

## Review

# Ligands for Intestinal Intraepithelial T Lymphocytes in Health and Disease

Akanksha Hada  and Zhengguo Xiao \*

Department of Animal and Avian Sciences, University of Maryland, College Park, MD 20742, USA; hada@umd.edu

\* Correspondence: xiao0028@umd.edu

**Abstract:** The intestinal tract is constantly exposed to a diverse mixture of luminal antigens, such as those derived from commensals, dietary substances, and potential pathogens. It also serves as a primary route of entry for pathogens. At the forefront of this intestinal defense is a single layer of epithelial cells that forms a critical barrier between the gastrointestinal (GI) lumen and the underlying host tissue. The intestinal intraepithelial T lymphocytes (T-IELs), one of the most abundant lymphocyte populations in the body, play a crucial role in actively surveilling and maintaining the integrity of this barrier by tolerating non-harmful factors such as commensal microbiota and dietary components, promoting epithelial turnover and renewal while also defending against pathogens. This immune balance is maintained through interactions between ligands in the GI microenvironment and receptors on T-IELs. This review provides a detailed examination of the ligands present in the intestinal epithelia and the corresponding receptors expressed on T-IELs, including T cell receptors (TCRs) and non-TCRs, as well as how these ligand-receptor interactions influence T-IEL functions under both steady-state and pathological conditions. By understanding these engagements, we aim to shed light on the mechanisms that govern T-IEL activities within the GI microenvironment. This knowledge may help in developing strategies to target GI ligands and modulate T-IEL receptor expression, offering precise approaches for treating intestinal disorders.

**Keywords:** mucosal immunity; intestinal immunity; intraepithelial lymphocytes; IELs; immune tolerance; gut epithelial barrier; ligand-receptor interactions; T cell receptor signaling; TCR; IL-15



Academic Editor: Subash Sad

Received: 26 December 2024

Revised: 17 January 2025

Accepted: 22 January 2025

Published: 23 January 2025

**Citation:** Hada, A.; Xiao, Z. Ligands for Intestinal Intraepithelial T Lymphocytes in Health and Disease. *Pathogens* **2025**, *14*, 109. <https://doi.org/10.3390/pathogens14020109>

**Copyright:** © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

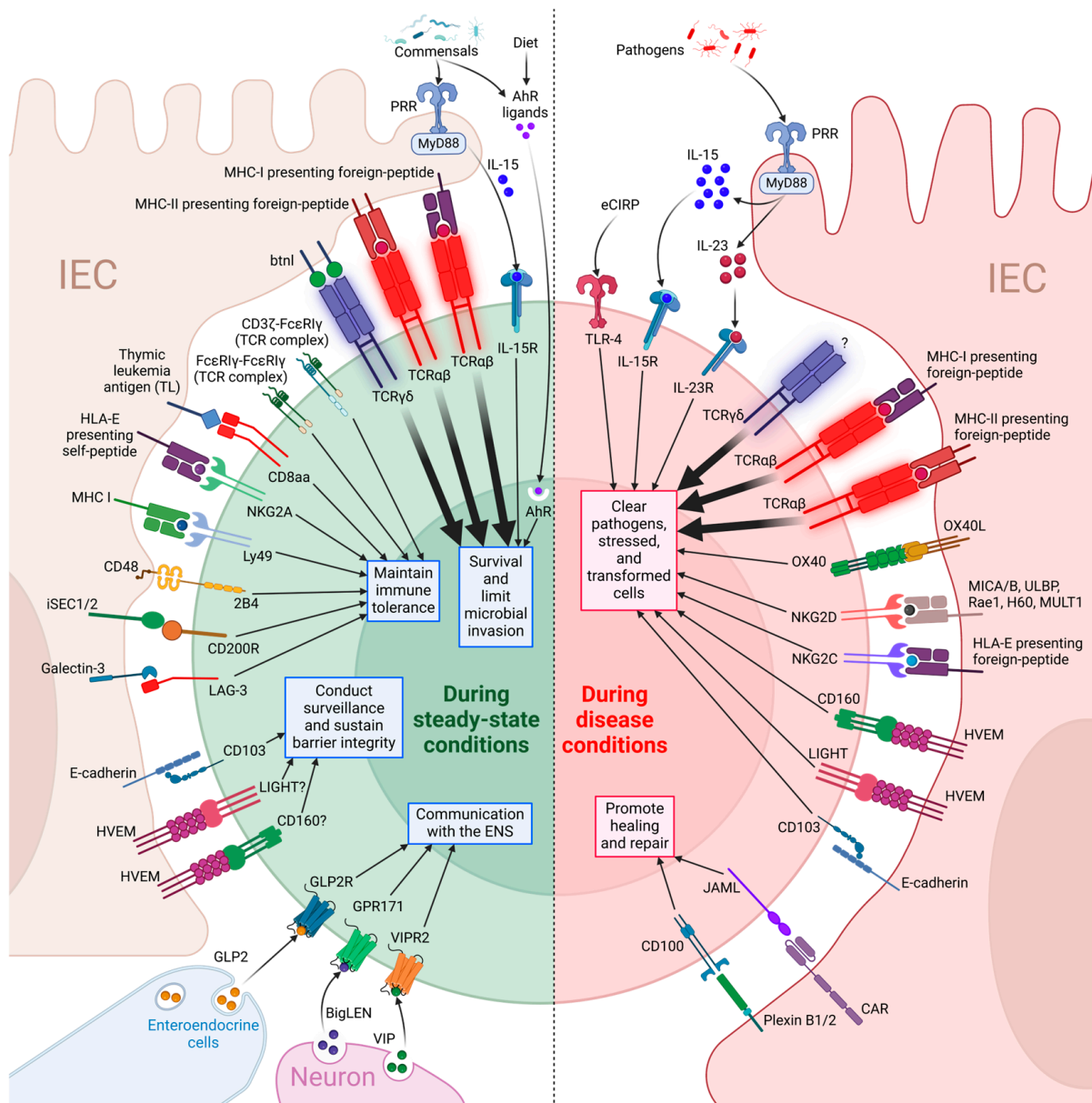
The epithelial tissues lining the body's surfaces form a vital barrier. They serve as the first line of protection against external threats while enabling the movement of ions, nutrients, and water across the epithelium. This role is especially critical in the intestinal epithelium, as it is continually exposed to a complex mixture of dietary antigens, commensal microorganisms, and potential pathogens [1,2]. In this challenging environment, the immune system must maintain tolerance to benign stimuli while mounting effective responses against the pathogens [2,3]. Among the key players are the intestinal intraepithelial T lymphocytes (T-IELs), a specialized population of immune cells residing within the intestinal epithelial layer [4]. These cells are strategically positioned and respond rapidly to changes in the gut microenvironment, playing an important role in maintaining intestinal homeostasis and coordinating immune responses [4,5].

T-IELs can be broadly categorized into two main subsets: natural T-IELs and induced T-IELs, each with distinct developmental origins and functional characteristics. Natural

T-IELs, which include  $\text{TCR}\alpha\beta+\text{CD8}\alpha\alpha+$  and  $\text{TCR}\gamma\delta+$  subsets, originate either directly from the thymus or extrathymic compartments [6]. Although the mechanisms underlying their development are not fully understood, these cells are considered pre-activated due to their recognition of self-antigen ligands and expression of activation markers during their ontogeny [7–10]. In contrast, induced T-IELs, comprising  $\text{TCR}\alpha\beta+\text{CD8}\alpha\beta+$  and  $\text{TCR}\alpha\beta+\text{CD4}+$  subsets, arise from conventional naive T cells that are activated by retinoic acid-producing dendritic cells in gut-associated lymphoid organs (GALT) such as mesenteric lymph nodes and Peyer's patches [11–17]. The activation of precursors of both natural and induced T-IELs upregulates gut-homing molecules such as  $\alpha 4\beta 7$  integrin and CCR9, facilitating their migration to the lamina propria of the intestinal mucosa [18–25]. Once there, these T-IELs undergo phenotypic and functional adaptations to establish their tissue-resident status within the epithelial layer. A key step in this process is a  $\text{TGF}\beta$ -mediated integrin switch, where the expression of  $\alpha 4\beta 7$  integrin is downregulated while  $\alpha \text{E}\beta 7$  is upregulated, facilitating their migration into the epithelium [11,18,26]. Concurrently, T-IELs undergo significant transcriptional reprogramming, which reduces the expression of genes involved in lymph node trafficking (e.g., CD62L and CCR7) and tissue egress (e.g., S1PR1, KLF2, and CXCR1) for their epithelial residency [27]. Additionally, T-IELs adopt a unique transcriptional profile driven by IL-15-regulated transcription factors, including T-bet and Runx3 [27]. These factors are crucial in programming the functional characteristics of T-IELs and guiding their terminal differentiation. As a result, T-IELs acquire the capacity to produce cytokines such as  $\text{IFN}\gamma$  and effector molecules such as granzymes, equipping them to fulfill their regulatory and protective roles in the GI microenvironment [27].

The functional diversity and specificity of T-IELs are fundamentally governed by real-time interactions between available ligands and expressed receptors on these cells within the epithelial layer, allowing them to rapidly adapt their responses to the dynamic intestinal milieu. As illustrated in Figure 1, the receptors on T-IELs can be broadly categorized into two main types: T cell receptors (TCRs) and non-TCR receptors. TCRs serve as the primary sensors for antigen recognition and initiating immune responses [28]. In peripheral T cells,  $\text{TCR}\alpha\beta$  recognizes pathogen-derived peptides presented by major histocompatibility complex (MHC) molecules [29,30], whereas  $\text{TCR}\gamma\delta$  is generally considered not to be restricted to MHC [31]. Within the intestinal epithelia, the specific ligands for the TCRs of natural T-IELs are not fully characterized. However, natural T-IELs with  $\text{TCR}\alpha\beta$  are known to interact with classical and non-classical MHC molecules [11,32–37], while  $\text{TCR}\gamma\delta$  in the  $\text{TCR}\gamma\delta+$  subset ( $\text{V}\gamma 7+$  in mice and  $\text{V}\gamma 4+$  in humans, which constitute the major population) binds to butyrophilin-like (btlnl) molecules expressed by IECs in an MHC-independent manner [10,11,26,38]. Although induced T-IELs are derived from conventional T cells, their TCRs are known to recognize not only pathogen-derived but also commensal-derived antigens presented by the MHCs, reflecting their unique functional adaptation to the gut [11,26]. Non-TCR receptors, on the other hand, fine-tune TCR signaling and can also potentially regulate T-IEL functions independently. These include cytokine receptors, co-stimulatory and co-inhibitory receptors, as well as pattern recognition receptors (PRRs), all depicted in Figure 1 along with their potential ligands in the GI microenvironment [4,26,39–45]. It is important to note that these receptors, reported in the literature, are dynamically regulated by signals within the GI microenvironment. Consequently, not all T-IELs express all these receptors simultaneously or at all times. The interaction of these receptors with specific ligands during various physiological and pathological states initiates cascades of intracellular signaling events [26,27,35,40]. The cumulative effect of these interactions collectively defines the specific functional profile of T-IELs, enabling them to respond appropriately to the ever-changing intestinal environment. This allows T-IELs to maintain intestinal

homeostasis under steady-state conditions and mount robust immune defenses during infections, while potentially contributing to pathogenesis in chronic illnesses.



**Figure 1.** Availability of potential ligands and the possible expression of T-IEL receptors in steady-state and disease conditions. This diagram illustrates the crucial roles of ligand availability and receptor expression on T-IELs within the intestinal epithelial environment under both steady-state (left) and disease (right) conditions. Under steady-state conditions, specific ligands and their corresponding receptors are essential for preserving T-IEL populations, preventing microbial invasion, maintaining immune tolerance, conducting immune surveillance, sustaining the integrity of the epithelial barrier, and communicating with the enteric nervous system, ultimately maintaining homeostasis. In contrast, during disease conditions, changes in ligand availability and ligand-receptor interactions aid in eliminating pathogens, stressed and malignantly transformed cells, as well as in promoting tissue repair and healing. TCR, T cell receptor; PRR, pattern recognition receptor; TLR, toll-like receptor; AhR, aryl hydrocarbon receptor; IL, interleukin; MHC, major histocompatibility complex; HVEM, herpesvirus entry mediator receptor; GLP, glucagon-like peptide; VIP, vasoactive intestinal peptide; eCIRP, extracellular cold-inducible RNA-binding protein; JAML, junctional adhesion molecule-like; CAR, coxsackievirus and adenovirus receptor. Created in BioRender.com (accessed on 15 January 2025).

Although ligand-receptor interactions are crucial for determining T-IEL function, there has been a lack of detailed analysis of the various ligands in the GI microenvironment and their specific interactions with T-IEL receptors. This review seeks to offer an overview of known ligand-receptor interactions and their impacts on T-IEL functions in both steady-state and pathological conditions, as illustrated in Figure 1. We explore T-IELs under steady-state conditions, focusing on how the availability of GI ligands and T-IEL receptor expression enable them to sense microbial components, modulate receptor expression and associated signaling pathways to promote immune tolerance, sustain survival, support immune surveillance, maintain epithelial barrier integrity, and potentially facilitate crosstalk with the enteric nervous system (Figure 1; left). In disease contexts, we examine GI ligands and T-IEL receptor expression, highlighting ligand-receptor interactions and their roles in pathogen clearance, the pathogenesis of chronic conditions such as celiac disease, inflammatory bowel disease, and cancer, as well as tissue repair (Figure 1; right).

## 2. T-IEL Ligands in Steady-State Condition

T-IELs are actively engaged in responding to ligands within the GI epithelia under steady-state conditions, which is crucial for maintaining intestinal homeostasis (Figure 1; left). These cells can selectively express a diverse array of receptors that interact with specific ligands available in the intestinal microenvironment during steady-state conditions. These interactions involve TCR signaling [27,46–48], microbial ligands signaling through Pattern Recognition Receptors (PRRs) on IECs [39,49], cytokine signaling via local mediators such as IL-15 [26,39,41–43], dietary influences from AhR ligands [26,35,43,50,51], and pathways mediated by other co-receptors [4,35]. T-IEL functions include preventing potential pathogen invasion by sensing microbial ligands, regulating receptor expression to promote immune tolerance, and supporting T-IEL survival. Additionally, they contribute to immune surveillance, maintain tight junctions, and facilitate communication with the enteric nervous system. Overall, the ligand-receptor interactions collectively create a complex network of signals that sustain intestinal homeostasis within the GI epithelia, as shown in Figure 1 (left).

### 2.1. Monitoring Lumen Microbes

Microbial components are sensed by T-IELs directly through their TCRs or indirectly via IECs [27,39,46–49]. These interactions drive cytokine production such as IL-15 by IECs [26,39,41–43], effector molecule expression, and antimicrobial peptide (AMP) release [52–55], ultimately strengthening the immune surveillance and regulatory functions of T-IELs to protect the epithelial barrier.

#### 2.1.1. Direct Sensing via TCRs

The sensing of microbial components through TCRs is a critical aspect of immune surveillance in the intestinal epithelium. Induced T-IELs, including both TCR $\alpha\beta$ +CD4+ and TCR $\alpha\beta$ +CD8 $\alpha\beta$ + subsets, appear to play a key role in this process by recognizing and responding to GI lumen microbial antigens that are constantly sampled by antigen-presenting cells (APCs) during steady-state conditions [27,46–48,56–58]. Despite the diverse luminal antigens at epithelial barriers, T-IELs exhibit limited TCR diversity, potentially allowing them to recognize conserved microbial or dietary components [27]. While some mechanisms and antigens are identified, our overall understanding of the process that drives this selective process is still limited. For instance, TCR $\alpha\beta$ +CD4+ T-IELs can recognize specific microbial enzymes such as  $\beta$ -N-acetylhexosaminidase from Bacteroidetes through their TCRs, which leads to proliferation and protection against inflammation in colitis models [27,46]. Furthermore, the interaction between TCRs on CD4+ T-IELs and MHC-

II molecules presenting microbial peptides on IECs is crucial for their activation and expansion. This interaction also supports subsequent IL-10 production, which is vital for effective microbial sensing and maintaining immune tolerance [47]. Additionally, MHC-II-dependent recognition of segmented filamentous bacteria (SFB) promotes the accumulation of SFB-specific TCR $\alpha\beta$ +CD4+ T-IELs. These T-IELs are also capable of producing Granzyme B, which aids in the turnover of IECs [48]. Similarly, TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs depend on TCR ligands for their survival and maintenance within the intestinal epithelium [56–58]. Although their specific ligands under normal physiological conditions remain undefined, variations in TCR V $\beta$  usage, particularly in V $\beta$ 6, V $\beta$ 7, and V $\beta$ 11, have been observed in these T-IELs from mice under specific pathogen-free (SPF), germ-free (GF), and antigen-free diet conditions [58]. These findings suggest that TCR signaling in steady-state conditions may be influenced by interactions with intestinal microbiota in TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs. Through these mechanisms, the TCRs on TCR $\alpha\beta$ +CD4+ and TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs seem to serve as crucial receptors for sensing microbial components in the intestinal environment, enabling a targeted immune response that maintains intestinal homeostasis under steady-state conditions.

### 2.1.2. Indirect Sensing Through IEC-Mediated Cytokine Signaling

The sensing of potential pathogens by T-IELs also occurs indirectly through interactions with IECs, providing effective protection for the epithelial barrier. IECs express pattern recognition receptors (PRRs) such as TLR-2 and TLR-4, which detect pathogen-associated molecular patterns (PAMPs) from microbes and trigger the production of cytokines, particularly IL-15. This cytokine plays a crucial role in activating, proliferating, and modulating the functions of T-IELs [11,26,39,41–43,55]. For instance, TLR-2 deficient mice exhibit reduced numbers of various T-IEL subsets, including TCR $\alpha\beta$ +CD8 $\alpha\alpha$ +, TCR $\alpha\beta$ +CD8 $\alpha\beta$ +, and TCR $\gamma\delta$ + T-IELs, with the remaining T-IELs showing decreased activation and proliferation, along with increased apoptosis [39]. In the colon, bacteria from the Bacteroidales order stimulate IL-6 production by T-IELs through NOD2 and MyD88 signaling in IECs, contributing to protection against *Citrobacter rodentium* infection [49]. PRR signaling-dependent IL-15 production by IECs also stimulates TCR $\gamma\delta$ + and TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs to produce antimicrobial peptides (AMPs) and enhances the cytotoxic activity of TCR $\alpha\beta$ + T-IELs [52–55]. This AMP production is crucial for preventing pathogen establishment and maintaining spatial separation between the microbiota and the intestinal epithelial surface under steady-state conditions [59]. While there is no evidence of T-IELs directly sensing microbes through their PRRs, TCR $\alpha\beta$ +CD4+CD8 $\alpha\alpha$ + T-IELs can eliminate infected IECs in response to damage-associated molecular patterns (DAMPs) such as extracellular cold-inducible RNA-binding protein (eCIRP) via TLR-4 [45]. However, it remains unclear whether these T-IELs can directly recognize microbial TLR-4 ligands such as lipopolysaccharide (LPS) [45]. This indirect sensing mechanism through cytokine receptor signaling complements the direct TCR-mediated recognition, providing a comprehensive surveillance system for maintaining intestinal homeostasis and protecting against potential pathogens.

Overall, the interaction of TCRs on T-IELs and PRRs on IECs with microbial-derived ligands provides essential signals that program T-IELs to prevent pathogen invasion. This programming modulates cytokine production, cytotoxic molecule release, and AMP secretion under steady-state conditions. While there is evidence of increased expression of intracellular peptidoglycan recognition receptors in TCR $\alpha\beta$ +CD8+ T-IELs during bacterial infection [43] and TLR-4 on TCR $\alpha\beta$ +CD4+CD8 $\alpha\alpha$ + T-IELs [45], comprehensive data on PRR expression and functionality across different T-IEL subsets remains limited, contrasting with the well-documented situation in peripheral T cells [60]. Interestingly, TCR $\alpha\beta$ +CD8 $\alpha\alpha$ +, TCR $\alpha\beta$ +CD8 $\alpha\beta$ +, and TCR $\gamma\delta$ + T-IELs appear to rely more heavily on



signals from IECs, particularly IL-15, while TCR $\alpha\beta$ +CD4+ T-IELs depend more on their TCR signaling during steady-state conditions [39,49,52–55]. To enhance our understanding of pathogen invasion prevention by T-IEL subsets, further research is needed in several areas. These include exploring TCR-mediated signaling pathways beyond TCR $\alpha\beta$ +CD4+ T-IELs, examining TLR expressions and functions within various T-IEL subsets, and investigating how different TLR stimulations in IECs indirectly activate distinct T-IEL populations.

## 2.2. Modulating Receptor Expression and Signaling Pathways for Immune Tolerance

T-IELs achieve immune tolerance in the GI epithelia through various means during steady-state conditions. Firstly, natural T-IELs in the intestine are equipped with tamed TCR signaling from birth, which is designed to balance responding to threats such as potential pathogens and tolerating normal gut contents such as commensals and dietary components. Modifications in the TCR complex and TCR signaling pathway reduce natural T-IELs' reactivity [38]. Secondly, the regulated expression of both inhibitory and stimulatory co-receptors on both natural and induced T-IELs as well as the availability of their respective ligands in the intestinal microenvironment establishes a finely tuned system for ensuring controlled immune responses that are crucial for immune tolerance by enabling T-IELs to maintain vigilance without overreacting under steady-state conditions [4,35].

### 2.2.1. Reduced TCR Signaling in Natural T-IELs

Natural T-IELs, including TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + and TCR $\gamma\delta$ +, are enriched in the intestine at birth, suggesting a pre-established role in neonatal immunity [61–68]. Although the intestinal microenvironment contains natural T-IELs' TCR ligands, such as classical and non-classical MHC-I molecules as well as MHC-II molecules presenting self-antigens ligands for TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + T-IELs, along with btl ligands on IECs for TCR $\gamma\delta$ + T-IELs, TCR signaling remains suppressed under steady-state conditions [10,11,32–38]. This reduced TCR signaling is primarily achieved through a distinct CD3 complex configuration, composed of either CD3 $\zeta$ -Fc $\epsilon$ RI $\gamma$  heterodimers or Fc $\epsilon$ RI $\gamma$ -Fc $\epsilon$ RI $\gamma$  homodimers, which differs from the CD3 $\zeta$ -CD3 $\zeta$  homodimers expressed by conventional T cells [69]. CD3 $\zeta$  typically has three immunoreceptor tyrosine-based activation motifs (ITAMs), whereas Fc $\epsilon$ RI $\gamma$  has only one. ITAMs are found in the cytoplasmic tails of the T cell receptor complex and are crucial for initiating signal transduction [70]. Therefore, the CD3 complex having Fc $\epsilon$ RI $\gamma$  has decreased potency and sensitivity of TCR signaling [71–73]. Moreover, natural T-IELs have significantly lower levels and reduced phosphorylation of Linker for Activation of T cells (LAT) protein as well as higher levels of LAT2, which further helps in decreasing the TCR signaling, compared to conventional TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T cells and induced T-IELs [38]. LAT is an adaptor protein essential for TCR-mediated signaling and T cell development, acting as an anchoring point for the assembly of TCR signaling complexes upon activation [74]. Restoring LAT expression in natural T-IELs rescues TCR signaling, as indicated by increased phosphorylation of downstream TCR signaling molecules [38]. Furthermore, LAT2 plays a dominant negative role in TCR signaling by competing with LAT for binding partners [38,75,76], and knocking out LAT2 leads to increased IFN $\gamma$  and TNF $\alpha$  production by natural T-IELs in vivo [38]. The reduced TCR signaling in natural T-IELs is functionally relevant, as it aligns with their tolerogenic role in the intestinal microenvironment. By suppressing TCR signaling, natural T-IELs might avoid unnecessary activation and pro-inflammatory responses to self-antigens and commensal microbiota, thus maintaining immune homeostasis and preventing tissue damage. For example, even though btl molecules interact with the TCRs of TCR $\gamma\delta$ + T-IELs, this interaction is known to diminish their TCR expression and decrease the production of pro-inflammatory cytokines such as IL-6 and IFN $\gamma$  [77,78]. Overall, the evidence suggests that even though natural

T-IELs' TCR ligands are present in the intestinal microenvironment, the modified TCR signalosome reduces their TCR signaling compared to peripheral T cells under steady-state conditions [38,79]. This mechanism should help natural T-IELs sustain self-tolerance within the gut and minimize the risk of inappropriate immune activation, harmful inflammation, or tissue damage in response to self-antigens and commensals.

### 2.2.2. Regulation of Co-Receptor Expression and Signaling Pathways

T-IELs regulate co-receptor expression and their signaling adapters to maintain immune tolerance. In mice, this regulation occurs through a coordinated process involving the upregulation of co-inhibitory receptors such as CD8 $\alpha\alpha$ , CD200R1, 2B4, LAG-3, Ly49 family members, PD-1, CD39, CD73, CD160, CD96, and CD161. Simultaneously, there is a reduction in expression and weakened signaling of activating co-receptors, including CD2, CD5, CD28, and NKG2D [4,35]. Furthermore, the availability of ligands for these receptors in the GI microenvironment is finely regulated. This intricate balance of receptor expression, signaling strength, and ligand availability collectively contributes to a state of immune tolerance under normal physiological conditions.

A major inhibitory co-receptor CD8 $\alpha\alpha$  is expressed in the majority of T-IELs in mice; however, it is negligibly expressed in human T-IELs [40,44,80–82], indicating unique regulatory mechanisms in these species. Tissue ligands such as TGF $\beta$ , IL-15, retinoic acid, IL-27, and IFN $\gamma$  help induce the CD8 $\alpha\alpha$  homodimer in T-IELs in mice [26,80,81,83]. The CD8 $\alpha\alpha$  binds Thymus-leukemia antigen (TL) that are highly expressed on IECs, and this CD8 $\alpha\alpha$ -TL interaction represses TCR signaling by redirecting CD8 $\alpha\alpha$  and its associated Lck tyrosine kinase away from the TCR [26], which prevents the full activation of T-IELs [84]. Additionally, TL also induces the death of activated CD8 $\alpha\beta$ + T-IELs that do not co-express CD8 $\alpha\alpha$  [84], thereby eliminating the chances of unwanted activation. The immunoregulatory effect of CD8 $\alpha\alpha$ -TL interaction is further supported by the fact that TL-deficient mice develop accelerated colitis in an inflammatory bowel disease (IBD) model [85], probably due to elevated TCR signaling in T-IELs. These effects likely control the balance between the anti-inflammatory and cytotoxic functions of T-IELs, especially during steady-state conditions, in mice. However, loss of TL expression only increases the proliferation of colonic CD8 $\alpha\alpha$ + T-IELs, but not on small intestinal T-IELs [43], which mandates further research. Furthermore, CD8 $\alpha\alpha$  expression is rarely detected in human T-IELs [86], indicating that human T-IELs are regulated through different mechanisms that are discussed below.

Under steady-state condition, both of the natural T-IELs and TCR $\alpha\beta$ +CD8 $\alpha\beta$ + induced T-IELs highly express inhibitory receptors, including CD200R1, 2B4/CD244, LAG-3, PD-1, CD39, CD73, CD160, CD96 and Natural Killer (NK) receptors, such as CD94/NKG2A, CD161, members of the Ly49 family. Natural T-IELs express a wider variety of these inhibitory receptors than induced T-IELs [35,75,87]. iSEC1 and iSEC2, ligands of CD200R, and CD48, a ligand of 2B4, are expressed exclusively by secretory cell lineages such as tuft cells, goblet cells, Paneth cells, enteroendocrine cells, etc., in the GI epithelium [75,88,89]. The binding of these ligands to their respective co-receptors on T-IELs could suppress inflammatory cytokine production and cytolytic activity by activating T-IELs. There is higher phosphorylation of intracellular adapters of CD200R1 and 2B4 in natural T-IELs compared to conventional CD8+ T cells, indicating their inhibitory role during steady-state conditions [38]. However, the suppressed feature is also observed in T-IEL cultures, and T-IELs do not express any CD200R1 and 2B4 ligands [38]. This raises uncertainty about whether CD200R1 and 2B4 ligands play an active role in T-IEL immune tolerance under steady-state conditions. Moreover, galectin-3, a ligand of LAG-3, is predominantly expressed in IECs of the villus tips [90]. Galectin-3 knockout mice suffer from a more severe disease progression in a DSS-induced colitis model [91], indicating their regulatory role in

the GI epithelia. Apart from galectin-3, IECs express MHC class II, which is also a canonical ligand of LAG-3 [92,93]. However, there was no difference in the response of T-IELs when cultured with or without anti-LAG-3 antibodies for 96 hours before stimulating it with anti-CD3 [38]. Additionally, ablation of TCR in  $\text{TCR}\alpha\beta+\text{CD8}\alpha\alpha+$  T-IELs decreased the expression of LAG-3 [50]. This suggests that TCR stimulation by their respective ligands might be required to increase the expression of LAG-3, which then responds to LAG-3 ligands to regulate the immune responses of these T-IELs. Furthermore, T-IELs in mice expressing the inhibitory Ly49 receptors such as Ly49A and Ly49G2, which bind to MHC-I molecules expressed on IECs, show diminished responses when their TCR is activated. This reduced responsiveness is indicated by an absence of CD69 upregulation and lower production of components that are vital for the activation and functionality of these immune cells, such as chemokines like MIP-1 $\alpha$  and lymphotactin [94]. On the other hand, T-IELs that do not express these Ly49 receptors, exhibit a more robust response to TCR stimulation [94]. These findings suggest that ligands for Ly49 inhibitory NK receptors are one of the candidates for being a regulating factor for T-IELs in steady-state conditions.

Apart from the expression of co-inhibitory receptors and the availability of their respective ligands, T-IELs typically lack or display reduced expression of co-stimulatory receptors. Additionally, their signaling is impaired and the co-stimulatory ligands are minimally expressed under steady-state conditions. For instance, natural T-IELs exhibit downregulated expression of TCR co-stimulatory molecules including CD2, CD5, and CD28 [27,69]. Moreover,  $\text{TCR}\alpha\beta+\text{CD8}\alpha\beta+$  T-IELs in mice do not express NKG2D, a type II transmembrane C-type lectin activating NK receptor, under steady-state conditions [40]. Distinct in humans, while  $\text{TCR}\alpha\beta+\text{CD8}\alpha\beta+$  T-IELs express NKG2D, they lack ITAM adapter molecules such as DAP12 that couple with NK receptors and mediate intracellular activation signals, thereby reducing their functional capabilities, including proliferation and cytokine secretion [95–98]. Thus, the TCR might be the only activating receptor capable of inducing cell proliferation and cytokine production in healthy human  $\text{TCR}\alpha\beta+\text{CD8}\alpha\beta+$  T-IELs [40]. Experimental evidence has shown that DAP12 expression is reduced under steady-state conditions but upregulated under pathological conditions [96,97]. Furthermore, under steady-state conditions, NKG2D receptor ligands such as MHC class I polypeptide-related sequence A/B (MICA/MICB) and UL16-Binding Proteins (ULBP) family members are either present at low levels or absent on IEC [99,100]. These lines of evidence suggest a distinct mechanism of tolerance in humans and mice.

In summary, the regulation of T-IELs through ligands that promote immune tolerance is crucial for maintaining gut homeostasis. Key to this regulation is the subdued TCR signaling in natural T-IELs. Moreover, the regulated expression of various co-receptors on T-IELs, influenced by the availability of specific ligands within the intestinal microenvironment, forms a sophisticated regulatory network. Additionally, significant species-specific differences in ligand interactions, particularly in the expression and function of co-receptors such as  $\text{CD8}\alpha\alpha$  and NKG2D in mice and humans, underscore unique regulatory mechanisms across species. Future research could focus on the roles and ligands of less understood co-receptors on T-IELs, including PD-1, CD39, CD73, CD160, CD96, and CD161 [35,75,87]. Additionally, investigating how altered TCR complex configurations affect TCR signaling in natural T-IELs during events such as pathogen invasion is important. Furthermore, it is important to explore the factors that regulate the expression of various inhibitory and stimulatory co-receptors on both natural and induced T-IELs, and the availability of their ligands under different conditions to understand their in-depth role in the intestine.



### 2.3. Essential Ligands for T-IEL Survival

The survival of T-IELs within the GI epithelia is facilitated by a complex network of ligand-receptor interactions, involving various receptors, such as TCRs [4,32–35,37], cytokine receptors [11,26,39,41–44,55,98,101], nuclear receptors [26,35,43,50,51], and other signaling co-receptors [102–106].

#### 2.3.1. TCR and Co-Receptor Signaling Requirements

Receptor-mediated signaling such as through TCR and  $\beta 1$  integrins has been known to be important in T-IEL maintenance. TCR ligands are dispensable for TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + and TCR $\alpha\beta$ +CD4+ T-IELs' survival but not for TCR $\gamma\delta$ + and TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs, in steady-state conditions. For instance, although TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + natural T-IELs have heterogenous TCRs that recognize either classical or nonclassical MHC molecules presenting self-antigen ligands [4,32–35,37], they can survive without surface expression of their TCR in the steady-state [20]. Moreover, the ablation of the TCR or long-term antigen withdrawal does not affect the maintenance of TCR $\alpha\beta$ +CD4+ T-IELs [79,107]. Therefore, these subsets do not require TCR signaling for their survival once they reach the GI epithelium [20,79,107]. In contrast, TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs do require TCR stimulation for their maintenance [56–58]. Their frequency decreases over time as encounters with antigen ligands on MHC molecules diminish, indicating their reliance on their TCRs for maintenance [56,57]. In the case of TCR $\gamma\delta$ + T-IELs, they are not restricted to any MHC molecules and their maintenance is independent of microbial or dietary antigens [11,43]. Instead, the predominant TCR $\gamma 7$ + subset in mice relies on their TCRs binding to btl1/6 on the IECs for their maintenance [108,109]. Similarly, in humans, the TCR $\gamma\delta$ + T-IEL population is primarily Vy4+ and is dependent on the expression of BTNL3/8 on IEC [110]. Btl expression on IECs is crucial for maintaining the TCR $\gamma 7$ + T-IEL subset, as its absence completely eliminates this population [108–111]. IECs express the HVEM (Herpesvirus Entry Mediator) receptor, a member of the TNF superfamily, whose ligand, LIGHT, enhances the epithelial production of basement membrane proteins, such as collagen IV. This basement membrane protein is known to interact with  $\beta 1$  integrins on T-IELs, promoting their survival in vitro [102]. This is further supported by the fact that the absence of  $\beta 1$  integrin leads to a reduction in TCR $\alpha\beta$ + T-IELs in vivo [102].

Overall, the TCRs and co-receptors are essential for sustaining T-IEL populations within the gut in a steady state. However, TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + and TCR $\alpha\beta$ +CD4+ T-IELs do not require TCR stimulation for their survival, making them notable exceptions.

#### 2.3.2. Microbial Signals and Cytokine-Dependent Survival Signals

Microbial components and local cytokines in the intestinal microenvironment are crucial for the maintenance of T-IELs. Microbial colonization seems to selectively induce clonal expansion of TCR $\alpha\beta$ + T-IEL subsets including TCR $\alpha\beta$ +CD8 $\alpha\alpha$ +, TCR $\alpha\beta$ +CD8 $\alpha\beta$ +, and TCR $\alpha\beta$ +CD4+, as oligoclonal TCRs are found on these T-IELs in the presence of microbes [40,112–116]. Additionally, the total number of T-IELs is significantly less in Germ-Free (GF) and antibiotic-treated mice as compared to that in specific pathogen-free (SPF) mice [55,117], and this number increases after microbial colonization in GF mice [118], which almost reaches the level in conventional mice [67]. This expansion of T-IELs relies on the expression of PRRs, including TLR and NOD-like Receptors on IECs [11,26,39,41,42]. These PRRs detect intestinal microbes, such as *Clostridia*, *Lactobacillus acidophilus*, *Bifidobacterium longum*, segmented filamentous bacteria (SFB), and viruses, which in turn result in the production of IL-15 by IECs, aiding in the maintenance of all T-IELs [11,26,39,41–44,55,101]. The critical role of IL-15 in T-IEL maintenance is evidenced by the marked reduction of T-IELs in IL-15 knockout mice [39,119]. The depletion

of TLR-2 or NOD2 decreases the number of these T-IEL subsets due to impaired IL-15 expression [39,119].

### 2.3.3. Dietary and Microbial-Derived Components

Dietary and microbial-derived components also play an essential role in the maintenance of T-IELs in GI epithelia. The Aryl Hydrocarbon Receptor (AhR) is a nuclear receptor that is highly expressed in T-IELs, and tryptophan-derived ligands, either dietary (indole-3-carbinol from cruciferous vegetables) or microbial (tryptophan metabolites from commensals such as *Lactobacillus reuteri*), are crucial for the survival and maintenance of all T-IEL subsets [26,35,43,50,51]. This is further supported by the fact that AhR-deficient mice or those lacking dietary AhR ligands have fewer T-IELs [120–122]. Moreover, AhR-deficient mice develop more severe colitis following DSS treatment than wild-type mice, and AhR ligands have been shown to relieve this colitis [120,123], possibly due to sustained T-IEL numbers. Additionally, dietary glutamine is known to aid in IL-22-AhR-dependent and IL-15-independent maintenance of the T-IEL population [124]. Furthermore, intestinal commensals convert dietary unsaturated fatty acids such as linoleic acid into conjugated linoleic acid isomers. These isomers help maintain TCR $\alpha\beta$ +CD4+CD8 $\alpha\alpha$ + T-IEL populations through unknown mechanisms, as evidenced by the decrease in this T-IEL subset when fatty acid isomerization pathways are genetically abolished [125].

Overall, T-IELs depend on a variety of receptors for their survival in the gut, including TCRs, membrane proteins, cytokines receptors, and nuclear receptors. Given the complexity of the intestinal epithelia, it is likely that additional receptors also contribute to the survival and maintenance of these T-IELs. Future research could focus on understanding if TCR $\alpha\beta$ +CD8 $\alpha\alpha$ + T-IELs recognize any specific self-peptide ligand in the gut and the effects of these interactions [38]. Conditional deletion of MHC molecules, specifically in the IEC in vivo, could shed more light on the functional outcomes of these interactions on TCR $\alpha\beta$ + T-IELs. Moreover, btl on IECs is essential for the survival of TCR $\gamma\delta$ + T-IELs. However, it is still unclear how btl expression is regulated on IECs, which signaling pathways it activates, whether other TCR ligands for TCR $\gamma\delta$ + T-IELs exist in the GI epithelia, and how these interactions affect TCR $\gamma\delta$ + T-IEL maintenance under steady-state conditions. Moreover, how T-IELs with specific TCR repertoires are selected for clonal expansion in different subsets of TCR $\alpha\beta$ + T-IELs, especially since this expansion seems to be TCR-independent is still not well characterized. Additionally, the maintenance mechanisms of TCR $\gamma\delta$ + T-IELs in the absence of microbes and microbe-dependent IEC-derived IL-15 remain poorly understood.

### 2.4. T-IEL Surveillance and Barrier Maintenance

Microbial signals mediate T-IEL's motility and patrolling behavior. This surveillance is supported by occludin, a tight junction component, along with E-cadherin and Ep-CAM, which are cell adhesion molecules on T-IELs that enable crucial interactions with IECs [5,75,106,126,127]. Glucose availability also ensures swift T-IEL migration, regulates barrier permeability, and maintains epithelial integrity [5]. HVEM ligands and GLP2R (glucagon-like peptide 2 receptors) on T-IELs further might enhance surveillance and repair functions, hinting at a sophisticated network that sustains immune homeostasis and intestinal integrity [75,102–104].

Local IL-15 produced by IECs in response to microbial PRR signals mediate TCR $\gamma\delta$ + T-IELs surveillance behavior. This is indicated by experiments where the inhibition of IL-2R $\beta$ , a subunit of IL-15 receptor, in these T-IELs significantly attenuated the basal motility of these cells, resulting in their idling within the lateral intercellular space during early invasion of pathogens such as *Salmonella typhimurium* [5,26,128]. Moreover, occludin expressed on TCR $\gamma\delta$ + T-IELs mediates their rapid migration within the space between

the epithelial layer and the basement membrane by contacting IECs, which helps them cover the entire intestinal epithelium within a few hours [5,106,126]. Occludin's tight associations with the presence of ZO-2 in T-IELs could enhance the intestinal epithelial barrier function by contributing to the formation and maintenance of tight junctions and could regulate the permeability of the epithelial barrier, ensuring proper movement of T-IELs between neighboring cells while also maintaining the integrity of the intestinal lining [75,129]. Additionally, this migration is dependent on glucose availability, as systemic administration of glucose-inhibitor, 2-deoxy-glucose, inhibited this movement in TCR $\gamma\delta$ + T-IELs [5]. Moreover, epithelial retention of TCR $\gamma\delta$ + T-IELs within the lateral intercellular spaces is partly mediated by the direct interaction of T-IEL's CD103 with E-cadherin on IEC [106]. E-cadherin and EpCAM, expressed on T-IELs, facilitate interactions with IECs and help in maintaining the structural integrity of epithelial tight junctions and regulating paracellular permeability across epithelial cells [75,127]. Furthermore, since T-IELs reside within the epithelial layer and do not recirculate, endothelial cell adhesion molecules such as PECAM-1 and CD62L, that facilitate naive T cell migration into secondary lymphoid organs, are not expressed on T-IELs [75].

Besides junction proteins, other receptors also seem crucial for T-IEL's surveillance and tight junction functions. HVEM expressed on IECs has been shown to be required for the surveillance behavior of CD8 $\alpha\alpha$ + T-IELs, which includes both natural and induced T-IELs in mice, as observed through intravital microscopy [102]. It is not clearly understood whether this occurs through direct binding of a receptor on T-IELs to HVEM on IECs. However, for direct binding, potential ligands for the HVEM receptors include CD160 and LIGHT expressed on T-IELs [102]. Additionally, T-IELs also express GLP2R, the receptor for glucagon-like peptide 2 (GLP2), which regulates intestinal barrier function and suggests a role in epithelial maintenance [75,103,104]. GLP2 is produced by enteroendocrine L cells in the intestines [130]. GLP2R expression on T-IELs links these cells to intestinal lining maintenance and repair by enhancing barrier function, promoting epithelial proliferation, reducing mitochondrial damage, and decreasing apoptosis [103–105]. However, further research is needed to clarify these interactions.

In conclusion, microbial stimuli, in conjunction with proteins such as occludin, E-cadherin, and EpCAM on T-IELs, are crucial for establishing interactions with IECs. These interactions promote rapid T-IEL migration, regulate barrier permeability, and maintain epithelial integrity. HVEM ligands and GLP2R add layers of complexity by enhancing surveillance and maintenance functions. The interactions and roles of cell adhesion molecules have been extensively studied in TCR $\gamma\delta$ + T-IELs; however, whether TCR $\alpha\beta$ + T-IELs engage in similar interactions and the full spectrum of functions mediated by these interactions across different T-IEL subsets remain unclear. Future research could explore the mechanisms through which T-IELs, particularly TCR $\alpha\beta$ + subsets, contribute to surveillance and barrier maintenance.

## 2.5. Crosstalk Between T-IELs and the Enteric Nervous System

The potential crosstalk between T-IELs and the enteric nervous system (ENS) via the expression of neuropeptide receptors such as GPR171 and VIPR2 (vasoactive intestinal peptide receptor 2) as well as through neural cell adhesion molecules such as NCAM1 and NrCAM on T-IELs underscores a complex neuroimmune interface within the gut mucosa, integrating immune responses with neural activities. T-IELs express neuropeptide receptors such as GPR171 and VIPR2, indicating their responsiveness to neuroactive peptides released by the ENS [75]. GPR171 and VIPR2, which are known to bind to BigLEN and vasoactive intestinal peptide (VIP) respectively, can modulate immune responses [75]. BigLEN suppresses T cell activation and anti-tumor responses [131]. VIP is known to activate the

cAMP/PKA pathway in conventional T cells for its anti-inflammatory properties [132–134]. Moreover, VIP is associated with Th2 cell development and inhibition of Th1 and Th17 responses [132–134]. The anti-inflammatory properties of VIP have been confirmed in knockout mouse models, where VIP-deficient mice develop lung inflammation [133,134]. Furthermore, TCR $\gamma\delta$ +CD8 $\alpha\alpha$ + T-IELs express the neural cell adhesion molecules NCAM1 (CD171) and NrCAM, which might play critical roles in homophilic adhesion and axonal guidance with the nerve endings [75]. Therefore, it is reasonable to speculate that these ligand-receptor interactions might be involved in communication between T-IELs and the enteric nervous system.

Future research could explore the interactions between T-IELs and the ENS, particularly focusing on how T-IEL-expressed neuropeptide receptors such as GPR171 and VIPR2 respond to neuroactive peptides and the importance of neural cell adhesion molecules such as NCAM1 (CD171) and NrCAM on T-IELs.

### 3. T-IEL Ligands in Diseases

During disease states, T-IELs engage with multiple ligand-receptor interactions in a sequential manner to mount effective immune responses (Figure 1; right). Initially, TCR $\gamma\delta$ + T-IELs respond through IEC's PRR-dependent pathways, where IEC-derived IL-15 trigger T-IELs to produce antimicrobial peptides (AMPs) such as RegIII $\gamma$  [5,26,53,106,126,135,136]. This early response is followed by TCR-dependent mechanisms, particularly in TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs, which recognize pathogen-derived peptides presented on MHC I molecules by IECs and respond with IFN $\gamma$  production and cytotoxicity [137–143]. Moreover, co-receptor interactions provide additional layers of immune defense. For instance, NK receptors such as NKG2D and NKG2C on T-IELs interact with stress-induced ligands such as MICA/MICB and ULBP family members on infected cells to initiate cytotoxicity, a process that may occur independently of or in conjunction with TCR signaling [4,96,136,144–146]. The HVEM-CD160 interaction activates cytotoxic responses and promotes RegIII $\gamma$  production through the HVEM-Stat3-REG3 pathway [147,148]. OX40-OX40L signaling enhances cytokine production and cytotoxicity during infection [149]. In chronic inflammatory diseases, persistent IL-15 production by IECs leads to increased expression of NK receptors (NKG2D, NKG2C) on TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs [96,98,150–153]. These T-IELs recognize stress ligands such as HLA-E and MICA on damaged epithelial cells, leading to cytotoxic activity that exacerbates epithelial damage and promotes further inflammation. Moreover, in cancers, TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs can eliminate tumor cells expressing NKG2D ligands, such as MICA and ULBP, through FasL-mediated cytotoxicity [154], with the CD103/E-cadherin interaction facilitating contact-dependent killing [155], thereby contributing to the control of tumor progression and invasion. Furthermore, TCR $\gamma\delta$ + T-IELs utilize junctional adhesion molecule-like protein (JAML)-coxsackievirus and adenovirus receptor (CAR) and CD100-mediated pathways to promote epithelial healing through Keratinocyte Growth Factor (KGF) production [69,156].

This complex network of ligand-receptor interactions allows T-IELs to respond appropriately to various pathological conditions, maintaining intestinal homeostasis and promoting tissue repair when needed, as illustrated in Figure 1 (right).

#### 3.1. Infectious Diseases

##### 3.1.1. Pathogen-Derived Ligands and T-IEL-Mediated Clearance

During infections, T-IELs coordinate IEC-dependent pathways [5,26,53,106,126,135,136], TCR-mediated responses [137–143,157], and co-receptor interactions [96,98,136,144,145,153] to effectively combat pathogens. The engagement of specific ligands with these correspond-

ing receptors plays an important role in shaping intestinal immune defenses and facilitating pathogen clearance, ultimately influencing disease progression and outcomes.

#### Early Responses Mediated by Cytokine Receptor Signaling

TCR $\gamma\delta$ + T-IELs are major players in early intestinal immune responses, utilizing IEC-dependent, TCR-independent mechanisms through interactions with various epithelial microenvironment ligands to prevent pathogen colonization and preserve epithelial barrier functions. These T-IELs preferentially migrate to mucosal sites where pathogens adhere and generate a robust immune response [5,26,53,106,126,135,136].

The initial response of TCR $\gamma\delta$ + T-IELs to infections such as *Salmonella Typhimurium* and *Toxoplasma gondii* is independent of TCR signaling, which is demonstrated by the inability of TCR signaling inhibitors, targeting Zap70/Syk kinases, and TCR signaling-blocking antibodies to alter their migration and defensive responses during the infections. Specifically, TCR $\gamma\delta$ + T-IELs limit these infections through indirect signaling involving the PRRs on IECs, which initiates a MyD88-dependent pathway on IECs to stimulate the production of the RegIII $\gamma$  AMP from these T-IELs in IL-15-independent manner [5,26,53,106,126,135,136]. Moreover, *Salmonella* triggers IL-23 production by IECs in a TLR-dependent manner, which in turn stimulates TCR $\gamma\delta$ + T-IELs to produce IL-22 to limit the bacterial invasion by inducing the secretion of bactericidal Angiogenin 4 AMP by Paneth cells [158]. Apart from bacterial defense, IL-22 produced by TCR $\gamma\delta$ + T-IELs could also help to activate goblet cells to secrete mucus, which could prevent GI epithelia from larger pathogens including helminths [159]. This is evidenced by experiments where IL-22-deficient mice are unable to expel the *Nippostrongylus brasiliensis* and *Trichuris muris* from their intestines due to a reduced number of goblet cells and intestinal mucins [159,160].

Overall, TCR $\gamma\delta$ + T-IELs play an indispensable role in early intestinal immune responses, leveraging intestinal ligands-mediated and TCR-independent mechanisms to prevent pathogen colonization and maintain epithelial barrier functions. Although human peripheral TCR $\gamma\delta$ + T cells respond directly to microbial products through their TLR stimulation and also recognize bacterial phosphoantigens via their TCR [43,161–163], it is yet to be determined whether TCR $\gamma\delta$ + T-IELs are also capable of directly recognizing luminal microbes in such manners. Furthermore, the mechanisms through which TCR $\alpha\beta$ + T-IELs sense pathogens and initiate their early responses, particularly those mediated by cytokine receptor signaling, remain poorly understood. While IL-15 signaling is known to enhance the effector functions of antigen-primed TCR $\alpha\beta$ + T-IELs, which are all induced T-IELs, its role in primary infections has not been thoroughly investigated [164–167]. Understanding the cytokine receptor-mediated mechanisms by which T-IELs detect pathogens during early infections can be crucial for developing targeted therapies to enhance mucosal immunity and combat intestinal infections.

#### TCR-Dependent Mechanisms

TCR being the major antigen receptor on T cells, it is likely that TCR ligands are critical players in responding to and eliminating pathogens. Induced T-IELs including both TCR $\alpha\beta$ +CD8 $\alpha\beta$ + and TCR $\alpha\beta$ +CD4+ subsets, are antigen-experienced and exhibit a strong response to TCR stimulation [38,75,168]. However, in case of new infections, antigen-specific T-IELs typically develop within at least seven days following the exposure [137–142,169]. Evidence predominantly supports the role of the TCR $\alpha\beta$ +CD8 $\alpha\beta$ + subset in recognizing intracellular pathogen-specific ligands and mediating immune responses [137–142,169]. TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs are the ones known to detect various pathogen ligands presented on MHC I molecules by cells such as IECs, responding in an antigen-specific manner by producing cytokines such as IFN $\gamma$  and resulting cytotoxic-



ity of infected cells [137–143]. For instance, during *Encephalitozoon cuniculi* infection, the depletion of TCR $\alpha\beta$ +CD4+ T-IELs does not alter the outcome, whereas the removal of TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs leads to mortality of the infected animals [138], indicating a critical role of these T-IELs in pathogen clearance. Additionally, during *Encephalitozoon cuniculi* oral infection, TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs demonstrate antigen-specific cytotoxicity as well as increased IFN $\gamma$  production [138]. A similar response occurs during *Toxoplasma gondii* infections [139]. Notably, adoptive transfer of TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs from infected mice provides long-lasting protection against *T. gondii* in an IFN $\gamma$ -independent manner [139,140], indicating antigen-specific protective capacity. This specificity is also observed in responses to *Cryptosporidium* [141] and acute LCMV infections [142]. Moreover, although TCR signaling plays a critical role in the differentiation of TCR $\alpha\beta$ +CD4+ T-IELs, their TCR-specific responses to pathogens remain unclear [79]. Once differentiated, these cells have been shown to react to antigens in the absence of cognate TCR interactions [79]. TCR $\gamma\delta$ + T-IELs have also been known to respond to virulent *Listeria monocytogenes* infections by producing IFN $\gamma$  and exhibiting cytotoxicity in a TCR-dependent way. To further confirm, the depletion of their TCR using anti- $\gamma\delta$  TCR monoclonal antibody GL3 diminished these responses [170]. However, the direct response of TCR $\gamma\delta$ + T-IELs to pathogens via TCR signaling remains less clear. Furthermore, peripheral TCR $\gamma\delta$ + T cells are known to directly recognize bacterial components such as Listeriolysin O and heat shock proteins through their TCR [31], suggesting a similar potential for TCR $\gamma\delta$ + T-IELs.

Overall, TCR signaling plays a crucial role in T-IEL-mediated pathogen clearance, with TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs being particularly effective in recognizing pathogen-specific ligands and mounting antigen-specific immune responses, while TCR $\gamma\delta$ + T-IELs also contribute to protection against certain infections through TCR-dependent mechanisms.

### Co-Receptor Interactions

Ligands for T-IEL co-receptors such as NK receptors, CD160, and OX40, are essential for immune defense within the GI epithelia. NK receptor ligands are upregulated on T-IELs in response to infections, cellular stress, or malignant transformations, including MICA/MICB and ULBP family members in humans, and retinoic acid early inducible-1 (Rae1), histocompatibility 60 (H60), and UL16-binding protein-like Transcript 1 (MULT1) in mice [4]. TLR signaling in IECs upregulates these NK ligands and increases the IL-15 production, which in turn boosts the expression of co-stimulatory NK receptors such as NKG2D and NKG2C on T-IELs [96,144,145]. Additionally, the presence of inflammatory cytokines such as IFN $\gamma$  and IL-1 $\beta$  in the GI microenvironment during infections further increases NKG2D expression on T-IELs [171,172]. Moreover, stress conditions upregulate the expression of NKG2C-associated DAP12 adapter protein in TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs, enhancing their functional capabilities [96]. The interaction of NKG2D with its ligands significantly lowers the TCR activation threshold, enabling the recognition of non-cognate, low-affinity antigens or allowing operation independently of the TCR [98,145,153]. For instance, oral infection with *Salmonella enterica* serovar Typhimurium triggers the expansion of TCR $\gamma\delta$ +CD8+ T-IELs. This response is accompanied by increased NKG2D expression on these T-IELs and upregulation of MULT1 on IECs, which enables T-IELs to destroy infected intestinal cells [136]. Blocking NKG2D or reducing these T-IELs decreases the clearance of pathogens such as *Salmonella* from the intestine and other tissues [136]. Furthermore, HLA-E molecules presenting pathogen-derived peptide ligands under stress conditions could further activate NKG2C, intensifying the cytotoxic responses of T-IELs during infections [146]. Apart from NK receptors, HVEM serves as a ligand for CD160 co-stimulatory receptor expressed on most T-IELs, the interaction that is essential for pathogen defense. This interaction induces a cytotoxic and inflammatory response in TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs

against pathogens in a TCR-dependent manner [147,173]. Moreover, this HVEM-CD160 interaction activates the HVEM-Stat3-REG3 pathway on IECs, which promotes the production of RegIII $\gamma$  AMP by IECs to eliminate pathogens [148]. Similarly, intestinal LIGHT ligand, which is also expressed on T-IELs, binding to HVEM has been known to be crucial in defense against pathogens such as *Salmonella Typhimurium* [102], probably also functioning through the HVEM-Stat3-REG3 pathway on IECs. Additionally, TCR signaling increases the expression of OX40 co-stimulatory receptor on CD8<sup>+</sup> T-IELs within 24 hours, otherwise absent under steady-state conditions, leading to pathogen clearance by enhancing cytokine production and cell-mediated cytotoxicity [149]. This study also noted the expression of both OX40 and its ligand OX40L on T-IELs [149], suggesting a mechanism to stimulate each other for pathogen clearance. Besides these co-receptors, it has been recently discovered that TCR $\alpha\beta$ +CD4+CD8 $\alpha\alpha$ + T-IELs can express TLR-4, a receptor not previously reported on T-IELs [45]. During sepsis, eCIRP from damaged cells binds to TLR-4, which triggers these T-IELs to produce granzyme B and perforins that lead to the death of IECs through cytotoxicity [45]. However, it is not yet known whether TLR-4 on TCR $\alpha\beta$ +CD4+CD8 $\alpha\alpha$ + T-IELs directly recognizes pathogen ligands such as LPS.

In summary, T-IELs play an important role in the immune defense of the intestinal mucosa by engaging IEC-dependent pathways, TCR-dependent mechanisms, and co-receptor interactions to combat various pathogens. Future research could focus on understanding the precise mechanisms by which T-IELs recognize pathogens, including the role of TCRs and the potential functions of PRRs on T-IELs in detecting specific PAMPs. Studies could also examine co-receptors activity in diseases, investigating changes in their expression during intestinal infections and inflammation, as well as the availability and distribution of their ligands in the intestinal epithelium. Investigating these mechanisms could lead to novel therapeutic strategies, including targeting TCR or co-receptor pathways and enhancing their ligand expression, to boost immune responses against intestinal pathogens and to improve clinical outcomes in GI diseases.

### 3.2. Chronic Diseases

T-IELs play a dual role in chronic illnesses, contributing to both disease progression and immune regulation depending on interactions between available ligands and expressed T-IEL receptors in the microenvironment. In chronic inflammatory conditions such as celiac disease (CeD) and inflammatory bowel disease (IBD), persistent IL-15 production by IECs induces major functional changes in T-IELs. IL-15 upregulates NK receptors, such as NKG2D, on TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs [96,98,152,153]. These receptors recognize stress-induced ligands, including HLA-E and MICA, expressed on damaged IECs. This ligand-receptor interaction drives TCR-independent T-IEL activation, triggering a cytotoxic response that exacerbates inflammation and tissue damage [174–179]. Recruitment of T-IELs to inflammatory sites via the CXCR3/CXCL10 chemokine axis further amplifies inflammation and perpetuates disease progression [180]. In cancer, T-IELs exhibit both protective and pathogenic roles depending on the microenvironment. Stress-induced ligands such as MICA and ULBP on tumor cells are recognized by NKG2D on T-IELs, initiating FasL-mediated cytotoxicity and direct tumor killing [154]. Additionally, CD103 on T-IELs interacts with E-cadherin on tumor cells to retain T-IELs in the tumor microenvironment and enhance their anti-tumor ability [155]. However, subsets of T-IELs expressing co-inhibitory receptors such as PD-1 or inhibited through galectin-3-mediated LAG-3 signaling may promote tumor growth [181,182]. The balance between T-IEL-mediated tumor suppression and immune evasion plays a critical role in shaping cancer progression. Overall, T-IELs play a central role in chronic disease pathogenesis, with their functions shifting between

maintaining immune homeostasis and driving pathology based on microenvironmental signals and ligand-receptor interactions.

### 3.2.1. Celiac Disease

Celiac disease (CeD) is an immune-mediated enteropathy driven by an aberrant immune response to dietary gluten. The disease is strongly associated with HLA-DQ2 and HLA-DQ8, which present deamidated gluten peptides as foreign ligands to CD4<sup>+</sup> T cells [183,184]. Once activated, these CD4<sup>+</sup> T cells secrete pro-inflammatory cytokines such as IFN $\gamma$ , leading to tissue destruction [150,151].

A hallmark of CeD is the chronic upregulation of IL-15 by IECs in the GI epithelia [150,151]. IL-15 promotes the activation of TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs by enhancing the expression of NK receptors, such as NKG2C and NKG2D, and increasing transcription of the DAP10 adapter protein [96,98,152,153]. Activation of these NK receptors enables T-IELs to kill neighboring IECs expressing stress-induced ligands, such as HLA-E and MICA [11,152,153,185]. Elevated IL-15 levels also drive the proliferation of TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs and increase their production of cytotoxic molecules, including granzyme A, granzyme B, and perforin [186]. These cytotoxic TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs are the primary mediators of tissue damage in CeD, whereas TCR $\alpha\beta$ +CD4<sup>+</sup> T-IELs do not contribute to this effect [175–179]. Notably, the cytotoxic activity of TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs in CeD does not rely on TCR recognition of gluten, as gluten-specific TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs are not observed [174]. Some TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs in CeD patients express the Nkp46, a receptor that interacts with ecto-calreticulin (ecto-CRT), a stress ligand translocated to the cell surface under endoplasmic reticulum stress. IL-15 may further upregulate Nkp46 expression on both TCR $\alpha\beta$ +CD8 $\alpha\beta$ + and TCR $\gamma\delta$ + T-IELs, amplifying their cytotoxic responses [4]. Additionally, IECs secrete CXCL10, a chemoattractant that recruits CXCR3-expressing TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs. This CXCR3/CXCL10 axis is hyperactivated in the small intestinal mucosa of untreated CeD patients, exacerbating tissue damage [180]. Moreover, CeD involves an increase in inflammatory TCR $\gamma\delta$ + T-IEL population [187,188]. Gluten exposure causes a loss of IEC-expressed BTNL3/8 and the resident V $\gamma$ 4<sup>+</sup> T-IELs, which are replaced by pro-inflammatory IFN $\gamma$ -producing V $\gamma$ 3<sup>+</sup> T-IELs [189,190]. This shift is not reversed by a gluten-free diet, indicating that chronic inflammation may permanently reconfigure the tissue-resident TCR $\gamma\delta$ + T-IEL compartment, disrupting protective IEC-T-IEL interactions [190]. In addition, TCR $\gamma\delta$ +CD8<sup>+</sup> T-IELs that suppress the cytotoxic programming of TCR $\alpha\beta$ + T-IELs via CD94/NKG2A engagement and TGF $\beta$  production, are reduced in active CeD [191].

Overall, CeD is characterized by complex immune responses driven by interactions between T-IELs and IECs. Key ligands contributing to intestinal damage and disease pathology include dietary gluten, cytokines such as IL-15, and stress-induced molecules like MICA. These factors collectively drive epithelial destruction, with TCR $\alpha\beta$ +CD8 $\alpha\beta$ + T-IELs playing a central role through their recognition and response to these ligands.

### 3.2.2. Inflammatory Bowel Disease

Inflammatory bowel disease (IBD) refers to Crohn's disease and ulcerative colitis, two chronic inflammatory diseases of the GI tract that may involve inappropriate immune responses against intestinal microbes [43]. As the lesions in IBD can be transmural, affecting the entire thickness of the bowel wall that completely removes the epithelial layer where T-IELs reside [173], it might be that the primary role of T-IELs is more in the prevention of this disease and may also contribute to its resolution, particularly through immunoregulation and healing functions. IBD shares many similarities with CeD such as overexpression of IL-15 from IECs [69]. Also, disease severity correlates with an increased number of

TCR $\gamma\delta$ <sup>+</sup> cells in the intestinal mucosa [192,193]. Moreover, decreased levels of Galectin-3 are associated with IBD [194–196], which may reduce LAG-3 co-inhibitory signaling from T-IELs, aiding inflammation. In addition, a low amount of Vitamin A, Vitamin D, and AhR has been associated with IBD [43,123]. These ligands are involved in the gut migration and maintenance of T-IELs, which indicates that the low numbers of T-IELs might lead to IBD. Despite these observations, the role of T-IELs in IBD remains complex and not fully elucidated. Recent studies have reported conflicting results regarding the presence of CD4<sup>+</sup>CD103<sup>+</sup> T cells in IBD patients [197,198], while a decrease in CD8 $\alpha\beta$ <sup>+</sup>CD103<sup>+</sup> T cells has been observed in inflamed mucosa of these patients [198]. CD4<sup>+</sup>CD103<sup>+</sup> cells in IBD patients exhibit increased production of pro-inflammatory cytokines, such as IFN $\gamma$ , IL-17A, and TNF $\alpha$  [199,200]. In contrast, CD8<sup>+</sup>CD103<sup>+</sup> cells, although reduced in number, express regulatory molecules such as IL-22, IL-26, and CCL20 [201], suggesting distinct functional roles. However, it is important to note that both T-IELs and lamina propria T cells can express CD103, making it challenging to distinguish between these populations. For instance, CD103 is expressed on > 90% of T-IELs and 45–50% of lamina propria T lymphocytes [202].

Future studies could investigate how T-IEL populations change during IBD compared to healthy conditions and how these shifts influence inflammation and tissue repair. Understanding the availability of key ligands and their roles in T-IEL development and function within inflamed tissue is essential. Additionally, examining the receptors expressed by T-IELs and their interactions with their ligands could provide insights into their functions. These findings could help develop targeted therapies to restore immune balance and promote tissue repair in IBD.

### 3.2.3. Cancer

The major ligands known to be responsible for triggering immune responses in intestinal cancers such as colon cancer are stress-induced ligands expressed on these cells, such as MICA and ULBP, which is similar to that in other chronic diseases such as CeD and IBD. This is why NK receptors play a crucial role in targeting and killing these malignant cells. For example, human TCR $\alpha\beta$ <sup>+</sup>CD8 $\alpha\beta$ <sup>+</sup> T-IELs can spontaneously kill colon cancer cell lines that express NKG2D ligands by inducing FasL-mediated cytotoxicity and producing inflammatory cytokines such as TNF $\alpha$  and IFN $\gamma$  [154]. Additionally, natural TCR $\alpha\beta$ <sup>+</sup>CD8 $\alpha\alpha$ <sup>+</sup> and TCR $\gamma\delta$ <sup>+</sup> T-IELs can kill tumor cells independently of TCR activation, through the stimulation of NKR-P1 cell receptors [203]. Moreover, TCR $\gamma\delta$ <sup>+</sup> T-IELs that express the Nkp46 receptor demonstrate strong cytotoxicity against colorectal cancer (CRC) and help reduce the risk of metastasis [204]. The use of co-receptors by T-IELs to eliminate the tumor is further supported by the fact that in a small intestinal tumor organoid model, the T-IELs eliminated these intestinal tumors in cell-to-cell contact dependent-manner through CD103/E-cadherin signal [155]. Additionally, Galectin-3, which is overexpressed in various types of epithelial cancers including CRC, is associated with decreased epithelium integrity in human colon cancer cells [182]. This may be partly due to co-inhibitory signaling through LAG-3 in T-IELs. In contrast, PD-1- and IL-17-expressing V $\gamma$ 4<sup>+</sup> and V $\gamma$ 6<sup>+</sup> T-IELs exhibit pro-tumorigenic functions, indicating the role of the PD-1 co-receptor and IL-17 in supporting cancer progression [181].

Overall, besides the stress ligands that engage NK receptors, the specific ligands of T-IELs that influence anti-tumor or pro-tumor conditions are largely unknown. Additionally, there is limited evidence of TCR $\alpha\beta$ <sup>+</sup>CD4<sup>+</sup> T-IELs' role in intestinal epithelial tumor microenvironment. Future research could focus on identifying the expression of different cancer-related receptors on T-IEL subsets and the ligands in the cancer microenvironment that can modulate the anti-tumorigenic and pro-tumorigenic activities of specific T-IEL

subsets. Clarifying these mechanisms could lead to targeted therapies that help manipulate these interactions to combat intestinal cancers more effectively.

### 3.3. Epithelial Repair and Healing

TCR $\gamma\delta$ + T-IELs utilize the junctional adhesion molecule-like protein (JAML) and CD100 pathways to stimulate key interactions crucial for epithelial healing. These ligand-receptor interactions facilitate Keratinocyte Growth Factor (KGF) production and maintenance of the epithelial barrier.

TCR $\gamma\delta$ + T-IELs express the co-stimulatory molecule, JAML, which binds to coxsackievirus and adenovirus receptor (CAR) on IECs, resulting in the upregulation of KGF, a critical factor for IEC proliferation and epithelial healing by facilitating the repair and maintenance of the intestinal epithelium [69]. CAR is part of the epithelial tight junction and aids interactions between IECs [205]. The condition under which CAR on the IECs binds to JAML on TCR $\gamma\delta$ + T-IELs to trigger epithelial repair and healing via KGF production remains unclear. Moreover, the expression of CD100, a co-receptor expressed on all colonic T-IELs, has been found to be important for KGF production, as TCR $\gamma\delta$ + T-IEL deficient in CD100 cannot produce this molecule [156]. However, KGF production could not be stimulated by CD100 cross-linking on TCR $\gamma\delta$ + T-IEL in vitro, suggesting that CD100 influences TCR $\gamma\delta$ + T-IEL functions in conjunction with other signals [156].

Overall, the restoration and maintenance of the intestinal epithelium are facilitated by critical ligand-receptor interactions. The JAML-CAR interaction notably stimulates KGF production, which is a key factor in epithelial healing [69]. Despite the importance of the CD100 co-receptor in this process, its full impact remains dependent on additional signaling pathways, which are yet unknown. Moreover, the healing and repair role of other T-IELs besides TCR $\gamma\delta$ + T-IELs has not been reported yet. Future studies could focus on understanding these complex ligand interactions in TCR $\gamma\delta$ + T-IELs and also understanding the role of TCR $\alpha\beta$ + T-IELs on healing and maintenance of the epithelial barrier to develop targeted therapies that enhance epithelial repair in diseases such as chronic intestinal inflammatory conditions.

## 4. Conclusions

In conclusion, this review highlights the complex interplay between T-IEL receptors and their corresponding ligands in the intestinal epithelium. The intricate network of ligand-receptor interactions, including TCRs and non-TCRs, such as co-inhibitory and co-stimulatory receptors, cytokine receptors, nuclear receptors, and various signaling molecules, directs T-IEL functions in both steady-state and pathological conditions. The availability of ligands, coupled with the differential expression of receptors on T-IELs within the GI microenvironment, enables these cells to maintain immune tolerance, effectively monitor and eliminate pathogens, infected or damaged cells, and malignancies, as well as contribute to tissue repair and healing.

**Author Contributions:** A.H. and Z.X. conceived and wrote this manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** This project received support from the USDA NIFA Grant 2016-67015-24948 (to Z.X.) and Grant 2019-67015-29831 (to Z.X.), the Jorgensen Foundation (to Z.X.), and the MAES program at the University of Maryland (to Z.X.).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data was created for this review.



**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

1. Soderholm, A.T.; Pedicord, V.A. Intestinal epithelial cells: At the interface of the microbiota and mucosal immunity. *Immunology* **2019**, *158*, 267–280. [\[CrossRef\]](#) [\[PubMed\]](#)
2. Xiong, Y.; Xu, G.; Chen, M.; Ma, H. Intestinal uptake and tolerance to food antigens. *Front. Immunol.* **2022**, *13*, 906122. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Ma, H.; Qiu, Y.; Yang, H. Intestinal intraepithelial lymphocytes: Maintainers of intestinal immune tolerance and regulators of intestinal immunity. *J. Leukoc. Biol.* **2021**, *109*, 339–347. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Vandereyken, M.; James, O.J.; Swamy, M. Mechanisms of activation of innate-like intraepithelial t lymphocytes. *Mucosal Immunol.* **2020**, *13*, 721–731. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Hoytema Van Konijnenburg, D.P.; Reis, B.S.; Pedicord, V.A.; Farache, J.; Victora, G.D.; Mucida, D. Intestinal epithelial and intraepithelial t cell crosstalk mediates a dynamic response to infection. *Cell* **2017**, *171*, 783–794.e713. [\[CrossRef\]](#)
6. Gui, Y.; Cheng, H.; Zhou, J.; Xu, H.; Han, J.; Zhang, D. Development and function of natural tcr+ cd8αα+ intraepithelial lymphocytes. *Front. Immunol.* **2022**, *13*, 1059042. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Pobezinsky, L.A.; Angelov, G.S.; Tai, X.; Jeurling, S.; Van Laethem, F.; Feigenbaum, L.; Park, J.-H.; Singer, A. Clonal deletion and the fate of autoreactive thymocytes that survive negative selection. *Nat. Immunol.* **2012**, *13*, 569–578. [\[CrossRef\]](#) [\[PubMed\]](#)
8. Leishman, A.J.; Gapin, L.; Capone, M.; Palmer, E.; Macdonald, H.R.; Kronenberg, M.; Cheroutre, H. Precursors of functional mhc class i- or class ii-restricted cd8αα+ t CELLS are positively selected in the thymus by agonist self-peptides. *Immunity* **2002**, *16*, 355–364. [\[CrossRef\]](#)
9. McDonald, B.D.; Bunker, J.J.; Ishizuka, I.E.; Jabri, B.; Bendelac, A. Elevated t cell receptor Signaling identifies a thymic precursor to the tcrαβ+cd4–cd8β– intraepithelial lymphocyte lineage. *Immunity* **2014**, *41*, 219–229. [\[CrossRef\]](#)
10. Di Marco Barros, R.; Roberts, N.A.; Dart, R.J.; Vantourout, P.; Jandke, A.; Nussbaumer, O.; Deban, L.; Cipolat, S.; Hart, R.; Iannitto, M.L.; et al. Epithelia use butyrophilin-like molecules to shape organ-specific γδ t cell compartments. *Cell* **2016**, *167*, 203–218.e217. [\[CrossRef\]](#) [\[PubMed\]](#)
11. McDonald, B.D.; Jabri, B.; Bendelac, A. Diverse developmental pathways of intestinal intraepithelial lymphocytes. *Nat. Rev. Immunol.* **2018**, *18*, 514–525. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Bono, M.; Tejon, G.; Flores-Santibañez, F.; Fernandez, D.; Roseblatt, M.; Sauma, D. Retinoic acid as a modulator of T cell immunity. *Nutrients* **2016**, *8*, 349. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Svensson, M.; Marsal, J.; Ericsson, A.; Carramolino, L.; Brodén, T.; Márquez, G.; Agace, W.W. Ccl25 mediates the localization of recently activated cd8αβ+ lymphocytes to the small-intestinal mucosa. *J. Clin. Investig.* **2002**, *110*, 1113–1121. [\[CrossRef\]](#)
14. Agace, W.W. T-cell recruitment to the intestinal mucosa. *Trends Immunol.* **2008**, *29*, 514–522. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Johansson-Lindbom, B.; Svensson, M.; Wurbel, M.-A.; Malissen, B.; Márquez, G.; Agace, W. Selective generation of gut tropic t cells in gut-associated lymphoid tissue (galt). *J. Exp. Med.* **2003**, *198*, 963–969. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Jaensson, E.; Uronen-Hansson, H.; Pabst, O.; Eksteen, B.; Tian, J.; Coombes, J.L.; Berg, P.-L.; Davidsson, T.; Powrie, F.; Johansson-Lindbom, B.; et al. Small intestinal cd103+ dendritic cells display unique functional properties that are conserved between mice and humans. *J. Exp. Med.* **2008**, *205*, 2139–2149. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Campbell, D.J.; Butcher, E.C. Rapid acquisition of tissue-specific homing phenotypes by cd4+ t cells activated in cutaneous or mucosal lymphoid tissues. *J. Exp. Med.* **2002**, *195*, 135–141. [\[CrossRef\]](#) [\[PubMed\]](#)
18. Staton, T.L.; Habtezion, A.; Winslow, M.M.; Sato, T.; Love, P.E.; Butcher, E.C. Cd8+ recent thymic emigrants home to and efficiently repopulate the small intestine epithelium. *Nat. Immunol.* **2006**, *7*, 482–488. [\[CrossRef\]](#) [\[PubMed\]](#)
19. Normont, A.M.; Bogatzki, L.Y.; Gantner, B.N.; Bevan, M.J. Murine ccr9, a chemokine receptor for thymus-expressed chemokine that is up-regulated following pre-tcr signaling. *J. Immunol.* **2000**, *164*, 639–648. [\[CrossRef\]](#) [\[PubMed\]](#)
20. Ruscher, R.; Lee, S.T.; Salgado, O.C.; Breed, E.R.; Osum, S.H.; Hogquist, K.A. Intestinal cd8αα iels derived from two distinct thymic precursors have staggered ontogeny. *J. Exp. Med.* **2020**, *217*, e20192336. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Siegers, G.M. Integral roles for integrins in γδ t cell function. *Front. Immunol.* **2018**, *9*, 521. [\[CrossRef\]](#)
22. Yu, S.; Bruce, D.; Froicu, M.; Weaver, V.; Cantorna, M.T. Failure of t cell homing, reduced cd4/cd8αα intraepithelial lymphocytes, and inflammation in the gut of vitamin d receptor ko mice. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 20834–20839. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Fu, H.; Ward, E.J.; Marelli-Berg, F.M. Mechanisms of t cell organotropism. *Cell Mol. Life Sci.* **2016**, *73*, 3009–3033. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Iwata, M.; Hirakiyama, A.; Eshima, Y.; Kagechika, H.; Kato, C.; Song, S.-Y. Retinoic acid imprints gut-homing specificity on t cells. *Immunity* **2004**, *21*, 527–538. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Hada, A.; Li, L.; Kandel, A.; Jin, Y.; Xiao, Z. Characterization of bovine intraepithelial t lymphocytes in the gut. *Pathogens* **2023**, *12*, 1173. [\[CrossRef\]](#) [\[PubMed\]](#)

26. Van Kaer, L.; Olivares-Villagómez, D. Development, homeostasis, and functions of intestinal intraepithelial lymphocytes. *J. Immunol.* **2018**, *200*, 2235–2244. [[CrossRef](#)] [[PubMed](#)]
27. Lockhart, A.; Mucida, D.; Bilate, A.M. Intraepithelial lymphocytes of the intestine. *Annu. Rev. Immunol.* **2024**, *42*, 289–316. [[CrossRef](#)] [[PubMed](#)]
28. Gascoigne, N.R.J.; Rybakina, V.; Acuto, O.; Brzostek, J. Tcr signal strength and t cell development. *Annu. Rev. Cell Dev. Biol.* **2016**, *32*, 327–348. [[CrossRef](#)] [[PubMed](#)]
29. Szeto, C.; Lobos, C.A.; Nguyen, A.T.; Gras, S. Tcr recognition of peptide–mhc-i: Rule makers and breakers. *Int. J. Mol. Sci.* **2020**, *22*, 68. [[CrossRef](#)]
30. Sundberg, E.J.; Deng, L.; Mariuzza, R.A. Tcr recognition of peptide/mhc class ii complexes and superantigens. *Semin. Immunol.* **2007**, *19*, 262–271. [[CrossRef](#)]
31. Deseke, M.; Prinz, I. Ligand recognition by the  $\gamma\delta$  tcr and discrimination between homeostasis and stress conditions. *Cell. Mol. Immunol.* **2020**, *17*, 914–924. [[CrossRef](#)]
32. Ruscher, R.; Kummer, R.L.; Lee, Y.J.; Jameson, S.C.; Hogquist, K.A. CD8 $\alpha\alpha$  intraepithelial lymphocytes arise from two main thymic precursors. *Nat. Immunol.* **2017**, *18*, 771–779. [[CrossRef](#)]
33. Das, G.; Gould, D.S.; Augustine, M.M.; Fragoso, G.; Scitto, E.; Stroynowski, I.; Van Kaer, L.; Schust, D.J.; Ploegh, H.; Janeway, C.A. Qa-2-dependent selection of cd8 $\alpha/\alpha$  t cell receptor  $\alpha/\beta$ + cells in murine intestinal intraepithelial lymphocytes. *J. Exp. Med.* **2000**, *192*, 1521–1528. [[CrossRef](#)] [[PubMed](#)]
34. Gapin, L.; Cheroutre, H.; Kronenberg, M. Cutting edge: Tcr alpha beta+ cd8 alpha alpha+ t cells are found in intestinal intraepithelial lymphocytes of mice that lack classical mhc class I molecules. *J. Immunol.* **1999**, *163*, 4100–4104. [[CrossRef](#)] [[PubMed](#)]
35. Olivares-Villagómez, D.; Van Kaer, L. Intestinal intraepithelial lymphocytes: Sentinels of the mucosal barrier. *Trends Immunol.* **2018**, *39*, 264–275. [[CrossRef](#)] [[PubMed](#)]
36. Ruscher, R.; Hogquist, K.A. Development, ontogeny, and maintenance of tcr $\alpha\beta$ (+) cd8 $\alpha\alpha$  iel. *Curr. Opin. Immunol.* **2019**, *58*, 83–88. [[CrossRef](#)] [[PubMed](#)]
37. Lin, T.; Matsuzaki, G.; Kenai, H.; Nomoto, K. Progenies of fetal thymocytes are the major source of cd4-cd8+ alpha alpha intestinal intraepithelial lymphocytes early in ontogeny. *Eur. J. Immunol.* **1994**, *24*, 1785–1791. [[CrossRef](#)]
38. Watt, H.J.; Chawla, A.S.; Lamoliatte, F.; Pryde, S.; Knatko, E.; Rasmussen, K.D.; Bending, D.; Swamy, M. Rewiring of the tcr signalosome in natural intestinal Intraepithelial t lymphocytes drives non-deletional tolerance. *bioRxiv* **2023**, preprint. [[CrossRef](#)]
39. Qiu, Y.; Pu, A.; Zheng, H.; Liu, M.; Chen, W.; Wang, W.; Xiao, W.; Yang, H. Tlr2-dependent signaling for il-15 production is essential for the homeostasis of intestinal intraepithelial lymphocytes. *Mediat. Inflamm.* **2016**, *2016*, 4281865. [[CrossRef](#)] [[PubMed](#)]
40. Mayassi, T.; Jabri, B. Human intraepithelial lymphocytes. *Mucosal Immunol.* **2018**, *11*, 1281–1289. [[CrossRef](#)] [[PubMed](#)]
41. Kaneko, M.; Mizunuma, T.; Takimoto, H.; Kumazawa, Y. Development of tcr $\alpha\beta$  cd8 $\alpha\alpha$  intestinal intraepithelial lymphocytes is promoted by interleukin-15-producing epithelial cells constitutively stimulated by gram-negative bacteria via tlr4. *Biol. Pharm. Bull.* **2004**, *27*, 883–889. [[CrossRef](#)]
42. Yu, Q.; Tang, C.; Xun, S.; Yajima, T.; Takeda, K.; Yoshikai, Y. Myd88-dependent signaling for il-15 production plays an important role in maintenance of cd8 $\alpha\alpha$  tcr $\alpha\beta$  and tcr $\gamma\delta$  intestinal intraepithelial lymphocytes. *J. Immunol.* **2006**, *176*, 6180–6185. [[CrossRef](#)] [[PubMed](#)]
43. Hu, M.D.; Edelblum, K.L. Sentinels at the frontline: The role of intraepithelial lymphocytes in inflammatory bowel disease. *Curr. Pharmacol. Rep.* **2017**, *3*, 321–334. [[CrossRef](#)] [[PubMed](#)]
44. Klose, C.S.N.; Blatz, K.; D’Hargues, Y.; Hernandez, P.P.; Kofoed-Nielsen, M.; Ripka, J.F.; Ebert, K.; Arnold, S.J.; Diefenbach, A.; Palmer, E.; et al. The transcription factor t-bet is induced by il-15 and thymic agonist selection and controls cd8 $\alpha\alpha$ + intraepithelial Lymphocyte development. *Immunity* **2014**, *41*, 230–243. [[CrossRef](#)]
45. Akama, Y.; Muraio, A.; Aziz, M.; Wang, P. Extracellular cirp induces cd4cd8 $\alpha\alpha$  intraepithelial lymphocyte cytotoxicity in sepsis. *Mol. Med.* **2024**, *30*, 17. [[CrossRef](#)] [[PubMed](#)]
46. Bousbaine, D.; Fisch, L.I.; London, M.; Bhagchandani, P.; Rezende de Castro, T.B.; Mimee, M.; Olesen, S.; Reis, B.S.; VanInsberghe, D.; Bortolatto, J.; et al. A conserved bacteroidetes antigen induces anti-inflammatory intestinal t lymphocytes. *Science* **2022**, *377*, 660–666. [[CrossRef](#)]
47. Tuganbaev, T.; Mor, U.; Bashiardes, S.; Liwinski, T.; Nobs, S.P.; Leshem, A.; Dori-Bachash, M.; Thaïss, C.A.; Pinker, E.Y.; Ratiner, K.; et al. Diet diurnally regulates small intestinal microbiome-epithelial-immune homeostasis and enteritis. *Cell* **2020**, *182*, 1441–1459.e1421. [[CrossRef](#)]
48. Brabec, T.; Schwarzer, M.; Kováčová, K.; Dobešová, M.; Schierová, D.; Březina, J.; Pacáková, I.; Šrůtková, D.; Ben-Nun, O.; Goldfarb, Y.; et al. Epithelial antigen presentation controls commensal-specific intraepithelial t-cells in the gut. *bioRxiv* **2022**, preprint. [[CrossRef](#)]

49. Kuhn, K.A.; Schulz, H.M.; Regner, E.H.; Severs, E.L.; Hendrickson, J.D.; Mehta, G.; Whitney, A.K.; Ir, D.; Ohri, N.; Robertson, C.E.; et al. Bacteroidales recruit il-6-producing intraepithelial lymphocytes in the colon to promote barrier integrity. *Mucosal Immunol.* **2018**, *11*, 357–368. [[CrossRef](#)] [[PubMed](#)]
50. Li, C.; Lanasa, D.; Park, J.-H. Pathways and mechanisms of cd4+cd8αα+ intraepithelial t cell development. *Trends Immunol.* **2024**, *45*, 288–302. [[CrossRef](#)]
51. Cervantes-Barragan, L.; Chai, J.N.; Tianero, M.D.; Di Luccia, B.; Ahern, P.P.; Merriman, J.; Cortez, V.S.; Caparon, M.G.; Donia, M.S.; Gilfillan, S.; et al. Lactobacillus reuteri induces gut intraepithelial cd4(+)cd8αα(+) t cells. *Science* **2017**, *357*, 806–810. [[CrossRef](#)]
52. Kawaguchi, M.; Nanno, M.; Umesaki, Y.; Matsumoto, S.; Okada, Y.; Cai, Z.; Shimamura, T.; Matsuoka, Y.; Ohwaki, M.; Ishikawa, H. Cytolytic activity of intestinal intraepithelial lymphocytes in germ-free mice is strain dependent and determined by t cells expressing gamma delta t-cell antigen receptors. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 8591–8594. [[CrossRef](#)] [[PubMed](#)]
53. Ismail, A.S.; Severson, K.M.; Vaishnav, S.; Behrendt, C.L.; Yu, X.; Benjamin, J.L.; Ruhn, K.A.; Hou, B.; Defranco, A.L.; Yarovinsky, E.; et al. γδ intraepithelial lymphocytes are essential mediators of host–microbial homeostasis at the intestinal mucosal surface. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 8743–8748. [[CrossRef](#)] [[PubMed](#)]
54. Ismail, A.S.; Behrendt, C.L.; Hooper, L.V. Reciprocal interactions between commensal bacteria and gamma delta intraepithelial lymphocytes during mucosal injury. *J. Immunol.* **2009**, *182*, 3047–3054. [[CrossRef](#)]
55. Chen, B.; Ni, X.; Sun, R.; Zeng, B.; Wei, H.; Tian, Z.; Wei, H. Commensal bacteria-dependent cd8αβ+ t cells in the intestinal epithelium produce antimicrobial peptides. *Front. Immunol.* **2018**, *9*, 1065. [[CrossRef](#)] [[PubMed](#)]
56. Masopust, D.; Jiang, J.; Shen, H.; Lefrançois, L. Direct analysis of the dynamics of the intestinal mucosa cd8 t cell response to systemic virus infection1. *J. Immunol.* **2001**, *166*, 2348–2356. [[CrossRef](#)]
57. Masopust, D.; Vezys, V.; Marzo, A.L.; Lefrançois, L. Preferential localization of effector memory cells in nonlymphoid tissue. *Science* **2001**, *291*, 2413–2417. [[CrossRef](#)] [[PubMed](#)]
58. Jung, J.; Surh, C.D.; Lee, Y.J. Microbial colonization at early life promotes the development of diet-induced cd8αβ Intraepithelial t cells. *Mol Cells* **2019**, *42*, 313–320. [[CrossRef](#)]
59. Vaishnav, S.; Yamamoto, M.; Severson, K.M.; Ruhn, K.A.; Yu, X.; Koren, O.; Ley, R.; Wakeland, E.K.; Hooper, L.V. The antibacterial lectin regiii promotes the spatial segregation of microbiota and host in the intestine. *Science* **2011**, *334*, 255–258. [[CrossRef](#)] [[PubMed](#)]
60. Stögerer, T.; Stäger, S. Innate immune sensing by cells of the adaptive immune system. *Front. Immunol.* **2020**, *11*, 1081. [[CrossRef](#)]
61. Latthe, M.; Terry, L.; Macdonald, T.T. High frequency of cd8αα homodimer-bearing t cells in human fetal intestine. *Eur. J. Immunol.* **1994**, *24*, 1703–1705. [[CrossRef](#)] [[PubMed](#)]
62. Kuo, S.; El Guindy, A.; Panwala, C.M.; Hagan, P.M.; Camerini, V. Differential appearance of t cell subsets in the large and small intestine of neonatal mice. *Pediatr. Res.* **2001**, *49*, 543–551. [[CrossRef](#)] [[PubMed](#)]
63. Torow, N.; Yu, K.; Hassani, K.; Freitag, J.; Schulz, O.; Basic, M.; Brennecke, A.; Sparwasser, T.; Wagner, N.; Bleich, A.; et al. Active suppression of intestinal cd4+ tcrαβ+ T-lymphocyte maturation during the postnatal period. *Nat. Commun.* **2015**, *6*, 7725. [[CrossRef](#)]
64. Manzano, M.; Abadía-Molina, A.C.; García-Olivares, E.; Gil, A.; Rueda, R. Absolute counts and distribution of lymphocyte subsets in small intestine of balb/c mice change during weaning. *J. Nutr.* **2002**, *132*, 2757–2762. [[CrossRef](#)] [[PubMed](#)]
65. Helgeland, L.; Brandtzaeg, P.; Rolstad, B.; Vaage, J.T. Sequential development of intraepithelial γδ and αβ t lymphocytes expressing cd8αβ in neonatal rat intestine: Requirement for the thymus. *Immunology* **1997**, *92*, 447–456. [[CrossRef](#)]
66. Steege, J.C.; Buurman, W.A.; Forget, P.P. The neonatal development of intraepithelial and lamina propria lymphocytes in the murine small intestine. *Dev. Immunol.* **1997**, *5*, 121–128. [[CrossRef](#)] [[PubMed](#)]
67. Umesaki, Y.; Setoyama, H.; Matsumoto, S.; Okada, Y. Expansion of alpha beta t-cell receptor-bearing intestinal intraepithelial lymphocytes after microbial colonization in germ-free mice and its independence from thymus. *Immunology* **1993**, *79*, 32–37.
68. Lockhart, A.; Reed, A.; Rezende De Castro, T.; Herman, C.; Campos Canesso, M.C.; Mucida, D. Dietary protein shapes the profile and repertoire of intestinal cd4+ t cells. *J. Exp. Med.* **2023**, *220*, e20221816. [[CrossRef](#)] [[PubMed](#)]
69. Cheroutre, H.; Lambolez, F.; Mucida, D. The light and dark sides of intestinal intraepithelial lymphocytes. *Nat. Rev. Immunol.* **2011**, *11*, 445–456. [[CrossRef](#)]
70. Love, P.E.; Hayes, S.M. Itam-mediated signaling by the t-cell antigen receptor. *Cold Spring Harb. Perspect. Biol.* **2010**, *2*, a002485. [[CrossRef](#)] [[PubMed](#)]
71. Nambiar, M.P.; Fisher, C.U.; Kumar, A.; Tsokos, C.G.; Warke, V.G.; Tsokos, G.C. Forced expression of the fc receptor gamma-chain renders human t cells hyperresponsive to tcr/cd3 stimulation. *J. Immunol.* **2003**, *170*, 2871–2876. [[CrossRef](#)] [[PubMed](#)]
72. Németh, T.; Futosi, K.; Szabó, M.; Aradi, P.; Saito, T.; Mócsai, A.; Jakus, Z. Importance of fc receptor γ-chain itam tyrosines in neutrophil activation and in vivo autoimmune arthritis. *Front. Immunol.* **2019**, *10*, 252. [[CrossRef](#)] [[PubMed](#)]
73. Meng, X.; Jing, R.; Qian, L.; Zhou, C.; Sun, J. Engineering cytoplasmic signaling of cd28ζ cars for improved therapeutic functions. *Front. Immunol.* **2020**, *11*, 1046. [[CrossRef](#)] [[PubMed](#)]

74. Balagopalan, L.; Kortum, R.L.; Coussens, N.P.; Barr, V.A.; Samelson, L.E. The linker for activation of t cells (lat) signaling hub: From signaling complexes to microclusters. *J. Biol. Chem.* **2015**, *290*, 26422–26429. [[CrossRef](#)] [[PubMed](#)]
75. Brenes, A.J.; Vandereyken, M.; James, O.J.; Watt, H.; Hukelmann, J.; Spinelli, L.; Dikovskaya, D.; Lamond, A.I.; Swamy, M. Tissue environment, not ontogeny, defines murine intestinal intraepithelial t lymphocytes. *eLife* **2021**, *10*, e70055. [[CrossRef](#)]
76. Denning, T.L.; Granger, S.W.; Mucida, D.; Graddy, R.; Leclercq, G.; Zhang, W.; Honey, K.; Rasmussen, J.P.; Cheroutre, H.; Rudensky, A.Y.; et al. Mouse tcr $\alpha$ beta+cd8 $\alpha$ alpha intraepithelial lymphocytes express genes that down-regulate their antigen reactivity and suppress immune responses. *J. Immunol.* **2007**, *178*, 4230–4239. [[CrossRef](#)] [[PubMed](#)]
77. Bas, A.; Swamy, M.; Abeler-Dörner, L.; Williams, G.; Pang, D.J.; Barbee, S.D.; Hayday, A.C. Butyrophilin-like 1 encodes an enterocyte protein that selectively regulates functional interactions with t lymphocytes. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 4376–4381. [[CrossRef](#)]
78. Melandri, D.; Zlatareva, I.; Chaleil, R.A.G.; Dart, R.J.; Chancellor, A.; Nussbaumer, O.; Polyakova, O.; Roberts, N.A.; Wesch, D.; Kabelitz, D.; et al. The  $\gamma\delta$ tr combines innate immunity with adaptive immunity by utilizing spatially distinct regions for agonist selection and antigen responsiveness. *Nat. Immunol.* **2018**, *19*, 1352–1365. [[CrossRef](#)]
79. Bilate, A.M.; London, M.; Castro, T.B.R.; Mesin, L.; Bortolatto, J.; Kongthong, S.; Harnagel, A.; Vitoria, G.D.; Mucida, D. T cell receptor is required for differentiation, but not maintenance, of intestinal cd4+ intraepithelial lymphocytes. *Immunity* **2020**, *53*, 1001–1014.e1020. [[CrossRef](#)] [[PubMed](#)]
80. Reis, B.S.; van Konijnenburg DP, H.; Grivennikov, S.I.; Mucida, D. Transcription factor t-bet regulates intraepithelial lymphocyte functional maturation. *Immunity* **2014**, *41*, 244–256. [[CrossRef](#)]
81. Konkel, J.E.; Maruyama, T.; Carpenter, A.C.; Xiong, Y.; Zamarron, B.F.; Hall, B.E.; Kulkarni, A.B.; Zhang, P.; Bosselut, R.; Chen, W. Control of the development of cd8 $\alpha\alpha$ + intestinal intraepithelial lymphocytes by tgf- $\beta$ . *Nat. Immunol.* **2011**, *12*, 312–319. [[CrossRef](#)] [[PubMed](#)]
82. Van Kaer, L.; Algood HM, S.; Singh, K.; Parekh, V.V.; Greer, M.J.; Piazuelo, M.B.; Weitkamp, J.-H.; Matta, P.; Chaturvedi, R.; Wilson, K.T.; et al. Cd8 $\alpha\alpha$ + innate-type lymphocytes in the intestinal epithelium mediate mucosal immunity. *Immunity* **2014**, *41*, 451–464. [[CrossRef](#)] [[PubMed](#)]
83. Reis, B.S.; Rogoz, A.; Costa-Pinto, F.A.; Taniuchi, I.; Mucida, D. Mutual expression of the transcription factors runx3 and thpok regulates intestinal cd4+ t cell immunity. *Nat. Immunol.* **2013**, *14*, 271–280. [[CrossRef](#)] [[PubMed](#)]
84. Huang, Y.; Park, Y.; Wang-Zhu, Y.; Larange, A.; Arens, R.; Bernardo, I.; Olivares-Villagómez, D.; Herndler-Brandstetter, D.; Abraham, N.; Grubeck-Loebenstien, B.; et al. Mucosal memory cd8+ t cells are selected in the periphery by an mhc class I molecule. *Nat. Immunol.* **2011**, *12*, 1086–1095. [[CrossRef](#)]
85. Olivares-Villagómez, D.; Mendez-Fernandez, Y.V.; Parekh, V.V.; Lalani, S.; Vincent, T.L.; Cheroutre, H.; Van Kaer, L. Thymus leukemia antigen controls intraepithelial lymphocyte function and inflammatory bowel disease. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17931–17936. [[CrossRef](#)] [[PubMed](#)]
86. Jarry, A.; Cerf-Bensussan, N.; Brousse, N.; Selz, F.; Guy-Grand, D. Subsets of cd3+ (t cell receptor alpha/beta or gamma/delta) and cd3- lymphocytes isolated from normal human gut epithelium display phenotypical features different from their counterparts in peripheral blood. *Eur. J. Immunol.* **1990**, *20*, 1097–1103. [[CrossRef](#)] [[PubMed](#)]
87. Beumer-Chuwonpad, A.; Behr, F.M.; Van Alphen, F.P.J.; Kragten, N.A.M.; Hoogendijk, A.J.; Van Den Biggelaar, M.; Van Gisbergen, K.P.J.M. Intestinal tissue-resident memory t cells maintain distinct identity from circulating memory T cells after in vitro restimulation. *Eur. J. Immunol.* **2024**, *54*, 2350873. [[CrossRef](#)] [[PubMed](#)]
88. Cabinian, A.; Sinsimer, D.; Tang, M.; Jang, Y.; Choi, B.; Laouar, Y.; Laouar, A. Gut symbiotic microbes imprint intestinal immune cells with the innate receptor slamf4 which contributes to gut immune protection against enteric pathogens. *Gut* **2018**, *67*, 847–859. [[CrossRef](#)]
89. Kojima, T.; Tsuchiya, K.; Ikemizu, S.; Yoshikawa, S.; Yamanishi, Y.; Watanabe, M.; Karasuyama, H. Novel cd200 homologues isec1 and isec2 are gastrointestinal secretory cell-specific ligands of inhibitory receptor cd200r. *Sci. Rep.* **2016**, *6*, 36457. [[CrossRef](#)]
90. Bülck, C.; Nyström, E.E.; Koudelka, T.; Mannbar-Frahm, M.; Andresen, G.; Radhouani, M.; Tran, F.; Scharfenberg, F.; Schrell, F.; Armbrust, F.; et al. Proteolytic processing of galectin-3 by meprin metalloproteases is crucial for host-microbiome homeostasis. *Sci. Adv.* **2023**, *9*, eadf4055. [[CrossRef](#)]
91. Tsai, H.-F.; Wu, C.-S.; Chen, Y.-L.; Liao, H.-J.; Chyuan, I.-T.; Hsu, P.-N. Galectin-3 suppresses mucosal inflammation and reduces disease severity in experimental colitis. *J. Mol. Med.* **2016**, *94*, 545–556. [[CrossRef](#)]
92. Shi, A.-P.; Tang, X.-Y.; Xiong, Y.-L.; Zheng, K.-F.; Liu, Y.-J.; Shi, X.-G.; Lv, Y.; Jiang, T.; Ma, N.; Zhao, J.-B. Immune checkpoint lag3 and its ligand fg11 in cancer. *Front. Immunol.* **2022**, *12*, 785091. [[CrossRef](#)]
93. Heuberger, C.; Pott, J.; Maloy, K.J. Why do intestinal epithelial cells express mhc class II? *Immunology* **2021**, *162*, 357–367. [[CrossRef](#)] [[PubMed](#)]
94. Taveirne, S.; Filtjens, J.; Van Ammel, E.; De Colvenaer, V.; Kerre, T.; Taghon, T.; Vandekerckhove, B.; Plum, J.; Held, W.; Leclercq, G. Inhibitory receptors specific for mhc class I educate murine nk cells but not cd8 $\alpha\alpha$  intestinal intraepithelial t lymphocytes. *Blood* **2011**, *118*, 339–347. [[CrossRef](#)] [[PubMed](#)]



95. Rosen, D.B.; Araki, M.; Hamerman, J.A.; Chen, T.; Yamamura, T.; Lanier, L.L. A Structural Basis for the Association of dap12 with mouse, but not human, nkg2d. *J. Immunol.* **2004**, *173*, 2470–2478. [[CrossRef](#)]
96. Meresse, B.; Curran, S.A.; Ciszewski, C.; Orbelyan, G.; Setty, M.; Bhagat, G.; Lee, L.; Tretiakova, M.; Semrad, C.; Kistner, E.; et al. Reprogramming of ctls into natural killer-like cells in celiac disease. *J. Exp. Med.* **2006**, *203*, 1343–1355. [[CrossRef](#)]
97. Wu, J.; Song, Y.; Bakker, A.B.H.; Bauer, S.; Spies, T.; Lanier, L.L.; Phillips, J.H. An activating immunoreceptor complex formed by nkg2d and dap10. *Science* **1999**, *285*, 730–732. [[CrossRef](#)] [[PubMed](#)]
98. Meresse, B.; Chen, Z.; Ciszewski, C.; Tretiakova, M.; Bhagat, G.; Krausz, T.N.; Raulet, D.H.; Lanier, L.L.; Groh, V.; Spies, T. Coordinated induction by il15 of a tcr-independent nkg2d signaling pathway converts ctl into lymphokine-activated killer cells in celiac disease. *Immunity* **2004**, *21*, 357–366. [[CrossRef](#)] [[PubMed](#)]
99. Tan, G.; Spillane, K.M.; Maher, J. The role and regulation of the nkg2d/nkg2d ligand system in cancer. *Biology* **2023**, *12*, 1079. [[CrossRef](#)]
100. Mistry, A.R.; O’Callaghan, C.A. Regulation of ligands for the activating receptor nkg2d. *Immunology* **2007**, *121*, 439–447. [[CrossRef](#)] [[PubMed](#)]
101. Lodolce, J.P.; Boone, D.L.; Chai, S.; Swain, R.E.; Dassopoulos, T.; Trettin, S.; Ma, A. Il-15 receptor maintains lymphoid homeostasis by supporting lymphocyte homing and proliferation. *Immunity* **1998**, *9*, 669–676. [[CrossRef](#)] [[PubMed](#)]
102. Seo, G.Y.; Takahashi, D.; Wang, Q.; Mikulski, Z.; Chen, A.; Chou, T.F.; Marcovecchio, P.; McArdle, S.; Sethi, A.; Shui, J.W.; et al. Epithelial hvem maintains intraepithelial t cell survival and contributes to host protection. *Sci. Immunol.* **2022**, *7*, eabm6931. [[CrossRef](#)]
103. Abdalqadir, N.; Adeli, K. Glp-1 and glp-2 orchestrate intestine integrity, gut microbiota, and immune system crosstalk. *Microorganisms* **2022**, *10*, 2061. [[CrossRef](#)] [[PubMed](#)]
104. Yusta, B.; Baggio, L.L.; Koehler, J.; Holland, D.; Cao, X.; Pinnell, L.J.; Johnson-Henry, K.C.; Yeung, W.; Surette, M.G.; Bang, K.W.A.; et al. Glp-1r agonists modulate enteric immune responses through the intestinal intraepithelial lymphocyte glp-1r. *Diabetes* **2015**, *64*, 2537–2549. [[CrossRef](#)]
105. He, S.; Kahles, F.; Rattik, S.; Nairz, M.; Mcalpine, C.S.; Anzai, A.; Selgrade, D.; Fenn, A.M.; Chan, C.T.; Mindur, J.E.; et al. Gut intraepithelial t cells calibrate metabolism and accelerate cardiovascular disease. *Nature* **2019**, *566*, 115–119. [[CrossRef](#)]
106. Edelblum, K.L.; Shen, L.; Weber, C.R.; Marchiando, A.M.; Clay, B.S.; Wang, Y.; Prinz, I.; Malissen, B.; Sperling, A.I.; Turner, J.R. Dynamic migration of  $\gamma\delta$  intraepithelial lymphocytes requires occludin. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 7097–7102. [[CrossRef](#)] [[PubMed](#)]
107. Parsa, R.; London, M.; Rezende de Castro, T.B.; Reis, B.; Buissant des Amorie, J.; Smith, J.G.; Mucida, D. Newly recruited intraepithelial ly6a(+)/ccr9(+)/cd4(+) t cells protect against enteric viral infection. *Immunity* **2022**, *55*, 1234–1249.e1236. [[CrossRef](#)] [[PubMed](#)]
108. Lebrero-Fernández, C.; Bergström, J.H.; Pelaseyed, T.; Bas-Forsberg, A. Murine butyrophilin-like 1 and btl6 Form heteromeric complexes in small intestinal epithelial cells and promote proliferation of local t lymphocytes. *Front. Immunol.* **2016**, *7*, 1. [[CrossRef](#)] [[PubMed](#)]
109. Lebrero-Fernández, C.; Bas-Forsberg, A. The ontogeny of butyrophilin-like (btl) 1 and btl6 in murine small intestine. *Sci. Rep.* **2016**, *6*, 31524. [[CrossRef](#)] [[PubMed](#)]
110. Herrmann, T.; Karunakaran, M.M. Butyrophilins:  $\gamma\delta$  t cell receptor ligands, immunomodulators and more. *Front. Immunol.* **2022**, *13*, 876493. [[CrossRef](#)] [[PubMed](#)]
111. Golovchenko, N.; Xu, W.; Galan, M.; Edelblum, K. Loss of  $\gamma\delta$  intraepithelial lymphocytes and reduced immunosurveillance of the epithelial barrier precedes the onset of crohn’s disease-like ileitis. *FASEB J.* **2022**, *36*, r4615. [[CrossRef](#)]
112. Helgeland, L.; Dissen, E.; Dai, K.-Z.; Midtvedt, T.; Brandtzaeg, P.; Vaage, J.T. Microbial colonization induces oligoclonal expansions of intraepithelial cd8 t cells in the gut. *Eur. J. Immunol.* **2004**, *34*, 3389–3400. [[CrossRef](#)] [[PubMed](#)]
113. Helgeland, L.; Vaage, J.T.; Rolstad, B.; Midtvedt, T.; Brandtzaeg, P. Microbial colonization influences composition and t-cell receptor  $v\beta$  repertoire of intraepithelial lymphocytes in rat intestine. *Immunology* **1996**, *89*, 494–501. [[CrossRef](#)] [[PubMed](#)]
114. Helgeland, L.; Johansen, F.-E.; Utgaard, J.O.; Vaage, J.T.; Brandtzaeg, P. Oligoclonality of rat intestinal intraepithelial t lymphocytes: Overlapping tcr  $\beta$ -chain repertoires in the cd4 single-positive and cd4/cd8 double-positive subsets. *J. Immunol.* **1999**, *162*, 2683–2692. [[CrossRef](#)]
115. Regnault, A.; Levraud, J.-P.; Lim, A.; Six, A.; Moreau, C.; Cumano, A.; Kourilsky, P. The expansion and selection of t cell receptor  $\alpha\beta$  intestinal intraepithelial t cell clones. *Eur. J. Immunol.* **1996**, *26*, 914–921. [[CrossRef](#)] [[PubMed](#)]
116. Regnault, A.; Cumano, A.; Vassalli, P.; Guy-Grand, D.; Kourilsky, P. Oligoclonal repertoire of the cd8 alpha alpha and the cd8 alpha beta tcr-alpha/beta murine intestinal intraepithelial t lymphocytes: Evidence for the random emergence of t cells. *J. Exp. Med.* **1994**, *180*, 1345–1358. [[CrossRef](#)]
117. Suzuki, H.; Jeong, K.I.; Itoh, K.; Doi, K. Regional variations in the distributions of small intestinal intraepithelial lymphocytes in germ-free and specific pathogen-free mice. *Exp. Mol. Pathol.* **2002**, *72*, 230–235. [[CrossRef](#)]



118. Imaoka, A.; Matsumoto, S.; Setoyama, H.; Okada, Y.; Umesaki, Y. Proliferative recruitment of intestinal intraepithelial lymphocytes after microbial colonization of germ-free mice. *Eur. J. Immunol.* **1996**, *26*, 945–948. [\[CrossRef\]](#)
119. Jiang, W.; Wang, X.; Zeng, B.; Liu, L.; Tardivel, A.; Wei, H.; Han, J.; Macdonald, H.R.; Tschopp, J.; Tian, Z.; et al. Recognition of gut microbiota by nod2 is essential for the homeostasis of intestinal intraepithelial lymphocytes. *J. Exp. Med.* **2013**, *210*, 2465–2476. [\[CrossRef\]](#)
120. Li, Y.; Innocentin, S.; Withers, D.R.; Roberts, N.A.; Gallagher, A.R.; Grigorieva, E.F.; Wilhelm, C.; Veldhoen, M. Exogenous stimuli maintain intraepithelial lymphocytes via aryl hydrocarbon receptor activation. *Cell* **2011**, *147*, 629–640. [\[CrossRef\]](#)
121. Boismenu, R.; Havran, W.L. Modulation of epithelial cell growth by intraepithelial  $\gamma\delta$  t cells. *Science* **1994**, *266*, 1253–1255. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Komano, H.; Fujiura, Y.; Kawaguchi, M.; Matsumoto, S.; Hashimoto, Y.; Obana, S.; Mombaerts, P.; Tonegawa, S.; Yamamoto, H.; Itohara, S. Homeostatic regulation of intestinal epithelia by intraepithelial gamma delta t cells. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 6147–6151. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Hou, J.-J.; Ma, A.-H.; Qin, Y.-H. Activation of the aryl hydrocarbon receptor in inflammatory bowel disease: Insights from gut microbiota. *Front. Cell. Infect. Microbiol.* **2023**, *13*, 1279172. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Chen, X.; Zhu, Y.; Wei, Y.; Fan, S.; Xia, L.; Chen, Q.; Lu, Y.; Wu, D.; Liu, X.; Peng, X. Glutamine alleviates intestinal injury in a murine burn sepsis model by maintaining intestinal intraepithelial lymphocyte homeostasis. *Eur. J. Pharmacol.* **2023**, *940*, 175480. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Song, X.; Zhang, H.; Zhang, Y.; Goh, B.; Bao, B.; Mello, S.S.; Sun, X.; Zheng, W.; Gazzaniga, F.S.; Wu, M.; et al. Gut microbial fatty acid isomerization modulates intraepithelial t cells. *Nature* **2023**, *619*, 837–843. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Edelblum, K.L.; Singh, G.; Odenwald, M.A.; Lingaraju, A.; El Bissati, K.; Mcleod, R.; Sperling, A.I.; Turner, J.R.  $\gamma\delta$  intraepithelial lymphocyte migration limits transepithelial pathogen invasion and systemic disease in mice. *Gastroenterology* **2015**, *148*, 1417–1426. [\[CrossRef\]](#)
127. Wu, C.-J.; Mannan, P.; Lu, M.; Udey, M.C. Epithelial cell adhesion molecule (epcam) regulates claudin dynamics and tight junctions. *J. Biol. Chem.* **2013**, *288*, 12253–12268. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Hu, M.D.; Ethridge, A.D.; Lipstein, R.; Kumar, S.; Wang, Y.; Jabri, B.; Turner, J.R.; Edelblum, K.L. Epithelial il-15 is a critical regulator of  $\gamma\delta$  intraepithelial lymphocyte motility within the intestinal mucosa. *J. Immunol.* **2018**, *201*, 747–756. [\[CrossRef\]](#)
129. Al-Sadi, R.; Khatib, K.; Guo, S.; Ye, D.; Youssef, M.; Ma, T. Occludin regulates macromolecule flux across the intestinal epithelial tight junction barrier. *Am. J. Physiol. Gastrointest. Liver. Physiol.* **2011**, *300*, G1054–G1064. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Grigoryan, M.; Kedees, M.H.; Charron, M.J.; Guz, Y.; Teitelman, G. Regulation of mouse intestinal l cell progenitors proliferation by the glucagon family of peptides. *Endocrinology* **2012**, *153*, 3076–3088. [\[CrossRef\]](#) [\[PubMed\]](#)
131. Fujiwara, Y.; Torphy, R.J.; Sun, Y.; Miller, E.N.; Ho, F.; Borchering, N.; Wu, T.; Torres, R.M.; Zhang, W.; Schulick, R.D.; et al. The gpr171 pathway suppresses t cell activation and limits antitumor immunity. *Nat. Commun.* **2021**, *12*, 5857. [\[CrossRef\]](#) [\[PubMed\]](#)
132. Ganea, D.; Hooper, K.M.; Kong, W. The neuropeptide vasoactive intestinal peptide: Direct effects on immune cells and involvement in inflammatory and autoimmune diseases. *Acta Physiol.* **2015**, *213*, 442–452. [\[CrossRef\]](#) [\[PubMed\]](#)
133. Smalley, S.G.R.; Barrow, P.A.; Foster, N. Immunomodulation of innate immune responses by vasoactive intestinal peptide (vip): Its therapeutic potential in inflammatory disease. *Clin. Exp. Immunol.* **2009**, *157*, 225–234. [\[CrossRef\]](#) [\[PubMed\]](#)
134. Martínez, C.; Juarranz, Y.; Gutiérrez-Cañas, I.; Carrión, M.; Pérez-García, S.; Villanueva-Romero, R.; Castro, D.; Lamana, A.; Mellado, M.; González-Álvaro, I.; et al. A clinical approach for the use of vip axis in inflammatory and autoimmune diseases. *Int. J. Mol. Sci.* **2019**, *21*, 65. [\[CrossRef\]](#) [\[PubMed\]](#)
135. Dalton, J.E.; Cruickshank, S.M.; Egan, C.E.; Mears, R.; Newton, D.J.; Andrew, E.M.; Lawrence, B.; Howell, G.; Else, K.J.; Gubbels, M.J.; et al. Intraepithelial gammadelta+ lymphocytes maintain the integrity of intestinal epithelial tight junctions in response to infection. *Gastroenterology* **2006**, *131*, 818–829. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Li, Z.; Zhang, C.; Zhou, Z.; Zhang, J.; Zhang, J.; Tian, Z. Small intestinal intraepithelial lymphocytes expressing cd8 and t cell receptor  $\gamma\delta$  are involved in bacterial clearance during salmonella enterica serovar typhimurium infection. *Infect. Immun.* **2012**, *80*, 565–574. [\[CrossRef\]](#)
137. Zufferey, C.; Erhart, D.; Saurer, L.; Mueller, C. Production of interferon- $\gamma$  by activated t-cell receptor- $\alpha\beta$  cd8 $\alpha\beta$  intestinal intraepithelial lymphocytes is required and sufficient for disruption of the intestinal barrier integrity. *Immunology* **2009**, *128*, 351–359. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Moretto, M.; Weiss, L.M.; Khan, I.A. Induction of a rapid and strong antigen-specific intraepithelial lymphocyte response during oral *Encephalitozoon cuniculi* Infection. *J. Immunol.* **2004**, *172*, 4402–4409. [\[CrossRef\]](#) [\[PubMed\]](#)
139. Chardès, T.; Buzoni-Gatel, D.; Lepage, A.; Bernard, F.; Bout, D. Toxoplasma gondii oral infection induces specific cytotoxic cd8 alpha/beta+ thy-1+ gut intraepithelial lymphocytes, lytic for parasite-infected enterocytes. *J. Immunol.* **1994**, *153*, 4596–4603. [\[CrossRef\]](#)
140. Lepage, A.C.; Buzoni-Gatel, D.; Bout, D.T.; Kasper, L.H. Gut-derived intraepithelial lymphocytes induce long term immunity against toxoplasma gondii1. *J. Immunol.* **1998**, *161*, 4902–4908. [\[CrossRef\]](#) [\[PubMed\]](#)

141. Culshaw, R.J.; Bancroft, G.J.; McDonald, V. Gut intraepithelial lymphocytes induce immunity against cryptosporidium infection through a mechanism involving gamma interferon production. *Infect. Immun.* **1997**, *65*, 3074–3079. [[CrossRef](#)] [[PubMed](#)]
142. Müller, S.; Bühler-Jungo, M.; Mueller, C. Intestinal intraepithelial lymphocytes exert potent protective cytotoxic activity during an acute virus infection. *J. Immunol.* **2000**, *164*, 1986–1994. [[CrossRef](#)] [[PubMed](#)]
143. Siebrecht, M.S.; Hsia, E.; Spychalski, C.; Nagler-Anderson, C. Stimulation of murine intestinal intraepithelial lymphocytes by the bacterial superantigen staphylococcal enterotoxin B. *Int. Immunol.* **1993**, *5*, 717–724. [[CrossRef](#)] [[PubMed](#)]
144. Zhou, R.; Wei, H.; Sun, R.; Zhang, J.; Tian, Z. NKG2D recognition mediates toll-like receptor 3 signaling-induced breakdown of epithelial homeostasis in the small intestines of mice. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 7512–7515. [[CrossRef](#)] [[PubMed](#)]
145. Tang, F.; Sally, B.; Ciszewski, C.; Abadie, V.; Curran, S.A.; Groh, V.; Fitzgerald, O.; Winchester, R.J.; Jabri, B. Interleukin 15 primes natural killer cells to kill via nkg2d and cpla2 and this pathway is active in psoriatic arthritis. *PLoS ONE* **2013**, *8*, e76292. [[CrossRef](#)] [[PubMed](#)]
146. Paterson, R.L.; La Manna, M.P.; Arena De Souza, V.; Walker, A.; Gibbs-Howe, D.; Kulkarni, R.; Fergusson, J.R.; Mulakkal, N.C.; Monteiro, M.; Bunjobpol, W.; et al. An hla-e-targeted tcr bispecific molecule redirects t cell immunity against *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. USA* **2024**, *121*, e2318003121. [[CrossRef](#)]
147. Tan, C.L.; Peluso, M.J.; Drijvers, J.M.; Mera, C.M.; Grande, S.M.; Brown, K.E.; Godec, J.; Freeman, G.J.; Sharpe, A.H. Cd160 stimulates cd8+ t cell responses and is required for optimal protective immunity to *Listeria monocytogenes*. *ImmunoHorizons* **2018**, *2*, 238–250. [[CrossRef](#)] [[PubMed](#)]
148. Shui, J.-W.; Larange, A.; Kim, G.; Vela, J.L.; Zahner, S.; Cheroutre, H.; Kronenberg, M. Hvem signalling at mucosal barriers provides host defence against pathogenic bacteria. *Nature* **2012**, *488*, 222–225. [[CrossRef](#)]
149. Wang, H.-C.; Klein, J.R. Multiple levels of activation of murine cd8+ intraepithelial lymphocytes defined by ox40 (cd134) expression: Effects on cell-mediated cytotoxicity, ifn- $\gamma$ , and il-10 regulation. *J. Immunol.* **2001**, *167*, 6717–6723. [[CrossRef](#)]
150. Voisine, J.; Abadie, V. Interplay between gluten, hla, innate and adaptive immunity orchestrates the development of coeliac disease. *Front. Immunol.* **2021**, *12*, 674313. [[CrossRef](#)] [[PubMed](#)]
151. Abadie, V.R.; Kim, S.M.; Lejeune, T.; Palanski, B.A.; Ernest, J.D.; Tastet, O.; Voisine, J.; Discepolo, V.; Marietta, E.V.; Hawash, M.B.F.; et al. IL-15, gluten and hla-dq8 drive tissue destruction in coeliac disease. *Nature* **2020**, *578*, 600–604. [[CrossRef](#)] [[PubMed](#)]
152. Jabri, B.; De Serre, N.P.M.; Cellier, C.; Evans, K.; Gache, C.; Carvalho, C.; Mougenot, J.F.; Allez, M.; Jian, R.; Desreumaux, P.; et al. Selective expansion of intraepithelial lymphocytes expressing the hla-e-specific natural killer receptor cd94 in celiac disease. *Gastroenterology* **2000**, *118*, 867–879. [[CrossRef](#)] [[PubMed](#)]
153. Hüe, S.; Mention, J.-J.; Monteiro, R.C.; Zhang, S.; Cellier, C.; Schmitz, J.; Verkarre, V.; Fodil, N.; Bahram, S.; Cerf-Bensussan, N.; et al. A Direct role for nkg2d/mica interaction in villous atrophy during celiac disease. *Immunity* **2004**, *21*, 367–377. [[CrossRef](#)] [[PubMed](#)]
154. Ebert, E.C.; Groh, V. Dissection of spontaneous cytotoxicity by human intestinal intraepithelial lymphocytes: Mic on colon cancer triggers nkg2d-mediated lysis through fas ligand. *Immunology* **2008**, *124*, 33–41. [[CrossRef](#)] [[PubMed](#)]
155. Morikawa, R.; Nemoto, Y.; Yonemoto, Y.; Tanaka, S.; Takei, Y.; Oshima, S.; Nagaishi, T.; Tsuchiya, K.; Nozaki, K.; Mizutani, T.; et al. Intraepithelial lymphocytes suppress intestinal tumor growth by cell-to-cell contact via cd103/E-cadherin signal. *Cell. Mol. Gastroenterol. Hepatol.* **2021**, *11*, 1483–1503. [[CrossRef](#)]
156. Meehan, T.F.; Witherden, D.A.; Kim, C.-H.; Sendaydiego, K.; Ye, I.; Garijo, O.; Komori, H.K.; Kumanogoh, A.; Kikutani, H.; Eckmann, L.; et al. Protection against colitis by cd100-dependent modulation of intraepithelial  $\gamma\delta$  t lymphocyte function. *Mucosal. Immunol.* **2014**, *7*, 134–142. [[CrossRef](#)]
157. Nakamura, T.; Matsuzaki, G.; Takimoto, H.; Nomoto, K. Age-associated changes in the proliferative response of rat intestinal intraepithelial leukocytes to bacterial antigens. *Gastroenterology* **1995**, *109*, 748–754. [[CrossRef](#)] [[PubMed](#)]
158. Walker, C.R.; Hautefort, I.; Dalton, J.E.; Overweg, K.; Egan, C.E.; Bongaerts, R.J.; Newton, D.J.; Cruickshank, S.M.; Andrew, E.M.; Carding, S.R. Intestinal intraepithelial lymphocyte-enterocyte crosstalk regulates production of bactericidal angiogenin 4 by paneth cells upon microbial challenge. *PLoS ONE* **2013**, *8*, e84553. [[CrossRef](#)] [[PubMed](#)]
159. Turner, J.-E.; Stockinger, B.; Helmby, H. IL-22 Mediates goblet cell hyperplasia and worm expulsion in intestinal helminth infection. *PLoS Pathog.* **2013**, *9*, e1003698. [[CrossRef](#)] [[PubMed](#)]
160. Inagaki-Ohara, K.; Sakamoto, Y.; Dohi, T.; Smith, A.L.  $\gamma\delta$  T cells play a protective role during infection with *Nippostrongylus brasiliensis* by promoting goblet cell function in the small intestine. *Immunology* **2011**, *134*, 448–458. [[CrossRef](#)] [[PubMed](#)]
161. Zheng, J.; Liu, Y.; Lau, Y.-L.; Tu, W.  $\gamma\delta$ -t cells: An unpolished sword in human anti-infection immunity. *Cell. Mol. Immunol.* **2013**, *10*, 50–57. [[CrossRef](#)] [[PubMed](#)]
162. Ribeiro, S.R.T.; Ribot, J.C.; Silva-Santos, B. Five layers of receptor signaling in  $\hat{\imath}\hat{\imath}'$  t-cell differentiation and activation. *Front. Immunol.* **2015**, *6*, 15. [[CrossRef](#)]
163. Serrano, R.; Wesch, D.; Kabelitz, D. Activation of human  $\gamma\delta$  t cells: Modulation by toll-like receptor 8 ligands and role of monocytes. *Cells* **2020**, *9*, 713. [[CrossRef](#)]

164. Lai, Y.-G.; Gelfanov, V.; Gelfanova, V.; Kulik, L.; Chu, C.-L.; Jeng, S.-W.; Liao, N.-S. Il-15 promotes survival but not effector function differentiation of cd8+ tcr $\alpha\beta$ + intestinal intraepithelial lymphocytes. *J. Immunol.* **1999**, *163*, 5843–5850. [[CrossRef](#)] [[PubMed](#)]
165. Ebert, E.C. Interleukin 15 is a potent stimulant of intraepithelial lymphocytes. *Gastroenterology* **1998**, *115*, 1439–1445. [[CrossRef](#)] [[PubMed](#)]
166. Ebert, E.C. Il-15 converts human intestinal intraepithelial lymphocytes to cd94 producers of ifn-gamma and il-10, the latter promoting Fas ligand-mediated cytotoxicity. *Immunology* **2005**, *115*, 118–126. [[CrossRef](#)] [[PubMed](#)]
167. Khan, I.A.; Casciotti, L. Il-15 prolongs the duration of cd8+ t cell-mediated immunity in mice infected with a vaccine strain of *Toxoplasma gondii*. *J. Immunol.* **1999**, *163*, 4503–4509. [[CrossRef](#)]
168. Montufar-Solis, D.; Garza, T.; Klein, J.R. T-cell activation in the intestinal mucosa. *Immunol. Rev.* **2007**, *215*, 189–201. [[CrossRef](#)]
169. Guk, S.-M.; Yong, T.-S.; Chai, J.-Y. Role of murine intestinal intraepithelial lymphocytes and lamina propria lymphocytes against primary and challenge infections with cryptosporidium parvum. *J. Parasitol.* **2003**, *89*, 270–275. [[CrossRef](#)]
170. Yamamoto, S.; Russ, F.; Teixeira, H.C.; Conradt, P.; Kaufmann, S.H. Listeria monocytogenes-induced gamma interferon secretion by intestinal intraepithelial gamma/delta t lymphocytes. *Infect. Immun.* **1993**, *61*, 2154–2161. [[CrossRef](#)]
171. Shimokawa, C.; Senba, M.; Kobayashi, S.; Kikuchi, M.; Obi, S.; Olia, A.; Hamano, S.; Hisaeda, H. Intestinal inflammation-mediated clearance of amebic parasites is dependent on ifn- $\gamma$ . *J. Immunol.* **2018**, *200*, 1101–1109. [[CrossRef](#)]
172. Li, Y.; Liu, M.; Zuo, Z.; Liu, J.; Yu, X.; Guan, Y.; Zhan, R.; Han, Q.; Zhang, J.; Zhou, R.; et al. Tlr9 regulates the nf- $\kappa$ B-nlrp3-il-1 $\beta$  pathway negatively in *Salmonella*-Induced nkg2d-mediated intestinal inflammation. *J. Immunol.* **2017**, *199*, 761–773. [[CrossRef](#)]
173. Jaeger, N.; Gamini, R.; Cella, M.; Schettini, J.L.; Bugatti, M.; Zhao, S.; Rosadini, C.V.; Esaulova, E.; Di Luccia, B.; Kinnett, B.; et al. Single-cell analyses of Crohn's disease tissues reveal intestinal intraepithelial t cells heterogeneity and altered subset distributions. *Nat. Commun.* **2021**, *12*, 1921. [[CrossRef](#)] [[PubMed](#)]
174. Sollid, L.M. Coeliac disease: Dissecting a complex inflammatory disorder. *Nat. Rev. Immunol.* **2002**, *2*, 647–655. [[CrossRef](#)] [[PubMed](#)]
175. De Kauwe, A.L.; Chen, Z.; Anderson, R.P.; Keech, C.L.; Price, J.D.; Wijburg, O.; Jackson, D.C.; Ladhams, J.; Allison, J.; McCluskey, J. Resistance to celiac disease in humanized hla-dr3-dq2-transgenic mice expressing specific anti-gliadin cd4+ t cells. *J. Immunol.* **2009**, *182*, 7440–7450. [[CrossRef](#)] [[PubMed](#)]
176. Marietta, E.; Black, K.; Camilleri, M.; Krause, P.; Rogers, R.S.; David, C.; Pittelkow, M.R.; Murray, J.A. A new model for dermatitis herpetiformis that uses hla-dq8 transgenic nod mice. *J. Clin. Investig.* **2004**, *114*, 1090–1097. [[CrossRef](#)] [[PubMed](#)]
177. Depaolo, R.W.; Abadie, V.; Tang, F.; Fehlner-Peach, H.; Hall, J.A.; Wang, W.; Marietta, E.V.; Kasarda, D.D.; Waldmann, T.A.; Murray, J.A.; et al. Co-adjuvant effects of retinoic acid and il-15 induce inflammatory immunity to dietary antigens. *Nature* **2011**, *471*, 220–224. [[CrossRef](#)] [[PubMed](#)]
178. Kaukinen, K.; Collin, P.; Maki, M. Latent coeliac disease or coeliac disease beyond villous atrophy? *Gut* **2007**, *56*, 1339–1340. [[CrossRef](#)] [[PubMed](#)]
179. Tosco, A.; Salvati, V.M.; Auricchio, R.; Maglio, M.; Borrelli, M.; Coruzzo, A.; Paparo, F.; Boffardi, M.; Esposito, A.; D'Adamo, G.; et al. Natural history of potential celiac disease in children. *Clin. Gastroenterol. Hepatol.* **2011**, *9*, 320–325, quiz e336. [[CrossRef](#)]
180. Sumida, H. Recent advances in roles of G-protein coupled receptors in intestinal intraepithelial lymphocytes. *Biosci. Microbiota Food Health* **2020**, *39*, 77–82. [[CrossRef](#)]
181. Reis, B.S.; Darcy, P.W.; Khan, I.Z.; Moon, C.S.; Kornberg, A.E.; Schneider, V.S.; Alvarez, Y.; Eleso, O.; Zhu, C.; Scherthanner, M.; et al. Tcr-v $\gamma\delta$  usage distinguishes protumor from antitumor intestinal  $\gamma\delta$  t cell subsets. *Science* **2022**, *377*, 276–284. [[CrossRef](#)] [[PubMed](#)]
182. Li, S.; Pritchard, D.M.; Yu, L.-G. Galectin-3 promotes secretion of proteases that decrease epithelium integrity in human colon cancer cells. *Cell Death Dis.* **2023**, *14*, 268. [[CrossRef](#)]
183. Qiao, S.-W.; Dahal-Koirala, S.; Eggesbø, L.M.; Lundin, K.E.A.; Sollid, L.M. Frequency of gluten-reactive t cells in active celiac lesions estimated by direct cell cloning. *Front. Immunol.* **2021**, *12*, 646163. [[CrossRef](#)] [[PubMed](#)]
184. Matysiak-Budnik, T.; Malamut, G.; De Serre, N.P.-M.; Grosdidier, E.; Segui, S.; Brousse, N.; Caillat-Zucman, S.; Cerf-Bensussan, N.; Schmitz, J.; Cellier, C. Long-term follow-up of 61 coeliac patients diagnosed in childhood: Evolution toward latency is possible on a normal diet. *Gut* **2007**, *56*, 1379–1386. [[CrossRef](#)]
185. Abadie, V.; Discepolo, V.; Jabri, B. Intraepithelial lymphocytes in celiac disease immunopathology. *Semin. Immunopathol.* **2012**, *34*, 551–566. [[CrossRef](#)] [[PubMed](#)]
186. James, O.J.; Vandereyken, M.; Marchingo, J.M.; Singh, F.; Bray, S.E.; Wilson, J.; Love, A.G.; Swamy, M. Il-15 and pim kinases direct the metabolic programming of intestinal intraepithelial lymphocytes. *Nat. Commun.* **2021**, *12*, 4290. [[CrossRef](#)] [[PubMed](#)]
187. Ferguson, A.; Murray, D. Quantitation of intraepithelial lymphocytes in human jejunum. *Gut* **1971**, *12*, 988–994. [[CrossRef](#)] [[PubMed](#)]
188. Marsh, M.; Heal, C. Evolutionary developments in interpreting the gluten-induced mucosal celiac lesion: An archimedean heuristic. *Nutrients* **2017**, *9*, 213. [[CrossRef](#)]

189. Eggesbø, L.M.; Risnes, L.F.; Neumann, R.S.; Lundin, K.E.A.; Christophersen, A.; Sollid, L.M. Single-cell tcr sequencing of gut intraepithelial  $\gamma\delta$  t cells reveals a vast and diverse repertoire in celiac disease. *Mucosal Immunol.* **2020**, *13*, 313–321. [[CrossRef](#)]
190. Mayassi, T.; Ladell, K.; Gudjonson, H.; McLaren, J.E.; Shaw, D.G.; Tran, M.T.; Rokicka, J.J.; Lawrence, I.; Grenier, J.-C.; Van Unen, V.; et al. Chronic inflammation permanently reshapes tissue-resident immunity in celiac disease. *Cell* **2019**, *176*, 967–981.e919. [[CrossRef](#)] [[PubMed](#)]
191. Bhagat, G.; Naiyer, A.J.; Shah, J.G.; Harper, J.; Jabri, B.; Wang, T.C.; Green, P.H.R.; Manavalan, J.S. Small intestinal cd8+tcry $\delta$ +nkg2a+ intraepithelial lymphocytes have attributes of regulatory cells in patients with celiac disease. *J. Clin. Investig.* **2008**, *118*, 281–293. [[CrossRef](#)] [[PubMed](#)]
192. Kanazawa, H.; Ishiguro, Y.; Munakata, A.; Morita, T. Multiple Accumulation of V $\delta$ 2+  $\gamma\delta$  t-cell clonotypes in intestinal mucosa from patients with Crohn’s disease. *Dig. Dis. Sci.* **2001**, *46*, 410–416. [[CrossRef](#)]
193. Yeung, M.M.; Melgar, S.; Baranov, V.; Öberg, Å.; Danielsson, Å.; Hammarström, S.; Hammarström, M.L. Characterisation of mucosal lymphoid aggregates in ulcerative colitis: Immune cell phenotype and tcr- $\gamma\delta$  expression. *Gut* **2000**, *47*, 215. [[CrossRef](#)]
194. Jensen-Jarolim, E.; Gscheidlinger, R.; Oberhuber, G.; Neuchrist, C.; Lucas, T.; Bises, G.; Radauer, C.; Willheim, M.; Scheiner, O.; Liu, F.-T.; et al. The constitutive expression of galectin-3 is downregulated in the intestinal epithelia of Crohn’s disease patients, and tumour necrosis factor alpha decreases the level of galectin-3-specific mRNA in hct-8 cells. *Eur. J. Gastroenterol. Hepatol.* **2002**, *14*, 145–152. [[CrossRef](#)] [[PubMed](#)]
195. Lippert, E.; Stieber-Gunckel, M.; Dunger, N.; Falk, W.; Obermeier, F.; Kunst, C. Galectin-3 modulates experimental colitis. *Digestion* **2015**, *92*, 45–53. [[CrossRef](#)] [[PubMed](#)]
196. Frol’Ová, L.; Smetana, K.; Borovská, D.; Kitanovičová, A.; Klimešová, K.; Janatková, I.; Malíčková, K.; Lukáš, M.; Drastich, P.; Beneš, Z.; et al. Detection of galectin-3 in patients with inflammatory bowel diseases: New serum marker of active forms of IBD? *Inflamm. Res.* **2009**, *58*, 503–512. [[CrossRef](#)]
197. Zundler, S.; Becker, E.; Spocinska, M.; Slawik, M.; Parga-Vidal, L.; Stark, R.; Wiendl, M.; Atreya, R.; Rath, T.; Leppkes, M.; et al. Hobit- and blimp-1-driven cd4+ tissue-resident memory t cells control chronic intestinal inflammation. *Nat. Immunol.* **2019**, *20*, 288–300. [[CrossRef](#)]
198. Roosenboom, B.; Wahab, P.J.; Smids, C.; Groenen, M.J.M.; Van Koolwijk, E.; Van Lochem, E.G.; Horjus Talabur Horje, C.S. Intestinal cd103+cd4+ and cd103+cd8+ t-cell Subsets in the gut of inflammatory bowel disease patients at diagnosis and during follow-up. *Inflamm. Bowel Dis.* **2019**, *25*, 1497–1509. [[CrossRef](#)]
199. Lamb, C.A.; Mansfield, J.C.; Tew, G.W.; Gibbons, D.; Long, A.K.; Irving, P.; Diehl, L.; Eastham-Anderson, J.; Price, M.B.; O’Boyle, G.; et al.  $\alpha$ E $\beta$ 7 Integrin identifies subsets of pro-inflammatory colonic cd4+ t lymphocytes in ulcerative colitis. *J. Crohn’s Colitis* **2016**, *11*, jjw189. [[CrossRef](#)]
200. Bishu, S.; El Zaatar, M.; Hayashi, A.; Hou, G.; Bowers, N.; Kinnucan, J.; Manoogian, B.; Muza-Moons, M.; Zhang, M.; Grasberger, H.; et al. Cd4+ tissue-resident memory t cells expand and are a major source of mucosal tumour necrosis factor  $\alpha$  in active crohn’s disease. *J. Crohn’s Colitis* **2019**, *13*, 905–915. [[CrossRef](#)] [[PubMed](#)]
201. Bottois, H.; Ngollo, M.; Hammoudi, N.; Courau, T.; Bonnereau, J.; Chardiny, V.; Grand, C.; Gergaud, B.; Allez, M.; Le Bourhis, L. Klr1 and cd103 expressions define distinct intestinal tissue-resident memory cd8 t cell subsets modulated in crohn’s disease. *Front. Immunol.* **2020**, *11*, 896. [[CrossRef](#)]
202. SchöN, M.P.; Arya, A.; Murphy, E.A.; Adams, C.M.; Strauch, U.G.; Agace, W.W.; Marsal, J.; Donohue, J.P.; Her, H.; Beier, D.R.; et al. Mucosal t lymphocyte numbers are selectively reduced in integrin  $\alpha$ e (cd103)-deficient mice. *J. Immunol.* **1999**, *162*, 6641–6649. [[CrossRef](#)] [[PubMed](#)]
203. Guy-Grand, D.; Cuénod-Jabri, B.; Malassis-Seris, M.; Selz, F.; Vassalli, P. Complexity of the mouse gut t cell immune system: Identification of two distinct natural killer t cell intraepithelial lineages. *Eur. J. Immunol.* **1996**, *26*, 2248–2256. [[CrossRef](#)] [[PubMed](#)]
204. Mikulak, J.; Oriolo, F.; Bruni, E.; Roberto, A.; Colombo, F.S.; Villa, A.; Bosticardo, M.; Bortolomai, I.; Lo Presti, E.; Meraviglia, S.; et al. Nkp46-expressing human gut-resident intraepithelial v $\delta$ 1 t cell subpopulation exhibits high antitumor activity against colorectal cancer. *JCI Insight* **2019**, *4*, e125884. [[CrossRef](#)] [[PubMed](#)]
205. Cohen, C.J.; Shieh, J.T.C.; Pickles, R.J.; Okegawa, T.; Hsieh, J.-T.; Bergelson, J.M. The coxsackievirus and adenovirus receptor is a transmembrane component of the tight junction. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 15191–15196. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.