



Complete Genome Sequence of Bovine Viral Diarrhea Virus 2 Japanese Reference and Vaccine Strain KZ-91CP

Asuka Sato,^a Ken-ichiro Kameyama,^c Makoto Nagai,^{a,b} Kentaro Tateishi,^a Keitaro Ohmori,^a Reiko Todaka,^d Kazuhiko Katayama,^d Tetsuya Mizutani,^b Makoto Yamakawa,^c Junsuke Shirai^{a,b}

Cooperative Department of Veterinary Medicine, Faculty of Agriculture, Tokyo University of Agriculture and Technology, Fuchu, Tokyo, Japan^a; Research and Education Center for Prevention of Global Infectious Diseases of Animals, Tokyo University of Agriculture and Technology, Fuchu, Tokyo, Japan^b; National Institute of Animal Health, National Agriculture and Food Research Organization, Tsukuba, Ibaraki, Japan^c; Department of Virology II, National Institute of Infectious Diseases, Musashimurayama, Tokyo, Japan^d

In this study, we report the complete genome sequence of the bovine viral diarrhea virus 2 Japanese reference strain KZ-91CP. The complete genome comprises 12,654 nucleotides and one open reading frame with 4,020 amino acids. A 369-nucleotide-long insertion encoding the chaperone protein DnaJ is found in the nonstructural 2 (NS2) coding region.

Received 27 December 2014 Accepted 5 January 2015 Published 12 February 2015

Citation Sato A, Kameyama K-I, Nagai M, Tateishi K, Ohmori K, Todaka R, Katayama K, Mizutani T, Yamakawa M, Shirai J. 2015. Complete genome sequence of bovine viral diarrhea virus 2 Japanese reference and vaccine strain KZ-91CP. Genome Announc 3(1):e01573-14. doi:10.1128/genomeA.01573-14.

Copyright © 2015 Sato et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Makoto Nagai, m-nagai@cc.tuat.ac.jp.

ovine viral diarrhea virus 2 (BVDV-2) is a member of the genus Pestivirus within the family Flaviviridae, together with the proposed species BVDV-1, classical swine fever virus, and border disease virus. BVDV-2 was first reported during an outbreak of bovine severe acute hemorrhagic syndrome in North America and Canada in the early 1990s (1, 2). Thereafter, BVDV-2 isolates were identified using samples collected worldwide from cattle with clinical signs similar to those accompanying BVDV-1 infection. KZ-91CP was isolated in 1991 from a Holstein cow age 30 months with mucosal disease and was recognized as the first BVDV-2 strain in Japan (3). This strain is used as an inactivated vaccine and a reference virus for laboratory diagnosis in Japan. Recently in Japan, of the two BVDV types (BVDV-1 and BVDV-2), BVDV-2a comprises 16% of all strains of BVDV (4); however, the complete genome sequences of Japanese BVDV-2a strains, including that of KZ-91CP, remained undetermined. This background prompted us to sequence the whole genome of KZ-91CP. In this study, we report the complete genome sequence of Japanese BVDV-2 reference strain KZ-91CP.

Total viral RNA was extracted from infected primary bovine fetal muscle cells using Isogen-LS (Nippon Gene, Toyama, Japan) and normalized to 50 ng using a Qubit 2.0 fluorometer (Invitrogen, Carlsbad, CA). Library construction for deep sequencing was performed using the NEBNext Ultra RNA library prep kit for Illumina version 2.0 (New England BioLabs, Ipswich, MA), according to the manufacturer's guidelines. After assessing the library quality and quantity on a bioanalyzer (Agilent Technologies, Santa Clara, CA) and Qubit 2.0 fluorometer, sequencing was conducted on a MiSeq benchtop sequencer. Sequence data analysis was conducted using the MiSeq reporter program to generate FastQ-formatted sequence data. The contig was assembled from the obtained sequence reads with the de novo assembly command in CLC Genomics Workbench 6.0 (CLC bio, Aarhus, Denmark). To confirm the sequences of the contig, Sanger sequencing of four overlapping reverse transcription-PCR (RT-PCR) products was

performed. The nucleotide sequences of the 5'-untranslated region (UTR) and 3'-UTR termini were determined using the rapid amplification of cDNA ends method (5, 6).

The complete genome of KZ-91CP comprises 12,654 nucleotides (nt), including a 386-nt-long 5' UTR, a 205-nt-long 3' UTR, and a large open reading frame that encompasses 4,020 amino acids (aa). The nonstructural 2 (NS2) coding region contains a 369-nt-long insertion (at amino acid position 1532). BVDVs are classified into two biotypes, cytopathogenic (cp) and noncytopathogenic (ncp), based on their effect on cultured cells (7). Many cp viruses carry host cell-derived insertions and/or duplicated rearranged viral genomic sequences (8, 9). KZ-91CP exhibits a cp phenotype and possesses an insertion encoding the chaperone protein DnaJ (10), which is most frequently observed in the vicinity of position A (aa position 1535) in BVDV-2 (11). KZ-91CP has higher homologies at the amino acid level with recent American and German BVDV-2 strains (98% to 99%) than with the BVDV-2 prototype strain 890 (2a) (91.3%).

Nucleotide sequence accession number. The complete genome sequence of BVDV strain KZ-91CP has been deposited in DDBJ/ENA/GenBank under the accession no. LC006970.

ACKNOWLEDGMENT

We received no financial support for this research.

REFERENCES

- Ridpath JF, Bolin SR, Dubovi EJ. 1994. Segregation of bovine viral diarrhea virus into genotypes. Virology 205:66–74. http://dx.doi.org/ 10.1006/viro.1994.1620.
- Pellerin C, van den Hurk J, Lecomte J, Tijssen P. 1994. Identification of a new group of bovine viral diarrhea virus strains associated with severe outbreaks and high mortalities. Virology 203:260–268. http://dx.doi.org/ 10.1006/viro.1994.1483.
- 3. Nagai M, Sato M, Nagano H, Pang H, Kong X, Murakami T, Ozawa T, Akashi H. 1998. Nucleotide sequence homology to bovine viral diarrhea

virus 2 (BVDV 2) in the 5' untranslated region of BVDVs from cattle with mucosal disease or persistent infection in Japan. Vet Microbiol **60**: 271–276. http://dx.doi.org/10.1016/S0378-1135(98)00158-8.

- Matsuno K, Sakoda Y, Kameyama K, Tamai K, Ito A, Kida H. 2007. Genetic and pathobiological characterization of bovine viral diarrhea viruses recently isolated from cattle in Japan. J Vet Med Sci 69:515–520. http://dx.doi.org/10.1292/jvms.69.515.
- Ishikawa K, Nagai H, Katayama K, Tsutsui M, Tanabayashi K, Takeuchi K, Hishiyama M, Saitoh A, Takagi M, Gotoh K, Muramatsu M, Yamada A. 1995. Comparison of the entire nucleotide and deduced amino acid sequences of the attenuated hog cholera vaccine strain GPEand the wild-type parental strain ALD. Arch Virol 140:1385–1391. http:// dx.doi.org/10.1007/BF01322665.
- Katayama K, Kurihara C, Fukushi S, Hoshino FB, Ishikawa K, Nagai H, Ando T, Oya A. 1995. Characterization of the hog cholera virus 5' terminus. Virus Genes 10:185–187. http://dx.doi.org/10.1007/BF01702600.

- Peterhans E, Bachofen C, Stalder H, Schweizer M. 2010. Cytopathic bovine viral diarrhea viruses (BVDV): emerging pestiviruses doomed to extinction. Vet Res 41:44. http://dx.doi.org/10.1051/vetres/2010016.
- 8. Ridpath JF, Qi F, Bolin SR, Berry ES. 1994. Natural recombination in bovine viral diarrhea viruses. Arch Virol Suppl 9:239–244.
- Vilcek S, Greiser-Wilke I, Nettleton P, Paton DJ. 2000. Cellular insertions in the NS2-3 genome region of cytopathic bovine viral diarrhoea virus (BVDV) isolates. Vet Microbiol 77:129–136. http://dx.doi.org/ 10.1016/S0378-1135(00)00269-8.
- Rinck G, Birghan C, Harada T, Meyers G, Thiel HJ, Tautz N. 2001. A cellular J-domain protein modulates polyprotein processing and cytopathogenicity of a pestivirus. J Virol 75:9470–9482. http://dx.doi.org/ 10.1128/JVI.75.19.9470-9482.2001.
- Ridpath JF, Neill JD. 2000. Detection and characterization of genetic recombination in cytopathic type 2 bovine viral diarrhea viruses. J Virol 74:8771–8774. http://dx.doi.org/10.1128/JVI.74.18.8771-8774.2000.