



Antibody Response and Safety After mRNA-1273 SARS-CoV-2 Vaccination in Peritoneal Dialysis Patients – the Vienna Cohort

Georg Beilhack¹, Rossella Monteforte¹, Florian Frommlet², Martina Gaggl¹, Robert Strassl³ and Andreas Vychytil^{1*}

¹ Division of Nephrology and Dialysis, Department of Medicine III, Medical University of Vienna, Vienna, Austria, ² Center for Medical Statistics, Informatics and Intelligent Systems, Medical University of Vienna, Vienna, Austria, ³ Division of Clinical Virology, Medical University of Vienna, Vienna, Austria

Background: Dialysis patients are at high risk for a severe clinical course after infection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Safety and early immune responses after mRNA-based vaccination have been reported mostly in patients on hemodialysis (HD), whereas reports of peritoneal dialysis (PD) patients remain rare.

OPEN ACCESS

Edited by:

Susu M. Zughaier, Qatar University, Qatar

Reviewed by:

Mohamed Emara, Qatar University, Qatar Luuk Hilbrands, Radboud University Nijmegen, Netherlands

*Correspondence: Andreas Vychytil andreas.vychytil@meduniwien.ac.at

Specialty section:

This article was submitted to Vaccines and Molecular Therapeutics, a section of the journal Frontiers in Immunology

> Received: 21 September 2021 Accepted: 11 November 2021 Published: 02 December 2021

Citation:

Beilhack G, Monteforte R, Frommlet F, Gaggl M, Strassl R and Vychytil A (2021) Antibody Response and Safety After mRNA-1273 SARS-CoV-2 Vaccination in Peritoneal Dialysis Patients – the Vienna Cohort. Front. Immunol. 12:780594. doi: 10.3389/fimmu.2021.780594 **Methods:** In this retrospective observational study, 39 PD patients had received two doses of the mRNA-1273 Moderna[®] vaccine. We analyzed SARS-CoV-2 Spike (S) antibody titers 4 weeks after each dose of mRNA-1273 and report local and systemic side effects in PD patients that occurred within one week after each mRNA-1273 dose. Using a quantile regression model we examined factors that might influence SARS-CoV-2 S antibody levels in PD patients.

Results: Four weeks after the first dose of mRNA-1273 vaccine 33 of 39 (84.6%) PD patients seroconverted and presented with 6.62 U/mL (median; IQR 1.57-22.5) anti-SARS-CoV-2 S antibody titers. After the second dose, 38 of 39 (97.4%) PD patients developed anti-SARS-CoV-2 S antibodies and titers increased significantly (median 968 U/mL; IQR 422.5-2500). Pain at the injection site was the most common local adverse event (AE) (71%). Systemic AEs occurring after the first dose were mostly fatigue (33%) and headache (20%). No severe systemic AEs were reported after the first injection. After the second dose the incidence and the severity of the systemic AEs increased. The most common systemic AEs were: fatigue (40.5%), headache (22.5%), joint pain (20%), myalgia (17.5%) and fever (13%). Lower Davies Comorbidity Score (p=0.04) and shorter dialysis vintage (p=0.017) were associated with higher antibody titers after the first dose. Patients with higher antibody titers after the first dose. Patients with higher antibody titers after the first dose. Patients with higher antibody titers after the first dose increased to have higher antibody titers after the second dose (p=1.53x10⁻⁰⁵).

Conclusions: Peritoneal dialysis patients in this cohort had a high seroconversion rate of 97.4%, showed high antibody titers after full vaccination and tolerated the anti-SARS-CoV-2 mRNA-1273 vaccine well without serious adverse events.

Keywords: COVID-19, dialysis, antibody response, safety, mRNA-1273

INTRODUCTION

In Austria, the first cases of the current pandemic of coronavirus disease (COVID-19) appeared on February 25th, 2020 when two individuals tested positive for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (1). As of November 7th 2021, 883887 SARS-CoV-2 cases and 11502 deaths due to COVID-19 were documented in Austria (population size 8.9 million) (2). Among the first vaccines available against SARS-CoV-2, the lipid-nanoparticleencapsulated mRNA-based vaccines mRNA-1273 and BNT162b2 proved to be safe and induce high levels of protection against COVID-19 in the general population (3, 4). Furthermore, it was shown that mRNA vaccines elicit serum neutralizing activity against wild-type SARS-CoV-2 and various emerging mutants (5). Dialysis patients are among the most vulnerable individuals susceptible to COVID-19, having a reported 28-day case-fatality rate up to 25% after infection with SARS-CoV-2 due to age and frailty (6). Furthermore, it is known that dialysis patients in general are immunocompromised and respond less effectively to many vaccines (7-10), although exceptions exist (11). In an effort to protect this vulnerable group, dialysis patients were recommended to be prioritized for vaccination against SARS-CoV-2 (12, 13).

According to the Austrian Dialysis and Transplantation Registry (ÖDTR) the mRNA-1273 vaccine was used in 52.5% of hemodialysis (HD) patients and 75.6% of peritoneal dialysis (PD) patients. The BNT162b2 vaccine was used in 46.6% of HD patients and 23.6% of PD patients, respectively (14).

Since the beginning of 2021, several studies in hemodialysis patients have reported varying seroconversion rates in mRNA vaccinated patients that range from 18% to 53% after one dose and from 70% to 96% after two doses, respectively (15). The humoral response to SARS-CoV-2 vaccination may differ between HD and PD patients (16). However, the majority of trials investigated these early antibody responses to anti-SARS-CoV-2 vaccines in patients on hemodialysis and only a few focused on peritoneal dialysis (17).

In this retrospective observational study, we report on 39 PD patients at our institution who received two doses of the mRNA-1273 anti-SARS-CoV-2 vaccine. We analyzed how many percent of these patients seroconverted after one or two vaccine doses and looked at tolerability of the vaccine in this patient population. Furthermore, we looked at antibody response after the first and second dose and analyzed factors which might be associated with anti-SARS-CoV-2 S antibody titers.

PATIENTS AND METHODS

Study Population

This retrospective, observational single-center study was performed at the Division of Nephrology and Dialysis, Department of Medicine III, Medical University of Vienna, Austria. Patients treated with regular peritoneal dialysis who were vaccinated against SARS-CoV-2 with Moderna®mRNA-1273 and received the full two-dose scheme were included. Patients with previous or active COVID-19 infection as well as those who received other vaccines against SARS-CoV-2 were excluded from the analysis. All patients attended our outpatient clinics on a regular basis, where they were tested for COVID-19 infection using the Abbott PanbioTM COVID-19 Rapid Antigen test (according to our hospital regulations) and were checked for COVID-19 specific symptoms. The study population was immunized with two doses of Moderna® mRNA-1273 28 days apart, as recommended by the manufacturer, on March 11th, 2021 (dose 1, 100µg mRNA-1273) and April 8th, 2021 (dose 2, 100µg mRNA-1273). Antibody levels were measured 4 weeks after each dose. Comorbidities of our PD patients were evaluated using the co-morbidity score published by Davies et al (18). Clinical records were reviewed in order to assess demographic data and laboratory values.

Serological Assessment

Blood was drawn from patients as part of their routine visits 4 weeks after the first and second vaccination, respectively. The antibody response against SARS-CoV-2 S was measured using the Roche Elecsys anti-SARS-CoV-2 S[®] assay on a Roche Cobas e801 platform according to manufacturer instructions (ROCHE[®] Diagnostics International Ltd.) (19). This assay detects antibodies against the receptor binding domain (RBD) of the SARS-Cov-2 spike (S) protein. RBD-targeting antibodies have been described to have a high neutralization activity against SARS-CoV-2 (20). The lower limit of detection was 0.4 U/mL, the upper detection limit was 2500 U/mL. The assay was calibrated to the current WHO International Standard for antibody detection against SARS-CoV-2 S. To detect whether asymptomatic patients had a previous infection with the SARS-CoV-2 virus, a serological assay (ROCHE[®] Elecsys anti-SARS-CoV-2 assay) was used to measure antibodies targeting the nucleocapsid (N) antigen.

Reporting of Adverse Events

Adverse events (AEs) occurring within 7 days from each vaccination were recorded using a standardized survey. AEs were categorized in local AEs (pain, swelling, bruising) and systemic AEs (fever, headache, fatigue, myalgia, joint pain, dizziness, vomiting). The patients were asked to grade their AEs using a scale from 0 to 4 (grade = 0: no event; grade = 1: mild, does not affect daily activities; grade = 2: moderate, interferes with activities of daily living; grade = 3: severe, interrupts usual activities of daily living; grade = 4: hospitalization=serious AE).

Statistical Analysis

Descriptive statistics are given as mean and standard deviation or median and interquartile range, as appropriate. Because of the

Abbreviations: AE, Adverse Event; BMI, Body Mass Index; CNI, Calcineurin Inhibitor; COVID-19, Corona Virus Disease 2019; FSGS, Focal Segmental Glomerulosclerosis; GFR, Glomerular Filtration Rate; GVHD, Graft *versus* Host Disease; HD, Hemodialysis; IQR, Interquartile Range; IS, Immunosuppression; Kt/V, Clearance of urea x time/volume; MMF, Mycophenolat-Mofetil; RAAS, Renin-Angiotensin-Aldosteron System; SARS-CoV-2, Severe Acute Respiratory Syndrome CoronaVirus 2; TMA, Thrombotic Microangiopathy; PD, Peritoneal dialysis.

skewed distribution of antibody titers and censoring of the titer levels at 2500 U/mL (technical maximum of titer level), quantile regression was used rather than least squares regression to analyze factors that influenced anti-SARS-CoV-2 S antibody levels using the R package quantreg (21). Titer levels were logtransformed both as predictor variable (after the first dose) and as outcome variable (after the second dose). The quantile regression analysis was performed at the log scale as it gave a much better fit to the data compared with regression models on the original data. Before- and after- second dose vaccination measurements of anti-SARS-CoV-2 S antibody levels were compared by Wilcoxon matched pairs signed rank test. Statistical analysis was performed with R version 4.1.1 (22). Since this is a single center study, the sample size (n=39) was limited by the number of patients treated in the PD unit of the Medical University of Vienna.

Ethical Considerations

Our study was approved by the ethics committee of the Medical University of Vienna (EK 1418/2021). All patients were informed about their antibody levels. Furthermore, we explained that the exact value of antibody titers which protects from COVID-19 infection is currently unknown. Patients were advised to continue keeping safety measures such as social distancing, hygiene standards and face masks after vaccination.

RESULTS

In total 40 PD patients received two doses of the Moderna[®]mRNA-1273 vaccine. One of these patients was excluded since he was retrospectively tested positive for antibodies targeting the nucleocapsid (N) antigen, confirming previous (asymptomatic) COVID-19 infection. The study cohort consisted of 26 men and 13 women. The mean age was 55.2 years (range 29-80 years) with a median dialysis vintage of 16.1 months (IQR 6-31.3 months). Seven of 39 patients received immunosuppressive therapy. Detailed characteristics of the study cohort are shown in **Table 1**.

Anti-SARS-CoV-2 S Antibody Responses to mRNA-1273 Vaccination

Our study cohort received two doses of mRNA-1273 vaccine 28 days apart, as recommended by the manufacturer. After the first dose 33 of 39 patients (84.6%) had detectable anti-SARS-CoV-2 S antibodies, but titers were rather low (median, 6.62 U/mL, IQR 1.57-22.5, lower detection limit 0.4 U/mL). Of the remaining six patients, five responded after the second dose. After two doses a total of 38 of 39 patients (97.4%) had seroconverted and titers increased significantly (median 968 U/mL, IQR 422.5-2500 U/mL, p = 8.057×10^{-08} , Wilcoxon matched pairs signed rank test) (**Figure 1**). Eleven of 39 patients (28.2%) reached an antibody titer of 2500 U/mL (upper detection limit of the assay).

The only patient who did not develop antibody titers within 4 weeks from the second dose was lung transplanted for cystic fibrosis and received immunosuppressive treatment (Figure 1 and Table 2). In our cohort six additional patients received immunosuppressive therapy. Five of these patients experienced

TABLE 1 | Characteristics of peritoneal dialysis patients vaccinated with Moderna $^{\textcircled{m}}$ mRNA-1273 vaccine.

Patient demographics 39 Total number of patients 39 Age (years, mean, range) 55.2 (29-80) Men 26 (66.7) Primary kidney disease 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 19 (48.7) A 2 (5.1) Laboratory (mean ± SD) 12 (30.8) Hemoglobin (g/dL) 26 ± 7.534 Alburnin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.5 ± 7.534 Alburnin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.5 ± 0.18 Phosphate (rmol/L) 2.7 ± 5.37 <		
Total number of patients 39 Age (years, mean, range) 55.2 (29-80) Men 26 (66.7) Primary kidney disease 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 7 0 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/dL) 2.6 ± 3.23 Potassium (mmol/L) 2.25 ± 7.5.34 Alburnin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.5 ± 0.18 Phosphate (mmol/L) <t< th=""><th>Patient demographics</th><th></th></t<>	Patient demographics	
Age (years, mean, range) 55.2 (29-80) Men 26 (66.7) Primary kidney disease 3 Diabetes 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 7 0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) Weekly total (renal-peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) Datoratory (mean ± SD) 12 (30.8) Hemoglobin (g/dL) 7.25 ± 7.5.34 Albumin (g/L) 2.6 ± 7.5.34 Albumin (g/L) 2.6 ± 3.45 Sodium (mmol/L) 2.5 ± 0.18 Phosphate (Total number of patients	39
Men 26 (66.7) Primary kidney disease 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dalysis vintage, months median (IQR) 1.61 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 0 0 19 (48.7) A 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/dL) 7.2 ± 2.11 Thrombocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 2.25 ± 7.5.34 Albumin (g/L) 2.65 ± 3.25 C-reactive protein (mpol/L) 2.7 ± 5.37 Obesity (BMI > 30	Age (years, mean, range)	55.2 (29-80)
Primary kidney disease 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 0 0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.6.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR" (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group ABO (%) 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/L) 1.06 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 2.25 ± 7.5.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.6 ± 3.23 Potassium (mmol/L) 2.7 ± 5.37 Ob	Men	26 (66.7)
Diabetes 11 Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 0 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.6.1 (6-31.3) Blood Group AB0 (%) 1.94 (1.78-2.35) GFR (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/dL) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 2.25 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 1.36 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 0.52 ± 0.69 <	Primary kidney disease	
Vascular Disease 3 Glomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.6.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/cL) 10.6 ± 3.423 Calcium (mmol/L) 2.25 ± 7.5.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.3	Diabetes	11
Giomerulonephritis 8 ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 0 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.6.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GR 6 (15.4) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) Hemoglobin (g/L) Hemoglobin (g/L) 2.6 ± 3.45 Sodium (mmol/L) 2.25 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30	Vascular Disease	3
ADPKD 4 Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) God Group AB0 (%) 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/L) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.7 ± 5.37 Obesity (BMI > 30 kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) 27 ± 5.37	Glomerulonephritis	8
Unknown 5 Other 8 Davies Comorbidity Score (%) 14 (36) 0 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.61.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 0 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/LL) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 2.25 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (B	ADPKD	4
Other 8 Davies Comorbidity Score (%) 14 (36) 0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 0 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/L) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.58 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m ²) 27 ± 5.37 Obesity (BMI > 30 kg/m ²) (%) 10 (25.6) Medication (n	Unknown	5
Davies Comorbidity Score (%) 14 (36) 0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 1.6.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 0 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/L) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 2.25 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.5 ± 0.18 Phosphate (mmol/L) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 20 Immunosup	Other	8
0 14 (36) 1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 0 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 2.25 ± 75.34 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.58 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 1 Immunosuppressive therapy 7 RAAS-inhi	Davies Comorbidity Score (%)	
1 14 (36) 2 7 (18) 3 3 (7.7) 4 0 5 1 (2.6) Dialysis vintage, months median (IQR) 16.1 (6-31.3) Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group ABO (%) 0 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/L) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.5.8 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 10 Immunosuppressive therapy 7 <	0	14 (36)
$\begin{array}{ccccccc} 2 & 7 & (18) \\ 3 & 3 & (7.7) \\ 4 & 0 \\ 5 & 1 & (2.6) \\ Dialysis vintage, months median (IQR) & 16.1 & (6-31.3) \\ Weekly total (renal+peritoneal) Kt/V median (IQR) & 1.94 & (1.78-2.35) \\ GFR* (mL/min) median (IQR) & 1.95 & (0.76-6.01) \\ Blood Group AB0 & (\%) & 0 \\ 0 & 19 & (48.7) \\ A & 12 & (30.8) \\ B & 6 & (15.4) \\ AB & 2 & (5.1) \\ Laboratory (mean \pm SD) \\ Hemoglobin (g/dL) & 10.6 \pm 1.39 \\ Leukocytes (G/L) & 7.2 \pm 2.11 \\ Thrombocytes (G/L) & 7.2 \pm 2.11 \\ Thrombocytes (G/L) & 225 \pm 75.34 \\ Albumin (g/L) & 3.6 \pm 3.45 \\ Sodium (mmol/L) & 136 \pm 3.23 \\ Potassium (mmol/L) & 136 \pm 3.23 \\ Potassium (mmol/L) & 2.55 \pm 0.18 \\ Phosphate (mmol/L) & 2.58 \pm 3.25 \\ C-reactive protein (mg/dL) & 0.52 \pm 0.69 \\ Body mass index (kg/m^2) & 27 \pm 5.37 \\ Obesity (BMI > 30 kg/m^2) & (\%) & 10 & (25.6) \\ Medication (n) & 1 \\ Immunosuppressive therapy & 7 \\ RAAS-inhibitor medication & 32 \\ Antibody titer (U/mL) median (IQR) \\ after 1st dose & 6.62 & (1.57-22.5) \\ \end{array}$	1	14 (36)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2	7 (18)
$\begin{array}{cccc} 4 & 0 \\ 5 & 1 (2.6) \\ \mbox{Dialysis vintage, months median (IQR)} & 16.1 (6-31.3) \\ \mbox{Weekly total (renal+peritoneal) Kt/V median (IQR)} & 1.94 (1.78-2.35) \\ \mbox{GFR* (mL/min) median (IQR)} & 1.95 (0.76-6.01) \\ \mbox{Blood Group AB0 (\%)} & & & & & & & & & & & & & & & & & & &$	3	3 (7.7)
$\begin{array}{cccc} 5 & 1 (2.6) \\ \mbox{Dialysis vintage, months median (IQR)} & 16.1 (6-31.3) \\ \mbox{Weekly total (renal+peritoneal) Kt/V median (IQR)} & 1.94 (1.78-2.35) \\ \mbox{GFR* (mL/min) median (IQR)} & 1.95 (0.76-6.01) \\ \mbox{Blood Group AB0 (%)} & & & & & & & & & & & & & & & & & & &$	4	0
Dialysis vintage, months median (IQR)16.1 (6-31.3)Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35)GFR* (mL/min) median (IQR) 1.95 (0.76-6.01)Blood Group AB0 (%) 0 0 19 (48.7)A 12 (30.8)B 6 (15.4)AB 2 (5.1)Laboratory (mean \pm SD) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.7 ± 0.42 Bicarbonate (mmol/L) 2.7 ± 0.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6)Medication (n) $10 (25.6)$ Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 6.62 (1.57-22.5)	5	1 (2.6)
Weekly total (renal+peritoneal) Kt/V median (IQR) 1.94 (1.78-2.35) GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 12 (30.8) 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean \pm SD) 10.6 \pm 1.39 Leukocytes (G/L) 7.2 \pm 2.11 Thrombocytes (G/L) 225 \pm 75.34 Albumin (g/L) 3.6 \pm 3.45 Sodium (mmol/L) 3.6 \pm 3.23 Potassium (mmol/L) 2.25 \pm 0.18 Phosphate (mmol/L) 2.7 \pm 0.42 Bicarbonate (mmol/L) 2.5.8 \pm 3.25 C-reactive protein (mg/dL) 0.52 \pm 0.69 Body mass index (kg/m ²) 27 \pm 5.37 Obesity (BMI > 30 kg/m ²) (%) 10 (25.6) Medication (n) 10 Immunosuppressive therapy 7 RAAS-inhibitor medication 32 Antibody titer (U/mL) median (IQR) 32	Dialysis vintage, months median (IQR)	16.1 (6-31.3)
GFR* (mL/min) median (IQR) 1.95 (0.76-6.01) Blood Group AB0 (%) 12 (30.8) 0 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.7 ± 0.42 Bicarbonate (mmol/L) 25.8 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 10 (25.6) Immunosuppressive therapy 7 RAAS-inhibitor medication 32 Antibody titer (U/mL) median (IQR) 32	Weekly total (renal+peritoneal) Kt/V median (IQR)	1.94 (1.78-2.35)
Blood Group AB0 (%) 19 (48.7) A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/dL) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 1.36 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 1.77 ± 0.42 Bicarbonate (mmol/L) 25.8 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 10 (25.6) Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 6.62 (1.57-22.5)	GFR* (mL/min) median (IQR)	1.95 (0.76-6.01)
$\begin{array}{cccc} 0 & 19 (48.7) \\ A & 12 (30.8) \\ B & 6 (15.4) \\ AB & 2 (5.1) \\ \\ Laboratory (mean \pm SD) \\ Hemoglobin (g/dL) & 10.6 \pm 1.39 \\ Leukocytes (G/L) & 7.2 \pm 2.11 \\ Thrombocytes (G/L) & 225 \pm 75.34 \\ \\ Albumin (g/L) & 3.6 \pm 3.45 \\ \\ Sodium (mmol/L) & 136 \pm 3.23 \\ \\ Potassium (mmol/L) & 136 \pm 3.23 \\ \\ Potassium (mmol/L) & 2.25 \pm 0.18 \\ \\ Phosphate (mmol/L) & 2.58 \pm 3.25 \\ \\ C-reactive protein (mg/dL) & 0.52 \pm 0.69 \\ \\ Body mass index (kg/m^2) & 27 \pm 5.37 \\ \\ Obesity (BM > 30 kg/m^2) (%) & 10 (25.6) \\ \\ Medication (n) & \\ \\ Immunosuppressive therapy & 7 \\ \\ RAAS-inhibitor medication & 20 \\ \\ \\ Vitamin D medication & 32 \\ \\ \\ Antibody titer (U/mL) median (IQR) \\ after 1st dose & 6.62 (1.57-22.5) \\ \end{array}$	Blood Group AB0 (%)	
A 12 (30.8) B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 25.8 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 1 Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 6.62 (1.57-22.5)	0	19 (48.7)
B 6 (15.4) AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 4.4 ± 0.63 Calcium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 25.8 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 1 Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 6.62 (1.57-22.5)	A	12 (30.8)
AB 2 (5.1) Laboratory (mean ± SD) 10.6 ± 1.39 Hemoglobin (g/dL) 10.6 ± 1.39 Leukocytes (G/L) 7.2 ± 2.11 Thrombocytes (G/L) 225 ± 75.34 Albumin (g/L) 3.6 ± 3.45 Sodium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 136 ± 3.23 Potassium (mmol/L) 2.25 ± 0.18 Phosphate (mmol/L) 2.58 ± 3.25 C-reactive protein (mg/dL) 0.52 ± 0.69 Body mass index (kg/m²) 27 ± 5.37 Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 1 Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 6.62 (1.57-22.5)	В	6 (15.4)
Laboratory (mean \pm SD)Hemoglobin (g/dL)10.6 \pm 1.39Leukocytes (G/L)7.2 \pm 2.11Thrombocytes (G/L)225 \pm 75.34Albumin (g/L)3.6 \pm 3.45Sodium (mmol/L)136 \pm 3.23Potassium (mmol/L)136 \pm 3.23Calcium (mmol/L)2.25 \pm 0.18Phosphate (mmol/L)2.7 \pm 0.42Bicarbonate (mmol/L)0.52 \pm 0.69Body mass index (kg/m²)27 \pm 5.37Obesity (BMI > 30 kg/m²) (%)10 (25.6)Medication (n)20Immunosuppressive therapy7RAAS-inhibitor medication20Vitamin D medication32Antibody titer (U/mL) median (IQR)6.62 (1.57-22.5)	AB	2 (5.1)
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Laboratory (mean \pm SD)	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	Hemoglobin (g/dL)	10.6 ± 1.39
$\begin{tabular}{lllllllllllllllllllllllllllllllllll$	Leukocytes (G/L)	7.2 ± 2.11
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Thrombocytes (G/L)	225 ± 75.34
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Albumin (g/L)	3.6 ± 3.45
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Sodium (mmol/L)	136 ± 3.23
$\begin{array}{ll} \mbox{Calcium (mmol/L)} & 2.25 \pm 0.18 \\ \mbox{Phosphate (mmol/L)} & 1.77 \pm 0.42 \\ \mbox{Bicarbonate (mmol/L)} & 25.8 \pm 3.25 \\ \mbox{C-reactive protein (mg/dL)} & 0.52 \pm 0.69 \\ \mbox{Body mass index (kg/m^2)} & 27 \pm 5.37 \\ \mbox{Obesity (BMI > 30 kg/m^2) (%)} & 10 (25.6) \\ \mbox{Medication (n)} & & \\ \mbox{Immunosuppressive therapy} & 7 \\ \mbox{RAAS-inhibitor medication} & 20 \\ \mbox{Vitamin D medication} & 32 \\ \mbox{Antibody titer (U/mL) median (IQR)} \\ \mbox{after 1st dose} & 6.62 (1.57-22.5) \\ \end{array}$	Potassium (mmol/L)	4.4 ± 0.63
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Calcium (mmol/L)	2.25 ± 0.18
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Phosphate (mmol/L)	1.77 ± 0.42
	Bicarbonate (mmol/L)	25.8 ± 3.25
Body mass index (kg/m²)27 ± 5.37Obesity (BMI > 30 kg/m²) (%)10 (25.6)Medication (n)10Immunosuppressive therapy7RAAS-inhibitor medication20Vitamin D medication32Antibody titer (U/mL) median (IQR)6.62 (1.57-22.5)	C-reactive protein (mg/dL)	0.52 ± 0.69
Obesity (BMI > 30 kg/m²) (%) 10 (25.6) Medication (n) 10 Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) after 1st dose 6.62 (1.57-22.5)	Body mass index (kg/m ²)	27 ± 5.37
Medication (n) Immunosuppressive therapy 7 Immunosuppressive therapy 7 RAAS-inhibitor medication 20 Vitamin D medication 32 Antibody titer (U/mL) median (IQR) 32 after 1st dose 6.62 (1.57-22.5)	Obesity (BMI > 30 kg/m ²) (%)	10 (25.6)
Immunosuppressive therapy7RAAS-inhibitor medication20Vitamin D medication32Antibody titer (U/mL) median (IQR)32after 1st dose6.62 (1.57-22.5)	Medication (n)	· · · · ·
RAAS-inhibitor medication20Vitamin D medication32Antibody titer (U/mL) median (IQR) after 1st dose6.62 (1.57-22.5)	Immunosuppressive therapy	7
Vitamin D medication32Antibody titer (U/mL) median (IQR) after 1st dose6.62 (1.57-22.5)	RAAS-inhibitor medication	20
Antibody titer (U/mL) median (IQR) after 1st dose 6.62 (1.57-22.5)	Vitamin D medication	32
after 1st dose 6.62 (1.57-22.5)	Antibody titer (U/mL) median (IQR)	
	after 1st dose	6.62 (1.57-22.5)
after 2nd dose 968 (422.5-2500)	after 2nd dose	968 (422.5-2500)

Data are presented as n (%), mean ± SD or as median (IQR); Other = Alport syndrome (1), GVHD-associated TMA (1), cardiorenal syndrome (1), secondary FSGS (1), AL-Amyloidosis (1), glomerulosclerosis (1), nephronophtisis (1), scleroderma (1): GVHD, graft versus host disease; TMA, thrombotic microangiopathy; FSGS, focal segmental glomerulosclerosis; BMI, body mass index; weekly Kt/V, clearance of urea x time/volume; GFR, residual glomerular filtration rate; RAAS, renin-angiotensin-aldosteron system; CNI, calcineurin inhibitors; MMF, mycophenolate-mofetil. *residual GFR was calculated as mean of renal creatinine and renal urea clearance using 24 h urine samples.

previous kidney graft failure and one patient had psoriasis (**Table 2**). Interestingly, patients who received mycophenolate mofetil (MMF) as part of the immunosuppressive therapy developed low antibody titers (below the 25% quantile), including the lung transplanted patient who received triple immunosuppressive therapy. In contrast, patients receiving prednisolon alone or in combination with a calcineurin inhibitor (CNI) had higher antibody titers reaching the 3rd quartile (**Table 2**).

Based on clinical significance and published data, antibody titers after the first dose, age, gender, immunosuppressive therapy, sodium, albumin, CRP, dialysis vintage, BMI, Kt/V, GFR and Davies Comorbidity Score were considered as potential predictors for humoral response after vaccination.

In the univariate analysis, shorter dialysis vintage and a lower Davies Comorbidity Score were associated with higher antibody titers after the first dose (p=0.017 and p=0.04, respectively) (Table 3). Patients who developed higher antibody titers after the first dose tended to have a higher antibody response after the second dose ($p=1.53 \times 10^{-05}$). Thus, titers after the first dose may predict antibody response after full vaccination as illustrated in Figure 1. The other variables tested as potential predictors did not meet statistical significance, although trends could be observed. Men tended to have lower antibody titers (p=0.06), whereas higher GFR (p=0.08) and younger age (p=0.09) seemed to be associated with higher antibody titers. When added to a model including titers after the first dose none of the additional covariates showed a significant effect on the antibody levels after the second dose in the multivariate analysis (Table 4).

Reported Adverse Events

Patients were asked to report AEs (local and systemic) that occurred within 7 days after the first and second vaccine dose. After the first dose the most common local AEs were pain (71%), swelling (10%) and bruising (10%) at the injection site. Systemic AEs occurring after the first dose were mostly fatigue (33%) and headache (20%). No severe systemic AEs were reported after the first injection. After the second dose patients reported local pain (71.5%) and swelling (13%) at the injection site. The incidence and the severity of the systemic AEs increased after the second



FIGURE 1 | Antibody titers in 39 peritoneal dialysis patients after the first and the second dose of the mRNA-1273 vaccine. Seropositivity was detected in 38 of 39 peritoneal dialysis patients (circles). Antibody titers increased significantly from a median of 6.62 U/mL (IQR 1.57-22.5) four weeks after the first dose to 968 U/mL (IQR 422.5-2500 U/mL) after the second dose (Wilcoxon matched pairs signed rank test, p = 8.057×10^{-08}). The technical upper limit of antibody detection was at 2500 U/mL and was reached in 11 patients (circles may overlap).

dose: the most common AEs were fatigue (40.5%), headache (22.5%), joint pain (20%), myalgia (17.5%) and fever (13%) and 7.7% of the patients graded the AEs as severe. All reported AEs were graded as mild, moderate or severe, no hospitalizations (serious AEs) were reported (**Figure 2**).

DISCUSSION

In this study we showed that 38 of 39 PD patients at our institution responded well to two doses of the anti-SARS-CoV-2 mRNA-1273 vaccine, reaching a seroconversion rate of 97.4%. The majority of the studies on humoral response to mRNA-based anti-SARS-CoV-2 vaccines included patients on hemodialysis. The seroconversion rates found in COVID-19 naive HD patients using predominantly the BNT162b2 vaccine were below 43% after a single dose of vaccination and between 82-97% after two doses (16, 23-42). Only three studies reported lower seroconversion rates (71-73%) in HD patients (36, 43, 44). Kaiser et al. described in a non-randomized retrospective study including 116 HD patients, that patients vaccinated with mRNA-1273 developed higher antibody titers than those that received BNT162b2 (32), whereas Broseta et al. reported similar seroconversion rates (45).

Few studies investigated the immune response of PD patients after SARS-CoV-2 vaccination, using predominantly the BNT162b2 vaccine (16, 23, 34, 39). These studies described seroconversion rates after the second dose between 85% and 100%, which are comparable with those found in the present study. In the only study analyzing the humoral response of PD patients to mRNA-1273 vaccine, published by Rodriguez-Espinosa, the seroconversion rate after two vaccine doses (97%) was similar to our results (97.4%) (46). However, in our study the percentage of patients with detectable humoral response after the first dose was higher than the one found by Rodriguez-Espinosa (86.4% vs 62.5%). Younger age (mean 55.2 vs 62.2 years) and shorter dialysis vintage (mean 22.5 vs 64.5 months) of our cohort could explain the higher seroconversion rate after one dose of vaccine observed in our PD patients.

Rodriguez-Espinosa et al. could not find any factors associated with the antibody response to the mRNA-1273 vaccine in PD patients (46). In an observational study including 21 PD and 35 HD patients, Tylicki et al. suggested that younger age, shorter dialysis vintage, better residual renal function and fewer comorbidities of PD patients lead to a higher antibody response after vaccination with the BNT162b2 vaccine in the PD cohort (16). In accordance with this hypothesis we found in the univariate analysis that shorter dialysis vintage and a lower Davies Comorbidity Score were associated with higher antibody titers after the first vaccine dose. Other studies reported that younger age (23, 39) and higher serum albumin (23) were factors that correlated with higher antibody levels in response to BNT162b2 vaccine in PD patients. Since both studies did not report the characteristics of included PD patients, any comparison with our population was unfeasible. In the multivariate analysis none of the factors (age, gender, immunosuppressive therapy, sodium, albumin, CRP, dialysis vintage, BMI, Kt/V, GFR and Davies

TABLE 2 | Antibody titers after mRNA-1273 vaccination in peritoneal dialysis patients with immunosuppressive therapy.

Patient (Sex, age)	IS	Dosage	Indication	Antibody titer (U/mL) 4 weeks after 1 st dose	Antibody titer (U/mL) 4 weeks after 2 nd dose
M, 43	CNI Prednisolon	2mg q.d. 5mg q.d.	Lung transplant	<0.40	<0.40
	MMF	250mg t.i.d.			
M, 43	MMF	500mg q.d.	Kidney graft failure	<0.40	234.00
F, 35	MMF	500mg b.i.d.	Kidney graft failure	27.10	250.00
F, 74	CNI Prednisolon	0.5mg q.d. 2.5mg q.d.	Kidney graft failure	17.70	1330.00
M, 54	Prednisolon	2.5mg q.d.	Kidney graft failure	13.90	1543.00
F, 55	Prednisolon	2.5mg q.d.	Kidney graft failure	3.58	2500.00
M, 76	Ustekinumab	90mg every 3 months	Psoriasis	20.00	677.00

M, male; F, female; IS, immunosuppressive therapy; CNI, calcineurin inhibitor; MMF, mycophenolate-mofetil; Ustekinumab (anti-IL-12/23); < 0,4 U/mL = negative (lower limit); 2500 U/mL = maximum (upper limit).

TABLE 3 | Effects of covariates on antibody titers after the first and the second dose of mRNA-1273 SARS-CoV-2 vaccine (univariate analysis).

	After 1 st dose			After 2 nd dose		
	β-coefficient	t value	p value	β-coefficient	t value	p value
Titer after 1 st dose	NA	NA	NA	0.44	4.97	1.53 x 10 ⁻⁵
Age	-0.05	-1.37	0.18	-0.03	-1.72	0.09
Gender	-1.48	-1.92	0.06	-0.80	-1.54	0.13
IS	0.74	0.57	0.57	-0.36	-0.39	0.70
Sodium	0.17	1.43	0.16	0.09	1.61	0.11
Albumin	0.12	1.92	0.36	0.09	1.16	0.25
CRP	-0.64	-0.92	0.36	-0.29	-0.62	0.53
Dialysis vintage	-0.04	-2.5	0.017	-0.02	-1.07	0.29
BMI	0.04	0.33	0.74	-0.05	-0.83	0.41
Weely total Kt/V	1.49	1.39	0.17	0.76	1.43	0.16
GFR*	0.23	1.78	0.08	0.08	1.30	0.20
Davies Score	-0.68	-2.12	0.04	-0.33	-2.01	0.051

Univariate analysis of the variables associated with antibody titers after the first and the second dose of mRNA-1273 vaccine using univariate quantile regression. P<0.05 was considered significant (significant associations marked in bold). IS, immunosuppression; BMI, body mass index; weekly Kt/V, clearance of urea x time/volume; *GFR, residual glomerular filtration rate (calculated as mean of renal creatinine and renal urea clearance using 24 h urine samples).

TABLE 4	Effects of the antibody	y titer after the 1 ³	st dose and other covariates or	1 the antibody titer after	er the 2 nd dose o	of mRNA-1273 SARS	-CoV-2 vaccine (multivariate analys	sis).
---------	-------------------------	----------------------------------	--	----------------------------	-------------------------------	-------------------	------------------	---------------------	-------

	After 2 nd dose			
	β-coefficient	t value	p value	
Age	-0.007	-0.69	0.49	
Gender	-0.31	-0.94	0.35	
IS	-0.11	-0.14	0.88	
Sodium	0.05	0.86	0.39	
Albumin	0.07	0.89	0.37	
CRP	0.02	0.09	0.92	
Dialysis vintage	0.001	0.09	0.92	
BMI	-0.03	-0.99	0.32	
Weekly total Kt/V	0.14	0.75	0.45	
GFR*	0.03	0.90	0.37	
Davies Score	-0.16	-0.91	0.37	

Results from multivariate quantile regression using the logarithm of the antibody titer after the first dose of mRNA-1273 plus an additional variable as predictors for the logarithm of antibody titer after the second dose. log(Titer 1) was always highly significant whereas none of the additional parameters became significant at the usual level α = 0.05. IS, immunosuppression; BMI, body mass index; weekly Kt/V, clearance of urea x time/volume per week; *GFR, residual glomerular filtration rate (calculated as mean of renal creatinine and renal urea clearance using 24 h urine samples).

Comorbidity Score) had a significant effect on the antibody titers in our PD cohort, possibly due to the limited number of included patients. However, we were able to observe some trends. Men tended to have lower antibody titers. Higher GFR and younger age seemed to be associated with higher antibody titers, confirming previous data with the BNT162b2 vaccine (16, 23, 39).

An important factor that interferes with the humoral response to the SARS-CoV-2 vaccines appears to be the



immunosuppressive therapy of kidney transplant and dialysis patients (34, 47-49). We showed that among patients with immunosuppressive therapy, those who were treated with MMF had a lower anti-SARS-CoV-2-S antibody response compared to the patients treated with prednisolon. MMF, a potent inhibitor of both T- and B- lymphocyte proliferation, is broadly used as immunosuppressive drug in transplant patients (50). In agreement with our findings, recently published data described a MMF-dose dependent reduction of humoral response after SARS-Cov-2 vaccination in kidney transplanted patients (47-49). Furthermore, MMF was associated with a significantly reduced BNT162b2-induced immunogenicity in patients with autoimmune inflammatory rheumatic diseases (51). These observations raise the question whether interrupting or reducing MMF dose before vaccination might improve the humoral response.

To our knowledge this is the first study describing the local and systemic adverse events in PD patients after the first and the second mRNA-1273 vaccination. The percentage of PD patients reporting AEs at the site of injection was lower than in the general population receiving the same vaccine: (79.5% vs. 84.3% after the first dose and 79.5% vs. 88.6% after the second dose, respectively) (3). As for the general population, the most common local AE was pain at the injection site. The occurrence of systemic AEs in our patients was less frequent in comparison to the general population (38.5% vs. 54.9% after the first dose and 51.3% vs. 79.4% after the second dose, respectively). These results could be explained by the fact that our study cohort consisted of 66.7% male participants compared to the 52.7% reported by Baden et al. Other studies have reported that women show a higher rate of adverse events in response to mRNA vaccines (52). Furthermore, the incidence of local and systemic adverse events in our study population was higher than in HD patients receiving the BNT162b2 vaccine (42, 52). The fact that our study participants were treated with PD, were on average 13 years younger than the participants of the other studies and received a different vaccine may explain these differences. A negative correlation between age and number of adverse events in response to mRNA vaccines has been reported previously (4, 42). Accordingly, systemic AEs in our study were more common in younger patients (87% of the patients reporting systemic AEs were younger than 65 years old). Furthermore, the severity of AEs increased after the second dose of vaccine, in consistency with data published by Baden et al. in the general population (3).

Our study had certain limitations. Most importantly, interpretation of our data may be limited by the fairly small sample size. Other limitations include the retrospective single

center design and lack of a control group. However, our data on the safety of the mRNA-1273 vaccine in PD patients represent an asset to the current knowledge, since, so far, safety data using this type of vaccine have not been reported in this patient group. The fact that the majority of adverse events were mild and no serious AEs occurred, might be helpful for PD patients to overcome their skepticism towards SARS-CoV-2 vaccination.

In conclusion, we found a high seroconversion rate of 97.4% and high median antibody titer levels in this cohort of PD patients after vaccination with two doses of mRNA-1273. Lower Davies Comorbidity Score and shorter dialysis vintage were associated with higher antibody titers after the first dose. Patients with higher antibody titers after the first dose tended to have higher antibody titers after the second dose. The two dose vaccination scheme was tolerated well and did not cause any serious AEs in this population.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

REFERENCES

- Nagel A, Laszewska A, Haidinger G, Simon J. The First 8 Weeks of the Austrian SARS-CoV-2 Epidemic. Wien Klin Wochenschr (2021) 133:364–76. doi: 10.1007/s00508-020-01804-9
- 2. Bundesministerium für Inneres Austria (2021). Available at: https://www.bmi. gv.at/news.aspx?id=4A7171477A51625143334D3D.
- Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine. N Engl J Med (2021) 384:403–16. doi: 10.1056/NEJMoa2035389
- Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. N Engl J Med (2020) 383:2603–15. doi: 10.1056/NEJMoa2034577
- Wu K, Werner AP, Koch M, Choi A, Narayanan E, Stewart-Jones GBE, et al. Serum Neutralizing Activity Elicited by mRNA-1273 Vaccine. N Engl J Med (2021) 384:1468–70. doi: 10.1056/NEJMc2102179
- Hilbrands LB, Duivenvoorden R, Vart P, Franssen CFM, Hemmelder MH, Jager KJ, et al. COVID-19-Related Mortality in Kidney Transplant and Dialysis Patients: Results of the ERACODA Collaboration. *Nephrol Dial Transplant* (2020) 35:1973–83. doi: 10.1093/ndt/gfaa261
- Girndt M, Pietsch M, Kohler H. Tetanus Immunization and Its Association to Hepatitis B Vaccination in Patients With Chronic Renal Failure. *Am J Kidney Dis* (1995) 26:454–60. doi: 10.1016/0272-6386(95)90491-3
- Kohler H, Arnold W, Renschin G, Dormeyer HH, Meyer zum Buschenfelde KH. Active Hepatitis B Vaccination of Dialysis Patients and Medical Staff. *Kidney Int* (1984) 25:124–8. doi: 10.1038/ki.1984.18
- Kreft B, Klouche M, Kreft R, Kirchner H, Sack K. Low Efficiency of Active Immunization Against Diphtheria in Chronic Hemodialysis Patients. *Kidney Int* (1997) 52:212–6. doi: 10.1038/ki.1997.322
- Rautenberg P, Proppe D, Schutte A, Ullmann U. Influenza Subtype-Specific Immunoglobulin A and G Responses After Booster Versus One Double-Dose Vaccination in Hemodialysis Patients. *Eur J Clin Microbiol Infect Dis* (1989) 8:897–900. doi: 10.1007/BF01963779
- Friedman EA, Beyer MM, Hirsch SR, Schiffman G. Intact Antibody Response to Pneumococcal Capsular Polysaccharides in Uremia and Diabetes. *JAMA* (1980) 244:2310–1. doi: 10.1001/jama.1980.03310200050027
- 12. Kronbichler A, Anders HJ, Fernandez-Juarez GM, Floege J, Goumenos D, Segelmark M, et al. Immunonephrology Working Group of the,

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Ethics Committee of the Medical University of Vienna EK 1418/2021. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

AUTHOR CONTRIBUTIONS

GB: contributed to conception and design of the study, analyzed data and wrote the manuscript. RM: analyzed data and wrote sections of the manuscript. FF: contributed to the design of the study and performed the statistical analysis. MG: contributed to conception and design of the study RS: performed antibody titer measurements. AV: contributed to conception and design of the study and wrote the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

ACKNOWLEDGMENTS

We sincerely thank our patients, our peritoneal dialysis team and our medical student Mr. Filip Zubic for their outstanding support.

Recommendations for the Use of COVID-19 Vaccines in Patients With Immune-Mediated Kidney Diseases. *Nephrol Dial Transplant* (2021) gfab064. doi: 10.1093/ndt/gfab064

- Windpessl M, Bruchfeld A, Anders HJ, Kramer H, Waldman M, Renia L, et al. COVID-19 Vaccines and Kidney Disease. *Nat Rev Nephrol* (2021) 17:291–3. doi: 10.1038/s41581-021-00406-6
- 14. Austrian Dialysis and Transplantation Registry (2021). Available at: https:// www.nephrologie.at/wp-content/uploads/COVID-19-OEDTR-240921-Tab.pdf.
- Ikizler TA, Coates PT, Rovin BH, Ronco P. Immune Response to SARS-CoV-2 Infection and Vaccination in Patients Receiving Kidney Replacement Therapy. *Kidney Int* (2021) 99:1275–9. doi: 10.1016/j.kint.2021.04.007
- Tylicki L, Piotrowska M, Biedunkiewicz B, Zielinski M, Dabrowska M, Tylicki P, et al. Humoral Response to COVID-19 Vaccination in Patients Treated With Peritoneal Dialysis: The COViNEPH Project. *Pol Arch Intern Med* (2021) 131(10). doi: 10.20452/pamw.16091
- Carr EJ, Kronbichler A, Graham-Brown M, Abra G, Argyropoulos C, Harper L, et al. Systematic Review of Early Immune Response to SARS-CoV-2 Vaccination Among Patients With Chronic Kidney Disease. *Kidney Int Rep* (2021) 6(9):2292–304. doi: 10.1016/j.ekir.2021.06.027
- Davies SJ, Phillips L, Naish PF, Russell GI. Quantifying Comorbidity in Peritoneal Dialysis Patients and Its Relationship to Other Predictors of Survival. *Nephrol Dial Transplant* (2002) 17:1085–92. doi: 10.1093/ndt/ 17.6.1085
- Roche. Available at: https://assets.cwp.roche.com/f/94122/x/379ebe6732/ factsheet-elecsys-anti-sars-cov-2-s_v1.pdf.
- Yuan M, Liu H, Wu NC, Wilson IA. Recognition of the SARS-CoV-2 Receptor Binding Domain by Neutralizing Antibodies. *Biochem Biophys Res Commun* (2021) 538:192–203. doi: 10.1016/j.bbrc.2020.10.012
- 21. Koenker R. Quantreg: Quantile Regression. R Package Version 5.86 (2021). Available at: https://CRAN.R-project.org/package=quantreg.
- 22. R Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing (2021). Available at: https://www.R-project.org/.
- Agur T, Ben-Dor N, Goldman S, Lichtenberg S, Herman-Edelstein M, Yahav D, et al. Antibody Response to mRNA SARS-CoV-2 Vaccine Among Dialysis Patients - A Prospectivecohort Study. *Nephrol Dial Transplant* (2021) gfab155. doi: 10.1093/ndt/gfab155

- Anand S, Montez-Rath M, Han J, Garcia P, Cadden L, Hunsader P, et al. Antibody Response to COVID-19 Vaccination in Patients Receiving Dialysis. J Am Soc Nephrol (2021) 32(10):2435–8. doi: 10.1101/2021.05.06.21256768
- Attias P, Sakhi H, Rieu P, Soorkia A, Assayag D, Bouhroum S, et al. Antibody Response to the BNT162b2 Vaccine in Maintenance Hemodialysis Patients. *Kidney Int* (2021) 99:1490–2. doi: 10.1016/j.kint.2021.04.009
- Billany RE, Selvaskandan H, Adenwalla SF, Hull KL, March DS, Burton JO, et al. Seroprevalence of Antibody to S1 Spike Protein Following Vaccination Against COVID-19 in Patients Receiving Hemodialysis: A Call to Arms. *Kidney Int* (2021) 99:1492–4. doi: 10.1016/j.kint.2021.04.008
- Chan L, Fuca N, Zeldis E, Campbell KN, Shaikh A. Antibody Response to mRNA-1273 SARS-CoV-2 Vaccine in Hemodialysis Patients With and Without Prior COVID-19. *Clin J Am Soc Nephrol* (2021) 16:1258–60. doi: 10.2215/CJN.04080321
- Frantzen L, Cavaille G, Thibeaut S, El-Haik Y. Efficacy of the BNT162b2 mRNA COVID-19 Vaccine in a Haemodialysis Cohort. Nephrol Dial Transplant (2021) 36:1756–7. doi: 10.1093/ndt/gfab165
- Goupil R, Benlarbi M, Beaubien-Souligny W, Nadeau-Fredette AC, Chatterjee D, Goyette G, et al. Short-Term Antibody Response After 1 Dose of BNT162b2 Vaccine in Patients Receiving Hemodialysis. *CMAJ* (2021) 193: E793–800. doi: 10.1503/cmaj.210673
- Grupper A, Sharon N, Finn T, Cohen R, Israel M, Agbaria A, et al. Humoral Response to the Pfizer BNT162b2 Vaccine in Patients Undergoing Maintenance Hemodialysis. *Clin J Am Soc Nephrol* (2021) 16(7):1037–42. doi: 10.2215/CJN.03500321
- 31. Jahn M, Korth J, Dorsch O, Anastasiou OE, Sorge-Hadicke B, Tyczynski B, et al. Humoral Response to SARS-CoV-2-Vaccination With BNT162b2 (Pfizer-BioNTech) in Patients on Hemodialysis. *Vaccines (Basel)* (2021) 9 (4):360. doi: 10.3390/vaccines9040360
- Kaiser RA, Haller MC, Apfalter P, Kerschner H, Cejka D. Comparison of BNT162b2 (Pfizer-BioNtech) and mRNA-1273 (Moderna) SARS-CoV-2 mRNA Vaccine Immunogenicity in Dialysis Patients. *Kidney Int* (2021) 100:697–8. doi: 10.1016/j.kint.2021.07.004
- Lacson E, Argyropoulos C, Manley H, Aweh G, Chin A, Salman L, et al. Immunogenicity of SARS-CoV-2 Vaccine in Dialysis. J Am Soc Nephrol (2021) 32(11):2735–42. doi: 10.1101/2021.04.08.21254779
- 34. Longlune N, Nogier MB, Miedouge M, Gabilan C, Cartou C, Seigneuric B, et al. High Immunogenicity of a Messenger RNA Based Vaccine Against SARS-CoV-2 in Chronic Dialysis Patients. *Nephrol Dial Transplant* (2021) 36 (9):1704–9. doi: 10.1093/ndt/gfab193
- Chrezenmeier E, Bergfeld L, Hillus D, Lippert JD, Weber U, Tober-Lau P, et al. Immunogenicity of COVID-19 Tozinameran Vaccination in Patients on Chronic Dialysis. Front Immunol (2021) 12:690698. doi: 10.3389/fimmu.2021.690698
- 36. Speer C, Goth D, Benning L, Buylaert M, Schaier M, Grenz J, et al. Early Humoral Responses of Hemodialysis Patients After COVID-19 Vaccination With BNT162b2. *Clin J Am Soc Nephrol* (2021) 16(7):1073–82. doi: 10.2215/ CJN.03700321
- 37. Strengert M, Becker M, Ramos GM, Dulovic A, Gruber J, Juengling J, et al. Cellular and Humoral Immunogenicity of a SARS-CoV-2 mRNA Vaccine in Patients on Haemodialysis. *EBioMedicine* (2021) 70:103524. doi: 10.1016/ j.ebiom.2021.103524
- Torreggiani M, Blanchi S, Fois A, Fessi H, Piccoli GB. Neutralizing SARS-CoV-2 Antibody Response in Dialysis Patients After the First Dose of the BNT162b2 mRNA COVID-19 Vaccine: The War Is Far From Being Won. *Kidney Int* (2021) 99:1494–6. doi: 10.1016/j.kint.2021.04.010
- Yanay NB, Freiman S, Shapira M, Wishahi S, Hamze M, Elhaj M, et al. Experience With SARS-CoV-2 BNT162b2 mRNA Vaccine in Dialysis Patients. *Kidney Int* (2021) 99:1496–8. doi: 10.1016/j.kint.2021.04.006
- Yau K, Abe KT, Naimark D, Oliver MJ, Perl J, Leis JA, et al. Evaluation of the SARS-CoV-2 Antibody Response to the BNT162b2 Vaccine in Patients Undergoing Hemodialysis. JAMA Netw Open (2021) 4:e2123622. doi: 10.1001/jamanetworkopen.2021.23622
- 41. Yi SG, Knight RJ, Graviss EA, Moore LW, Nguyen DT, Ghobrial RM, et al. Kidney Transplant Recipients Rarely Show an Early Antibody Response

Following the First COVID-19 Vaccine Administration. *Transplantation* (2021) 105:e72–3. doi: 10.1097/TP.00000000003764

- 42. Zitt E, Davidovic T, Schimpf J, Abbassi-Nik A, Mutschlechner B, Ulmer H, et al. The Safety and Immunogenicity of the mRNA-BNT162b2 SARS-CoV-2 Vaccine in Hemodialysis Patients. *Front Immunol* (2021) 12:704773. doi: 10.3389/fimmu.2021.704773
- Rincon-Arevalo H, Choi M, Stefanski AL, Halleck F, Weber U, Szelinski F, et al. Impaired Humoral Immunity to SARS-CoV-2 BNT162b2 Vaccine in Kidney Transplant Recipients and Dialysis Patients. *Sci Immunol* (2021) 6 (60):eabj1031. doi: 10.1126/sciimmunol.abj1031
- 44. Simon B, Rubey H, Treipl A, Gromann M, Hemedi B, Zehetmayer S, et al. Haemodialysis Patients Show a Highly Diminished Antibody Response After COVID-19 mRNA Vaccination Compared to Healthy Controls. *Nephrol Dial Transplant* (2021) 36(9):1709–16. doi: 10.1101/2021.03.26.21254259
- 45. Broseta JJ, Rodriguez-Espinosa D, Rodriguez N, Mosquera MDM, Marcos MA, Egri N, et al. Humoral and Cellular Responses to mRNA-1273 and BNT162b2 SARS-CoV-2 Vaccines Administered to Hemodialysis Patients. *Am J Kidney Dis* (2021) 78(4):571–81. doi: 10.1053/j.ajkd.2021.06.002
- Rodriguez-Espinosa D, Broseta JJ, Maduell F, Bedini JL, Vera M. Humoral Response of the mRNA-1273 SARS-CoV-2 Vaccine in Peritoneal Dialysis Patients. *Kidney Int* (2021) 100:476–7. doi: 10.1016/j.kint. 2021.05.018
- 47. Grupper A, Rabinowich I, Schwartz D, Schwartz IF, Ben-Yehoyada M, Shashar M, et al. Reduced Humoral Response to mRNA SARS-CoV-2 BNT162b2 Vaccine in Kidney Transplant Recipients Without Prior Exposure to the Virus. Am J Transplant (2021) 21:2719–26. doi: 10.1111/ajt.16615
- 48. Kantauskaite M, Muller L, Kolb T, Fischer S, Hillebrandt J, Ivens K, et al. Intensity of Mycophenolate Mofetil Treatment Is Associated With an Impaired Immune Response to SARS-CoV-2 Vaccination in Kidney Transplant Recipients. Am J Transplant (2021). doi: 10.1111/ajt.16851
- Rozen-Zvi B, Yahav D, Agur T, Zingerman B, Ben-Zvi H, Atamna A, et al. Antibody Response to SARS-CoV-2 mRNA Vaccine Among Kidney Transplant Recipients: A Prospective Cohort Study. *Clin Microbiol Infect* (2021) 27:1173.e1. doi: 10.1016/j.cmi.2021.04.028
- Ritter ML, Pirofski L. Mycophenolate Mofetil: Effects on Cellular Immune Subsets, Infectious Complications, and Antimicrobial Activity. *Transpl Infect Dis* (2009) 11:290–7. doi: 10.1111/j.1399-3062.2009.00407.x
- 51. Furer V, Eviatar T, Zisman D, Peleg H, Paran D, Levartovsky D, et al. Immunogenicity and Safety of the BNT162b2 mRNA COVID-19 Vaccine in Adult Patients With Autoimmune Inflammatory Rheumatic Diseases and in the General Population: A Multicentre Study. Ann Rheum Dis (2021) 80:1330–8. doi: 10.1136/annrheumdis-2021-220647
- Polewska K, Tylicki P, Biedunkiewicz B, Rucinska A, Szydlowska A, Kubanek A, et al. Safety and Tolerability of the BNT162b2 mRNA COVID-19 Vaccine in Dialyzed Patients. COViNEPH Project. *Medicina (Kaunas)* (2021) 57 (7):732. doi: 10.3390/medicina57070732

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Beilhack, Monteforte, Frommlet, Gaggl, Strassl and Vychytil. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.