


Advances in Patient-Specific Induced Pluripotent Stem Cells Shed Light on Drug Discovery for Amyotrophic Lateral Sclerosis

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Abstract

Induced pluripotent stem cells (iPSCs), which are generated through reprogramming adult somatic cells by expressing specific transcription factors, can differentiate into derivatives of the three embryonic germ layers and accelerate rapid advances in stem cell research. Neurological diseases such as amyotrophic lateral sclerosis (ALS) have benefited enormously from iPSC technology. This approach can be particularly important for creating iPSCs from patients with familial or sporadic forms of ALS. Motor neurons differentiated from the ALS-patient-derived iPSC can help to determine the relationship between cellular phenotype and genotype. Patient-derived iPSCs facilitate the development of new drugs and/or drug screening for ALS treatment and allow the exploration of the possible mechanism of ALS disease. In this article, we reviewed ALS-patient-specific iPSCs with various genetic mutations, progress in drug development for ALS disease, functional assays showing the differentiation of iPSCs into mature motor neurons, and promising biomarkers in ALS patients for the evaluation of drug candidates.

Keywords

ALS, biomarker, drug screening, iPSC, motor neuron, neurodegenerative disease

Introduction

Amyotrophic lateral sclerosis (ALS), also known as Lou Gehrig's disease, is a fatal disease with a survival time of less than 5 years and prevalence of 3–4 per 100,000 persons in the United States^{1,2}. The neurodegenerative disorder was first described in the late 19th century and is characterized by the death of motor neurons in the brain, brainstem, and spinal cord, leading to paralysis^{3,4}. Although ALS has been studied for many decades, scientific and clinical results have only partially revealed that genetic mutation may play a role in the onset of disease^{5–15}. In addition to hereditary factors, an unregulated neural transmitter system and environmental factors are the major factors influencing the progression of ALS¹⁶. These broad and general causes have confounded the investigation of therapies.

Although most cases of ALS are the sporadic form (sALS) of unknown etiology, ~25% of ALS cases are the familial form (fALS), which is clinically and pathologically indistinguishable from sALS¹⁷. More detailed studies have been conducted on fALS than on sALS. Dominantly inherited gene mutations associated with fALS such as cytosolic copper-zinc superoxide dismutase 1 (*SOD1*), hexanucleotide

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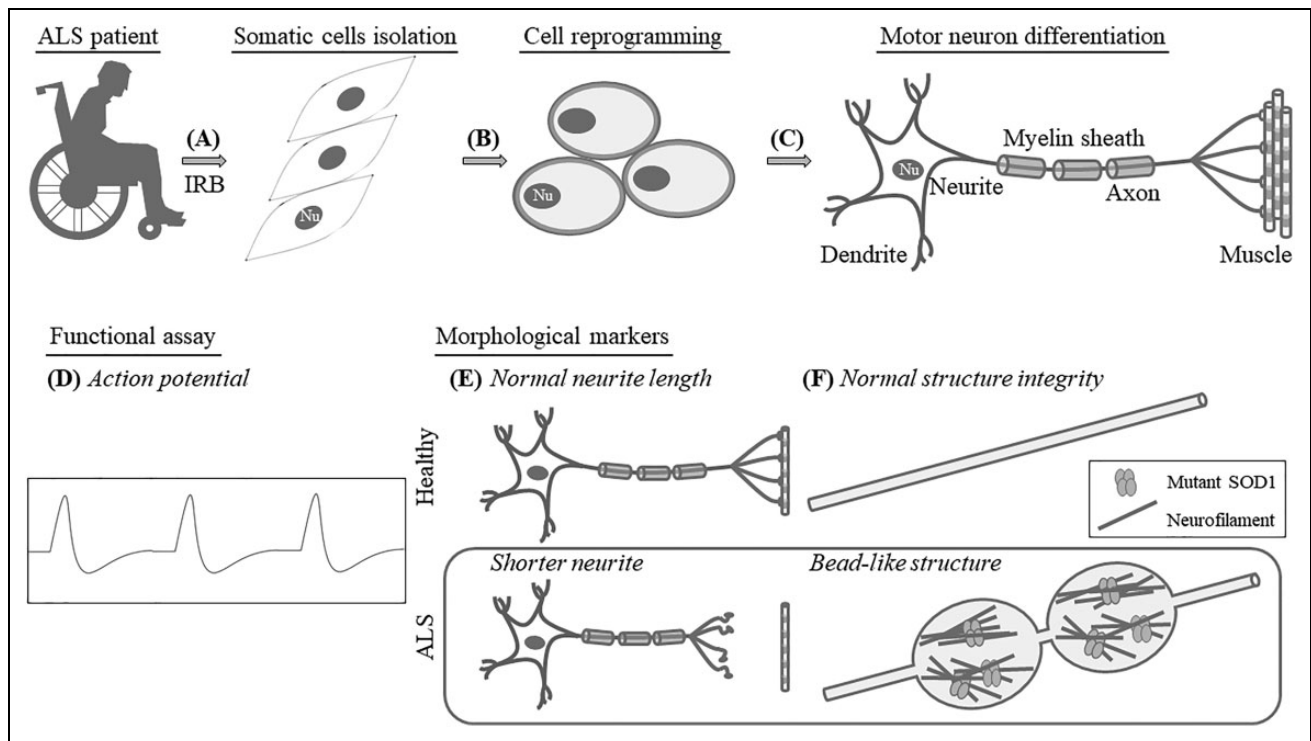


Fig. 1. Schematic illustration showing that iPSCs generated from ALS patients differentiate into motor neurons for functional and morphological analysis. (A) ALS patients' somatic cells are collected with donors' informed consent under institutional review board monitoring (A) and the cells can be reprogrammed to the induced pluripotent state (B). After motor neuron differentiation of ALS patient's iPSCs (C), further studies of the disease can be performed to assess physiological properties (D), providing a link to the level of maturation. The examination of morphological changes such as neurite length (E) subcellular aggregate (F), and patient-specific samples can be used to characterize the pathogenesis of ALS in patient-derived iPSCs. Nu: nucleus.

repeat (chromosome 9 open reading frame 72, *C9ORF72*), tar DNA-binding protein 43 (*TDP-43*), fused in sarcoma (*FUS*), and other less frequent genes contribute to considerable genetic heterogeneity^{6,17-23}. Among the ALS-related gene mutations, SOD1 mutants are responsible for 20% of fALS²⁴ and the synergistic effect of the mutant Alsln (also known as *ALS2*) and SOD1 are linked to the early onset of ALS^{7,25}. Genetically engineered cell lines or animal models for these genetic mutations are used to investigate the development of new drugs and to verify drug efficacy.

ALS modeling rodents or embryonic stem cells (ESCs) expressing ALS-causing genes have been generated to represent a specific subset of the syndrome in an effort to understand the onset and progression of ALS disease^{22,26-32}. However, the transgene method alone cannot be used for mimicking the real physiology of motor neurons in sALS patients because it cannot completely correlate the genotype with the neuronal phenotype^{26,33}. Because animal and stem cell models for elucidating ALS pathogenesis have demonstrated shortcomings in recapitulating sALS patients, their capacity and application for the evaluation of therapeutic efficacy remain unmet needs. As an alternative, researchers have obtained induced pluripotent stem cells (iPSCs) from patients with neurodegenerative diseases, which have allowed the study of these cells with specific cell differentiation^{2,34-36}.

Human iPSCs can be produced by reprogramming the somatic cells for a state of embryonic-like stem cell via delivering the required transcription factors (Fig. 1A and B)³⁷⁻³⁹. Such iPSCs can be directed to generate different kinds of neural cell types. iPSCs obtained from patients with ALS symptoms can be investigated at the molecular level to probe the changes in degeneration/death of motor neurons that trigger destruction of the neuromuscular junction⁴⁰⁻⁴². Therefore, differentiated cells from ALS patients, such as those of the motor neuron type, can be used to identify drug candidates for ALS treatment. The present review focuses on existing medication and medical needs of ALS patients, advantages of using patients' iPSCs, scope for improvement in the approaches of motor neuron differentiation and functional assays, and strategies for the evaluation of drug efficacy in ALS disease.

Currently Available Treatments and Potential Small Molecules

Drugs Approved by the U.S. Food and Drug Administration for ALS Treatment

Riluzole is the first available compound known to attenuate the progression and partly relieve the symptoms of ALS via depressing N-methyl-D-aspartate (NMDA) receptor- and

α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptor-mediated signaling^{43,44}. Riluzole has several side effects and extends survival only by an average of 2–3 months, so more effective ALS therapy is urgently needed^{45,46}. The recently approved molecule edaravone is a small-radical scavenger that can inhibit oxidative stress. Edaravone was initially used to treat acute ischemic stroke⁴⁷. It quenches the hydroxyl radical functional group and down-regulates both radical-dependent and radical-independent lipid peroxidation⁴⁸. Previous results showed that edaravone could delay the degeneration of motor neurons in a murine model with mutated *SOD1* expression⁴⁹ and data from a phase 2 trial in ALS patients consistently reported the effect of the drug in reducing oxidative stress⁵⁰. Although evidence supported these positive outcomes in a phase 3 trial of edaravone, the inconvenience of intravenous administration and the extremely restricted population of ALS patients caused physicians not to prescribe the drug⁵¹. In addition to the inconvenience and high treatment cost, the long-term efficacy and toxicity profile of edaravone have not yet been established. Therefore, the clinical needs and challenges in clinical trials for ALS patients still exist and much more efficient protocols for drug screening are required.

Small-Molecule Development for Clinical Application Targeting ALS

Because of extensive studies investigating small molecules, exploring mechanisms, and synthesizing new compounds, the scope of drug discovery is changing and scientists can combine the power of synthetic chemistry with advanced screening technologies for small-molecule development. These small molecules can be covalent bonding inhibitors, protein–protein interaction modulators, protein conformational allosteric regulators, or epigenetic controllers and they sometimes have effects on multiple targets at one time^{52–54}. For example, olaparib, an ADP-ribose polymerase inhibitor, and imatinib, a tyrosine kinase inhibitor, can significantly reduce ovarian cancer and chronic myeloid leukemia cells, respectively^{55,56}. Furthermore, small molecules can potentially penetrate the blood–brain barrier into the central nervous system to treat patients with brain tumors or neurological diseases⁵⁷. Creating small molecules that target mutant proteins associated with ALS may be promising candidates for the treatment of ALS.

Drug development based on neuroprotective properties such as reducing oxidative stress and excitotoxicity and eliminating toxic aggregates/fragments may benefit ALS patients. Recently, several groups have shown that autophagy plays an important role in ALS pathogenesis^{58–60}. The autophagy–lysosome machinery is the major system for removing abnormal aggregates through cytoplasmic protein degradation⁶¹. One of these molecules that has been preferentially examined using *in vitro* and *in vivo* ALS models and shows promise is a purified compound from a plant used in traditional Chinese medicine (*Angelica sinensis*),

n-butylidenephthalide (*n*-BP). Evidence has suggested that accumulated mutant SOD1 can initiate autophagy in an animal model⁵⁹ and *n*-BP can extend the lifespans of the *SOD1* murine model through modulating cellular autophagy and apoptosis in the spinal cord⁶⁰. Furthermore, findings also indicated that *n*-BP could reduce the death of motor neurons, relieve muscular atrophy, suppress neuroinflammation, and recover gastrocnemius muscle⁵⁹. Although the functions of *n*-BP in anti-apoptosis, anti-inflammation, anti-oxidation, and autophagy modulation have been revealed, the data were collected from mutated *SOD1* in *in vitro* and *in vivo* models. Investigations based on ALS patients' iPSCs may provide further information for the development of this drug candidate.

iPSCs Differentiate into Functional Motor Neuron Units

Differentiation into Motor Neurons

iPSCs can generate different subtypes of neural cells, such as neurons, motor neurons, astrocytes, and other cell types^{35,62–64}; therefore, the characterization of these *in vitro* cells recapitulating *in vivo* ALS progression can be achieved. Previous studies reported that human ESCs have been differentiated successfully into motor neurons by exogenously expressing the mutated genes of ALS. The approaches of motor neuron differentiation have also been applied to iPSCs. fALS/sALS-derived motor neurons can be harvested and investigated for the development of new drugs (Fig. 1C). Crucial secreted signaling factors, such as bone morphogenetic protein (BMP), fibroblast growth factor (FGF), retinoic acid (RA), Sonic hedgehog (Shh), and Wnts and transforming growth factor-beta (TGF β) signaling, which are involved in the patterning neural axis during brain development, have been used to direct iPSCs toward motor neurons^{65–68}.

Motor neurons are classified as corticospinal upper motor neurons and spinal cord lower motor neurons (LMNs). The degeneration and death of motor neurons lead to the onset of neurodegenerative diseases such as ALS^{69–71}. According to results obtained in different species^{67,72–75}, signaling in the development of neuroectoderm has been shown to specify LMN fate^{66,68}. The process of motor neuron differentiation is time consuming, expensive, and inefficient in the acquisition of cells. After making efforts to fine-tune the protocols, two small molecules, LDN193189 and SB431542, were found to accelerate the differentiation of motor neurons through the inhibition of both BMP and TGF β signaling and blocking mesoderm and endoderm fates⁷⁶. Therefore, more abundant motor neurons can be obtained from iPSCs in a shorter time period.

Functional Assay and Morphological Marker for Motor Neurons

The most extensive functional examination of differentiated motor neurons is detection of the electrophysiological action

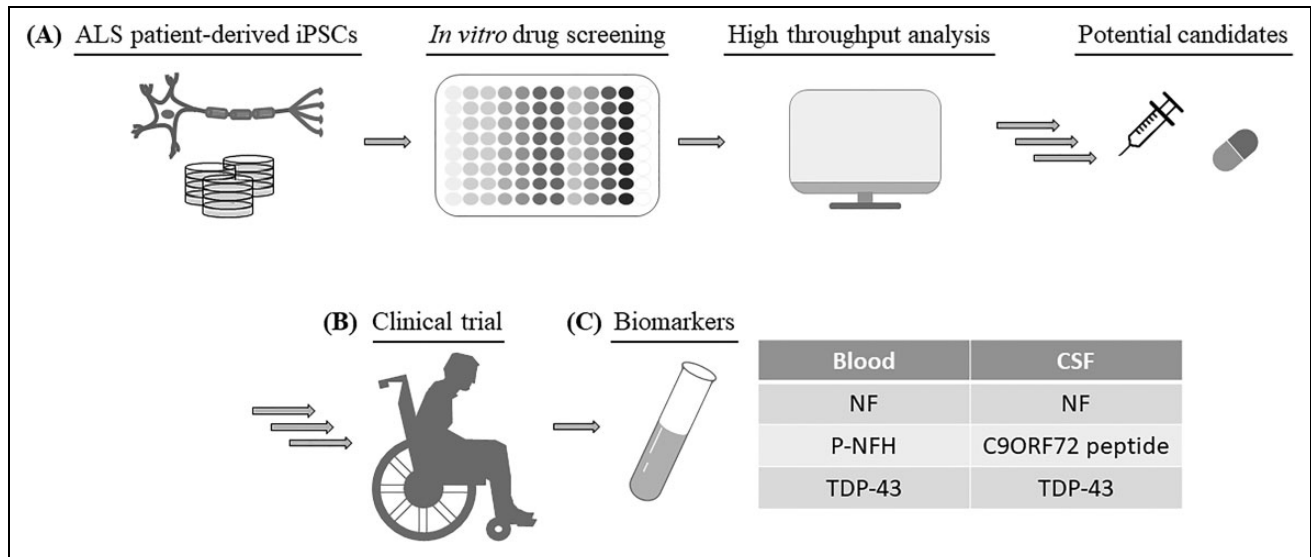


Fig. 2. Combination of human iPSC approach, high-throughput drug screening, and biomarker application can potentiate clinical candidate selection. (A) iPSCs derived from patients with ALS can be used for high-content drug screening to identify pathways and targets of small molecules with therapeutic potential. After preclinical studies, these small molecules may enter clinical trials with suitable administration routes (B) and the use of biomarkers can be used to calculate and determine drug efficacy (C). P-NFH: phosphorylated heavy chain of NF.

potential for evaluating the maturation of motor neurons *in vitro*⁷⁶ (Fig. 1D). During the early stages of neural differentiation, voltage-gated ion channels need to be induced to generate an action potential. In addition, spontaneous action potentials require time to develop after the maturation of motor neurons⁷⁷. To achieve a high level of mature motor neurons, the transition between induced and spontaneously occurred action potentials requires the involvement of various factors such as synapse formation^{78,79}. To determine the propagation of electrical signals, protocols to direct the differentiation of motor neurons from iPSCs with the capability of synapse formation is critical.

Because ALS pathogenesis presents heterogeneous causes accompanied by distinct mechanisms linked to the onset and progression of ALS, imaging-based approaches are required to determine cellular/molecular changes and drug effects on ALS disease⁸⁰. For evaluation, information on cellular morphology can be obtained by automated data acquisition, thereby lowering the cost of drug screening in terms of labor. Both the fragmentation of neurites with reduced length and morphological swelling, the so-called bead-like structure resulting from abnormal protein aggregation (Fig. 1E and F)^{22,81–83}, can be analyzed under the microscope. Such findings suggest that specific antibodies can be also used to label mutants for understanding the disease progression of ALS⁸⁴. These protein aggregates are expressed *in vivo* in native form, so the antibodies may not be available to recognize targets correctly and represent the real distribution or localization.

Use of ALS Patient-Derived iPSCs for Drug Development

Drug-Screening Platform via iPSCs from ALS Patients

During the process of novel drug development, safety and efficacy testing are costly and time consuming. Many clinical trials of ALS have been withdrawn or failed due to unexpected toxicity or lack of efficacy⁸⁵. The potential toxicities challenging clinical translation of drug candidates suggest that *in vitro* and *in vivo* nonhuman models might not be sufficient. To select effective therapies for ALS patients and to improve the success rate of drug selection, iPSCs have been used as models in many outstanding studies (Fig. 2A and Table 1)^{2,23,34,35,52,58,83,86–90}. Egawa et al. recently reported an iPSC drug-screening assay in which they used several compounds targeting the epigenetic/transcriptional level of iPSC-derived motor neurons from ALS patients with the *TDP-43* mutation². One of the compounds, anacardic acid, was shown to reverse part of the ALS phenotype found in motor neurons. This work can be a stepping stone to toward using ALS-specific motor neurons to verify candidate drugs.

Motor neurons derived from patients with fALS or sALS were used to screen various kinase inhibitors, epigenetic modulators, anti-sense oligo-nucleotides (ASOs), or approved molecules for any other indication (Table 1). Furthermore, Wainger et al. used a high-throughput electrode array to detect the action potentials of variant ALS patients' motor neurons (*SOD1*, *C9ORF72*, and *FUS* mutation) and revealed that retigabine could reduce the death of motor neurons by blocking hyperexcitability⁸⁹. The drug

Table 1. ALS Cell Models from Disease-Derived iPSCs.

ALS patient (responsible gene)	Neural cell type (differentiation marker)	Functional assay	Potential candidate	Author and year [reference number]
<ul style="list-style-type: none"> TDP-43 (Q343R, M337V, G298S) 	<ul style="list-style-type: none"> Motor neuron (Isl-1, HB9, SMI-32, CHAT) 	<ul style="list-style-type: none"> Motor neuron markers similar to healthy Shorter neurite TDP-43 aggregation (WB and IF) 	<ul style="list-style-type: none"> Trichostatin A Spliceostatin A Anacardic acid Garcinol 	<ul style="list-style-type: none"> Egawa et al., 2012²
<ul style="list-style-type: none"> SOD1 (L144F) TDP-43 (M337V) 	<ul style="list-style-type: none"> Motor neuron (Isl-1, Tuj-1) 	<ul style="list-style-type: none"> Action potential SOD1 aggregation (WB and IF) Synapse and motor neuron number 	<ul style="list-style-type: none"> Kenpaullone 	<ul style="list-style-type: none"> Yang et al., 2013⁵²
<ul style="list-style-type: none"> C9ORF72 	<ul style="list-style-type: none"> Motor neuron (Tuj-1, SMI-32, CHAT) 	<ul style="list-style-type: none"> Action potential C9ORF72 aggregation (WB) C9ORF72 RNA foci (FISH) 	<ul style="list-style-type: none"> Small RNA molecule: C9ORF72 ASO 	<ul style="list-style-type: none"> Sareen et al., 2013³⁵
<ul style="list-style-type: none"> TDP-43 	<ul style="list-style-type: none"> Motor neuron (Isl-1, HB9, SMI-31) 	<ul style="list-style-type: none"> Action potential TDP-43 aggregation (IF) 	<ul style="list-style-type: none"> CDK inhibitor JNK inhibitor Triptolide FDA-approved cardiac glycosides Digoxin Lanatoside C Proscillaridin A 	<ul style="list-style-type: none"> Burkhardt et al., 2013³⁴
<ul style="list-style-type: none"> C9ORF72 	<ul style="list-style-type: none"> Motor neuron (Tuj-1) 	<ul style="list-style-type: none"> Glutamate insult C9ORF72 aggregation (WB and Q-PCR) C9ORF72 RNA foci (FISH) 	<ul style="list-style-type: none"> Small RNA molecules: ADARB2 siRNA Small RNA molecules: C9ORF72 ASO 	<ul style="list-style-type: none"> Donnelly et al., 2013⁸⁸
<ul style="list-style-type: none"> SOD1 (A4V, D90A, G85S) C9ORF72 FUS (M511FS, H517Q) C9ORF72 	<ul style="list-style-type: none"> Motor neuron (Isl-1, HB9) 	<ul style="list-style-type: none"> Action potential Hyper-excitability Motor neuron number 	<ul style="list-style-type: none"> Retigabine 	<ul style="list-style-type: none"> Wainger et al., 2014⁸⁹
<ul style="list-style-type: none"> SOD1 (A4V, D90A, G85S) C9ORF72 	<ul style="list-style-type: none"> Motor neuron (Isl-1, HB9, Tuj1, SMI32, CHAT, VGLUT1, NMDAR1, SYTI, SYP) Astrocyte (ALDH1, GFAP) 	<ul style="list-style-type: none"> Action potential C9ORF72 RNA foci (FISH) Intracellular protein transport (FRAP) RanGAP1 aggregation (IHC) 	<ul style="list-style-type: none"> Small RNA molecule: C9ORF72 ASO Porphyrin (TMPyP4 and KPT-276) 	<ul style="list-style-type: none"> Zhang et al., 2015⁹⁰
<ul style="list-style-type: none"> SOD1 (D90A, A4V) 	<ul style="list-style-type: none"> Motor neuron (Isl-1, Tuj1, CHAT) Astrocyte 	<ul style="list-style-type: none"> Action potential NF aggregation (Q-PCR) Shorter neurite Luciferase activity 	<ul style="list-style-type: none"> Riluzole Kenpaullone EphA inhibitor 	<ul style="list-style-type: none"> Du et al., 2015⁸³
<ul style="list-style-type: none"> SOD1 (L144FVX, G93S) TDP-43 (M337V, Q343R, G298S) C9ORF72 sALS sALS 	<ul style="list-style-type: none"> Motor neuron (HB9, SMI-32, CHAT) 	<ul style="list-style-type: none"> Action potential SOD1 aggregation (WB) Autophagy Motor neuron number 	<ul style="list-style-type: none"> Bosutinib Desatinib 	<ul style="list-style-type: none"> Imamura et al., 2017⁵⁸

(continued)

Table 1. (continued)

ALS patient (responsible gene)	Neural cell type (differentiation marker)	Functional assay	Potential candidate	Author and year [reference number]
<ul style="list-style-type: none"> FUS (R521H, P525L) 	<ul style="list-style-type: none"> Motor neuron (HB9, Isl-1, CHAT, SMI-32, Synapsin1, Tuj1) 	<ul style="list-style-type: none"> Action potential Hypo-excitability FUS localization (IF) Axonal transport 	<ul style="list-style-type: none"> Small RNA molecule: HDAC6 ASO ACY-738 and Tubastatin A 	<ul style="list-style-type: none"> Guo et al., 2017²³

WB: Western blotting analysis; IF: immunofluorescence assay; FISH: fluorescence *in situ* hybridization; IHC: immunohistochemistry; FRAP: fluorescence recovery after photobleaching; Isl-1: Islet-1; HB9: Homeobox 9; Tuj1: beta III tubulin; SMI32: neurofilament H non-phosphorylated; CHAT: choline acetyltransferase; VGLUT1: vesicular glutamate transporter 1; NMDAR1: NMDA receptor 1; SYTI: synaptotagmin I, SYP: synaptophysin; ALDH1: aldehyde dehydrogenase 1; GFAP: glial fibrillary acidic protein; ASO: anti-sense oligo-nucleotide; CDK: cyclin-dependent kinase; JNK: c-Jun N-terminal kinase

candidates identified by such methods can confirm the toxicity and efficacy of drugs in humans and retigabine has entered a phase 2 trial for efficacy evaluation (NCT02450552; estimated completion date: February 2018). Importantly, ALS-patient-specific iPSCs present many benefits, including the correct genetic background and pluripotency. The results from the above-mentioned drug-screening studies have shown that high-throughput approaches can produce abundant and useful information that can expedite and increase the probability of success for drug candidates entering clinical trials⁹¹ (Fig. 2B).

Although iPSCs are mainly generated from skin fibroblasts or peripheral blood from ALS patients, the pathogenic phenotypes may disappear. These iPSCs may be reverted to the early stage pluripotent cells. Thus, the recapitulation of a late onset of neurodegenerative disease cannot be fully achieved⁹². To generate iPSCs associated with the characterization of aging, oxidative stress, DNA-damaging and proteasome modulator agents are being tested to produce high-level maturation of patient-specific iPSC derivatives^{93–95}. Previous reports indicated that AMPA receptor, a glutamate receptor, could mediate excitatory transmission in the brain; therefore, glutamate insult accelerates the aging process by allowing high level of calcium ions to enter the neurons^{15,96}. Such induction can be used to change the phenotypes of patient-derived iPSCs to mimic neurodegeneration. In addition, downregulation of phosphatase and tensin homolog deleted on chromosome 10 (PTEN) can attenuate neuron death by manipulating the trafficking of the GluR2 subunit of the AMPA receptor within the cell membrane⁹⁷. Many small molecules modulating PTEN have been synthesized, so it may be a potential therapeutic strategy for medical intervention in ALS patients.

To verify the use of screening approaches and to develop reliable platforms for exploring the molecular underpinnings of ALS, multiple iPSC clones from different ALS patients and several markers are urgently required. Using at least two iPSC lines from patients with ALS in combination with automated and imaging-based techniques would acquire considerable information on cell morphology, formation of protein aggregates, and distribution/patterns of specific

molecules by immunocytochemistry/immunocytochemistry or immunofluorescence staining. Moreover, high-throughput equipment has been used for examining small-molecule compounds in several studies. Nevertheless, there is no clear mechanism of neuron death that can clearly distinguish sALS from fALS. Large sample sizes are necessary to reveal the detailed and unique ALS pathogenesis, but the cost of reprogramming ALS patients' fibroblasts for iPSCs remains high.

Biomarkers of Motor Neurons

Throughout the drug development process, biologically validated biomarkers are used to assess the subject's response to a specific treatment. Biomarker analysis demonstrating the effect of the drug candidates on ALS targets is also important (Fig. 2C). Previous studies showed that CD40L, which is expressed by activated T cells and required for induced immune response, could be a treatment target for ALS patients. Moreover, inhibiting CD40L-mediated inflammatory responses can improve survival in animal studies^{98,99}. In addition to altered inflammatory gene expression, monocyte-expressed, ALS-specific, inflammation-related microRNA or the unregulated complement system can be treated with specific antibodies or inhibitors to delay ALS progression, extend survival, and improve motor function in animal models^{100,101}.

Abundant neurofilament (NF) proteins have been detected in the cerebral spinal fluid (CSF) or blood from *SOD1*^{G93A} mice^{102–104} (Fig. 2C). Among them, plasma containing the phosphorylated heavy chain of NF was thought to be associated with motor neuron death^{105,106} (Fig. 2C). Moreover, NF aggregates (bead-like structure) can also be observed in ALS-patient-specific iPSCs with mutation of *SOD1*^{A4V} or *SOD1*^{D90A}⁸¹ (Fig. 1F).

Several studies have revealed that translational products of *C9ORF72* repeats resulted in aggregation and were localized in the neurons of the central nervous system. This phenotype was seen in iPSC models^{107,108}. When small molecules bind to the GGGGCC repeat sequence, they can inhibit the formation of translational aggregates in the

differentiated motor neurons derived from iPSCs (Table 1). In addition, peptide aggregation has been found in CSF from ALS patients with the *C9ORF72* mutation¹⁰⁸ (Fig. 2C); therefore, this measurement is available for evaluating therapeutic efficacy.

Other reports about genetic mutations also found the formation of TDP-43 inclusion bodies, which was used as one of the biomarkers^{2,34,109}. Using enzyme-linked immunosorbent assay-based approaches, the TDP-43 level was found to be increased significantly in plasma and CSF from patients with ALS compared with healthy individuals. Although the discovery of biomarkers to distinguish ALS patients and healthy people are highly challenging, much more effective therapies may be investigated and developed with more biomarkers

Conclusion

Neurodegenerative diseases have a potentially disastrous impact on quality of life, so ongoing efforts to develop novel therapeutic drug candidates are of utmost importance^{99,110–113}. The use of genetic manipulation in cell systems and nonhuman animal models may not predict the potential toxicity and lack of efficacy accurately in humans⁸⁵. Recent achievements in differentiation of patient-specific iPSCs into motor neurons have provided the basis to establish a platform to screen drug candidates, test toxicity, and confirm the efficacy of drugs in complicated fALS/sALS populations (Table 1). Although some constraints, such as the labor-intensive and high-cost procedure, protocols for generating fully mature motor neurons, detection methods of protein aggregates, applicable biomarkers, and basic mechanism of motor neuron death in sALS, need to be improved, the use of iPSC-dependent screening may accelerate the process of drug development. Following a systemic assessment (Fig. 2), studies of ALS may shed light on causes of disease and help to improve drug development in the future.

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