



Draft Genome Sequence of *Pantoea ananatis* GB1, a Plant-Growth-Promoting Hydrocarbonoclastic Root Endophyte, Isolated at a Diesel Fuel Phytoremediation Site Planted with *Populus*

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We report the 4.76-Mb draft genome of *Pantoea ananatis* GB1, a Gram-negative bacterium of the family *Enterobacteriaceae*, isolated from the roots of poplars planted for phytoremediation of a diesel-contaminated plume at the Ford Motor Company site in Genk, Belgium. Strain GB1 promotes plant growth in various hosts and metabolizes hydrocarbons.

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The genus *Pantoea* is known for its metabolic versatility (1), and *P. ananatis* has been globally implicated as a pathogen in significant agricultural crops and forest tree species (2). More recently, *P. ananatis* strains have been reported to exhibit a range of host interactions including beneficial, weakly pathogenic, commensal, and mutualistic (3, 4). We found that *P. ananatis* GB1 promotes the growth of hybrid poplar cuttings (*Populus deltoides* × [*trichocarpa* × *deltoides*] cv. Grimminge), as well as *Lupinus luteus* and *Cytisus striatus*, even though 16S rRNA gene sequence data along with phenotypic profiling indicate that GB1 is closely related to the rice pathogen *Pantoea ananatis* PA13 (5).

An Ion Torrent PGM (Life Technologies, Inc., Carlsbad, CA) was used for sequencing after extracting DNA from stationary-phase cells of GB1 using standard techniques as described by Thijs et al. (6). In total, 1.52 million reads (mean length 265 bases) generated 403 Mb of data (>362 M Q20 bases) in Torrent Suite 4.2.1. These were assembled using SPAdes 3.1.0 (7, 8) (uniform coverage mode; k-mers 21, 33, 55, 77, 99) into 33 contigs greater than 500 bp, giving a consensus length of 4,765,050 bp at 51.0× coverage (largest contig 780,856 bp; $N_{50} = 363,336$ bp). The Rapid Annotations using Subsystems Technology (RAST) server (9) was used to identify closely related organisms with complete genomes in NCBI RefSeq. *Pantoea ananatis* PA13 (GenBank accession no. CP003085) was used as a reference to reorder the GB1 contigs in Mauve (10).

Genome annotation was completed in the PGAP (NCBI) pipeline (11). The *P. ananatis* GB1 genome is 4.76 Mb and consists of one chromosome (55.2 % G+C) which includes 4,433 coding genes that were arranged into pathways using Pathway Tools (12, 13), 273 pseudogenes, 13 rRNAs (5S, 16S, 23S), 53 tRNAs, and 6 noncoding RNAs (ncRNAs).

Homologues of genes related to plant growth promoting traits such as inorganic phosphorus solubilization, nitrogen fixation,

and siderophore production were found, along with multiple copies of genes for alkanes utilization such as: alkane monooxygenase genes (*alkB*), medium-chain acyl-CoA dehydrogenase (*acdA*) genes, and long-chain acyl-CoA dehydrogenase (*acdB*) genes. Interestingly, GB1 carries a bacterial laccase gene, in agreement with reports of a novel bacterial laccase, Lac4, produced by *P. ananatis* Sd-1, a strain that is able to oxidize nonphenolic and phenolic compounds (14).

The use of *P. ananatis* GB1 plant-growth-promoting inoculant for improving phytoremediation of diesel-contaminated sites is being explored.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number JYGW00000000. The version described in this paper is version JYGW01000000.

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REFERENCES

- Walterson AM, Stavrinides J. 2015. *Pantoea*: insights into a highly versatile and diverse genus within the *Enterobacteriaceae*. FEMS Microbiol Rev 39:968–984. http://dx.doi.org/10.1093/femsre/fuv027.
- Coutinho TA, Venter SN. 2009. Pantoea ananatis: an unconventional plant pathogen. Mol Plant Pathol 10:325–335. http://dx.doi.org/10.1111/ j.1364-3703.2009.00542.x.
- Sheibani-Tezerji R, Naveed M, Jehl M-A, Sessitsch A, Rattei T, Mitter B. 2015. The genomes of closely related *Pantoea ananatis* maize seed endophytes having different effects on the host plant differ in secretion system genes and mobile genetic elements. Front Microbiol 6:440. http:// dx.doi.org/10.3389/fmicb.2015.00440.
- 4. De Maayer P, Chan WY Rubagotti E, Venter SN, Toth IK, Birch PRJ, Coutinho TA. 2014. Analysis of the *Pantoea ananatis* pan-genome reveals

factors underlying its ability to colonize and interact with plant, insect and vertebrate hosts. BMC Genomics 15:404. http://dx.doi.org/10.1186/1471 -2164-15-404.

- Choi O, Lim JY, Seo Y-S, Hwang I, Kim J. 2012. Complete genome sequence of the rice pathogen *Pantoea ananatis* strain PA13. J Bacteriol 194:531. http://dx.doi.org/10.1128/JB.06450-11.
- Thijs S, Van Hamme J, Gkorezis P, Rineau F, Weyens N, Vangronsveld J. 2014. Draft genome sequence of *Raoultella ornithinolytica* TNT, a trinitrotoluene-denitrating and plant growth-promoting strain isolated from explosive-contaminated soil. Genome Announc 2(3):e00431-14. http://dx.doi.org/10.1128/genomeA.00491-14.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/ cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. http://dx.doi.org/10.1093/bioinformatics/btt086.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using

Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.

- Rissman AI, Mau B, Biehl BS, Darling AE, Glasner JD, Perna NT. 2009. Reordering contigs of draft genomes using the mauve aligner. Bioinformatics 25:2071–2073. http://dx.doi.org/10.1093/bioinformatics/btp356.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta) genomic annotation. Omics J Integr Biol 12:137–141. http://dx.doi.org/10.1089/omi.2008.0017.
- 12. Caspi R, Altman T, Billington R, Dreher K, Foerster H, Fulcher CA, Holland TA, Keseler IM, Kothari A, Kubo A, Krummenacker M, Latendresse M, Mueller LA, Ong Q, Paley S, Subhraveti P, Weaver DS, Weerasinghe D, Zhang P, Karp PD. 2014. The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of pathway/ genome databases. Nucleic Acids Res 42:D459–D471. http://dx.doi.org/ 10.1093/nar/gkt1103.
- Karp PD, Paley S, Romero P. 2002. The pathway tools software. Bioinformatics 18:S225–S232. http://dx.doi.org/10.1093/bioinformatics/ 18.suppl_1.S225.
- 14. Shi X, Liu Q, Ma J, Liao H, Xiong X, Zhang K, Wang T, Liu X, Xu T, Yuan S, Zhang X, Zhu Y. 2015. An acid-stable bacterial laccase identified from the endophyte *Pantoea ananatis* Sd-1 genome exhibiting lignin degradation and dye decolorization abilities. Biotechnol Lett 37:2279–2288. http://dx.doi.org/10.1007/s10529-015-1914-1.