



# Draft Genome Sequence of *Lactobacillus rhamnosus* NCB 441, Isolated from Egyptian White Domiati Cheese

Madison A. Moore,<sup>a</sup> Hunter D. Whittington,<sup>a</sup>  M. Andrea Azcarate-Peril,<sup>b,c</sup>  José M. Bruno-Bárcena<sup>a</sup>

<sup>a</sup>Department of Plant and Microbial Biology, North Carolina State University, Raleigh, North Carolina, USA

<sup>b</sup>Department of Medicine, Division of Gastroenterology and Hepatology, School of Medicine, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA

<sup>c</sup>UNC Microbiome Core, Center for Gastrointestinal Biology and Disease, School of Medicine, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA

**ABSTRACT** Here, we report the draft genome sequence of *Lactobacillus rhamnosus* NCB 441, which was isolated from pickled white cheese samples gathered at Farafra Oasis in New Valley Governorate, Egypt. The genome size is 2,969,245 bp with a G+C content of 46.7%.

Lactic acid bacteria inhabit a wide variety of ecological niches in addition to being members of a phylogenetically diverse group of organisms (1, 2). Of those, *Lactobacillus* is one of the more diverse genera and contains multiple commercially exploited *Lactobacillus rhamnosus* strains. Here, we report the draft genome sequence of *L. rhamnosus* NCB 441, which was isolated from pickled white cheese (Egyptian white Domiati cheese) samples gathered at Farafra Oasis in New Valley Governorate, Egypt (27.0567°N, 27.9703°E), where daily ambient temperatures reach upwards of 40°C (104°F).

Five samples of Domiati cheese were used to isolate lactic acid bacteria. Twenty-five grams of each cheese sample was homogenized with 225 ml of 0.85% NaCl, plated onto MRS agar, and incubated anaerobically at 30°C for 48 h for colony selection. Isolated colonies were inoculated into MRS broth and cultivated at 30°C prior to genomic DNA extraction. Extraction was carried out according to the method described by Hoffman and Winston (3). Briefly, cells were harvested in the early logarithmic stage of growth at an optical density at 600 nm ( $OD_{600}$ ) of approximately 0.4 and then were homogenized with a bead beater. The DNA was subjected to chloroform extraction and ethanol precipitation prior to library preparation. The sequencing library was produced using the 454 FLX Titanium rapid library kit according to the manufacturer's instructions (Roche, Indianapolis, IN, USA). The Microbiome Core at the University of North Carolina at Chapel Hill generated shotgun sequencing data for the strain using a 454 GS FLX Titanium+ system (Roche).

For all software, default parameters were used except where otherwise noted. Sequencing data generated 2,607,325 raw reads which were quality filtered for a minimum 500-bp read length. Reads were assembled *de novo* using Newbler v2.6 (4) to produce 108 contigs with an  $N_{50}$  value of 59,695 bp and a coverage depth of 36 $\times$ . The 108 contigs were subjected to a second round of assembly using SPAdes v3.14.1 (5) to produce 45 contigs with an  $N_{50}$  value of 111,618 bp. The resulting genome assembly of *L. rhamnosus* NCB 441 is 2,969,245 bp, with a G+C content of 46.7%. Genome completeness was assessed using CheckM v1.0.18 (6) and was determined to be 98.91%.

Species assignment was based on the average nucleotide identity (ANI) (7) of NCB 441 being over 97% with respect to all 192 *L. rhamnosus* species sequenced and deposited in the NCBI GenBank database to date. All genomes were downloaded using the command line package Pyani (8) and later were compared to NCB 441 using the

**Citation** Moore MA, Whittington HD, Azcarate-Peril MA, Bruno-Bárcena JM. 2020. Draft genome sequence of *Lactobacillus rhamnosus* NCB 441, isolated from Egyptian white Domiati cheese. *Microbiol Resour Announc* 9:e01191-20. <https://doi.org/10.1128/MRA.01191-20>.

**Editor** Kenneth M. Stedman, Portland State University

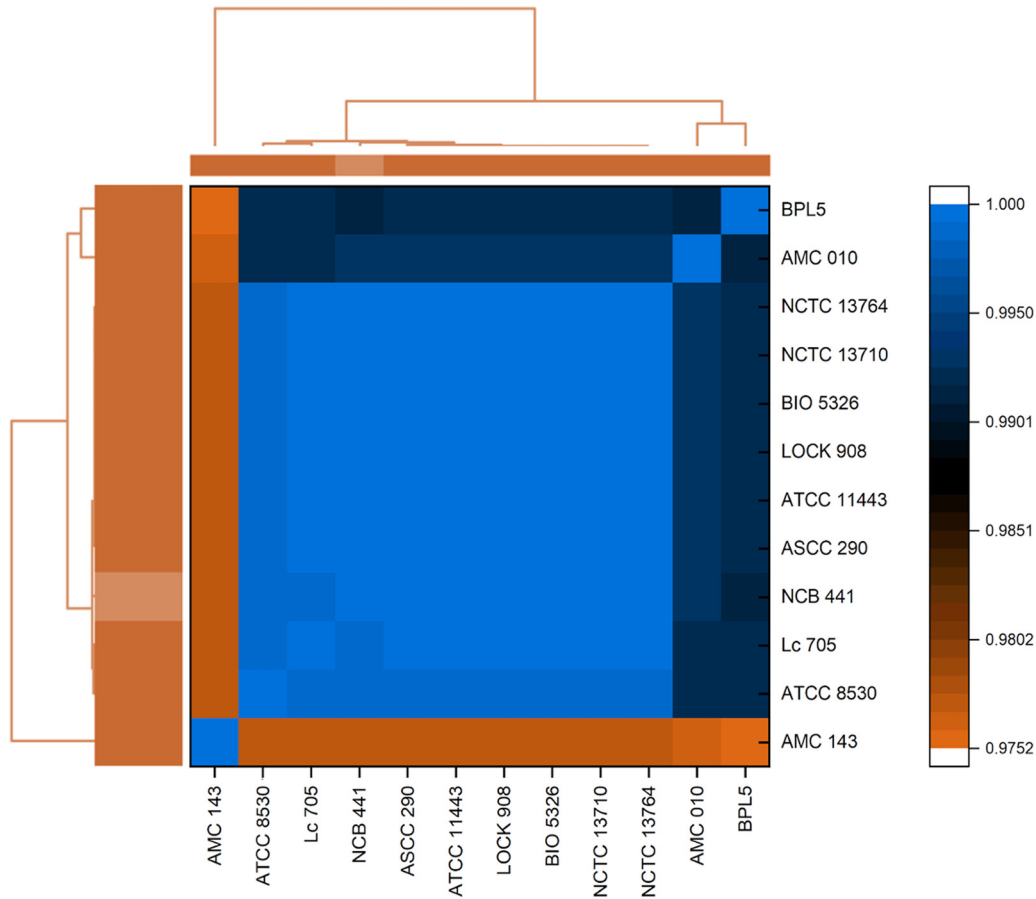
**Copyright** © 2020 Moore et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to José M. Bruno-Bárcena, [jbbarcen@ncsu.edu](mailto:jbbarcen@ncsu.edu).

**Received** 20 October 2020

**Accepted** 5 November 2020

**Published** 3 December 2020



**FIG 1** Heatmap display of ANI values for 12 *Lactobacillus rhamnosus* strains. The heatmap was generated with Origin v2018b using ANI values for 12 of 192 *L. rhamnosus* strains, including ATCC 8530 (GenBank accession no. [CP003094.1](#)), Lc 705 ([FM179323.1](#)), ASCC 290 ([CP014645.1](#)), ATCC 11443 ([CP022109.1](#)), LOCK908 ([CP005485.1](#)), BIO5326 ([CP046267.1](#)), NCTC13710 ([LR134322.1](#)), NCTC13764 ([LR134331.1](#)), BPL5 ([LT220504.1](#)), AMC010 ([MSTC00000000.1](#)), and AMC143 ([MSTB00000000.1](#)), for which sequences were obtained from NCBI. Values range from 0.9752 (97.52%) to 1 (100% ANI); orange represents <98.51% ANI, while blue represents highly similar strains exhibiting ANI values of >99%. Dendrograms link ANI percentages to construct a hierarchical clustering inferring phylogeny, thus confirming NCB 441 as a *L. rhamnosus* strain.

MUMmer nucmer algorithm to calculate ANI values (9). A condensed heatmap was generated using Origin v2018b (OriginLab Corp., Northampton, MA) (Fig. 1) with only the top ANI values from nine fully sequenced *L. rhamnosus* genomes (ATCC 8530, Lc 705, ASCC 290, ATCC 11443, LOCK908, BIO5326, NCTC13710, NCTC13764, and BPL5), as well as two strains previously sequenced by our research group (AMC010 and AMC143) (10, 11). ANI values indicate over 97% nucleotide identity to strains BPL5, AMC010, and AMC143, as well as over 99% nucleotide identity to all other strains, which provides evidence for NCB 441 to be classified as a *L. rhamnosus* strain.

**Data availability.** The genome sequence of *L. rhamnosus* NCB 441 has been deposited in DDBJ/EMBL/GenBank under the accession no. [JACSDP00000000](#). The version described in this paper is the first version. Raw sequencing data have been deposited with SRA accession no. [SRR12515116](#).

#### ACKNOWLEDGMENTS

This project was supported by the North Carolina State University Department of Plant and Microbial Biology. The UNC Microbiome Core is supported in part by project P30 DK034987 (Center for Gastrointestinal Biology and Disease) and project P30 DK056350 (UNC Nutrition Obesity Research Center). M.A.M. is the recipient of a

Genetics and Genomics Provost's doctoral research fellowship. H.D.W. is supported by the NC State Graduate Student Support Plan.

## REFERENCES

1. Ceapa C, Davids M, Ritari J, Lambert J, Wels M, Douillard FP, Smokvina T, de Vos WM, Knol J, Kleerebezem M. 2016. The variable regions of *Lactobacillus rhamnosus* genomes reveal the dynamic evolution of metabolic and host-adaptation repertoires. *Genome Biol Evol* 8:1889–1905. <https://doi.org/10.1093/gbe/evw123>.
2. Douillard FP, Ribbera A, Kant R, Pietila TE, Jarvinen HM, Messing M, Randazzo CL, Paulin L, Laine P, Ritari J, Caggia C, Lahtinen T, Brouns SJ, Satokari R, von Ossowski I, Reunanen J, Palva A, de Vos WM. 2013. Comparative genomic and functional analysis of 100 *Lactobacillus rhamnosus* strains and their comparison with strain GG. *PLoS Genet* 9:e1003683. <https://doi.org/10.1371/journal.pgen.1003683>.
3. Hoffman CS, Winston F. 1987. A ten-minute DNA preparation from yeast efficiently releases autonomous plasmids for transformation of *Escherichia coli*. *Gene* 57:267–272. [https://doi.org/10.1016/0378-1119\(87\)90131-4](https://doi.org/10.1016/0378-1119(87)90131-4).
4. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Irzyk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. *Nature* 437:376–380. <https://doi.org/10.1038/nature03959>.
5. Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Pribelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20:714–737. <https://doi.org/10.1089/cmb.2013.0084>.
6. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
7. Richter M, Rossello-Mora R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.
8. Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. *Anal Methods* 8:12–24. <https://doi.org/10.1039/C5AY02550H>.
9. Delcher AL, Phillippy A, Carlton J, Salzberg SL. 2002. Fast algorithms for large-scale genome alignment and comparison. *Nucleic Acids Res* 30:2478–2483. <https://doi.org/10.1093/nar/30.11.2478>.
10. Arnold JW, Monteagudo-Mera A, Altermann E, Cadenas MB, Thompson AL, Azcarate-Peril MA. 2017. Genome sequences of potential probiotic *Lactobacillus rhamnosus* isolates from human infants. *Genome Announc* 5:e00107-17. <https://doi.org/10.1128/genomeA.00107-17>.
11. Arnold JW, Simpson JB, Roach J, Bruno-Barcena JM, Azcarate-Peril MA. 2018. Prebiotics for lactose intolerance: variability in galacto-oligosaccharide utilization by intestinal *Lactobacillus rhamnosus*. *Nutrients* 10:1517. <https://doi.org/10.3390/nu10101517>.