





## Whole-Genome Sequence of Brevibacillus borstelensis SDM, Isolated from a Sorghum-Adapted Microbial Community

Martina Aulitto, a,b Lauren M. Tom, a,b Javier A. Ceja-Navarro, Blake A. Simmons, a,b 📵 Steven W. Singera,b

<sup>a</sup>Joint BioEnergy Institute, Emeryville, California, USA

<sup>b</sup>Biological Systems and Engineering Division, Lawrence Berkeley National Laboratory, Berkeley, California, USA

**ABSTRACT** The isolation of novel microbes from environmental samples continues to be a key strategy for the discovery of new metabolic capacities for the degradation and transformation of lignocellulose. We report the draft genome sequence of a new strain of Brevibacillus borstelensis isolated from a sorghum-adapted microbial community derived from a compost sample.

oil contains an untapped diversity of microbial communities with equally diverse metabolic capacities (1). Researchers around the world continue developing strategies for isolating microorganisms that can efficiently degrade lignocellulosic biomass, and most rely on the isolation of these microorganisms from environmental samples (2). In the present study, a new strain of Brevibacillus borstelensis was isolated from a compost-derived microbial community enriched on untreated sorghum biomass. To date, several B. borstelensis organisms have been sequenced, but only a few have available whole-genome sequences (BioProject numbers PRJDB5988, PRJNA191598, PRJNA229942, PRJNA233554, PRJNA348753, and PRJNA498706) (3, 4). B. borstelensis is a Gram-positive, aerobic, thermophilic, endospore-forming bacterium of the family Paenibacillaceae. Other studies have shown the importance of B. borstelensis in different environmental applications, such as the capability to degrade polyethylene by 30% (5) and to enhance the transformation of food waste into biofertilizer (6).

B. borstelensis SDM was isolated by spreading an enrichment culture of green waste compost obtained from the city of Berkeley, California, incubated with sorghum in M9TE (7) on Luria Bertani (LB) solid medium and incubating it overnight at 50°C. After several isolation steps, six colonies were randomly picked and identified through 16S rRNA-encoding gene sequencing as Brevibacillus borstelensis (99% identical). B. borstelensis was cultivated overnight in LB medium (at 50°C and 200 rpm), and genomic DNA was extracted using a DNeasy PowerSoil kit (Qiagen). The library was prepared using a KAPA HyperPrep kit for DNA (KK8504) and then sequenced with a NovaSeq S4 platform (Illumina). The raw sequences were uploaded to KBase (https://narrative.kbase.us/ narrative/ws.39490.obj) (8), and default parameters were used for all software unless otherwise noted. The read quality was assessed using FastQC v 0.11.5 (9), and a quality score above Q30 for 97.9% of the bases was reported. A total of 611,322,296 reads (average length, 151 bp) were assembled using (3) different assemblers (SPAdes v 3.13.0 [10], IDBA-UD v 1.1.3 [11], and MEGAHIT v 1.2.9 [12]) and compared using QUAST v 4.4 (13). The best assembly was obtained using MEGAHIT and comprised 128 contigs, all of them with  $\geq$ 1,000 bp, with a total length of 5,246,051 bp, an  $N_{50}$  value of 82,015 bp, and an average GC content of 51.62%. The genome coverage was 116.5×. Furthermore, the assembly was evaluated for completeness (99.73%) and contamination (2.35%) using CheckM v 1.0.18 (14).

The genome annotation of B. borstelensis SDM was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP), where a total of 4,853 coding

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Address correspondence to Steven W. Singer, swsinger@lbl.gov.

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sequences (CDS) were identified (15). Moreover, for the annotation of carbohydrate-active enzymes (CAZYmes), all proteins were mapped against the dbCAN database v 8 (16) using hmmscan (HMMER 3.1b2) with an E value cutoff of 1e-5. This analysis revealed the existence of 266 CAZYmes, suggesting that *B. borstelensis* SDM has the potential to degrade and transform lignocellulosic biomass.

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number JABTBQ000000000, and the raw reads have been deposited in the Sequence Read Archive under the accession number PRJNA633907.

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## **REFERENCES**

- Deltedesco E, Keiblinger KM, Piepho HP, Antonielli L, Pötsch EM, Zechmeister-Boltenstern S, Gorfer M. 2020. Soil microbial community structure and function mainly respond to indirect effects in a multifactorial climate manipulation experiment. Soil Biol Biochem 142:107704. https://doi.org/10.1016/j.soilbio.2020.107704.
- Muaaz-Us-Salam S, Cleall PJ, Harbottle MJ. 2020. Application of enzymatic and bacterial biodelignification systems for enhanced breakdown of model lignocellulosic wastes. Sci Total Environ 728:138741. https://doi. org/10.1016/j.scitotenv.2020.138741.
- Khalil AB, Sivakumar N, Arslan M, Saleem H, Qarawi S. 2018. Insights into Brevibacillus borstelensis AK1 through whole genome sequencing: a thermophilic bacterium isolated from a hot spring in Saudi Arabia. Biomed Res Int 2018:5862437. https://doi.org/10.1155/2018/5862437.
- Tripathy S, Padhi SK, Sen R, Maji U, Samanta M, Mohanty S, Maiti NK. 2016. Draft genome sequence of Brevibacillus borstelensis cifa\_chp40, a thermophilic strain having biotechnological importance. J Genomics 4:4–6. https://doi.org/10.7150/jgen.14036.
- Hadad D, Geresh S, Sivan A. 2005. Biodegradation of polyethylene by the thermophilic bacterium Brevibacillus borstelensis. J Appl Microbiol 98:1093–1100. https://doi.org/10.1111/j.1365-2672.2005.02553.x.
- Arya R, Kumar N, Anil M. 2016. Brevibacillus borstelensis and Streptomyces albogriseolus have roles to play in degradation of herbicide, sulfosulfuron. 3 Biotech 6:1–7. https://doi.org/10.1007/s13205-016-0562-z.
- Gladden JM, Allgaier M, Miller CS, Hazen TC, VanderGheynst JS, Hugenholtz P, Simmons BA, Singer SW. 2011. Glycoside hydrolase activities of thermophilic bacterial consortia adapted to switchgrass. Appl Environ Microbiol 77:5804–5812. https://doi.org/10.1128/AEM.00032-11.
- 8. Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM,

- Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, et al. 2018. KBase: the United States Department of Energy systems biology knowledgebase. Nat Biotechnol 36:566–569. https://doi.org/10.1038/nbt.4163.
- Brown J, Pirrung M, McCue LA. 2017. FQC Dashboard: integrates FastQC results into a Web-based, interactive, and extensible FASTQ quality control tool. Bioinformatics 33:3137–3139. https://doi.org/10.1093/bioinformatics/ btx373
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Peng Y, Leung HCM, Yiu SM, Chin FYL. 2012. IDBA-UD: a de novo assembler for single-cell and metagenomic sequencing data with highly uneven depth. Bioinformatics 28:1420–1428. https://doi.org/10.1093/bioinformatics/bts174.
- Li D, Liu CM, Luo R, Sadakane K, Lam TW. 2015. MEGAHIT: an ultra-fast single-node solution for large and complex metagenomics assembly via succinct de Bruijn graph. Bioinformatics 31:1674–1676. https://doi.org/10.1093/bioinformatics/btv033.
- Mikheenko A, Valin G, Prjibelski A, Saveliev V, Gurevich A. 2016. Icarus: visualizer for de novo assembly evaluation. Bioinformatics 32:3321–3323. https://doi.org/10.1093/bioinformatics/btw379.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https://doi.org/10.1101/gr.186072.114.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.
- Yin Y, Mao X, Yang J, Chen X, Mao F, Xu Y. 2012. dbCAN: a Web resource for automated carbohydrate-active enzyme annotation. Nucleic Acids Res 40:W445–W451. https://doi.org/10.1093/nar/gks479.