. Telford, S.R.; Dawson, J.E.; Katavolos, P.; Warner, C.K.; Kolbert, C.P.; Persing, D.H. Perpetuation of the agent of human granulocytic ehrlichiosis in a deer tick-rodent cycle. *Proc. Natl. Acad. Sci. USA* **1996**, 93, 6209–6214. [CrossRef]
- 302. Walls, J.J.; Greig, B.; Neitzel, D.F.; Dumler, J.S. Natural infection of small mammal species in Minnesota with the agent of human granulocytic ehrlichiosis. *J. Clin. Microbiol.* **1997**, *35*, 853–855. [CrossRef] [PubMed]
- 303. Foley, J.; Foley, P.; Brown, R.N.; Lane, R.S.; Dumlers, J.S.; E Madigan, J. Ecology of *Anaplasma phagocytophilum* and Borrelia burgdorferi in the western United States. *J. Vector Ecol.* **2004**, *29*, 41–50. [PubMed]
- 304. Woldehiwet, Z. *Anaplasma phagocytophilum* in Ruminants in Europe. *Ann. N. Y. Acad. Sci.* **2006**, 1078, 446–460. [CrossRef] [PubMed]
- 305. Daniels, T.J.; Falco, R.C.; Schwartz, I.; Varde, S.; Robbins, R.G. Deer ticks (*Ixodes scapularis*) and the agents of Lyme disease and human granulocytic ehrlichiosis in a New York City park. *Emerg. Infect. Dis.* **1997**, 3, 353–355. [CrossRef]
- 306. A Magnarelli, L.; Stafford, K.C.; Mather, T.N.; Yeh, M.T.; Horn, K.D.; Dumler, J.S. Hemocytic rickettsia-like organisms in ticks: Serologic reactivity with antisera to Ehrlichiae and detection of DNA of agent of human granulocytic ehrlichiosis by PCR. *J. Clin. Microbiol.* **1995**, *33*, 2710–2714. [CrossRef]
- 307. Fritz, C.L.; Bronson, L.R.; Smith, C.R.; Crawford-Miksza, L.; Yeh, E.; Schnurr, D. Clinical, epidemiologic andenvironmental surveillance for ehrlichiosis and anaplasmosis in an endemic area of northern California. *J. Vector Ecol.* **2005**, *30*, 4–10.

- 308. Burgdorfer, W.; Lackman, D. Identification of *Rickettsia rckettsii* in the Wood Tick, *Dermacentor andersoni* byMeans of Fluorescent Antibody. *J. Infect. Dis.* 1960, 107, 241–244. [CrossRef]
- 309. Johnson, A.J.; Karabatsos, N.; Lanciotti, R.S. Detection of Colorado tick fever virus by using reverse transcriptase PCR and application of the technique in laboratory diagnosis. *J. Clin. Microbiol.* **1997**, 35, 1203–1208. [CrossRef]
- 310. Burgdorfer, W.; Eklund, C.M. Studies on the ecology of Colorado tick fever virus in Western Montana. *Am. J. Epidemiol.* **1959**, *69*, 127–137. [CrossRef]
- 311. Burgdorfer, W.; Eklund, C.M. Colorado Tick Fever I. Further Ecological Studies in Western Montana. *J. Infect. Dis.* **1960**, 107, 379–383. [CrossRef]
- 312. Geissler, A.L.; Thorp, E.; Van Houten, C.; Lanciotti, R.S.; Panella, N.; Cadwell, B.L.; Murphy, T.; Staples, J.E. Infection with Colorado Tick Fever Virus Among Humans and Ticks in a National Park and Forest, Wyoming. *Vector-Borne Zoonotic Dis.* **2014**, *14*, 675–680. [CrossRef] [PubMed]
- 313. Williamson, B.N.; Fischer, R.J.; Lopez, J.E.; Ebihara, H.; Schwan, T.G. Prevalence and Strains of Colorado Tick Fever Virus in Rocky Mountain Wood Ticks in the Bitterroot Valley, Montana. *Vector-Borne Zoonotic Dis.* **2019**, *19*, 694–702. [CrossRef] [PubMed]
- 314. Brown, S.E.; Miller, B.R.; McLean, R.G.; Knudson, D.L. Co-Circulation of Multiple Colorado Tick Fever Virus Genotypes. *Am. J. Trop. Med. Hyg.* **1989**, *40*, 94–101. [CrossRef] [PubMed]
- 315. Elston, D.M.; Hivnor, C. What's eating you? Dermacentor andersoni. Cutis 2001, 67, 113–115.
- 316. Dworkin, M.S.; Shoemaker, P.C.; Anderson, J.D.E. Tick Paralysis: 33 Human Cases in Washington State, 1946–1996. *Clin. Infect. Dis.* 1999, 29, 1435–1439. [CrossRef]
- 317. Krawczak, F.S.; Nieri-Bastos, F.A.; Nunes, F.P.; Soares, J.F.; Moraes-Filho, J.; Labruna, M.B. Rickettsial infection in *Amblyomma cajennense* ticks and capybaras (*Hydrochoerus hydrochaeris*) in a Brazilian spotted fever-endemic area. *Parasites Vectors* **2014**, *7*, 7. [CrossRef]
- 318. Biggs, H.M.; Behravesh, C.B.; Bradley, K.K.; Dahlgren, F.S.; Drexler, N.A.; Dumler, J.S.; Folk, S.M.; Kato, C.Y.; Lash, R.R.; Levin, M.L.; et al. Diagnosis and Management of Tickborne Rickettsial Diseases: Rocky Mountain Spotted Fever and Other Spotted Fever Group Rickettsioses, Ehrlichioses and Anaplasmosis—United States. *MMWR Recomm. Rep.* **2016**, *65*, 1–44. [CrossRef]
- 319. Soares, J.F.; Soares, H.S.; Barbieri, A.M.; Labruna, M.B. Experimental infection of the tick *Amblyomma cajennense*, Cayenne tick with *Rickettsia rickettsii* theagent of Rocky Mountain spotted fever. *Med. Vet. Entomol.* **2011**, *26*, 139–151. [CrossRef]
- 320. Labruna, M.B. Comparative susceptibility of larval stages of *Amblyomma aureolatum*, *Amblyomma cajennense* and *Rhipicephalus sanguineus* to infection by *Rickettsia rickettsii*. *J. Med. Entomol.* **2008**, 45, 1156–1159. [CrossRef]
- 321. Grattan-Smith, P.J.; Morris, J.G.; Johnston, H.M.; Yiannikas, C.; Malik, R.; Russell, R.; Ouvrier, R. Clinical and neurophysiological features of tick paralysis. *Brain* 1997, 120, 1975–1987. [CrossRef]
- 322. Goodrich, B.; Murray, M. Factors influencing the toxicity of salivary gland extracts of *Ixodes holocyclus* Neumann. *Int. J. Parasitol.* **1978**, *8*, 313–320. [CrossRef]
- 323. Eppleston, K.; Kelman, M.; Ward, M.P. Distribution, seasonality and risk factors for tick paralysis in Australian dogs and cats. *Vet. Parasitol.* **2013**, *196*, 460–468. [CrossRef] [PubMed]
- 324. Edlow, J.A.; McGillicuddy, D.C. Tick Paralysis. *Infect. Dis. Clin. North. Am.* 2008, 22, 397–413. [CrossRef] [PubMed]
- 325. Barker, S.C.; Walker, A.R.; Campelo, D. A list of the 70 species of Australian ticks; diagnostic guides to and species accounts of *Ixodes holocyclus* (paralysis tick), *Ixodes cornuatus* (southern paralysis tick) and *Rhipicephalus australis* (Australian cattle tick); and consideration of the place of Australia in the evolution of ticks with comments on four controversial ideas. *Int. J. Parasitol.* **2014**, 44, 941–953. [CrossRef] [PubMed]
- 326. Logan, T.M.; Watts, D.M.; Linthicum, K.J.; Moulton, J.R.; Bailey, C.L. Experimental Transmission of Crimean-Congo Hemorrhagic Fever Virus by *Hyalomma truncatum* Koch. *Am. J. Trop. Med. Hyg.* **1989**, 40, 207–212. [CrossRef] [PubMed]
- 327. Gonzalez, J.; Camicas, J.; Cornet, J.; Faye, O.; Wilson, M. Sexual and transovarian transmission of Crimean-Congo haemorrhagic fever virus in *Hyalomma truncatum* ticks. *Res. Virol.* **1992**, 143, 23–28. [CrossRef]

- 328. Wilson, M.; Gonzalez, J.-P.; Cornet, J.-P.; Camicas, J.-L. Transmission of Crimean-Congo haemorrhagic fever virus from experimentally infected sheep to *Hyalomma truncatum* ticks. *Res. Virol.* **1991**, 142, 395–404. [CrossRef]
- 329. Linthicum, K.J.; Logan, T.M.; Bailey, C.L.; Dohm, D.J.; Moulton, J.R. Transstadial and Horizontal Transmission of Rift Valley Fever Virus in *Hyalomma truncatum*. *Am. J. Trop. Med. Hyg.* **1989**, 41, 491–496. [CrossRef]
- 330. Nchu, F.; Rand, A. Rift Valley fever outbreaks: Possible implication of *Hyalomma truncatum* (Acari: Ixodidae). *Afr. J. Microbiol. Res.* **2013**, *7*, 3891–3894.
- 331. Kumsa, B.; Socolovschi, C.; Raoult, D.; Parola, P. Spotted fever group rickettsiae in ixodid ticks in Oromia, Ethiopia. *Ticks Tick-Borne Dis.* **2015**, *6*, 8–15. [CrossRef]
- 332. Mediannikov, O.; Diatta, G.; Fenollar, F.; Sokhna, C.; Trape, J.-F.; Raoult, D. Tick-Borne Rickettsioses, Neglected Emerging Diseases in Rural Senegal. *PLoS Neglected Trop. Dis.* **2010**, *4*, e821. [CrossRef] [PubMed]
- 333. Morita, C.; El Hussein, A.R.M.; Matsuda, E.; Gabbar, K.M.A.A.; Muramatsu, Y.; Rahman, M.B.A.; Eleragi, A.M.H.; Hassan, S.M.; Chitambo, A.M.; Ueno, H. Spotted fever group rickettsiae from ticks captured in Sudan. *Jpn. J. Infect. Dis.* **2004**, *57*, 107–109.
- 334. Mutai, B.K.; Wainaina, J.M.; Magiri, C.G.; Nganga, J.K.; Ithondeka, P.M.; Njagi, O.N.; Jiang, J.; Richards, A.L.; Waitumbi, J.N. Zoonotic Surveillance for Rickettsiae in Domestic Animals in Kenya. *Vector-Borne Zoonotic Dis.* **2013**, *13*, 360–366. [CrossRef] [PubMed]
- 335. Tomassone, L.; De Meneghi, D.; Adakal, H.; Rodighiero, P.; Pressi, G.; Grego, E. Detection of *Rickettsia aeschlimannii* and *Rickettsia africae* in ixodid ticks from Burkina Faso and Somali Region of Ethiopia by new real-time PCR assays. *Ticks Tick-Borne Dis.* **2016**, *7*, 1082–1088. [CrossRef]
- 336. Perry, B.; Kruska, R.; Lessard, P.; Norval, R.; Kundert, K. Estimating the distribution and abundance of *Rhipicephalus appendiculatus* in Africa. *Prev. Vet. Med.* **1991**, *11*, 261–268. [CrossRef]
- 337. Mahara, F. Japanese Spotted Fever: Report of 31 Cases and Review of the Literature. *Emerg. Infect. Dis.* **1997**, 3, 105–111. [CrossRef] [PubMed]
- 338. Lu, M.; Tian, J.-H.; Yu, B.; Guo, W.-P.; Holmes, E.C.; Zhang, Y.-Z. Extensive diversity of rickettsiales bacteria in ticks from Wuhan, China. *Ticks Tick-Borne Dis.* **2017**, *8*, 574–580. [CrossRef]
- 339. Seki, M.; Ikari, N.; Yamamoto, S.; Yamagata, Y.; Kosai, K.; Yanagihara, K.; Kakugawa, T.; Kurihara, S.; Izumikawa, K.; Miyazaki, Y.; et al. Severe Japanese Spotted Fever Successfully Treated with Fluoroquinolone. *Intern. Med.* 2006, 45, 1323–1326. [CrossRef]
- 340. Arthan, W.; Sumrandee, C.; Hirunkanokpun, S.; Kitthawee, S.; Baimai, V.; Trinachartvanit, W.; Ahantarig, A. Detection of *Coxiella*-like endosymbiont in *Haemaphysalis* tick in Thailand. *Ticks Tick-Borne Dis.* **2015**, *6*, 63–68. [CrossRef]
- 341. Khoo, J.-J.; Lim, F.-S.; Chen, F.; Phoon, W.-H.; Khor, C.-S.; Pike, B.L.; Chang, L.-Y.; Abubakar, S. *Coxiella* Detection in Ticks from Wildlife and Livestock in Malaysia. *Vector-Borne Zoonotic Dis.* **2016**, *16*, 744–751. [CrossRef]
- 342. Tan, D.S.K.; Smith, C.E.G.; McMahon, D.A.; Bowen, E.T.W. Lanjan Virus aNew Agent isolated from *Dermacentor auratus* in Malaya. *Nat. Cell Biol.* **1967**, 214, 1154–1155. [CrossRef]
- 343. Adamantos, S. Australian tick paralysis in a dog imported into the UK. Vet. Rec. 2005, 83, 327. [CrossRef]
- 344. Attipa, C.; Maguire, D.; Solano-Gallego, L.; Szladovits, B.; Barker, E.N.; Farr, A.; Baneth, G.; Tasker, S. Hepatozoon canis in three imported dogs: A new tickborne disease reaching the United Kingdom. *Vet. Rec.* **2018**, *183*, 716. [CrossRef] [PubMed]
- 345. Hansford, K.M.; Pietzsch, M.; Cull, B.; Medlock, J.M.; Wall, R. Overwintering of the brown dog tick in residential properties in England–raising awareness. *Vet. Rec.* **2015**, *177*, 156. [CrossRef]
- 346. Hansford, K.M.; Medlock, J.M.; Atkinson, B.; Santos-Silva, M.M. Importation of a *Hyalomma lusitanicum* tick into the UK on a dog. *Vet. Rec.* **2016**, *179*, 415. [CrossRef] [PubMed]
- 347. Abdullah, S.; Helps, C.; Tasker, S.; Newbury, H.; Wall, R. Ticks infesting domestic dogs in the UK: A large-scale surveillance programme. *Parasites Vectors* **2016**, *9*, 1–9. [CrossRef]
- 348. Hansford, K.M.; Pietzsch, M.; Cull, B.; Medlock, J.M. Brown dog tick infestation of a home in England. *Vet. Rec.* **2015**, *176*, 129–130. [CrossRef] [PubMed]
- 349. Hansford, K.M.; E Pietzsch, M.; Cull, B.; Medlock, J.M. Importation of *R sanguineus* into the UK via dogs: Tickborne diseases. *Vet. Rec.* **2014**, *175*, 385–386. [CrossRef]
- 350. Featherstone, C.; Phipps, P.; Pietzsch, M.; Hansford, K.; Medlock, J. Tick surveillance in the UK. *Vet. Rec.* **2012**, *171*. [CrossRef]

- 351. Hansford, K.M.; E Pietzsch, M.; Cull, B.; Gillingham, E.L.; Medlock, J.M. Potential risk posed by the importation of ticks into the UK on animals: Records from the Tick Surveillance Scheme. *Vet. Rec.* **2017**, *182*, 107. [CrossRef]
- 352. Bates, P.; Rankin, M.; Shickle, L. Importation of the brown dog or kennel tick (*Rhipicephalus sanguineus*) into the UK. *Vet. Rec.* **2002**, *150*, 224.
- 353. Hansford, K.M.; Carter, D.; Gillingham, E.L.; Hernandez-Triana, L.M.; Chamberlain, J.; Cull, B.; McGinley, L.; Phipps, L.P.; Medlock, J.M. *Hyalomma rufipes* on an untraveled horse: Is this the first evidence of *Hyalomma* nymphs successfully moulting in the United Kingdom? *Ticks Tick-Borne Dis.* **2019**, *10*, 704–708. [CrossRef] [PubMed]
- 354. Jameson, L.J.; Morgan, P.J.; Medlock, J.M.; Watola, G.; Vaux, A.G. Importation of *Hyalomma marginatum*, vector of Crimean-Congo haemorrhagic fever virus into the United Kingdom by migratory birds. *Ticks Tick-Borne Dis.* **2012**, *3*, 95–99. [CrossRef]
- 355. Pietzsch, M.; Mitchell, R.; Jameson, L.J.; Morgan, C.; Medlock, J.; Collins, D.; Chamberlain, J.; Gould, E.; Hewson, R.; Taylor, M.; et al. Preliminary evaluation of exotic tick species and exotic pathogens imported on migratory birds into the British Isles. *Vet. Parasitol.* **2008**, *155*, 328–332. [CrossRef]
- 356. Mihalca, A.D. Ticks imported to Europe with exotic reptiles. *Vet. Parasitol.* **2015**, 213, 67–71. [CrossRef] [PubMed]
- 357. Pietzsch, M.; Quest, R.; Hillyard, P.D.; Medlock, J.M.; Leach, S. Importation of Exotic Ticks into the United Kingdom via the International Trade in Reptiles. *Exp. Appl. Acarol.* **2006**, *38*, 59–65. [CrossRef]
- 358. Kenny, M.J.; Shaw, S.E.; Hillyard, P.D.; Forbes, A.B. Ectoparasite and haemoparasite risks associated with imported exotic reptiles. *Vet. Rec.* **2004**, *154*, 434–435. [CrossRef] [PubMed]
- 359. Pietzsch, M.E. Detection of *Dermacentor marginatus* and a possible *Rickettsia slovaca* case in the United Kingdom—The risk of the visiting traveller. *Travel Med. Infect. Dis.* **2015**, *13*, 200–201. [CrossRef] [PubMed]
- 360. Office for National Statistics. Overseas Travel and Tourism, Monthly. 2019. Available online: https://www.ons.gov.uk/peoplepopulationandcommunity/leisureandtourism/datasets/monthlyoverseastravelandtourismreferencetables (accessed on 1 September 2020).
- 361. Dobson, A.D.M.; Taylor, J.L.; Randolph, S.E. Tick (*Ixodes ricinus*) abundance and seasonality at recreational sites in the UK: Hazards in relation to fine-scale habitat types revealed by complementary sampling methods. *Ticks Tick-Borne Dis.* **2011**, *2*, 67–74. [CrossRef]
- 362. Pérez, D.; Kneubühler, Y.; Rais, O.; Gern, L. Seasonality of *Ixodes ricinus* Ticks on Vegetation and on Rodents and *Borrelia burgdorferi* sensu lato Genospecies Diversity in Two Lyme Borreliosis–Endemic Areas in Switzerland. *Vector-Borne Zoonotic Dis.* **2012**, *12*, 633–644. [CrossRef]
- 363. Schulz, M.; Mahling, M.; Pfister, K. Abundance and seasonal activity of questing *Ixodes ricinus* ticks in their natural habitats in southern Germany in 2011. *J. Vector Ecol.* **2014**, *39*, 56–65. [CrossRef] [PubMed]
- 364. Hubálek, Z. Epidemiology of Lyme Borreliosis. Curr. Probl. Dermatol. 2009, 37, 31–50. [CrossRef] [PubMed]
- 365. Rizzoli, A.; Hauffe, H.; Carpi, G.; Neteler, M.; Rosa, R. Lyme borreliosis in Europe. Eurosurveillance 2011, 16, 1–8.
- 366. Stanek, G. Lyme borreliosis. *Lancet* **2012**, *379*, 461–473. [CrossRef]
- 367. Hofhuis, A. Lyme borreliosis in the Netherlands: Strong increase in GP consultations and hospital admissions in the past ten years. *Eurosurveillance* **2006**, *11*, e060622.2. [CrossRef]
- 368. Vanthomme, K.; Bossuyt, N.; Boffin, N.; van Casteren, V. Incidence and management of presumption of Lyme borreliosis in Belgium: Recent data from the sentinel network of general practitioners. *Eur. J. Clin. Microbiol. Infect. Dis.* 2012, 31, 2385–2390. [CrossRef]
- 369. Letrilliart, L.; Ragon, B.; Hanslik, T.; Flahault, A. Lyme disease in France: A primary care-based prospective study. *Epidemiol. Infect.* **2005**, *133*, 935–942. [CrossRef]
- 370. Smith, R.; Takkinen, J.; Collective Editorial Team. Lyme borreliosis: Europe-wide coordinated surveillance and action needed? *Eurosurveillance* 2006, 11, 2977. [CrossRef]
- 371. Sykes, R.A.; Makiello, P. An estimate of Lyme borreliosis incidence in Western Europe. *J. Public Health* **2016**, 39, 76–87. [CrossRef]
- 372. Granger, D.M. Tick-borne encephalitis among US travelers to Europe and Asia 2000–2009. *Morb. Mortal. Wkly. Rep.* **2010**, *59*, 335–338.
- 373. Reusken, C.B.; Reimerink, J.; Verduin, C.; Sabbe, L.; Cleton, N.; Koopmans, M. Case report: Tick-borne encephalitis in two Dutch travellers returning from Austria, Netherlands Julyand August 2011. *Eurosurveillance* 2011, 16, 20003. [PubMed]

- 374. Rendi-Wagner, P. Risk and Prevention of Tick-borne Encephalitis in Travelers. *J. Travel Med.* **2006**, *11*, 307–312. [CrossRef]
- 375. Haditsch, M.; Kunze, U. Tick-borne encephalitis: A disease neglected by travel medicine. *Travel Med. Infect. Dis.* **2013**, *11*, 295–300. [CrossRef]
- 376. CDC. Anaplasmosis. 2019. Available online: https://www.cdc.gov/anaplasmosis/stats/index.html (accessed on 8 August 2019).
- 377. Dumler, J.S.; Choi, K.-S.; Garcia-Garcia, J.C.; Barat, N.S.; Scorpio, D.G.; Garyu, J.W.; Grab, D.J.; Bakken, J.S. Human Granulocytic Anaplasmosis and *Anaplasma phagocytophilum*. *Emerg. Infect. Dis.* **2005**, *11*, 1828–1834. [CrossRef] [PubMed]
- 378. Blanco, J.; Oteo, J.A. Human granulocytic ehrlichiosis in Europe. *Clin. Microbiol. Infect.* **2002**, *8*, 763–772. [CrossRef] [PubMed]

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