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α -SNAP inhibits AMPK signaling to reduce mitochondrial biogenesis and dephosphorylates Thr172 in AMPK α *in vitro*

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Abstract

The AMP-activated protein kinase (AMPK) regulates metabolism in normal and pathological conditions and responds to nutrients, hormones, anti-diabetic drugs and physical exercise. AMPK is activated by kinase LKB1 and inactivated by phosphatases whose identities remain uncertain. Here we show that AMPK associates with α -SNAP, an adapter that enables disassembly of cis-SNARE complexes formed during membrane fusion. Knockdown of α -SNAP activates AMPK to phosphorylate its endogenous substrates ACC and Raptor and provokes mitochondrial biogenesis. AMPK phosphorylation is rescued from α -SNAP RNAi by LKB1 knockdown or expression of wild type but not mutated α -SNAP. Recombinant wild type but not mutated α -SNAP dephosphorylates pThr172 in AMPK α *in vitro*. Over-expression of wild-type but not mutated α -SNAP prevents AMPK activation in cells treated with agents to elevate AMP concentration. The mouse α -SNAP mutant *hyh* (*hydrocephalus with hop gait*) shows enhanced binding and inhibition of AMPK. By negatively controlling AMPK, α -SNAP therefore potentially coordinates membrane trafficking and metabolism.

Introduction

Cellular energy in the form of ATP drives biosynthetic pathways and supports essential processes such as ion transport, cytoskeletal integrity and membrane trafficking^{1,2}. Homeostasis requires throttling these demands for energy while regenerating ATP, predominantly by mitochondrial respiration. Central to this balancing act is the 5'-AMP-activated protein kinase (AMPK), an enzyme comprised of catalytic α subunit plus regulatory β and γ subunits, the latter that alternatively binds ATP, ADP or AMP to gauge metabolic status^{1,2}. In terms of animal physiology AMPK is a critical node of regulation, for it controls fat and carbohydrate metabolism, and responds to nutrients, hormones, drugs and physical exercise in both normal and pathological conditions³. The rise in ratio of [AMP+ADP] vs. [ATP] due to energy expenditure or inhibition of respiration activates

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Contributions

D.L.B and L.W. designed the study; L.W. performed experiments; D.L.B and L.W. wrote the paper.

Competing financial interests

The authors declare no competing financial interests.

AMPK predominantly by phosphorylation of T172 in the kinase activation loop in the α subunit⁴. The pT172 levels depend on a balance between the primary AMPK activating kinase LKB1^{5,6} and dephosphorylation by protein phosphatases⁷. Dephosphorylation of pT172 is affected by AMP and ADP^{8,9}, however, the identity of the physiologically relevant AMPK phosphatase(s) remains unclear.

Here we discovered that inactive AMPK bound to ATP stably associates with the protein α -SNAP that acts as a negative regulator of the kinase. Knockdown of α -SNAP resulted in enhanced phosphorylation of T172 in AMPK, phosphorylation of AMPK substrates, and increased mitochondrial biogenesis and respiration. Effects of α -SNAP knockdown were rescued by over expression of α -SNAP and complemented by knockdown of LKB1, showing the effects were due to T172 phosphorylation. Furthermore, recombinant α -SNAP dephosphorylated AMPK *in vitro*, even though the sequence and structure of α -SNAP does not resemble any known protein phosphatase. Mutations in α -SNAP are shown to provide either loss-of-function or gain-of-function phenotypes relative to AMPK inactivation. Multiple actions of α -SNAP provide a connection between membrane fusion events and metabolism required to fuel these processes.

Results

Binding of AMPK to α -SNAP is responsive to ATP

To explore regulation of AMPK we immunoprecipitated FLAG-tagged AMPK- γ 1 from stably transfected HEK293 cells. Silver staining after SDS-PAGE revealed the AMPK γ subunit co-precipitated endogenous α and β subunits, with or without oligomycin addition to the cells to activate AMPK (Fig. 1a). In addition, a 35 kDa protein associated with inactive (lane 2), but not active AMPK (lane 3). This band was excised and LC-MS/MS identified it as α -SNAP (soluble NSF (*N-ethylmaleimide sensitive factor*) attachment protein; *a.k.a.* NAPA; NSF attachment protein, alpha). For many years α -SNAP has been studied as factor required for activation of the NSF ATPase for resolution of cis-SNARE (SNAP receptor) complexes formed during membrane fusion¹⁰. The activity of NSF allows repeated rounds of membrane fusion, therefore α -SNAP regulates a critical limiting step in intracellular membrane trafficking and exocytosis¹¹. We confirmed the association of α -SNAP with AMPK by immunoprecipitation of endogenous α -SNAP with endogenous AMPK that was recovered from unstressed cells (Fig. 1b). Addition of 2-deoxyglucose or phenformin to metabolically stress cells and elevate [AMP] activated AMPK, evident from the increase in phosphorylation of T172 in AMPK α and robust phosphorylation of S79 in the endogenous AMPK substrate, Acetyl CoA Carboxylase (ACC). Activation of AMPK substantially decreased its co-immunoprecipitation with endogenous α -SNAP (Fig. 1b). This reduced binding to AMPK in cells with elevated [AMP] indicated that association with α -SNAP did not depend on the phosphorylation of T172 in AMPK, but was sensitive to the conformation of the kinase. Co-expression of HA- α -SNAP with FLAG-AMPK $\alpha\beta\gamma$ allowed reciprocal anti-HA or anti-FLAG co-precipitation of the proteins (Fig. 1c.), even though artificial binding can occur due to overexpression of proteins. We think that α -SNAP contacts AMPK at multiple sites, in different subunits of the heterotrimer, just as α -SNAP simultaneously binds both NSF and SNARE proteins¹¹.

Purified, recombinant α -SNAP bound directly to FLAG-AMPK γ in an *in vitro* assay and binding was enhanced by addition of ATP (Fig. 1d). AMPK is allosterically regulated by binding of either ATP or AMP/ADP to the γ subunit, at sites formed by four CBS (cystathionine beta synthase) domains³. The assay revealed a two-fold increase in binding to AMPK γ in the presence of ATP or non-hydrolyzable derivatives of ATP. In contrast, there was no ATP-dependent increase in α -SNAP binding with FLAG-AMPK γ R299G (equivalent to R531G in γ 2), a single residue substitution in the CBS4 domain that is linked to human Wolf-Parkinson-White cardiomyopathy and known to affect nucleotide binding¹². We concluded that α -SNAP preferentially interacts with the ATP-bound conformation of the γ subunit, which would account for the differential co-immunoprecipitation from cells, depending on metabolic stress-induced increases in [AMP].

AMPK phosphorylation at T172 is negatively regulated by α -SNAP

We knocked down α -SNAP in cells by RNAi using two different sequences of short hairpin RNAs (shRNA) and examined effects on AMPK phosphorylation and signaling (Figure 2). Both of these shRNA effectively reduced α -SNAP to low levels and resulted in a significant 4 to 5-fold increase in phosphorylation of T172 in AMPK, relative to controls expressing shRNA for a sequence in GFP (Fig. 2a). Knockdown of α -SNAP significantly increased phosphorylation of S79 in ACC and S792 in Raptor, an essential subunit of the mTORC1 complex, showing activation of AMPK led to phosphorylation of its endogenous substrates to suppress anabolic metabolism. Phosphorylation of Raptor S792 negatively regulates mTORC1 activity¹³, and indeed phosphorylation of T389 in the mTORC1 substrate p70S6K was reduced by knockdown of α -SNAP (Fig. 2a). As an additional control we used RNAi to knock down NSF, the known partner of α -SNAP. Knockdown of NSF did not affect the basal phosphorylation of AMPK α , activation of AMPK by 2-deoxyglucose, or AMPK signaling to endogenous ACC, Raptor and p70S6K (Fig. 2a). Therefore, activation of AMPK signaling in response to knockdown of α -SNAP was not secondary to NSF-dependent actions in cells. Activation of AMPK due to knockdown of α -SNAP was not secondary to elevation of the [AMP+ADP]/[ATP] nucleotide ratio. Knockdown of α -SNAP actually reduced the cellular ratio of [AMP+ADP]/[ATP] (Supplementary Table S1). On the other hand, over-expression of α -SNAP significantly blunted the activation of AMPK by metabolic stress induced by oligomycin, phenformin, or 2-deoxyglucose relative to controls, and reduced AMPK-dependent signaling to ACC and Raptor (Fig. 2b). These results showed α -SNAP imposes a major inhibitory restraint on AMPK activation and anabolic signaling in living cells.

We observed similar effects of α -SNAP knockdown on AMPK phosphorylation and signaling in a second cell line, Madin-Darby Canine Kidney (MDCK) epithelial cells. Knockdown of α -SNAP using canine shRNA elevated the level of pT172 in AMPK, and increased phosphorylation of ACC at S79 in both unstimulated cells and cells treated with 2-deoxyglucose (Fig. 2c). The increased phosphorylation of AMPK and ACC due to knockdown of α -SNAP was reduced to levels in control cells by stable expression of human α -SNAP that was not targeted by canine shRNA (Fig. 2c). Furthermore, increased phosphorylation of AMPK and ACC in response to α -SNAP knockdown was suppressed by the coincident knockdown of LKB1 using either of two independent shRNA sequences (Fig.

2d). The requirement for the AMPK activating kinase LKB1 for activation of AMPK showed that α -SNAP was controlling phosphorylation of T172.

Knockdown of α -SNAP enhances oxygen consumption and mitochondrial biogenesis

To gauge the physiological and metabolic effects of α -SNAP knockdown we used the Seahorse analyzer to measure the cellular oxygen consumption rate (OCR). Knockdown of α -SNAP significantly elevated the OCR over the control (Fig. 3a), and LKB1 knockdown itself did not alter the OCR but did prevent the increase in OCR due to α -SNAP knockdown, showing the responses involved regulation of AMPK. As controls, in time course experiments, the addition of complex V inhibitor oligomycin reduced and complex I inhibitor rotenone fully extinguished the OCR and conversely, FCCP as an uncoupler of oxidative-phosphorylation increased the OCR (Fig. 3b). In contrast, the extracellular acidification rate, an indicator of anaerobic glycolytic lactate production, did not change significantly in α -SNAP knockdown cells relative to controls (Supplementary Fig. S1a). AMPK is known to induce mitochondrial biogenesis via PGC-1 α and SIRT1¹⁴. We quantified mitochondrial biogenesis by citrate synthase activity and quantitative immunoblotting for mitochondrial proteins such as ATP synthase, complex III and cytochrome oxidase subunit IV. Knockdown α -SNAP by either of two shRNA activated AMPK and significantly increased mitochondrial biogenesis (Fig. 3c, 3d and Supplementary Fig. S1b). The increase in mitochondrial content was suppressed by knockdown of LKB1, by either of two independent shRNA (Fig. 3e and Supplementary Fig. S1c). This showed that the responses to α -SNAP knockdown depended on the phosphorylation of T172 in AMPK.

α -SNAP dephosphorylates T172 in AMPK *in vitro*

How does α -SNAP reduce AMPK activation? The T172 site in AMPK α is phosphorylated by LKB1, which is thought to be a constitutively active AMPK kinase¹⁵. Therefore, we reasoned that α -SNAP was promoting dephosphorylation of AMPK, either as an adapter recruiting a protein Ser/Thr phosphatase, or alternatively α -SNAP could have phosphatase activity. To test this latter idea we expressed four versions of recombinant α -SNAP in bacteria and purified the proteins to homogeneity (Fig. 4a). These were 1) wild type (WT), 2) the *hyh* mutant M105I, 3) L294A, (substitution of the only strictly conserved residue in the C-terminal of SNAPs), and 4) α -SNAP (1-285) deleted of the C-terminal 10 residues. Previous studies showed the L294A and 1-285 mutants of α -SNAP bound to NSF the same as wild type, but did not activate NSF and therefore acted as dominant negative proteins in cells, blocking resolution of SNARE complexes and exocytosis¹⁶. We found that recombinant WT α -SNAP but not L294A or 1-285 exhibited phosphatase activity with the chemical substrate DiFMUP and pSer/pThr peptides (unpublished). We measured *in vitro* dephosphorylation of AMPK recombinant protein by purified WT α -SNAP (Fig. 4b, c). GST-AMPK α (1-392) was purified from bacteria and ³²P-labeled at T172 with purified CaMKK II for use as substrate. The phosphatase reaction was visualized by the loss of ³²P from the acid precipitated substrate recovered from the assay (Fig. 4b). The release of ³²P from this substrate showed a linear concentration dependence on the amount of α -SNAP added to the assay (Fig. 4c). The specific activity was calculated as 24.9 pmol phosphate released/min/ μ g enzyme, within the range of other known protein phosphatases. In addition,

we assayed the four different forms of recombinant α -SNAP for dephosphorylation of pT172 in AMPK α that was immunoprecipitated from extracts of stimulated cells. Both the WT and M105I α -SNAP efficiently and completely dephosphorylated T172 in AMPK, but neither the L294A nor 1-285 mutant of α -SNAP dephosphorylated AMPK under the same conditions (Fig. 4d). As a test for substrate specificity, we compared pT308 in Akt and pS792 in Raptor as alternative phosphoprotein substrates in the assay with recombinant WT α -SNAP, but these proteins were not dephosphorylated (Fig. 4e). The results showed that α -SNAP could act as a protein phosphatase in biochemical *in vitro* assays to dephosphorylate T172 in AMPK.

Effects of overexpression of WT and α -SNAP mutants L294A and 1-285 in cells

Overexpression of WT α -SNAP reduced basal phosphorylation of T172 in AMPK in unstimulated cells and significantly reduced activation of AMPK and downstream signaling to S79 in ACC in response to addition of 2-deoxyglucose to the cells (Fig. 5a). In contrast, expression of the same levels of the L294A or 1-285 forms of α -SNAP did not significantly reduce the basal level of T172 phosphorylation or affect the activation in response to addition of 2-deoxyglucose to the cells (Fig. 5a). Moreover, only the WT α -SNAP, but not the L294A or 1-285 forms, rescued cells from α -SNAP knockdown, in terms of increased AMPK, ACC and Raptor phosphorylation (Fig. 5b). The significant increase in mitochondrial biogenesis due to knockdown of α -SNAP, measured by citrate synthase activity, was prevented by overexpression of WT, but not by L294A or 1-285 forms of α -SNAP (Fig. 5c). The HA-tagged L294A and 1-285 mutated α -SNAPs co-immunoprecipitated the FLAG-tagged α subunit of AMPK comparable to WT α -SNAP, indicating the mutations in the C-terminal region of the protein did not eliminate binding to AMPK (Fig. 5d). The results suggest that the negative regulation of AMPK by α -SNAP in living cells was not simply due to protein-protein association, but probably depended on the phosphatase activity of α -SNAP.

Evidence for gain-of-function by *hyh* mutant α -SNAP M105I

The *hyh* (*hydrocephalus with hop gait*) mouse has structural and behavioral defects and abnormal localization of apical proteins in neuroepithelial cells, attributed to the homozygous point mutation Met105Ile (M105I) in α -SNAP¹⁷. The mechanistic basis for the *hyh* phenotype has been an unsolved mystery for over a decade. This single residue substitution was shown to not alter α -SNAP binding to NSF, or NSF-mediated disassembly of SNARE complexes¹⁷. LKB1 and AMPK have been assigned a role in establishing cell polarity^{18,19}, and this raised the possibility that the M105I mutation in α -SNAP might produce the *hyh* phenotype by affecting AMPK. Over-expression of α -SNAP (M105I) was more effective than equivalent levels of WT α -SNAP in preventing AMPK activation and phosphorylation of its substrates ACC and Raptor in response to addition of 2-deoxyglucose to the cells (Fig. 6a). This was manifested over a range of 2-deoxyglucose concentrations, where the M105I was consistently more effective than WT α -SNAP at reducing the level of T172 phosphorylation of AMPK (Fig. 6b). Furthermore, we observed a significant, ~2-fold increase in M105I α -SNAP co-immunoprecipitation with AMPK α compared to WT α -SNAP (Fig. 6c). These data suggest the *hyh* mutation in α -SNAP is a gain-of-function

substitution that enhances binding to AMPK to reduce LKB1-AMPK signaling. How lower AMPK activity is involved in the *hyh* phenotype merits further investigation.

Discussion

Our studies reveal α -SNAP as a negative regulator of pT172 in AMPK, limiting activation of the kinase in response to various stimuli, and thereby restricting AMPK suppression of anabolic signaling and induction of mitochondrial biogenesis. We demonstrate direct binding of α -SNAP to AMPK that is affected by ATP, making the binding potentially sensitive to the adenylate energy charge in cells. This is consistent with previous reports of adenine nucleotide effects on AMPK dephosphorylation^{8, 9}. Because recombinant α -SNAP *in vitro* dephosphorylated GST-AMPK we favor a model wherein α -SNAP acts as an AMPK phosphatase in cells, but it is possible other protein phosphatases also are involved in control of AMPK. Okadaic acid (OA), an inhibitor of all the PPP family phosphatases (e.g. PP1, PP2A, PP4, PP5, PP6), did not alter basal AMPK activity in rat hepatocytes, arguing against PPPs as AMPK phosphatases²⁰. However, OA was reported to attenuate palmitate-induced inhibition of AMPK activity²¹. Previous results using RNAi suggested that PP1 α and PP1 β , or PP2A, both of the PPP family, or PPM1E/F of a different phosphatase family could affect pT172 in AMPK α ²¹⁻²³. Recent genetic results in yeast implicate Sit4 phosphatase (mammalian PP6) in control of SNF1, the yeast homologue of AMPK²⁴. We suggest that perhaps multiple phosphatases can act on AMPK, and there may be some specificity of different phosphatases for individual AMPK isoform combinations.

The sequence of α -SNAP does not align with any other known Ser/Thr phosphatases. The properties of α -SNAP as a phosphatase distinguish it from the PPP, PPM and FCP/SCP families of phosphatases²⁵ (unpublished). Human α -SNAP is related in sequence to γ -SNAP and Sec17 (the yeast homologue), whose 3D structures consists of N-terminal helical repeats, mostly arranged anti-parallel into an extended twisted sheet, and a C-terminal helical bundle folded back against the sheet^{26, 27}. It is curious that the C termini of these proteins are flexible and the last 30 residues of γ -SNAP and the C terminal residues of Sec17 were not visualized in the crystal structures. Investigating the structural basis of α -SNAP as a phosphatase will require additional experimentation.

Our studies demonstrate that α -SNAP negatively regulates AMPK activation and thereby could affect cell growth, metabolism, polarity and development. Knockout of α -SNAP in mice is embryonic lethal and α -SNAP has been associated with pathological conditions²⁸. Articles published while this work was in progress reported knockdown of α -SNAP provokes apoptosis in epithelial cells²⁹ and induces p53 and sensitizes HEK293 cells to cisplatin³⁰. Interestingly, AMPK activation also is reported to induce p53 accumulation and inhibit cancer cell proliferation³¹. Because of the reports of apoptosis we tested whether RNAi knockdown of α -SNAP activates AMPK by inducing apoptosis. Extended knockdown of α -SNAP did induce caspase activation, shown by an increase of cleaved PARP, and this was blocked by co-treatment with Z-VAD, a caspase inhibitor (supplementary Fig. S2a). There was no effect +/- Z-VAD on activation of AMPK in response to α -SNAP knockdown. In addition, we found other agents that stimulated apoptosis, such as cisplatin, doxorubicin and TRAIL did not elicit an increase in AMPK

activity (supplementary Fig. S2b). Because α -SNAP knockdown activates AMPK, maybe AMPK is involved in induction of p53. The effects of α -SNAP knockdown on apoptosis and p53 were proposed to be independent of NSF, the canonical partner of α -SNAP^{29,30}. We also found that effects of α -SNAP on AMPK did not seem to depend on or involve NSF, so a reevaluation of the function of α -SNAP in cells seems to be warranted. Our discovery of *in vitro* phosphatase activity by recombinant α -SNAP WT, but not the previously known inactive L294A and 1-285 forms, suggests a broader repertoire of possible biological functions for this protein, long thought to merely act as an adapter. Whether there are other substrates of the α -SNAP phosphatase besides AMPK or whether α -SNAP serves as an adapter to direct AMPK to phosphorylate substrates are possibilities that deserve further study. We expect our finding of a physical and functional relationship between α -SNAP and AMPK provides interconnections between signaling pathways and intracellular processes that will impact systems biology analyses of network relationships.

Methods

Antibodies and reagents

The antibodies were obtained from the following sources: AMPK α 1/2, β 1/2, pAMPK α T172, pACC S79, pRaptor S792, pS6K T389, cleaved PARP, NSF and COXIV from Cell Signaling Technology, FLAG, FLAG M2 bead, HA and β -actin from Sigma, α -SNAP from Exalpa Biologicals, pAkt T308 from Merck, CV- α and CIII-core2 from Mitosciences. Other reagents obtained: Ni-NTA agarose from Qiagen, 2-deoxyglucose, phenformin, oligomycin, ATP, AMP, ATP- γ -S, PNP-AMP, FCCP and rotenone from Sigma, microcystin-LR, okadaic acid, calyculin A and protease inhibitor cocktail from Calbiochem, ZVAD-fmk from MP Biomedical, Citrate synthase activity assay kit from Sigma, protein G agarose, DiFMUP, Lipofectamine 2000 and all reagents for cell culture from Invitrogen, Seahorse assay reagents from Seahorse Biosciences, purified PP2A from Merck (Upstate). Other chemicals were from Fisher Scientific.

Plasmids

The constructs of AMPK α 2, β 1 and γ 1 were gifts from B. E. Kemp (University of Melbourne). α -SNAP human cDNA was obtained from Open Biosystems. The cDNAs of AMPK α 2, β 1, γ 1 and α -SNAP were amplified by PCR and subcloned into epitope tagged vectors by standard protocols. AMPK α 2, β 1 and γ 1 were fused with FLAG tag at N-terminus in pCMV tag-2B. α -SNAP was inserted into pcDNA3 with HA tag at C-terminus and bacterial expression vector pET28a with His tag at N-terminus. Site directed mutagenesis (AMPK γ 1 R299G, α -SNAP M105I and L294A) was performed using Quick Change II XL (Stratagene) according to the manufacturer's instructions. α -SNAP truncation from amino acid 1 to 285 (1 - 285) was obtained by PCR.

Cell culture and transfection

HEK293, HEK293T and MDCK cells were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS). HEK293T or HEK293 cells were seeded in 6-well plate, 6-cm dish and 10-cm dish. 18 h later plasmids were transfected using Lipofectamine 2000 at 1:1~3 ratio (plasmid:lipofectamine). The cells were harvested and

analyzed at 36 h after transfection. For stable transfection, cells were selected by G418 with concentration up to 1.5 mg/ml and maintained in 80 µg/ml of G418 after stable selection.

For analyzing AMPK activity with α -SNAP, HEK293T cells were grown in DMEM containing 5% FBS, 2 mM sodium pyruvate, 5 mM MgCl₂ and 50 mM glucose for 16 hrs to keep low basal AMPK activity. Cells were treated in this medium containing 5 mM glucose.

Cell extract and immunoprecipitation

Cells were rinsed once with chilled phosphate-buffered saline (145 mM NaCl, 5.4 mM KCl, and 10 mM sodium phosphate, 15 mM glucose, pH 7.4) and then scraped and homogenized by repeated passage through a 20-gauge needle. Homogenization buffer contained 25 mM HEPES, pH 7.4, 50 mM β -glycerophosphate, 10 mM sodium phosphate, 10 mM NaF, 2 mM EDTA, 2 mM EGTA, 1 mM dithiothreitol, 0.2% Tween 20 supplemented with 0.5 µM microcystin LR and protease inhibitor cocktail. Homogenates were centrifuged at 12,000 \times g for 10 min, and the supernatants were retained for analyses. For immunoblotting, samples were resolved by SDS-PAGE, transferred into nitrocellulose membrane and quantified with an Odyssey fluorescence scanner (Licor Industries) after immunoblotting. For immunoprecipitation, cell extracts (800 µl for 10-cm dish) were incubated with antibodies (2 µg) bound to protein G-agarose beads (10 µl) at room temperature for 1.5 h with constant mixing. The beads were then washed 4 times with homogenization buffer.

Lentiviral shRNA

Oligonucleotides encoding the short-hairpin RNA were annealed and subcloned into pLKO.1 (#8453, Addgene). HEK293LT cells (6-cm dish) were co-transfected with 2 µg of plasmids expressing shRNA, 1.5 µg of pCMV-dR8.2dvpr, and 0.5 µg pCMV-VSVG plasmids using Lipofectamine 2000. Virus-containing supernatants were collected and combined from 36 to 60 h after transfection and used to infect cells in the presence of 8 µg/ml polybrene (Sigma). Infected cells were selected with 4 µg/ml puromycin (Sigma) and analyzed on the 4-8th day after infection.

Mass spectrometry

Samples were separated by SDS-PAGE with 6-12.5% gradient gel and visualized by silver staining. Protein bands were excised and protein identification was performed at the Vincent Coates Foundation Mass Spectrometry Laboratory, Stanford University. After in-gel trypsin digestion, trypsin-generated peptides were analyzed by liquid chromatography-coupled tandem mass spectrometry (LC-MS/MS). Proteins were identified by comparing the mass spectrometry data against the NCBI nr protein database through the Mascot search engine.

Purification of recombinant proteins

His-tagged forms of α -SNAP wild type (WT), M105I, L294A and 1-285 and GST-tagged AMPK α 1-392 were expressed in *Escherichia coli* BL21-RIPL and purified. Briefly, 1 liter cultures were grown at 37°C for 12-16 hr to $A_{600} = 1.5$ and induced with 0.1 mM isopropyl-1-thio- β -D-galactopyranoside at 14°C for 20-24 hr. Bacterial pellets collected by centrifugation were resuspended in lysis buffer (20 mM Tris, 100 mM NaCl, 1 mM phenylmethylsulfonyl fluoride, 10 mM imidazole, 0.5% Triton X-100, pH 8.0). Cells were

broken by freeze-thaw for 2 cycles and sonicated (60 s), and lysates were centrifuged for 20 min at $4,000 \times g$. α -SNAP was purified from the supernatant by binding to Ni-NTA agarose and elution with imidazole (200 mM). Proteins were dialyzed against buffer (20 mM Tris, 100 mM NaCl, 1 mM dithiothreitol, pH 7.4) for 2-3 times. To assess purity and to confirm protein concentrations, samples were subjected to SDS-PAGE and then stained with Coomassie Blue. We found that concentrated solutions of α -SNAP tended to undergo aggregation, with loss of phosphatase activity. After testing a variety of conditions we found that dilution in pH 4.0 buffers with 0.5% lauryldimethylamide oxide provided consistently higher activity.

Binding assay of α -SNAP and AMPK γ

FLAG-tagged AMPK γ was expressed in HEK 293 T cells and isolated with anti-FLAG antibody coupled to Protein G-Agarose beads. One third of beads was mixed with 1 μ g purified recombinant α -SNAP in 100 μ l assay buffer (50 mM Tris pH 7.4, 100 mM NaCl, 1 mM DTT, 0.2% Triton X-100 and 1% BSA) supplemented with or without nucleotides, and incubated at room temperature for 20 min. The beads were washed by centrifugation four times with assay buffer and bound proteins analyzed by fluorescent immunoblotting and quantification with an Odyssey 2D IR scanner (Licor Inds.).

Phosphatase activity assay

GST-AMPK α 1-392 recombinant protein was phosphorylated *in vitro* by CaMKK II (a generous gift from Art Edelman) with 32 P-ATP. The reaction included 50 mM Tris pH 7.4, 1mM DTT, 10 mM MgSO₄, 1mM MnCl₂, 1 mM CaCl₂, 10 unit/ μ l calmodulin, 2 mM ATP (200 μ Ci/mM 32 P-ATP), 2 mg GST-AMPK α 1-392 protein and 10 μ g CaMKK II, incubating at 37°C for 24 hours. The reaction was terminated by adding 1/9 volume of 100% TCA, followed by dialysis of precipitated proteins in the buffer of 50 mM Tris pH 7.4, 5 mM DTT and 0.01% Brij 35 for 16 hours. Phosphatase assay with phosphoprotein was performed using trichloroacetic acid precipitation according to the procedure described previously¹

For *in vitro* phosphatase assay with AMPK, FLAG-AMPK α 2 was expressed in HEK293T cells that were stimulated with 25 mM 2-DG to activate the AMPK. FLAG-M2 beads were used to recover the phosphorylated FLAG-AMPK α 2 that was eluted by FLAG peptide (0.5 μ g/ μ l). The purified phosphorylated proteins were incubated for 30 min with 0.5 μ g of α -SNAP WT, M105I, L294A, 1-285 recombinant proteins and GST (as control) in assay buffer (50 mM Tris, 1 mM dithiothreitol, 10 mM MgCl₂, 10 μ M ATP, pH 7.4). AMPK α 2 pT172 was examined by fluorescent immunoblotting.

Measurements of oxygen consumption rate and extracellular acidification rate

A Seahorse Bioscience XF24-3 Extracellular Flux Analyzer was used to measure oxygen consumption rate (OCR) and extracellular acidification rate (ECAR). HEK293T cells (25,000 cells each well) were seeded in poly-L-lysine coated XF24 cell culture plate for 18-24 hours. The media was replaced with XF assay media (143 mM NaCl, 5.4 mM KCl, 0.8 mM MgSO₄, 1.8 mM CaCl₂, 0.91 mM NaH₂PO₄, 15 mg/ml phenol red supplemented with 15 mM glucose, 2 mM sodium pyruvate, 1 mM glutamine and 2% FBS) and

equilibrated in non-CO₂ incubator for 1 hr. Seahorse analyzer uses a cartridge with 24 optical fluorescent O₂ and pH sensors to measure OCR (pmol/min) and ECAR (mPh/min). Basal OCR and ECAR were measured for 3 min every 10 min for 4 points, followed by sequential injection of oligomycin (3 μM), 2-DG (25 mM), FCCP (1 μM) and Rotenone (1 μM). Each treatment was measured for 3 min every 10 min for 2 points. After measurement of OCR and ECAR, media is removed carefully and completely and 20 μl lysis buffer (20 mM Tris, 0.1% Triton X-100, protease inhibitor cocktail, pH 7.4) was added and protein concentration was analyzed to normalize OCR and ECAR.

Citrate synthase activity assay

HEK293T cells were homogenized in buffer containing 0.2% Tween-20 and 0.1% Triton X-100, and freeze-thawed once. Homogenates were centrifuged at 12,000 × *g* for 10 min, and the supernatants were retained for analyses. Samples of 3 μg protein were used for assay in 96-well microplate. The assay was performed following to manufacturer's instructions. Plate was read at 412 nm on kinetic program in linear response range (up to 10 min) and enzyme activity units (μmol/min/mg protein) calculated from the change in absorbance (A₄₁₂)/min according to the instructions.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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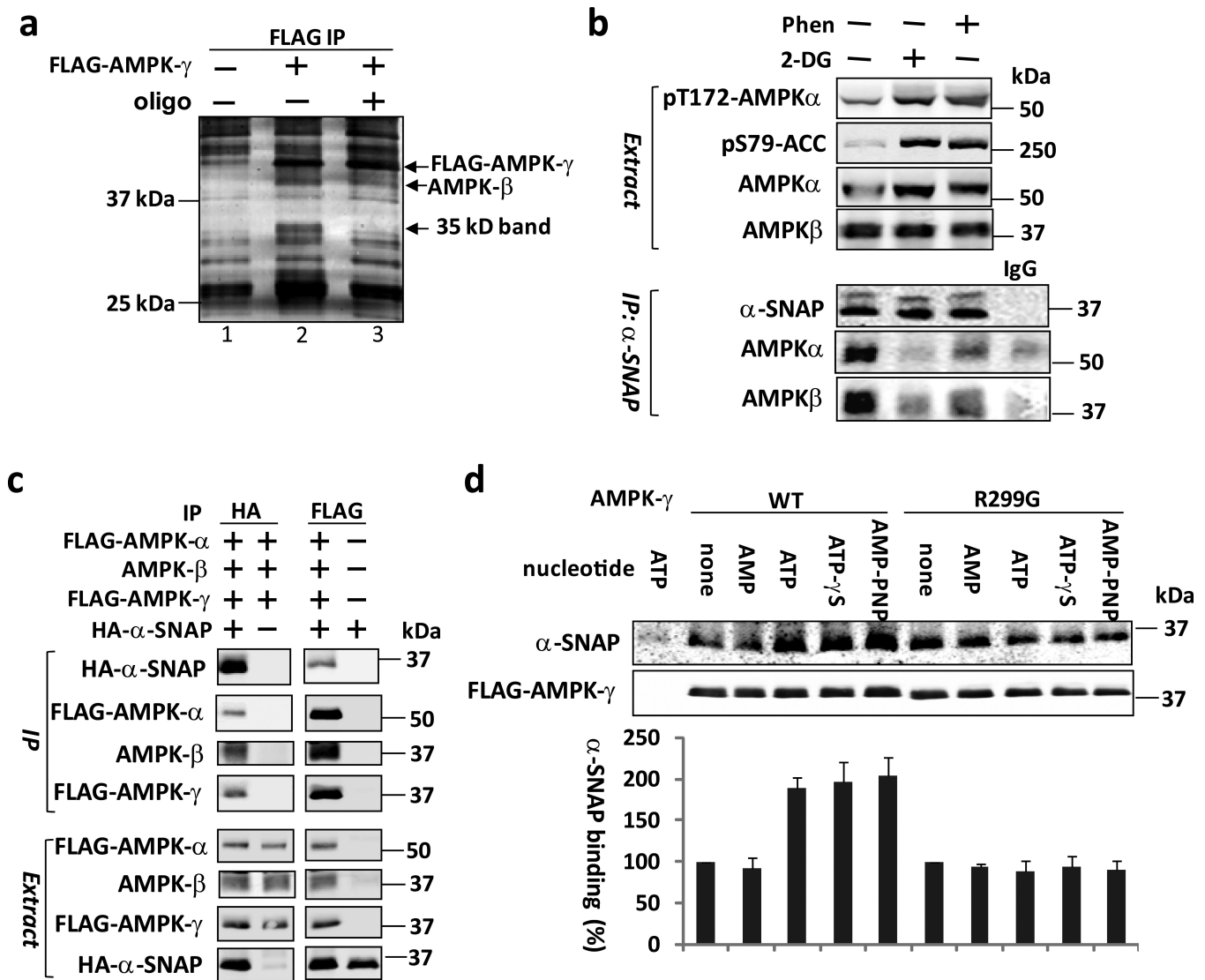


Figure 1. α -SNAP associates with AMPK bound to ATP

(a) HEK293 cells as control (lane 1) and cells stably expressing FLAG-AMPK γ 1 were treated for 20 min with vehicle (lane 2) or 2 μ M oligomycin (oligo) (lane 3). AMPK was isolated by FLAG immunoprecipitation (IP) and proteins resolved by SDS-PAGE, followed by silver-staining. The AMPK β and γ subunits (lane 2 and 3) are marked with arrows, and the prominent 35 kDa band (lane 2) was excised and analyzed by LC/MS/MS mass spectrometry. (b) Immunoblots of cell extracts (upper panels) and immunoprecipitates (IP, lower panels) of endogenous α -SNAP, compared to non-specific IgG as a control. HEK293T cells were untreated as control or treated with 25 mM 2-deoxyglucose (2-DG), or 3 mM phenformin (phen). (c) FLAG-AMPK α 2, β 1, γ 1 were co-expressed with HA- α -SNAP in HEK293T cells. Cell extracts and HA and FLAG immunoprecipitates were immunoblotted. (d) FLAG-AMPK γ 1 wild type (WT) and R299G were expressed in HEK293T cells and recovered on beads as FLAG immunoprecipitates. Beads were incubated with purified recombinant α -SNAP protein without or with addition of the indicated nucleotides. Samples were immunoblotted and staining intensity normalized

relative to samples with no nucleotide added. Data represent means \pm SEM for $n = 3$ and statistical analysis of ANOVA followed by multiple Fisher's test was performed (WT NONE versus ATP, ATP- γ S and AMP-PNP: $p < 0.02$; WT NONE versus AMP: $p = 0.997$; R299G variant, $p = 0.927$).

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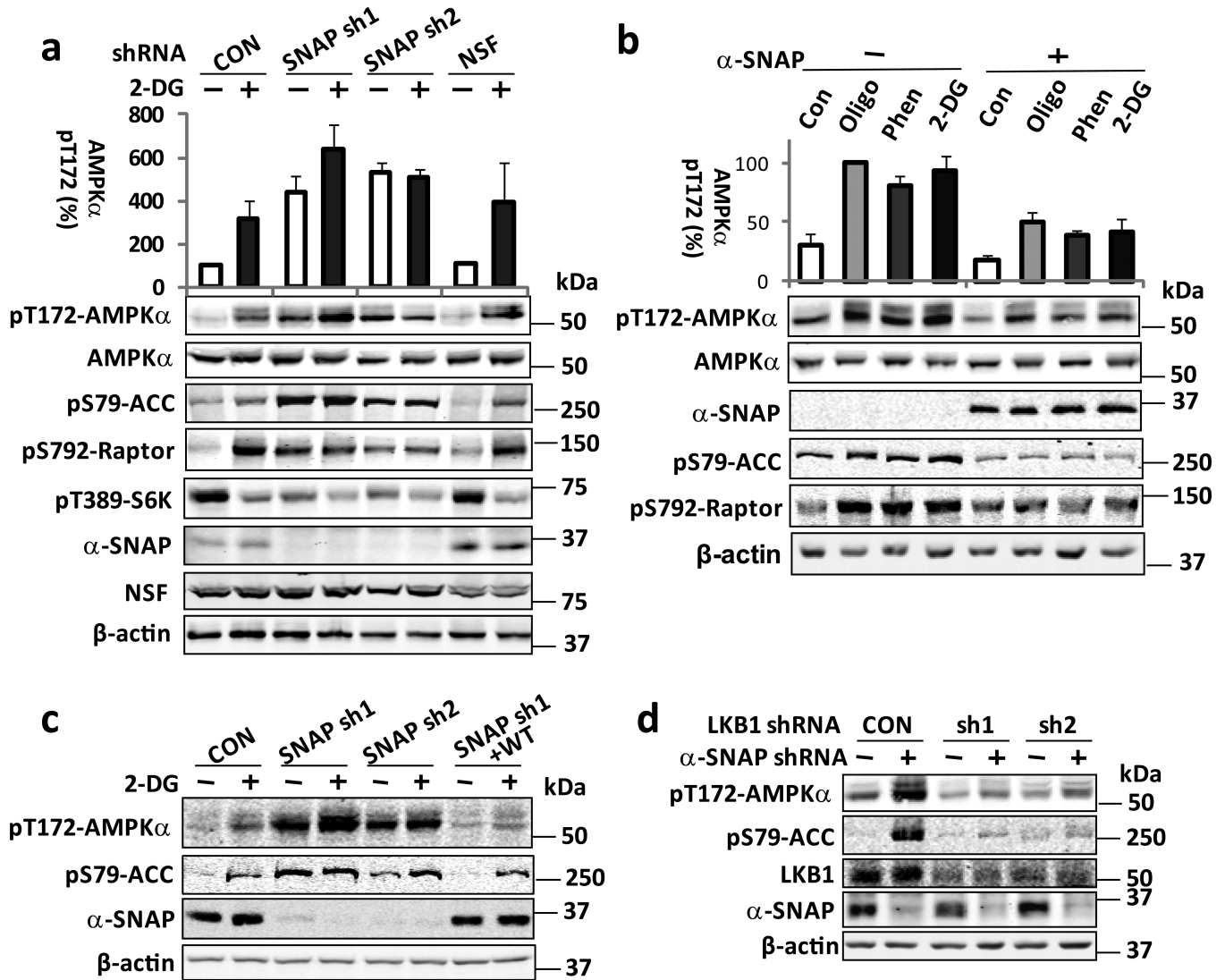


Figure 2. α -SNAP negatively regulates AMPK signaling in living cells

(a) HEK293T cells expressing lentiviral shRNA targeting GFP (control, CON), α -SNAP (SNAP sh1 and sh2), or NSF were treated +/- 25 mM 2-deoxyglucose (2-DG) for 20 min and cell extracts analyzed by immunoblotting, with AMPK α and β -actin as loading controls. Data represent means \pm SEM for $n = 3$ and statistical analysis of ANOVA followed by multiple Fisher's test was performed (basal CON versus basal α -SNAP sh1, sh2: $p < 0.001$; basal CON versus basal NSF sh: $p = 0.903$). (b) HEK293T cells were transfected in 6-well plates for 36 hrs with empty vector or vector expressing α -SNAP. Cells were treated with 2 μ M oligomycin (oligo), 3 mM phenformin (phen) or 25 mM 2-DG for 20 min and cell extracts analyzed by immunoblotting, with AMPK α and β -actin as loading controls. Data represent means \pm SEM for $n = 3$ and statistical analysis of two way ANOVA test was performed (without versus with α -SNAP expression: $p < 0.001$). (c) Madin Darby canine kidney (MDCK) cells were infected with lentivirus encoding shRNA targeting GFP (CON), dog α -SNAP (SNAP sh1 and sh2) or SNAP sh1 together with expression vector for human α -SNAP. Cells were treated (-/+) with 25 mM 2-DG for 20 min and cell extracts analyzed

by immunoblotting. **(d)** HEK293T cells were knocked down by infection with lentiviral shRNA targeting GFP (CON) or LKB1 (sh1 and sh2). Cells were subsequently infected with lentiviral shRNA targeting GFP (-) or α -SNAP sh1 (+) and extracts analyzed by immunoblotting.

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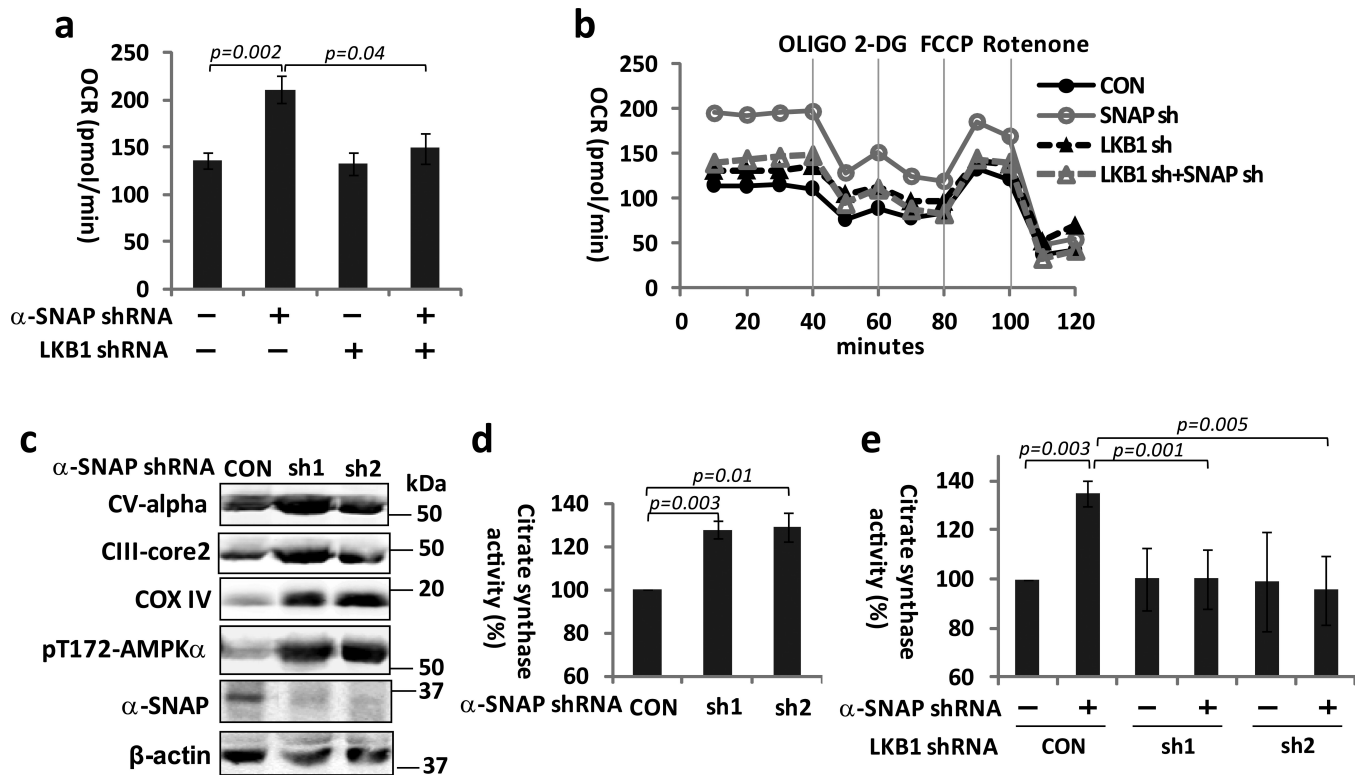


Figure 3. Knockdown of α -SNAP elevates mitochondrial respiration and stimulates mitochondrial biogenesis

(a) Oxygen consumption rate (OCR, pmol/min) in HEK293T cells measured by a Seahorse Analyzer. Lentiviral shRNA infected cells are indicated and double knockdown was performed as Fig. 2d. Basal measurements were normalized for protein concentrations and the mean \pm SEM from 4 independent experiments are plotted. Student's t test was used to calculate significance. (b) Seahorse continuous measurement of OCR. (c) HEK293T cells were infected with lentiviral shRNA targeting GFP (control, CON) or α -SNAP (SNAP sh1 and sh2). Mitochondrial proteins were detected by immunoblotting: CV- α , ATP synthase subunit alpha; CIII-core2, Complex III subunit Core 2; COXIV, cytochrome c oxidase subunit IV. (d and e) Citrate synthase activity as a measure of mitochondrial content was assayed and normalized relative to controls. Data represents mean \pm SEM (n = 4) and p values from Student's t-test of significance are indicated. (d) Cells prepared as c. e, α -SNAP and LKB1 were knocked down as described in Fig. 2d.

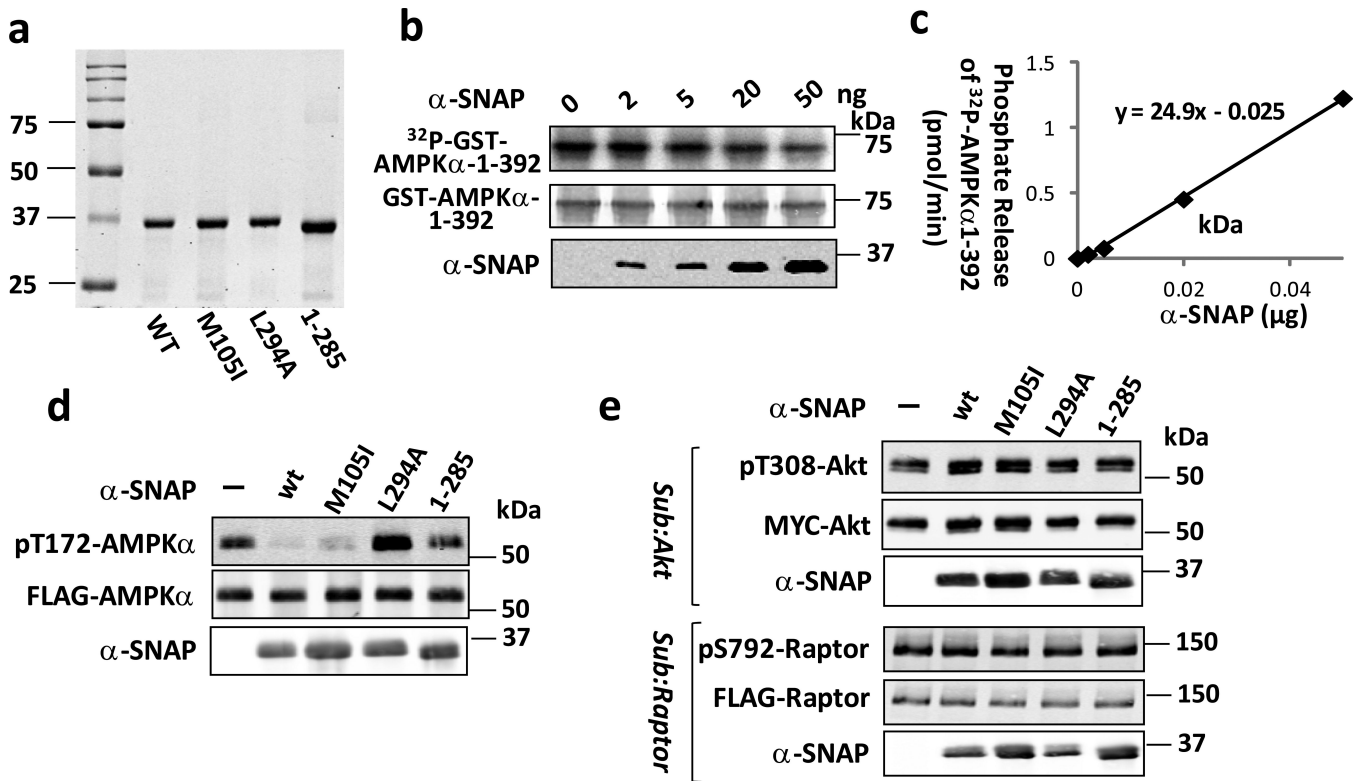


Figure 4. α-SNAP dephosphorylates pT172 in AMPKα

(a) Coomassie blue staining of purified wild type (WT) α-SNAP, mutants M105I, L294A and 1-285 were purified, used in assays for phosphatase activity. (b and c) ³²P labeled-GST-AMPKα1-392 recombinant protein was used as substrate of α-SNAP phosphatase. α-SNAP (2, 5, 10 and 50 ng) was incubated with ³²P-GST-AMPKα-1-392 (b) ³²P-GST-AMPKα-1-392 recovered by trichloroacetic acid precipitation was resolved in SDS-PAGE and detected by phosphor imager and GST-AMPKα-1-392 and α-SNAP were examined by immunoblotting (c) ³²P phosphate release was measured by scintillation counting and specific activity of different amounts of α-SNAP calculated. (d) Phosphorylated FLAG-AMPKα2 was purified from activated HEK293T cells and incubated for 30 min with GST (as control, CON), or α-SNAP WT, M105I, L294A or 1-285 (0.5 μg each). Phosphorylated T172 of AMPKα2 was analyzed by immunoblotting. (e) Raptor and Akt were expressed in HEK293T cells, purified by FLAG or MYC immunoprecipitation and incubated with GST (as control, CON), or α-SNAP WT, M105I, L294A or 1-285 (0.5 μg each). Phosphorylation of Akt T308 and Raptor S792 were analyzed by immunoblotting.

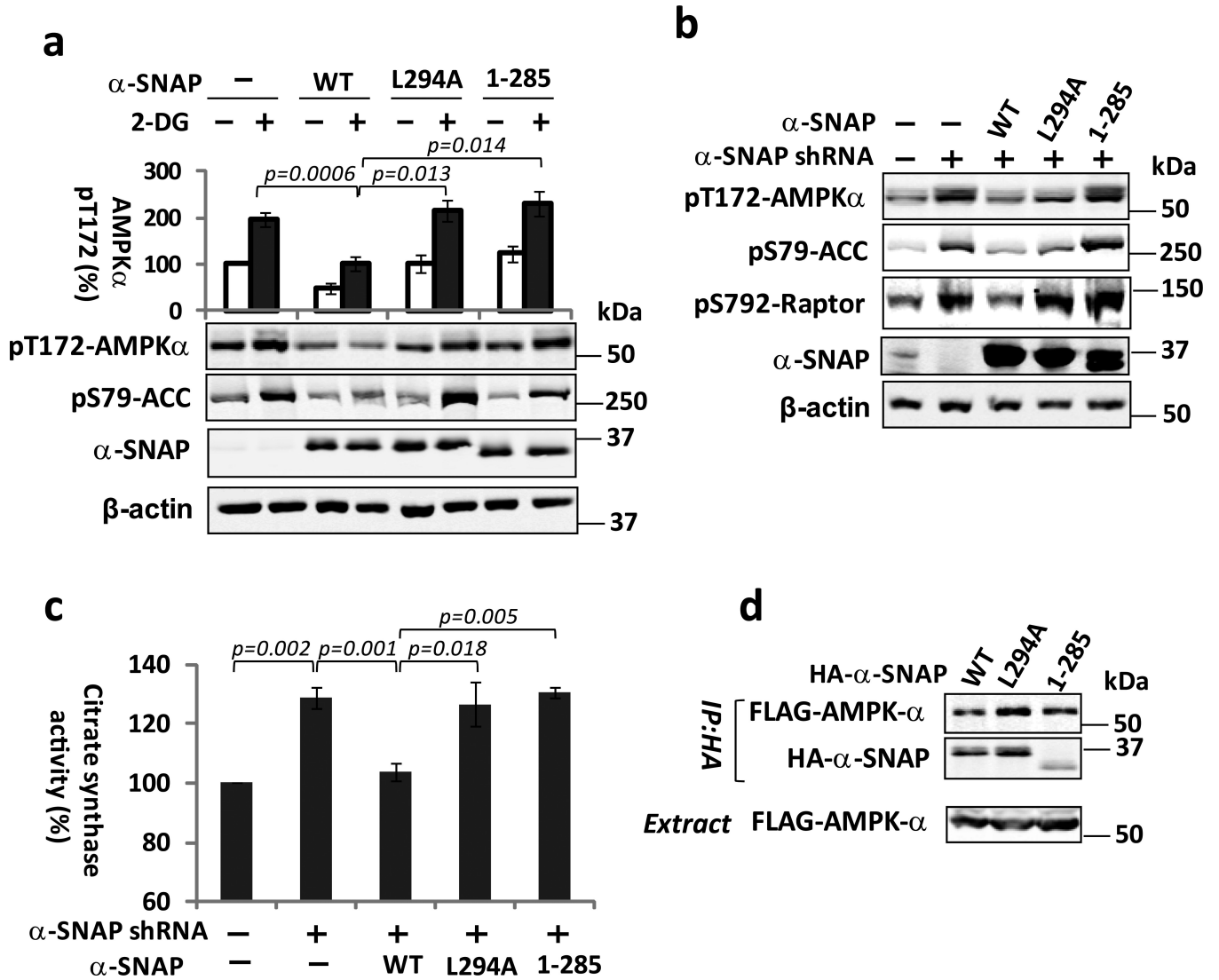


Figure 5. Effects of α-SNAP mutations on inactivation of AMPK

(a) The α-SNAP WT, L294A and 1-285 proteins were expressed in HEK293T cells that were treated (-/+) 25 mM 2-DG. Data represent mean ± SEM for n = 3 and statistical analysis of paired T test was performed. (b) HEK293 cells were infected with lentiviral shRNA targeting GFP (as control) or α-SNAP (SNAP sh1), followed by expression of α-SNAP WT, L294A and 1-285, as indicated. (c) Citrate synthase activity was measured in the cells prepared as in b. Data represents mean ±SEM (n = 4) and p values from Student's t-test of significance are indicated. (d) FLAG-AMPKα2 and γ1 were co-expressed with HA-α-SNAP WT, L294A and 1-285 in HEK293T cells as indicated. HA immunoprecipitates from cell extracts were analyzed by immunoblotting.

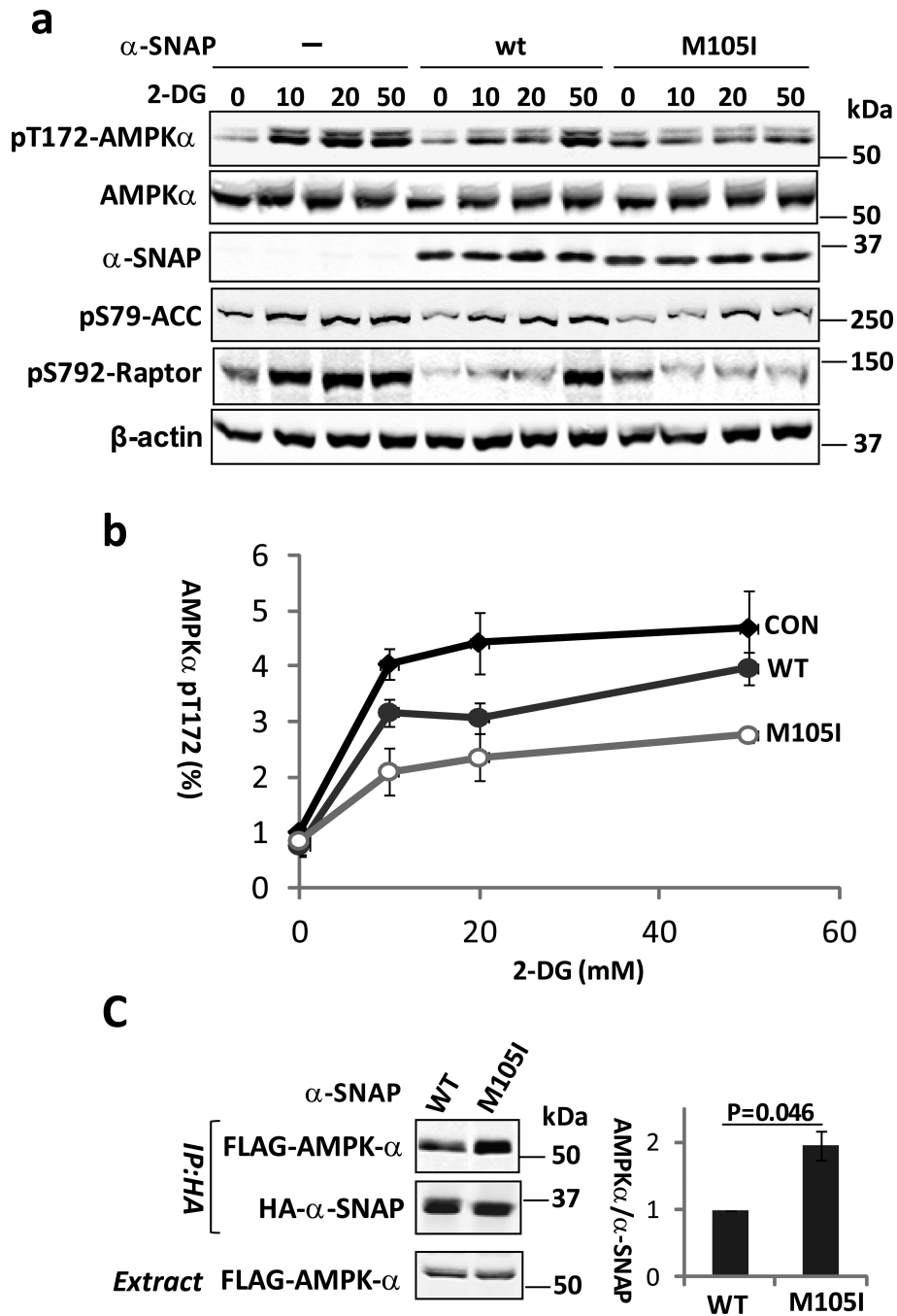


Figure 6. Gain-of-function by α -SNAP hyh mutant M105I in binding and inactivation of AMPK
(a) Vector control, α -SNAP WT and M105I were expressed in HEK293T cells. Cells were treated with 10, 20 and 50 mM 2-DG. Cell extracts were analyzed by immunoblotting. **(b)** Quantification of **a**. Data represents mean \pm SEM for 3 independent experiments. **(c)** FLAG-AMPK α 2 and γ 1 were co-expressed with HA- α -SNAP WT or M105I in HEK293T cells. HA immunoprecipitation was performed with cell extract. Data represent mean \pm SEM (n = 3).