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Data Article

Salivary lipocalin family proteins from *Panstrongylus chinai*, a vector of Chagas disease

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ARTICLE INFO

Article history:

Received 24 August 2017

Received in revised form

7 September 2017

Accepted 19 September 2017

Available online 22 September 2017

Keywords:

Panstrongylus chinai

Saliva

Lipocalin

Transcriptome

ABSTRACT

The dataset in this report is related to the research article with the title: “Salivary gland transcripts of the kissing bug, *Panstrongylus chinai*, a vector of Chagas disease” (Kato et al., 2017) [1]. Lipocalin family proteins were identified as the dominant component in *P. chinai* saliva, and phylogenetic analysis of the salivary lipocalins resulted in the formation of five major clades. For further characterization, each clade of *P. chinai* lipocalin was s alignment and phylogenetic analyses together with homologous triatomine lipocalins; pallidipin 2, an inhibitor of collagen-induced platelet aggregation identified from saliva of *Triatoma pallidipennis* (clade I), pallidipin-like salivary lipocalin from *Triatoma dimidiata* (clade II), salivary lipocalin from *T. dimidiata* (clade III), triatin-like salivary lipocalin identified in the saliva of *T. dimidiata* (clade IV), and lipocalin-like TiLipo37 from *Triatoma infestans* (clade V).

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DOI of original article: <http://dx.doi.org/10.1016/j.actatropica.2017.06.022>

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<http://dx.doi.org/10.1016/j.dib.2017.09.039>

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Specifications Table

Subject area	Biology
More specific subject area	Salivary lipocalins of a hematophagous insect
Type of data	Table, figure
How data was acquired	Transcriptome, alignment and phylogenetic analyses
Data format	Analyzed
Experimental factors	A dataset of transcripts from salivary glands of <i>Panstrongylus chinai</i>
Experimental features	Alignment and phylogenetic analyses of salivary lipocalins from <i>Panstrongylus chinai</i>
Data source location	Ecuador
Data accessibility	Accession numbers of the sequence data are available in the reference [1].

Value of the data

- The data is the second report of the salivary lipocalins in a *Panstrongylus* species.
- The result will provide further information into the salivary biochemical and pharmacological complexity of triatomine bugs and the evolution of salivary components in blood sucking arthropods.
- cDNAs and recombinant proteins prepared from these transcripts will result in the discovery of novel pharmacologically active compounds, as well as the development of biomarkers following exposure to *Panstrongylus chinai*.

1. Data

The dominant transcripts of *Panstrongylus chinai* salivary glands were analyzed by sequence analysis of the cDNA library, and 73.7% of transcripts encoding the putative secreted proteins coded for the lipocalin family of proteins [1]. Table 1 shows the grouping of transcripts coding for lipocalin family proteins in *P. chinai* salivary glands obtained by the phylogenetic analysis [1]. Figs. 1–5 represent alignment and phylogenetic analyses of each clade of *P. chinai* salivary lipocalins together with homologous proteins; pallidipin 2, a platelet aggregation inhibitor identified from *Triatoma pallidipennis* saliva (clade I), pallidipin-like salivary lipocalin from *Triatoma dimidiata* saliva with unknown function (clade II), salivary lipocalin from *T. dimidiata* saliva with unknown function (clade III), triatin-like salivary lipocalin identified in the saliva of *T. dimidiata* with unknown function (clade IV), and lipocalin-like TiLipo37 from *Triatoma infestans* saliva with unknown function (clade V), showing their structural similarity and diversity.

2. Experimental design, materials and methods

The sequences of *P. chinai* salivary lipocalins were obtained in the study “Salivary gland transcripts of the kissing bug, *Panstrongylus chinai*, a vector of Chagas disease” [1]. The sequences coding for lipocalin family of proteins by BLASTx analyses were aligned with CLUSTAL W software [2] and examined using Molecular Evolutionary Genetics Analysis (MEGA) version 5.2 [3]. Phylogenetic trees by the neighbor-joining method were constructed with the distance algorithms available in the MEGA package. Bootstrap values were determined on 1000 replicates of the data sets. Accession numbers of the sequence data are available in “Salivary gland transcripts of the kissing bug, *Panstrongylus chinai*, a vector of Chagas disease” [1].

Table 1
Transcripts coding for lipocalin family proteins in *Panstrongylus chinai* salivary glands.

Clade	Similar to	No. of clusters	No. of seq	% seq
Clade I	pallidipin 2 (<i>Triatoma pallidipennis</i>): AAA30329	16	151	46.9
	Td42, similar to pallidipin 2 (<i>Triatoma dimidiata</i>): BAI50842	1	1	0.3
Clade II	Td38, similar to pallidipin-like salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50839	2	5	1.6
	Td33, similar to pallidipin-like salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50837	1	1	0.3
Clade III	Td26, similar to salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50831	2	11	3.4
	salivary lipocalin (<i>Triatoma infestans</i>): ABR27920	1	8	2.4
	Td40, similar to triabin-like lipocalin 4a precursor (<i>Triatoma dimidiata</i>): BAI50840	1	3	0.9
	triabin-like lipocalin 4a precursor (<i>Triatoma infestans</i>): ABR27959	1	2	0.7
Clade IV	salivary lipocalin (<i>Triatoma infestans</i>): ABR27868	1	1	0.3
	Td18, similar to triatin-like salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50824	4	66	20.5
	Td11, similar to triatin-like salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50818	1	43	13.3
	Td45, similar to pallidipin-like salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50844	1	8	2.5
Clade V	lipocalin-like TiLipo37 (<i>Triatoma infestans</i>): AAQ68063	2	10	3.1
	salivary lipocalin (<i>Triatoma brasiliensis</i>): ABH09436	1	6	1.9
Others	Td24, similar to salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50829	2	2	0.7
	triabin-like salivary lipocalin (<i>Triatoma infestans</i>): ABR27927	1	1	0.3
	salivary lipocalin (<i>Triatoma infestans</i>): ABR27831	1	1	0.3
	salivary lipocalin 1 (<i>Triatoma brasiliensis</i>): ABH09421	1	1	0.3
	Td23, similar to salivary lipocalin (<i>Triatoma dimidiata</i>): BAI50828	1	1	0.3
Total		41	322	100.0

A

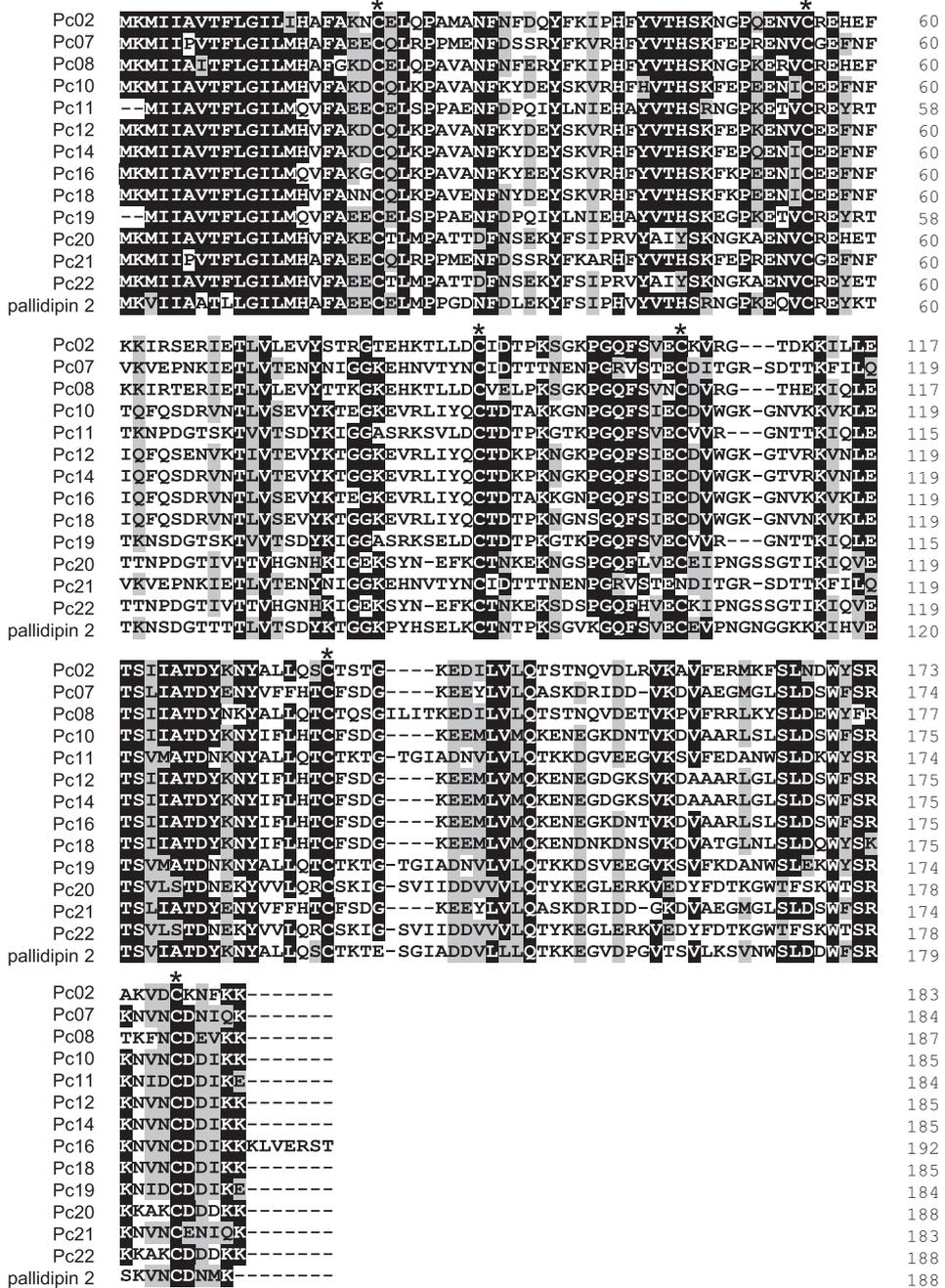


Fig. 1. (A) Sequence alignment of pallidipin 2-like proteins from *Panstrongylus chinai* (Pc02, Pc07, Pc08, Pc10–Pc12, Pc14, Pc16 and Pc18–Pc22) together with pallidipin 2 from *Triatoma pallidipennis* (accession number: AAA30329). Black-shaded amino acids represent identical amino acids and gray-shaded amino acids represent conserved amino acids. Dashes indicate gaps introduced for maximal alignment. Asterisks at the top of the amino acids denote conserved cysteine residues. (B) Phylogenetic tree analysis of pallidipin 2-like proteins (Pc02, Pc07, Pc08, Pc10–Pc12, Pc14, Pc16 and Pc18–Pc22) from *P. chinai* with pallidipin 2 from *T. pallidipennis*. The numbers in parentheses indicate the number of transcripts of each contig. The scale bar represents 0.1% divergence. Bootstrap values are shown above branches.

B

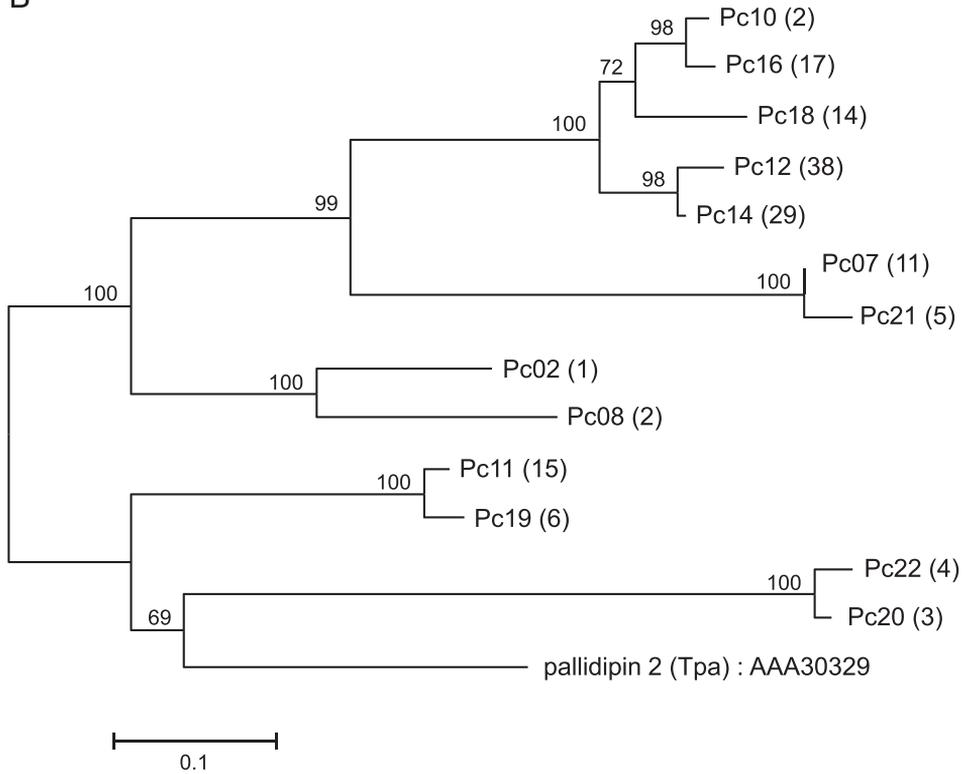


Fig. 1. (continued)

Pc85	MKTIILAVTFI GILTYA CDEKSSERGPTE C NVTAMANFDSEKYLKFFPAYT THSK -YKG-I	58
Pc156	MKTIILAVAF TAILTYA FAQKPSQS--NRCKHKAMONFDS T KYLQI PRAYATHSK DEI PYL	58
Td38	MNTIIL ALTFI GILTYAFAE EP CDK--TE CK HEP MANFNPT KYLQ MTPLYSTHAK -YSSNI	57
Pc85	T VCRL F KSAT K SCDTADTTII G YY Q LS D V G GIQ NY EMV C ST S EEFYK G Q F TE C KT V RD	118
Pc156	T VC G VLETT Q RS D STVD T LMY G YY Q DS--G K TH Y SE A QC N T K VES V KT G F Y ED C IV I ND	116
Td38	T VC R V F TS I T I SCDA V NI N I H GY N KS--G N I Y D Y EM F C N T S FD K AE Y L A D C K I V K D	115
Pc85	SL F CD L R N SS--LS I K L Y M SV D TD Y EN Y A I RY N C L KE E K--G I LD N Y E V L Q K N P T A CG E IN	175
Pc156	TL F ED P K I Y K D I V H K V Y V SV I DT D Y E NY S IL Y R C LE D EV D CG F Q D TI L V L K I D K ---D A ND	173
Td38	AQ F ED G R I P--EP F K L Y M SV I DT D Y E NY A IL L Y L C I ND E Y--G L ED N F E V L Q K N P ---G T SN	169
Pc85	EL I KE A L N K R C E N F DE F IS-M N T Y C Q K H -D I S---	207
Pc156	DL V KE A L G K V G M D L Q S EN P Y T N N T Y C D D N K N I K S K N	209
Td38	E A V S T A L K N N E M D L E K F F -R D Y T Y C Q S G-N K D E E Q	203

Fig. 2. Sequence alignment of Td38-like proteins from *Panstrongylus chinai* (Pc85 and Pc156) together with Td38 from *Triatoma dimidiata* (accession number: AB470389). Black-shaded amino acids represent identical amino acids and gray-shaded amino acids represent conserved amino acids. Dashes indicate gaps introduced for maximal alignment. Asterisks at the top of the amino acids denote conserved cysteine residues.

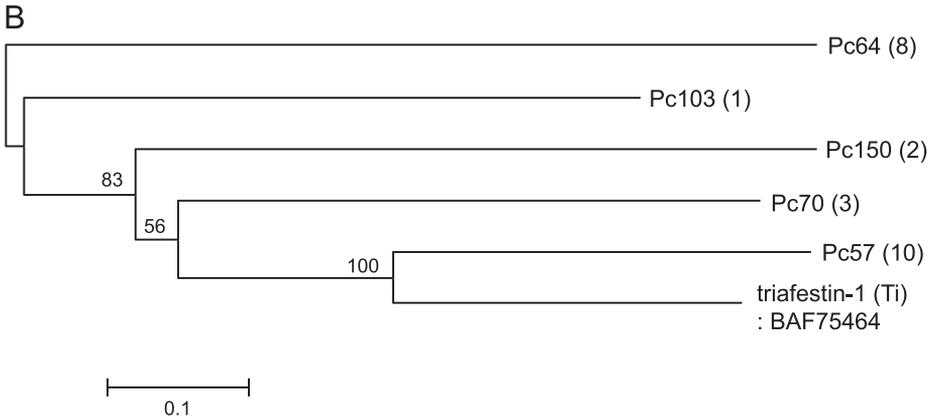
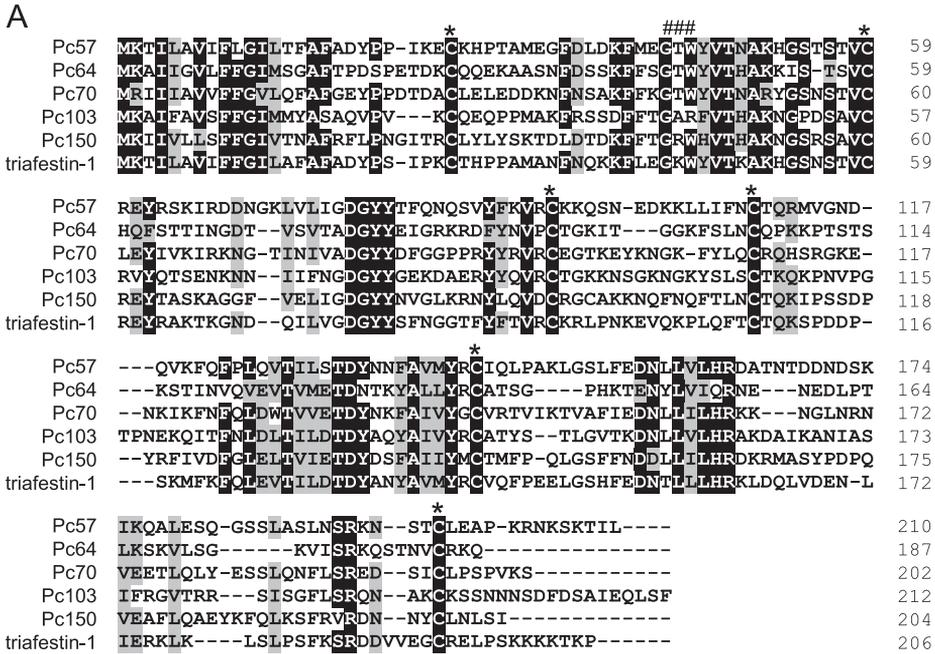


Fig. 3. (A) Sequence alignment of triaefestin-1-like proteins from *Panstrongylus chinai* (Pc57, Pc64, Pc70, Pc103 and Pc150) together with triaefestin-1 from *Triatoma infestans* (accession number: BAF75464). Black-shaded amino acids represent identical amino acids and gray-shaded amino acids represent conserved amino acids. Dashes indicate gaps introduced for maximal alignment. Asterisks at the top of the amino acids denote conserved cysteine residues, and the GXW motif is indicated by ###. (B) Phylogenetic tree analysis of triaefestin-1-like proteins from *P. chinai* (Pc57, Pc64, Pc70, Pc103 and Pc150) with triaefestin-1 from *T. infestans*. The numbers in parentheses indicate the number of transcripts of each contig. The scale bar represents 0.1% divergence. Bootstrap values are shown above branches.

A

Pc13	MKTLTAVTFFGILTYAFAEELKYGGDACCIDNVDCLVNLDPEKFFTGTWYLTHATPSRV	60
Pc15	MKTLTAVTFFGILTYAFAEELKYGGDACCIDNVDCLVNLDPEKFFTGTWYLTHATPSARV	60
Pc58	MKTLTAVTFFGILTYTSAETIOYCKGSCQT--VKCMKNFDAAOFFSGPWSLTHATPSARV	58
Pc59	MKTLTAVTFFGILTYTSAETIOYCKGSCQT--VKCMKNFDAAOFFSGPWSLTHATPSARV	58
Td18	MKTLTAVAFFGILTYAFADTVVYKVKVCOIDNNDGCVNLDPKRFESGTWYLTHATQSTRV	60
Pc13	TKSTICRDYAPTL-TNGK-VDVITYSYNENGASGNRYMFCNGAKNEKRDIFNFICKSEN	118
Pc15	TKSTICRDYAATL-TNGQ-VDVITYSYNENGASGNRYMDFKNGVKNKREDIFNFICKSEN	118
Pc58	TKSTICRDYALSTGNDGK-LFGTYGYENGASGNYYDIHCSGSKSTTKEDLYGFDCELTN	117
Pc59	TKSTICRDYALSTGNDGK-LFGTYGYENGASGNYYDIHCSGSKSTTKEDLYGFDCELTN	117
Td18	TLSTICRDFEPKLNANGKTLGVITYGYENGCTDNRMEVSCSGTQNTTRRDI FNFDCSSNN	120
Pc13	ERGETASFQVDVSEFLATDYTKYGVLYRCVKIDTFEDNVFLIFRQKEPPDENKVEELKHHY	178
Pc15	GRGENASFQVDVSEFLATDYTKYGVLYRCVKIDTFEDNVFLIFRQKEPPDENKVEELKHHY	178
Pc58	ARGEKTHTHIDAYFLATDYTNGLIYRCVKSDTQFEDNVFVIYRNKNPSDDKVKELLSPI	177
Pc59	DRGEKTHTHIDVYFLATDYKNYGLIYRCVKSDTQFEDNVFVIYRNKNPSDDKVKELLSPI	177
Td18	GRGETTNFHDGSEFLATDYDSYGVLYRCVTTGTLTEDNVFLIHRQKNPSDDEVKNNLKHV	180
Pc13	GLN--SLISRKEATCKI-	193
Pc15	GLN--SLISRKEANCKN-	193
Pc58	GLDLGKFI SRKEATCTTK	195
Pc59	GLDLGKFI SRKEATCTTK	195
Td18	GLDLKKFISRQKATCSN-	197

B

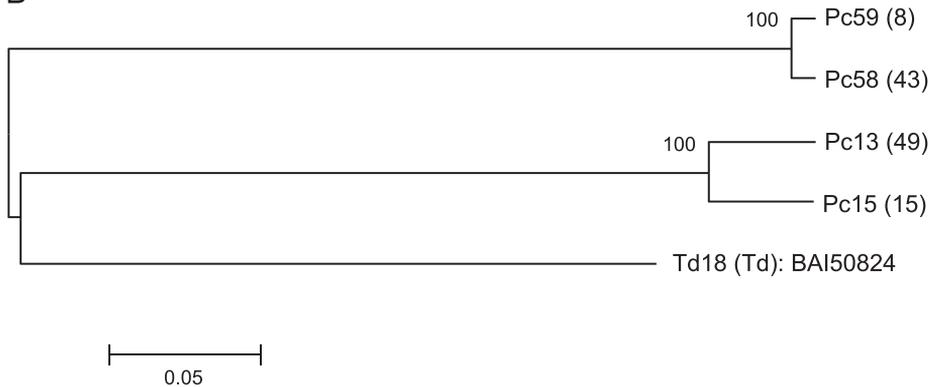
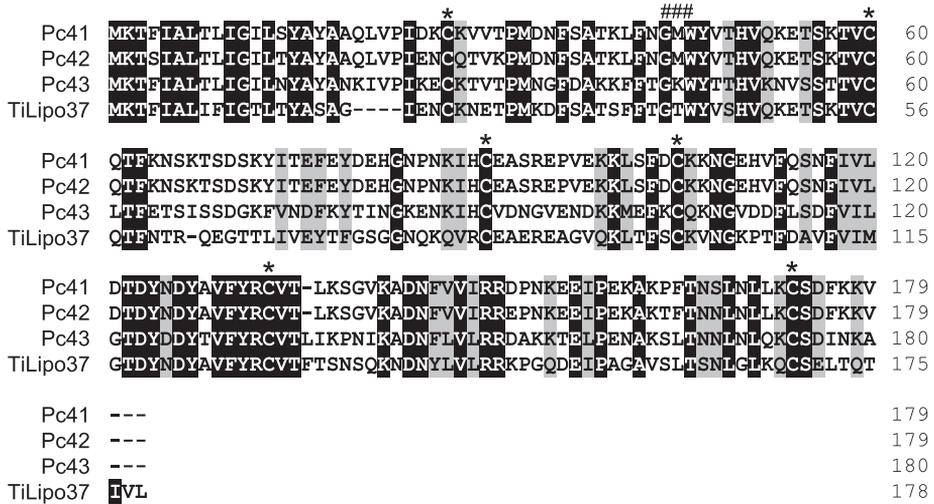


Fig. 4. (A) Sequence alignment of Td18-like proteins from *Panstrongylus chinai* (Pc13, Pc15, Pc58 and Pc59) together with Td18 from *Triatoma dimidiata* (accession number: BAI50824). Black-shaded amino acids represent identical amino acids and gray-shaded amino acids represent conserved amino acids. Dashes indicate gaps introduced for maximal alignment. Asterisks at the top of the amino acids denote conserved cysteine residues, and the GXW motif is indicated by ###. (B) Phylogenetic tree analysis of Td18-like proteins from *P. chinai* (Pc13, Pc15, Pc58 and Pc59) together with Td18 from *T. dimidiata*. The numbers in parentheses indicate the number of transcripts of each contig. The scale bar represents 0.05% divergence. Bootstrap values are shown above branches.

A



B

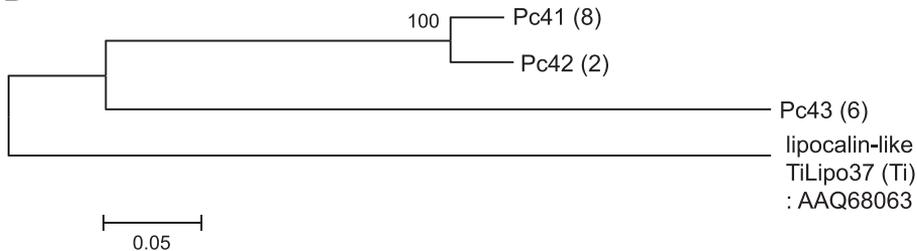


Fig. 5. (A) Sequence alignment of lipocalin-like TiLipo37-like proteins from *Panstrongylus chinai* (Pc41–Pc43) together with lipocalin-like TiLipo37 from *Triatoma infestans* (accession number: AAQ68063). Black-shaded amino acids represent identical amino acids and gray-shaded amino acids represent conserved amino acids. Dashes indicate gaps introduced for maximal alignment. Asterisks at the top of the amino acids denote conserved cysteine residues, and the GXW motif is indicated by ###. (B) Phylogenetic tree analysis of lipocalin-like TiLipo37-like proteins from *P. chinai* (Pc41–Pc43) together with lipocalin-like TiLipo37 from *T. infestans*. The numbers in parentheses indicate the number of transcripts of each contig. The scale bar represents 0.05% divergence. Bootstrap values are shown above branches.

Acknowledgements

We are grateful to Dr. José M. C. Ribeiro (Vector Biology Section, Laboratory of Malaria and Vector Research, NIAID, NIH, USA) for the development and training of all custom bioinformatics programs used in this research. This study was supported by the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan (Grant nos. 25257501 and 15H04588).

Transparency document. Supplementary material

Transparency document associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2017.09.039>.

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