Research Article

A Binary Supramolecular Assembly with Intense Fluorescence Emission, High pH Stability, and Cation Selectivity: Supramolecular Assembly-Induced Emission Materials

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We construct a fluorescent supramolecular system (TPE- $Q_4 \,\subset$ DSP5) of excellent tolerance to a wide range of pH by the facile selfassembly of a new pillar[5]arene bearing disulfonated arms (DSP5) with an AIE-active tetraphenylethene-based tetratopic guest bearing four quaternary ammonium binding sites (TPE- Q_4), which exhibits strong blue emission even in dilute aqueous solutions along with much higher quantum yield and longer fluorescence lifetime than TPE- Q_4 itself. This appreciable property can be attributed to the supramolecular assembly-induced emission (SAIE) mechanism endowed by the host-guest inclusion complexation based on synthetic macrocycles. Remarkably, the enhanced fluorescence of the supramolecular assembly is quenched efficiently and exclusively by ferric ions in water with a high Stern–Volmer formula constant of $1.3 \times 10^5 \text{ mol}^{-1}$, demonstrating the excellent cation selectivity and visualized responsiveness in ion sensing and detection.

1. Introduction

Fluorescent supramolecular assemblies represent an extraordinarily attractive class of functional systems, due to their integrated functionalities, highly tunable fluorescence properties, and good responsiveness towards external stimuli [1]. The inherent features of their fluorescent entities that can be further mediated by the rational design of supramolecular building blocks endow them extensive potentials in smart materials and light-emitting devices [2]. In 2001, Tang and coworkers discovered some propeller-like molecules with π -conjugated structures that exhibited weak fluorescence in solution but bright emission in aggregated states in sharp contrast to traditional fluorescent molecules of aggregation caused quenching (ACQ) and creatively named this phenomenon aggregation-induced emission (AIE) [3]. Many fluorophores with AIE properties (AIEgens) have been discovered and extensively investigated for applications in chemistry and materials science [4, 5]. The mechanism of fluorescent emission enhancement of AIEgens in the aggregated states mainly lies in the restriction of intramolecular motion

(RIM) and the blockade of nonradiation energy dissipation channel from the excited state to ground state [4, 5].

Inspired by the mechanism of AIE, researchers have successfully developed a supramolecular pathway, namely, supramolecular assembly-induced emission (SAIE), to achieve effective emission enhancement of AIEgens in the ordered assembly states no matter in dilute solutions or aggregated solid states [2, 6-19]. In 2014, Tang and coworkers covalently linked a cyclodextrin with tetraphenylethene (TPE) and utilized self-inclusion complexation for RIM to achieve enhanced emission in aqueous solutions, which represented the prototype SAIE study and paved the way of recruiting supramolecular macrocycles to manipulate the fluorescent properties of AIEgens [6]. Considerable effort has been made in the rapid development of SAIE recently, accompanied by the synthesis of functional supramolecular assemblies with tunable emission [7-19]. Various types of supramolecular fluorescent assemblies based on AIEgens and functional synthetic macrocycles have been reported for applications in cell imaging, [11] fluorescent supramolecular polymers and gels [7-10, 13, 14],



Scheme 1: Chemical structures and cartoons of DSP5 and TPE-Q₄ and the schematic presentation of their self-assembly into a fluorescent supramolecular system for the selective detection of Fe^{3+} ions.

room-temperature phosphorescence [15], controlled drug delivery [16], and other stimuli-responsive materials [17–19]. But nonetheless, challenges in their preparation and application remain.

On the other hand, pillarenes, as an important class of macrocyclic hosts in the realm of supramolecular chemistry and advanced functional materials, are attracting more and more attention due to the unique pillar-like rigid structure, electron-rich cavity, well-studied host-guest properties, and facile modification methodology [20–22]. Functionalized pillarenes have played important roles in ion detection [23], antibacterial therapeutics [24], supramolecular nanovalves

[25–29], and so on [30]. Significantly, by elaborate functionalization, pillarenes have been utilized as an important candidate of synthetic macrocycles in the construction of stimuliresponsive fluorescent materials through the marriage with AIEgens [2].

Herein, we report on the fabrication of a pH-tolerant fluorescent system for specific cation recognition based on supramolecular self-assembly of disulfonated pillar[5]arene (DSP5) and a water-soluble TPE-based tetratopic guest (TPE-Q₄) with four quaternary ammonium binding sites (Scheme 1). Because of the high binding affinity of DSP5 towards TPE-Q₄, a supramolecular assembly (TPE-Q₄) \subset

DSP5) was fabricated in aqueous solution via host-guest interaction, exhibiting bright blue emission in a wide range of pH conditions. Meanwhile, this supramolecular assembly shows a specific response of fluorescence quenching to Fe^{3+} through the coordination between the sulfonate arms of DSP5 and Fe^{3+} ions, endowing the material with smart cation responsiveness.

2. Results

2.1. Host-Guest Complexation. DSP5 was successfully designed and synthesized for the first time via a Williamson ether-type synthetic approach (Scheme S1). To investigate the host-guest property of DSP5 toward TPE-Q4, the following experiments have been conducted. A TPE-based monotopic guest (TPE-Q) with single quaternary ammonium binding site was chosen as a model guest for the study of the host-guest binding between DSP5 and the guest linker moiety on TPE in deionized water at 298 K via fluorescence titration (Figure S1). The stoichiometric ratio of DSP5 and the monotopic model guest TPE-Q was determined to be 1:1 and their association constant (K_a) was calculated to be (2.68 \pm 0.24) \times 10 5 $M^{-1}.$ The 2D NOESY spectrum (Figures S2 and S3) of the 4:1 mixture of DSP5 (4.0 mM) and TPE-Q₄ (1.0 mM) in D_2O showed obvious correlations between the phenyl protons H_a of DSP5 and the alkyl chains protons H1-H4 and methyl protons H9 of TPE-Q4, indicating the existence of strong host-guest interactions between DSP5 and TPE-Q₄ and the inclusion of alkyl chains of TPE-Q₄ into the cavities of DSP5 macrocycles. Furthermore, DOSY experiments were employed to investigate the formation of TPE-Q₄ \subset DSP5 in water (see Figure S4). The measured weight-average diffusion coefficient (D) of TPE-Q₄ decreased from 9.66 \times 10⁻¹⁰ to 8.85 \times 10⁻¹⁰ m²/s with the addition of 4 equiv. of DSP5, suggesting the formation of supramolecular assemblies. Experimental evidence given above has demonstrated the successful construction of the 1:4 supramolecular inclusion complex of TPE-Q₄ \subset DSP5.

2.2. Fluorescent Properties and Morphology. Fluorescent changes of TPE-Q₄ (1 μ M) in water in the presence of different concentrations of DSP5 were monitored by fluorescence spectroscopy (Figure 1). The emission of TPE-Q₄ solution is negligible; however upon addition of DSP5, the whole system exhibited green emission color and the fluorescence intensity dramatically increased, reaching the maxima when 4 eq. of DSP5 was added ([DSP5]/[TPE-Q₄] = 4:1) (Figure 1(a)). The rational explanation is that the intramolecular rotation of phenyl rings of TPE-Q₄ was restricted by DSP5 upon hostguest inclusion complexation and supramolecular assembly (SAIE). The maximum fluorescence enhancement at the hostguest molar ratio of 4:1 indicated a complete formation of TPE-Q₄ \subset DSP5; thus this ratio was chosen for the following studies. Additional supporting data including UV-Vis absorption spectra have also been obtained (Figure S5). It is worth mentioning that negligible changes (Figure 1(b)) could be observed in the fluorescence intensity of TPE-Q₄ \subset

DSP5 over a wide range of pH (3-13), proving its great pH tolerance and system robustness.

To further investigate the emissive characteristics of TPE- $Q_4 \in DSP5$, time-resolved fluorescence data of TPE-Q₄ and TPE-Q₄ \subset DSP5 were collected (Figures 1(c) and 1(d), Table S1). The absolute fluorescence quantum yield of TPE-Q₄ \subset DSP5 was 54.40%, which is much higher than that of bare TPE-Q₄ (0.68%). The photoluminescence decay profiles of TPE-Q₄ and TPE-Q₄ \subset DSP5 suggested that the fluorescence lifetime of TPE-Q4 itself is 1.14 ns, while the excited state of TPE-Q₄ \subset DSP5 relaxed mainly through a slow pathway with a lifetime of 5.64 ns. Interestingly, TPE-Q₄ \subset DSP5 showed one order of magnitude smaller nonradiative decay rate constant $(8.08 \times 10^7 \text{ s}^{-1})$ and ca. 16 times bigger radiative decay rate constant $(9.64 \times 10^7 \text{ s}^{-1})$ as compared to bare TPE-Q₄ $(8.68 \times 10^8 \text{ s}^{-1} \text{ and } 5.90 \times 10^6 \text{ s}^{-1}, \text{ respectively})$ (Table S1). These experimental results further support the mechanism of SAIE in the system that the intramolecular motions of the TPE-Q₄ units are restricted by DSP5 macrocycles, which block the nonradiative relaxation pathway and populated excitons that underwent radiative decay. The above results evidenced that TPE-Q₄ \subset DSP5 can emit strong fluorescence even in a dilute solution and DSP5 synthetic macrocycle plays a crucial role in the fluorescence enhancement of this supramolecular assembled system.

Based on the aforementioned knowledge, we deduced that the amphiphilic nature of TPE-Q₄ would lead to its selfassembly in aqueous solution, while the ensembles might undergo observable alteration by the host-guest complexation upon addition of DSP5. To obtain direct evidence, the size and morphology of the supramolecular assemblies were studied by scanning electron microscope (SEM) and dynamic light scattering (DLS). Interestingly, cube-shaped assemblies of TPE-Q4 with a diameter of ~300 nm and spherical micelles of DSP5 with a diameter of ~20 nm were observed, respectively (Figures 2(a1), 2(a2), 2(b1), and 2(b2)). However, as we envisioned, TPE-Q₄ \subset DSP5 exhibited bulky cube-like morphology, yet it possessed a larger diameter of ~ 500 nm as compared with the nanostructures formed by TPE- Q_4 or DSP5 alone (Figures 2(cl) and 2(c2)). The formation of supramolecular host-guest complex between DSP5 and TPE-Q₄ affected the overall amphiphilic nature including the size of building blocks (TPE-Q₄ \subset DSP5 versus TPE-Q₄ or DSP5) and introduced the inter-building block electrostatic repulsions, simultaneously contributing to the increased size of the assembled architectures.

2.3. Selective Detection of Fe^{3+} Ion. Inspired by the above advances, fluorescence experiments were conducted to investigate the responsiveness of TPE-Q₄ \subset DSP5 toward different analytes such as ions. Fluorescence quenching degrees and the Stern–Volmer formula constants of TPE-Q₄ \subset DSP5 upon addition of different cations and anions at the same concentration were provided (Figures 3(a) and 3(b)), with Cl⁻ and Na⁺ as the counterions, respectively. Surprisingly, only Fe³⁺ ions caused a remarkable fluorescence quenching (1 - $I/I_0 = 0.84$) with a considerably high Stern–Volmer formula constant ($K_{sy} = 2.7 \times 10^5 \text{ mol}^{-1}$). To further confirm this



FIGURE 1: Fluorescence characterizations of TPE-Q₄ and TPE-Q₄ \subset DSP5. (a) Fluorescence spectra of TPE-Q₄ (1 μ M) in water with different concentrations of DSP5 (inset images: (A) DSP5; (B) TPE-Q₄ \subset DSP5; (C) TPE-Q₄ in water); (b) fluorescence spectra of TPE-Q₄ \subset DSP5 in PBS of different pH values. Experimental conditions: $\lambda_{ex} = 330$ nm; $\lambda_{em} = 475$ nm; slit widths: Ex. 5 nm, Em. 5 nm; 298 K; [TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M. (c) Fluorescence decay profiles and (d) absolute fluorescence quantum yields of TPE-Q₄ and TPE-Q₄ \subset DSP5. Experimental conditions: $\lambda_{ex} = 365$ nm, 298 K, [TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M in water.

ion sensing specificity, experiments on competitive cations were performed and the results showed that fluorescence quenching degrees are as high (~1 - $I/I_0 > 0.75$) with or without competition cations (Figure S6). The limit of detection (LOD) of TPE-Q₄ \subset DSP5 to Fe³⁺ was calculated to be 8.6 $\times 10^{-7}$ M on the basis of 3 δ /S, indicating that the

material possesses a highly sensitive responsiveness to Fe³⁺ (Figure S8). Considering that the material exhibits enhanced fluorescence with good pH stability, the detection of Fe³⁺ by TPE-Q₄ \subset DSP5 was further testified under varied pH circumstances, and efficient fluorescence quenching was observed within the wide pH range in the presence of Fe³⁺

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FIGURE 2: Morphology and size characterizations. SEM images and DLS measurements of (a1) and (a2) TPE-Q₄, (b1) and (b2) DSP5, (c1) and (c2) TPE-Q₄ \subset DSP5, and (d1) and (d2) TPE-Q₄ \subset DSP5 with the addition of Fe³⁺ ([TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M).

(Figure S9), which denoted that the material can be applied for the selective detection of Fe^{3+} in aqueous conditions without the interference of pH levels.

2.4. Investigation of the Quenching Mechanism. To further explore the quenching mechanism, a series of measurements were carried out. SEM image of TPE-Q₄ \subset DSP5 in the presence of Fe³⁺ showed irregular morphology, which was different from the assembly structure of TPE-Q₄ \subset DSP5 itself (Figures 2(d1) and 2(d2)); thus we consider that a complexation between DSP5 and Fe³⁺ occurred. In order to exclude the influence of the electron-rich cavity of DSP5, monomer molecule (DSNa) with sulfonate groups was synthesized. SEM study demonstrated that Fe^{3+} could coordinate with the sulfonate groups of DSNa (Figure S10). Moreover, ¹H NMR experiments were carried out to confirm the coordination of DSP5 and Fe³⁺ (Figure S11). Because TPE-Q₄ \subset DSP5 precipitated from D₂O in the NMR tube, no peaks could be observed in the ¹H NMR spectrum. However, the proton peaks of TPE-Q₄ reappeared after the addition of Fe^{3+} but the proton peaks of DSP5 were still absent in the spectrum, suggesting that DSP5 coordinated with Fe³⁺, forming waterinsoluble coordination polymers. Fluorescent images were also obtained to further confirm the existence of TPE-Q₄ in the aqueous media, where the fluorescence recovered when DSP5 was added after the precipitation (Figure S12).

The ion-responsive property of TPE-Q₄ \subset DSP5 and the interactions of DSP5 and Fe³⁺ were described in a more intuitive way in Figure 3(c). The TPE-Q₄ \subset DSP5 water solution (B) was clear and emitted intensely under 365 nm irradiation, yet precipitation and quenching occurred after adding Fe³⁺ (C) at a higher concentration. After centrifugation, the precipitates were separated from the solutions (Figure S13). However, A and B solutions still showed Tyndall effect but C and D solutions did not, indicating larger supramolecular assemblies in C and D than in A and B solutions. Importantly, the precipitation in D further confirmed that DSP5 interacted with Fe^{3+} , leaving TPE-Q₄ free in water, where the fluorescence was greatly quenched.

To prove the coordination effect between sulfonate groups of DSP5 and Fe³⁺, we used X-ray photoelectron spectroscopy (XPS) to record the valence state changes of DSP5 before and after the addition of Fe³⁺ (Figures 4(a)–4(d)). The XPS spectra demonstrated that the binding energies of the Cls peaks at 284.7 eV, 290.0 eV, and 286.2 eV underwent no obvious changes. But the binding energy of the S2p peak at 168.4 eV, attributing to the sulfonate group, changed to 168.8 eV upon the addition of Fe³⁺ [31]. Moreover, the binding energy of Fe2p (712.2 eV) is different from the ferric chloride (711.5 eV) according to previous literature [32]. Besides, solid-state CP-MAS ¹³C NMR of the DSP5 and DSP5+Fe³⁺ powder also provided convincing evidences for the coordination of DSP5 and Fe³⁺ (Figure 4(e)).

3. Discussion

In conclusion, we have successfully synthesized a novel watersoluble disulfonate-functionalized pillar[5]arene (DSP5), based on which a pH-stable fluorescent material (TPE- $Q_4 \subset$ DSP5) was facilely designed and constructed, exhibiting SAIE properties. The as-prepared supramolecular material showed excellent photoluminescent characteristics of emitting switchable bright blue fluorescence even in dilute aqueous solution. In addition, the supramolecular assembly possessed superior responsiveness to Fe³⁺ in water with high selectivity and sensitivity, since the enhanced fluorescence arising from TPE- $Q_4 \subset$ DSP5 could be efficiently quenched









FIGURE 3: Highly selective detection of Fe³⁺ by TPE-Q₄ \subset DSP5. (a) Fluorescence quenching degrees of TPE-Q₄ \subset DSP5 in the presence of different cations and anions in water. (b) The Stern–Volmer formula constants of TPE-Q₄ \subset DSP5 upon interacting with different cations and anions in water. (c) The pictures of (A) TPE-Q₄ \subset DSP5, (C) TPE-Q₄ \subset DSP5 + Fe³⁺, and (D) DSP5 + Fe³⁺ in water. Experimental conditions: $\lambda_{ex} = 330$ nm; $\lambda_{em} = 475$ nm; slit widths: Ex. 5 nm, Em. 5 nm; 298 K; [TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M, [Fe³⁺] = 20 μ M, [Ions] = 20 μ M.

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FIGURE 4: Coordination effect between DSP5 and Fe³⁺. (a) C1s, (b) O1s, and (c) S2p regions of XPS spectra of DSP5 before (up) and after (down) the addition of Fe³⁺, and (d) Fe2p regions of XPS spectrum of DSP5+Fe³⁺ powder. (e) Solid-state CP-MAS ¹³C NMR characterization of DSP5 and DSP5 + Fe³⁺ powder.

by the coordination effect of DSP5 and Fe³⁺. We envision that this new type of supramolecular fluorescent materials will find great applications in various fields, including fluorescent probing, cell imaging, and visualized monitoring of metal ions.

4. Materials and Methods

4.1. Materials and Characterization. All reagents were commercially available and used as supplied without further purification. ¹H NMR spectra were collected on a temperature-controlled 300 MHz spectrometer. ¹³C NMR spectra were recorded on a DMX-500 spectrometer. Solidstate cross-polarization magic angle spinning (CP/MAS) ¹³C NMR spectra of polymers were measured on a Bruker Digital Avance III HD 400 WB (400 MHz) NMR spectrometer at ambient temperature with a magic angle spinning rate of 7.0 kHz internal reference. 2D diffusion-ordered NMR spectroscopy (DOSY) and 2D nuclear Overhauser effect NMR spectroscopy (NOESY) were recorded on a Bruker Avance DMX 600 spectrophotometer. High-resolution electrospray ionization mass spectra (HRESI-MS) were obtained on a Bruker 7-Tesla FT-ICR mass spectrometer equipped with an electrospray source (Billerica, MA, USA). The UVvis spectra were collected via UV-vis spectroscopy on a Shimadzu UV-1800 spectrophotometer. Deionized water, purifying by Experimental Water System (Lab-UV-20), was used in relevant experiments. Scanning electron microscope (SEM) investigations were carried out on a HITACHI-SU8020 instrument. Dynamic light scattering (DLS) results were obtained on a Zetasizer Nano ZS instrument. The fluorescence experiments were conducted on a RF-5301 spectrofluorometer (Shimadzu Corporation, Japan). The surface compositions of polymers were determined by X-ray photoelectron spectroscopy (XPS) measured on a PREVAC XPS/UPS System using monochromatic Al K_{α} (1486.7 eV) radiation. The time-resolved fluorescence decay curves and absolute fluorescence quantum yields were obtained on a FLS920 instrument (Edinburgh Instrument) with the excitation of 330 nm.

4.2. Syntheses of Compound. The detailed synthesis procedures and characterizations of the studied compound (DSP5, TPE-Q₄, TPE-Q, and DSNa) can be found in the Supplementary Materials.

4.3. Detection of Fe^{3+} . Experiments were carried out to examine the potential for detecting cations and anions. Firstly, we prepared the DSP5 and TPE-Q₄ \subset DSP5 aqueous solution ([TPE-Q₄] = 4 μ M, [DSP5] = 1 μ M). 40 μ L M(Cl)_x aqueous solution (M^{x+} = Na⁺, Fe³⁺, Fe²⁺, Co²⁺, Ni²⁺, Cu²⁺, Cu⁺, Zn²⁺, Cd²⁺, Hg²⁺, Mg²⁺, K⁺, Ca²⁺, NH₄⁺, Pr³⁺, Nd³⁺, Sm³⁺, Eu³⁺, Gd³⁺, Tb³⁺, Dy³⁺, Ho³⁺, Er³⁺, Tm³⁺, Yb³⁺, Lu³⁺, and Al³⁺, [M^{x+}] = 2 mM) and 40 μ L (Na)_yN aqueous solution (N^{y-} = Cl⁻, F⁻, OH⁻, ClO⁻, CH₃COO⁻, NO₂⁻, HPO₄²⁻, H₂PO₄⁻, PO₄³⁻, S₂O₃²⁻, HSO₃⁻, SO₄²⁻, S₂O₄²⁻, HCO₃⁻, CO₃²⁻, [N^{y-}] = 2 mM) were added into the above TPE-Q₄ \subset DSP5 solution (3.96 mL), respectively. Then, the fluorescence

of each solution was recorded immediately. Similarly, interference metal ion with the same concentration as Fe³⁺ was added into the TPE-Q₄ \subset DSP5 solution containing Fe³⁺ to further investigate the fluorescence recognition of Fe³⁺. The fluorescence quenching induced by Fe³⁺ was investigated to confirm whether the developed TPE-Q₄ \subset DSP5 could be used to quantitative detection of Fe³⁺. Thus, different volumes of Fe³⁺ solutions (1 mM, 0~28 μ L) were added into TPE-Q₄ \subset DSP5 solution (1 μ M) and the fluorescence intensities were recorded, respectively.

4.4. Preparation of DSP5 + Fe^{3+} Powder. FeCl₃ aqueous solution (1 mL, 0.1 mM) was added to DSP5 aqueous solution (0.1 mM, 8 mL) and the precipitates were generated immediately. Let the mixture stood for about 24 h and the precipitates were isolated by centrifuge at 10000 rpm. Then the product was washed with deionized water for three times to remove the FeCl₃ residue and dried under high vacuum oven to get a brown powder.

Conflicts of Interest

The authors declare no financial conflicts of interest.

Authors' Contributions

Conceptualization: Xu Wang, Xiao-Yu Jin, and Ying-Wei Yang. Methodology: Xu Wang, Xiao-Yu Jin, and Ying-Wei Yang. Investigation: Xu Wang and Xiao-Yu Jin. Writing original draft: Xu Wang and Xin-Yue Lou. Writing review & editing: Feng Liang and Ying-Wei Yang. Funding acquisition: Ying-Wei Yang. Resources: Feng Liang and Ying-Wei Yang. Supervision: Ying-Wei Yang. All the authors have approved the final version of the manuscript.

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Supplementary Materials

Scheme S1. Synthetic route of the targeted compound DSP5. Scheme S2. Synthetic route of the targeted compound TPE- Q_4 . Scheme S3. Synthetic route of the targeted compound TPE-Q. Scheme S4. Synthetic route of the targeted compound DSNa. Figure S1. The study of (a) and (b) association constant (K_a), and (c) stoichiometric ratio between DSP5 and TPE-Q. Figure S2. NOESY spectrum (600 MHz, D₂O, 298 K) of a mixture of TPE-Q₄ (1.0 mM) and DSP5 (4.00 mM). Figure S3. Partial NOESY spectrum (600 MHz, D₂O, 298 K) of a mixture of TPE-Q₄ (1.0 mM) and DSP5 (4.00 mM). Figure S4. DOSY spectrum (600 MHz, D₂O, 298 K) of (a) TPE-Q₄ (1.00 mM) and (b) a mixture of TPE-Q₄ (1.0 mM) and

DSP5 (4.00 mM). Figure S5. UV-vis spectra of TPE-Q₄ (1 μ M) in water with different concentrations of DSP5. Figure S6. Fluorescence quenching degrees in competition experiments of TPE-Q₄ \subset DSP5 in water. Front: TPE-Q₄ \subset DSP5 with different competing cations; back: TPE-Q₄ \subset DSP5 with Fe³⁺ and the same equiv of cations. (Experimental conditions: λ_{ex} = 330 nm; λ_{em} = 475 nm; slit widths: Ex. 5 nm, Em. 5 nm; 298 K; [TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M, [Fe³⁺] = 20 μ M, $[Ions] = 20 \ \mu M$). Figure S7. Fluorescence quenching degrees and the Stern–Volmer formula constants of TPE-Q₄ \subset DSP5 in the presence of different cations and anions in water. 2D plots for all the M^{x+} with Cl^- (a, c) and Na^+ with N^{y-} (b, d). Figure S8. (a) Fluorescence spectra of supramolecular assemblies TPE-Q₄ \subset DSP5 aqueous solution in the presence of increasing amounts of Fe^{3+} ; (b) the photograph of the linear range (experimental conditions: $\lambda_{ex} = 330$ nm; $\lambda_{em} =$ 475 nm; slit widths: Ex. 5 nm, Em. 5 nm; 298 K; [TPE-Q₄] = 1 μ M, [DSP5] = 4 μ M). Figure S9. Fluorescence spectra of TPE-Q4 \subset DSP5 in the presence of Fe³⁺ under different pH values. Figure S10. SEM images of (a) DSP5+Fe³⁺ and (b) DSNa+Fe³⁺; TEM images of (c) DSP5+Fe³⁺ and (d) TPE- $Q_4 \subset DSP5+Fe^{3+}$ ([DSP5] = 10 μ M, [DSNa] = 50 μ M, $[DSP5]/[Fe^{3+}] = 1:20$, $[DSNa]/[Fe^{3+}] = 1:20$). Figure S11. ¹H NMR spectra of TPE-Q₄, TPE-Q₄ \subset DSP5 and TPE-Q₄ \subset $DSP5+Fe^{3+}$ ([TPE-Q₄] = 1.00 mM, [DSP5] = 4.00 mM, [Fe^{3+}] = 20.00 mM, 300 MHz, D₂O, 298 K). Figure S12. The pictures of (A) TPE-Q₄, (B) TPE-Q₄ \subset DSP5, (C) TPE-Q₄ \subset DSP5 + Fe³⁺ and (D) supernatant upon addition of DSP5 again after removing the DSP5 + Fe^{3+} precipitate. Figure S13. The pictures of (A) TPE-Q₄, (B) TPE-Q₄ ⊂ DSP5, (C) TPE- $Q_4 \in DSP5 + Fe^{3+}$, and (D) DSP5 + Fe³⁺ in water (after centrifugation). Figure S14. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 1. Figure S15. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 2. Figure S16. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 3. Figure S17. ¹³C NMR spectrum (126 MHz, D₂O, 298 K) of DSP5. Figure S18. ¹H NMR spectrum (300 MHz, D₂O, 298 K) of DSP5. Figure S19. HRESI-MS m/z [M-Na]⁻ calcd. for C₅₁H₆₀O₁₆S₂Na⁻ 1015.3215, found 1015.3226. Figure S20. ¹H NMR spectrum (300 MHz, DMSO- d_6 , 298 K) of compound 4. Figure S21. ¹H NMR spectrum (300 MHz, D₂O, 298 K) of TPE-Q₄. Figure S22. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 5. Figure S23. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 6. Figure S24. ¹H NMR spectrum (300 MHz, CDCl₃, 298 K) of compound 7. Figure S25. ¹H NMR spectrum (300 MHz, DMSO- d_6 , 298 K) of TPE-Q. Figure S26. ¹H NMR spectrum (300 MHz, D₂O, 298 K) of DSNa. Table S1. Emission lifetimes (τ), radiative decay rate constant (k_{rad}) , and nonradiative decay rate constant (k_{nrad}) of bare TPE-Q₄ and TPE-Q₄ \subset DSP5 in water. (Supplementary Materials)

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