# **Review** Article

# Specific Roles of MicroRNAs in Their Interactions with Environmental Factors

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MicroRNAs (miRNAs) have emerged as critical regulators of gene expression by modulating numerous target mRNAs expression at posttranscriptional level. Extensive studies have shown that miRNAs are critical in various important biological processes, including cell growth, proliferation, differentiation, development, and apoptosis. In terms of their importance, miRNA dysfunction has been associated with a broad range of diseases. Increased number of studies have shown that miRNAs can functionally interact with a wide spectrum of environmental factors (EFs) including drugs, industrial materials, virus and bacterial pathogens, cigarette smoking, alcohol, nutrition, sleep, exercise, stress, and radiation. More importantly, the interactions between miRNAs and EFs have been shown to play critical roles in determining abnormal phenotypes and diseases. In this paper, we propose an outline of the current knowledge about specific roles of miRNAs in their interactions with various EFs and analyze the literatures detailing miRNAs-EFs interactions in the context of various of diseases.

# 1. Introduction

MicroRNAs (miRNAs) are regulatory RNAs that are 20-30 nucleotides long that bind the 3'-untranslated regions of target mRNAs [1-3]. miRNAs have emerged as critical regulators of gene expression by modulating the expression of numerous target mRNAs mainly at the posttranscriptional level [4]. Since partial or imperfect complementarity of an miRNA to a target mRNA can lead to translational repression, a single miRNA has the ability of regulating a large number of genes [5]. miRNAs have been shown to play a role in regulating a wide range of biological processes, such as cell growth, proliferation, differentiation, development, and apoptosis [6]. In terms of their importance, dysfunction of miRNAs has been associated with various diseases [7-9]. In contrast to the wealth of publications about their biological effects, the information about specific regulations of miRNAs has comparatively lagged behind.

The phenotype of an organism is determined by the complex interactions between genetic factors (GFs) and environmental factors (EFs). EFs have been shown to

contribute tremendously to the formation and development of many diseases, especially complex diseases [10-12]. The interactions between GFs and EFs, often hypothesized to be mediated by epigenetic mechanisms, modulate the reproductive fitness of an organism, its response to external stimuli and health [13]. Similar to other GFs, miRNAs have complex interactions with a wide spectrum of EFs [14]. Recently, increased number of studies have shown that miRNAs can functionally interact with a variety of EFs including drugs [15, 16], industrial materials [17, 18], virus and bacterial pathogens [19, 20], alcohol [21-23], cigarette smoking [24-26], nutrition [27, 28], sleep [29], exercise [30, 31], stress [32, 33], and radiation [34]. The interactions between miRNAs and these EFs play critical roles in determining phenotypes. The illustration of the specific roles of miRNAs and the patterns of miRNA deregulation in the miRNAs-EFs interactions can help to discover the mechanisms of diseases and drug response therefore providing insights into disease diagnosis, prognosis and novel pharmacologic approaches (Figure 1). In this paper, we propose an outline of the current understanding about specific roles of miRNAs when they



FIGURE 1: Significant roles of miRNAs-EFs interactions.

are interacting with various EFs and analyze the literatures studying the miRNAs-EFs interactions in the context of diseases.

#### 2. Drugs

Previous studies had highlighted the post-transcriptional modifications mediated by miRNAs in drug treatment, drug sensitivity, and drug addiction.

Several groups have shown the altered miRNAs profiles in anticancer drugs treatment, including all-trans-retinoic acid [35, 36], hydroxycamptothecin [37, 38], imatinib [16, 39-41], 5-fluorouracil [42, 43], gemcitabine [44-47], cisplatin [48–52], fludarabine [53], lithium [54–56], epigallocatechin gallate [57], and curcumin [58–60]. Garzon et al. studied the effects of all-trans-retinoic acid (ATRA) treatment on the expression of miRNAs in acute promyelocytic leukemia patients as well as in cell lines by using a miRNA microarray platform and quantitative real-time polymerase chain reaction (qRT-PCR). Their results showed upregulation of miR-15a, 15b, 16-1, 223, 342 107 and let-7a-3, let-7c, let-7d and downregulation of miR-181b in ATRA treatment. Among the upregulated miRNAs, miR-107 was predicted to target NFI-A, a gene involved in the regulatory loop with the participation of miR-223 and C/EBPa during granulocytic differentiation [35].

Drug insensitivity or resistance is a major obstacle for successful cancer therapy. Increasing evidence suggested that miRNAs modulate cellular sensitivity to anticancer reagents. Wu et al. showed significantly different sensitivities of the six gastric cancer cell lines to hydroxycamptothecin (HCPT). The levels of 25 miRNAs were shown to be deregulated in the HCPT-resistant gastric cancer cells, including miR-200 family, miR-196a, 338, 126, 31, 98, 7 and let-7 g. The target genes of these miRNAs are involved in cancer development, progression, and chemosensitivity [37]. Ali et al. evaluated the effects of curcumin or its analogue difluorinated-curcumin (CDF) alone or in combination with gemcitabine on cell viability and apoptosis in gemcitabinesensitive as well as gemcitabine-resistant pancreatic cancer (PC) cell lines [58]. They showed downregulation of miR-200 and upregulation of miR-21 (an indicator of tumor aggressiveness) in gemcitabine-resistant cells in comparison to gemcitabine-sensitive cells. In addition, gemcitabine sensitivity can be induced in PC cells through the modulation of miR-200 and miR-21 expression by curcumin or its analogue CDF. With the application of anti-miR-21 oligonucleotide, Li et al. were able to use anti-miR-21 oligonucleotide to enhance the chemosensitivity of leukemic HL60 cells to arabinosylcytosine [61]. In the study performed by Ji et al. in 2009, the expression patterns of miRNAs in patients with hepatocellular carcinoma (HCC), their survival, and response to interferon alfa were examined [62]. They found a higher expression of miR-26a and miR-26b in nontumor liver tissue in female than in male HCC patients. They also showed significantly decreased levels of miR-26 expression in comparison to the paired noncancerous tissues. Furthermore, cancer patients with tumors of low miR-26 level had shorter overall survival but better response to interferon therapy than those with tumors of high levels of miR-26. These results suggested the association between the miR-26 expression level and the survival of HCC patients as well as their response to adjuvant interferon alfa therapy [62].

Drug abuse induces persistent structural and functional changes in the mesolimbic dopaminergic system and leads to the progression of patients toward the high-risk drug-seeking behavior and relapse. Much emerging evidence suggests that drug-induced neuroplasticity is regulated by gene expression and post-transcriptional regulation [63, 64]. Chandrasekar and Dreyer had shown that a chronic usage of cocaine decreased miR-124 and let-7d levels and increased the expression of miR-181a in mesolimbic pathway [65]. They also proposed a complex regulatory pathway involving

miRNAs in cocaine-mediated neuronal adaptations [66]. Evidence has suggested that cross-regulation between transcription factors and miR-212/132 played a role in drug addiction [67]. MiR-132 and miR-212 are tandem miRNAs whose expression is necessary for the proper development, maturation, and function of neurons. Deregulation of miR-212/132 had been associated with several neurological disorders [67]. It has been shown that overexpression of miR-212 induces CREB activity, the negative modulator of the reward response to cocaine, therefore exerts protective function against cocaine addiction by reducing the responsiveness to the motivational properties of the drug [68]. The X-linked transcriptional repressor methyl CpG binding protein 2 (MeCP2), known for its role in the neurodevelopmental disorder Rett syndrome, is emerging as an important regulator of neuroplasticity in postmitotic neurons. MeCP2 regulates cocaine intake through homeostatic interactions with miR-212 to control the effects of cocaine on striatal brainderived neurotrophic factor (BDNF) levels. As MeCP2 levels are correlated to BDNF expression, which controls cocaine intake, miR-212 could fine-tune the responses of patients to drug abuse by both increasing CREB signaling and decreasing BDNF expression, leading to decreased cocaine intake [69].

#### 3. Industrial Materials

Many industrial materials have toxicity and may alter developmental pathways and cell processes through epigenetic mechanisms. Bisphenol A (BPA) is an industrial plasticizer widely used as coating for food cans and plastic water bottles. The frequency of BPA exposure has steadily increased during recent years [18]. Whiting and colleagues investigated the effects of BPA exposure on miRNAs in human placental cell lines using miRNA microarray and qRT-PCR analysis. MiR-146a was shown to be the only miRNA validated by qRT-PCR as being significantly upregulated in both 3A and HTR-8 cell lines with BPA treatment [17]. It was also demonstrated that stable overexpression of miR-146a in 3A placental cell line leads to a significant decrease in cell proliferation, a result consistent with several other studies [70, 71]. In addition, over-expression of miR-146a in placental cell lines led to increased sensitivity of these cells to bleomycin, the DNA damaging agent, suggesting that the targets of miR-146a may be involved in the DNA damage response.

Jardim et al. investigated the pollutant-mediated regulations of miRNAs in airway epithelial cells with diesel exhaust particles (DEP), the largest source of emitted airborne particulate matter (PM).They showed that DEP exposure significantly modulated the miRNA expression profile in human airway epithelial cell lines. Specifically, 197 out of 313 (62.9%) detectable miRNAs had been either upregulated or downregulated by 1.5 fold. Molecular network analysis of putative targets of the 12 mostly altered miRNAs suggested that DEP exposure was associated with inflammatory responsive pathways and a strong tumorigenic disease signature. These results clearly demonstrated that alteration of miRNA expression profiles by environmental pollutants such as DEP could modify cellular processes by regulation of gene expression and subsequently lead to disease pathogenesis [72]. In a similar study, Bollati et al. discovered the significant increases of the expression of miR-222 and miR-21 in peripheral blood leukocytes from foundry workers with exposure to metal-rich particulate matter [73].

#### 4. Virus and Other Pathogens

miRNAs have been known to play a critical role in the life cycle of retroviruses and a few oncogenic viruses, including HIV [74, 75], reticuloendotheliosis virus strain T (REV-T) [76], influenza A virus [77], Epstein-Barr virus [78, 79], human T-cell leukemia virus type 1 (HTLV-1) [80, 81], gammaretrovirus XMRV [82], cytomegalovirus (HCMV) [83, 84], Hepatitis B virus (HBV) [85, 86], Hepatitis C virus (HCV) [87, 88], and human papilloma virus (HPV) [89]. These viruses regulate viral replication in the host cells with the mediation of specific miRNAs. HCC is the fifth most common cancer worldwide. The greatest risk factor for HCC is the existence of cirrhosis, most commonly induced by HBV or HCV infections as well as factors such as alcohol consumption, oxidative stress, and aflatoxin or arsenic exposure. Specific miRNAs have been shown to be involved in subgroups of HCC caused by different environmental risk factors. Examinations of miRNAs functions and their expression are important for HCC diagnosis, treatment, and prognosis (Figure 2). Several host cell factors including liver-specific miR-122 have been shown to be involved in virus translation, replication, and production. Jopling et al. observed that sequestration of miR-122 in liver cells could result in marked loss of autonomously replicating hepatitis C viral RNAs. MiR-122 has been shown to increase HCV abundance in HCV replication models by interacting with the 5' noncoding region (NCR) of the HCV genome [90]. MiR-122 has also been shown to stimulate HCV translation by enhancing the association of ribosome with viral RNA at an early initiation stage [91]. Studies by Lanford and colleagues suggested that miR-122 was essential for the accumulation of HCV RNA in chronically HCV-infected chimpanzees by inhibiting miR-122 through a specific locked nucleic acid (LNA) oligonucleotide [87]. Qiu et al. also suggested a role of miR-122 in the regulation of HBV replication by downregulating the Heme oxygenase-1 (HO-1) [92]. These results suggest that miR-122 may serve as a target for antiviral intervention.

Human T-lymphotropic virus type 1 (HTLV-1) is the etiologic agent to which the prolonged exposure can cause the adult T-cell leukemia of CD4+ T-cell origin, a severe and fatal lymphoproliferative disease. Pichler et al. reported the deregulation of several miRNAs, including miR-21, 24, 146a, 155, and 223 in HTLV-1-transformed cells. The expression pattern of these miRNAs presented a uniform phenotype in HTLV-transformed cells in comparison to HTLV-negative control cells. Studies suggested miR-146a could be directly attributed to oncoprotein Tax via NF- $\kappa$ B-mediated transactivation in HTLV signaling pathways [81].



FIGURE 2: Specific miRNAs are associated with different subgroups of HCC associated with various environmental risk factors. † indicates up-regulation; ↓ indicates down-regulation; HCC: hepatocellular carcinoma; HCV: hepatitis C virus; HBV: hepatitis B virus; AFB1: aflatoxin B1; ROS: reactive oxygen species.

Recent studies characterized modifications of host miR-NAs expression following infection by exclusively extracellular (*Helicobacter pylori*) or intracellular (*Salmonella enterica*) gram-negative bacteria, gram-positive bacteria (*Listeria monocytogenes*) and other pathogens such as *Mycobacterium*, *Francisella* species, and *Plasmodium berghei* [20, 93, 94].

# 5. Cigarette Smoking

Throughout one's life course, a number of exposures can affect an individual's development, health, and overall quality of life. One of the most common, potentially hazardous environmental exposures that negatively influence health and development is cigarette smoke exposure [24]. Maccani et al. investigated the effects of maternal cigarette smoking during pregnancy on differential expression of miRNAs in the placenta. The data suggested that maternal cigarette smoking during pregnancy was associated with the down-regulation of miR-16, 21, and 146a. They further identified two components, nicotine and benzo [a]pyrene as the specific miRNA modulators in cigarette smoking [26, 95].

Xi et al. investigated the effects of cigarette smoke condensate (CSC) on miRNA expression and function in both normal human respiratory epithelial cells and lung cancer cells [96]. Their results showed that the exposure to CSC induced the expression of miR-31 in both normal respiratory epithelial cells and lung cancer cells. Further studies showed that the overexpression of miR-31 led to increased proliferation and tumorigenicity in lung cancer cells, whereas knockdown of miR-31 expression significantly decreased lung cancer cell proliferation [96]. These results suggested that miR-31 was responsive to cigarette smoke exposure in both normal respiratory epithelia and lung cancer cells and might play a role as an oncomir in lung cancer carcinogenesis pathway [96]. Schembri et al. investigated whole-genome miRNA expression in bronchial airway epithelium from regular smokers and nonsmokers. They observed that 28 miRNAs were differentially expressed, the majority of which were downregulated in the airway epithelium of regular smokers including miR-218 [97]. Several studies have also been performed to characterize the effects of environmental cigarette smoke (ECS) (also called "passive," "secondhand," or "sidestream" cigarette smoke) on miRNA expression. Izzotti et al. found that the most greatly downregulated miRNAs belonged to miRNA families which had been previously shown to regulate a number of key biological processes, including stress response, proliferation, angiogenesis, and apoptosis [98]. Furthermore, they showed that exposure to ECS upregulated 2.9% of genes and 9.7% of proteins in the same tissue, suggesting that the ECS exposure-induced down-regulation of miRNAs may lead to increased protein levels of genes which are negatively regulated by these miRNAs [98].

# 6. Alcohol

Neurotoxicity-mediated pathophysiological conditions could manifest diseases or disabilities such as Parkinson's and Alzheimer's which have debilitating implications. Neurotoxicity as well as brain injury can be caused by external agents including drugs and alcohol, and miRNAs have been shown to play a pivotal role in these processes [99]. Maternal ethanol consumption during pregnancy can lead to a stereotypic cluster of fetal craniofacial, cardiovascular, skeletal, and neurological defects that are collectively termed the fetal alcohol spectrum disorder (FASD) [22]. The developing brain is an important and vulnerable target for ethanol. In 2007, Sathyan et al. obtained the first evidence showing that miRNAs mediated the teratogenic effects by ethanol treatment [100]. The accumulated data support the following conclusions. (1) Ethanol appears to affect a relatively small subset of expressed miRNAs. (2) Ethanol appears to affect miRNAs which are normal components of the cellular repertoire for a given stage of differentiation. As an example, miR-124 is sufficient to direct fetal neural stem cells (NSCs) toward a neuronal lineage (22,101). (3) Ethanol appears to affect increasing numbers of miRNAs along the developmental stage. (4) MiRNAs targeted by ethanol exhibit developmental stage-specific sensitivity. (5) Some ethanol-sensitive miRNAs exert their functions over multiple developmental stages, for example, miR-9 exhibits ethanol sensitivity at multiple stages of development, expanding from the embryo through fetus to the adulthood [22, 101, 102]. In order to illustrate the mechanisms of ethanol regulation of miRNAs, Sathyan et al. reported that GABAA receptors and nicotinic acetylcholine receptors could mediate certain ethanol effects on miRNAs expression, respectively [100, 101]. It has been shown that several ethanol-sensitive miRNAs are localized on chromosomal regions that are susceptible to epigenetic modifications. Ligand-gated ion channel receptors and epigenetic modifications have been shown to be potential mediators of the effects of ethanol on fetal miRNAs [22]. Ethanol and nicotine are often coabused. Balaraman et al. studied their combined effects on fetal neural development, particularly on NSCs. Their results showed that ethanol suppressed the expression of known ethanol-sensitive miRNAs (miR-9, 21, 153, and 335) and miR-140-3p, while nicotine at the concentrations attained by cigarette smokers could induce a dose-related induction in these miRNAs. These data suggest that concurrent exposure to ethanol and nicotine disrupts miRNA regulatory networks that are important for NSC maturation [101].

Alcohol-dependence can lead to long-term changes of gene expression in brain. Tapocik et al. examined the potential roles of miRNAs in persistent gene expression regulation of the medial prefrontal cortex (mPFC) of the rats with a history of alcohol-dependence. Their results showed that more than 41 rat miRNAs and 165 mRNAs in the mPFC were significantly altered after chronic alcohol exposure. Categories of gene ontology differential expression indicated that these miRNAs served in functional processes commonly associated with neurotransmission, neuroadaptation, and synaptic plasticity. Their results demonstrated a significant shift in miRNA expression pattern in the mPFC following a history of alcohol-dependence. MiRNAs may play a pivotal role in the reorganization of synaptic connections and long-term neuroadaptations in alcohol-dependence due to their functions of global regulation of multiple target transcripts [23].

Chronic alcohol abuse can cause liver damage, including inflammation, fatty liver, fibrosis, cirrhosis, and HCC. Bala and Szabo showed that alcohol could induce miR-155 and miR-132 expression in liver as well as in isolated hepatocytes and Kupffer cells of alcohol-fed mice [103]. Yin et al. showed that miR-217 could promote ethanol-induced fat accumulation in hepatocytes by downregulating SIRT1 [104]. Ladeiro et al. suggested the miRNA deregulations may be associated with alcohol consumption and HBV infection in HCC. Their studies indicated the correlation between miR-126 down-regulation and alcohol abuse-related HCC in comparison to the other HCCs (Figure 2). Their miRNAs profiling is the first global genomic approach that enables us to differentiate the alcohol abuse-related HCC from the other HCC tumors [21].

# 7. Stress

Cells frequently encounter conditions that can lead to stress. A few well-characterized stressors include hyperthermia, hypoxia, ATP depletion, and oxidative stress [105, 106]. In order to survive and adapt to stressful conditions, all mammalian cells have evolved a molecular defense reaction termed the cellular stress response (CSR). Recent studies have suggested that miRNAs participate in the CSR, such as hypothermia [107], hyperthermia [108], hypoxia [109–112], folate deficiency, and arsenic exposure [33].

Wilmink et al. studied the specific group of deregulated miRNAs in the cells exposed to hyperthermia. Their results showed that the heat could induce upregulation of miRNAs including miR-125b, 382, and 452 and downregulation of miRNAs including miR-138, 7, and 196b. Several thermally-regulated miRNAs (miR-452, 382, and 378) expresses only in cells exposed to hyperthermia [108].

Hypoxia is an essential feature of the neoplastic microenvironment. Tumors with extensive low oxygen level tend to exhibit poor prognosis and resistance to conventional therapy [109]. Ivan's group identified a set of hypoxiaregulated miRNAs (HRMs), including miR-21, 23a, 23b, 24, 26a, 26b, 27a, 30b, 93, 103, 103, 106a, 107, 125b, 181a, 181b, 181c, 192, 195, 210, and 213 [112]. In addition, a set of miRNAs were down-regulated in hypoxic cells, including miR-15b, 16, 19a, 20a, 20b, 29b, 30b, 30e-5p, 101, 141, 122a, 186, 320, and 197 [113, 114]. Saito et al. suggested the involvement of the miR-130 family in hypoxia-induced expression of hypoxia inducible factor  $1\alpha$  (HIF- $1\alpha$ ). MiR-130 family has been identified to target DDX6 mRNA, which is a component of the P-bodies, and facilitate the translation of HIF- $1\alpha$  during hypoxia [110].

Dietary folate deficiency has been linked to developmental anomalies as well as increased risk for a number of cancers [115]. Low dietary folate levels lead to decreased DNA stability through misincorporation of uracil and subsequent DNA damage during the repair process as well as to increased genomic hypomethylation [116]. Arsenic exposure, similar to ionizing radiation and folate deficiency, has been linked to a variety of human cancers [117]. Marsit et al. examined the in vitro effect of folate deficiency and arsenic exposure on the expression pattern of 385 known human miRNAs using microarray analysis and they discovered alterations of specific miRNA profiles. The miRNAs that were significantly altered by 6-day folate deficiency treatment included miR-181b, 182, 222, 345, 181a, 205, 145, 99a, 125b, 130b, 221, 22, 191, 103, 107, 34a, 198, 183, 146, 422b, 210, 24, and 361 [33]. These results suggested that aberrant phenotypes might result from alterations in key miRNAs expression at critical stages in development and tumorigenesis due to altered folate status. Reduced dietary folate could lead to decreased level of S-adenosyl methionine, which is the required methylgroup donor for all cellular methylation reactions, including those for DNA and proteins such as histones [33]. The altered miRNAs expression profiles may be related to the altered methyl-donor pool and the subsequent epigenetic alterations at the DNA or histone code level, resulting in changes of miRNA and gene expression [33]. The miRNAs that were significantly altered by 6-day arsenic exposure included miR-210, 22, 34a, 221, and 222 [33], all of which were also altered by folate deficiency. Similar effects on genomic methylation status have also been reported for arsenic [118]. These results supported the hypothesis that arsenic may alter one-carbon metabolism and therefore cause downstream epigenetic effects.

#### 8. Radiation

DNA damage can be caused by multiple stressors including ionizing radiation, UV exposure, reactive oxygen species, and many DNA damaging chemicals, such as doxorubicin and camptothecin [119-121]. MiRNA-mediated gene regulation has been contributed to the mechanisms of DNA-damage caused by UV exposure and ionizing radiation. Studies by Pothof et al. suggested that the cellular responses to UVinduced DNA damage were also regulated at the posttranscriptional level by miRNAs. UV damage has been shown to cause cell-cycle-dependent relocalization of Ago2 into stress granules and changes of miRNA expression. Both events of miRNA-expression alteration and stress-granule formation occurred within the first hour following genotoxic stress, which suggested miRNAs-mediated gene regulation happened earlier than most transcriptional responses. The functionality of the miRNAs response was illustrated by the UV-inducible miR-16 which down-regulated checkpointgene CDC25a and regulated cell proliferation [122].

Ionizing radiation (IR) has been widely used in cancer therapy and biological studies. It disrupts cellular homeostasis through multiple mechanisms including the alteration of gene expression profile [123]. Shin et al. investigated the effects of IR on miRNA expression profile in the human lung carcinoma cell line A549. Microarray analysis identified 12 and 18 miRNAs in 20- and 40-Gy-exposed A549 cells, respectively, with more than 2-fold changes in their expression levels. Target prediction for IR-responsive miRNAs suggested that these miRNAs might target genes which are related to apoptosis, regulation of cell cycle, and DNA damage and repair. The results suggested that the expression levels of miRNAs could be affected by radiation and they might be involved in the regulation of radiation responses [123]. Niemoeller et al. investigated IR-induced miRNA expression profiles in six malignant cell lines. Their results showed that IR could cause 2-3 fold change of the expression level of miRNAs known to be involved in the regulation of cellular processes such as apoptosis, proliferation, invasion, local immune response, and radioresistance (e.g., miR-1285, 24-1, 151-5p, and let-7i). Moreover, they discovered several new miRNAs that are radiation-responsive (e.g., miR-144) [34].

Exposure to high-dose radiation causes ionization in the molecules of living cells, leads to DNA damage, and promotes tumor formation. On the other hand, low-dose radiation induces the beneficial effects on organisms, named as hormesis. Cha et al. identified responses of miRNAs to low- or high-dose *y*-irradiation in the human lymphoblast line IM9. They discovered that IR exposure could induce changes in specific miRNA expression in a dose-dependent manner and provided evidence that low-dose radiation could suppress the progression of malignant cancer by controlling miRNA expression [124].

#### 9. Conclusion and Future Directions

Environmental exposures including drugs, industrial materials, cigarette smoking, alcohol, stress, and radiation may cause a number of deleterious effects on health, development, and survival. Increased research has discovered the roles of miRNAs in the responses to environmental exposures under various conditions. The interactions of miRNAs and EFs have been associated with abnormal phenotypes and diseases. For example, specific miRNAs have been identified for different subgroups of HCC with various environmental risk factors. Investigation into the specific roles of miRNAs in the interactions between miRNAs and EFs may help uncovering the mechanisms of disease. These studies may potentially lead to the development of useful indicators of toxic exposure or novel biomarkers for diseases diagnosis. Studies of miRNAs in the disease context may also contribute to the development of novel pharmacologic approaches. The recent development of miRNA derivatives with increased stability and binding efficiency, including anti-microRNA oligonucleotides (AMOs) and LNAs served such purpose [87, 109]. As an example, targeting an HRM which plays a survival role in hypoxia may provide a new angle in targeting a notoriously refractory fraction of tumor cells [109]. Anti-miR-21 oligonucleotide has been shown to enhance the chemosensitivity of leukemic HL60 cells to arabinosylcytosine by inducing apoptosis [61]. Treatment of chronically infected chimpanzees with a LNA-modified oligonucleotide (SPC3649) complementary to miR-122 led to long-lasting suppression of HCV viremia [87]. Furthermore, manipulations of selected miRNAs could synergize with conventional therapies in the treatment of various diseases. Finally, insilico analysis and modeling may help studying patterns among miRNAs, EFs, and diseases and identifying novel strategies for disease diagnosis, treatment, and prognosis [125].

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### References

- R. C. Lee, R. L. Feinbaum, and V. Ambros, "The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14," *Cell*, vol. 75, no. 5, pp. 843–854, 1993.
- [2] R. C. Lee and V. Ambros, "An extensive class of small RNAs in *Caenorhabditis elegans*," *Science*, vol. 294, no. 5543, pp. 862– 864, 2001.
- [3] Y. Lee, K. Jeon, J. T. Lee, S. Kim, and V. N. Kim, "MicroRNA maturation: stepwise processing and subcellular localization," *EMBO Journal*, vol. 21, no. 17, pp. 4663–4670, 2002.
- [4] D. V. Dugas and B. Bartel, "MicroRNA regulation of gene expression in plants," *Current Opinion in Plant Biology*, vol. 7, no. 5, pp. 512–520, 2004.
- [5] W. Filipowicz, S. N. Bhattacharyya, and N. Sonenberg, "Mechanisms of post-transcriptional regulation by microR-NAs: are the answers in sight?" *Nature Reviews Genetics*, vol. 9, no. 2, pp. 102–114, 2008.
- [6] D. P. Bartel, "MicroRNAs: genomics, biogenesis, mechanism, and function," *Cell*, vol. 116, no. 2, pp. 281–297, 2004.
- [7] A. Esquela-Kerscher and F. J. Slack, "Oncomirs—microRNAs with a role in cancer," *Nature Reviews Cancer*, vol. 6, no. 4, pp. 259–269, 2006.
- [8] C. L. Bartels and G. J. Tsongalis, "Mini-reviews micrornas:novel biomarkers for human cancer," *Clinical Chemistry*, vol. 55, no. 4, pp. 623–631, 2009.
- [9] M. Lu, Q. Zhang, M. Deng et al., "An analysis of human microRNA and disease associations," *PLoS ONE*, vol. 3, no. 10, Article ID e3420, 2008.
- [10] A. M. Soto and C. Sonnenschein, "Environmental causes of cancer: endocrine disruptors as carcinogens," *Nature Reviews Endocrinology*, vol. 6, no. 7, pp. 363–370, 2010.
- [11] U. N. Das, "Obesity: genes, brain, gut, and environment," *Nutrition*, vol. 26, no. 5, pp. 459–473, 2010.
- [12] W. H. Chow, L. M. Dong, and S. S. Devesa, "Epidemiology and risk factors for kidney cancer," *Nature Reviews Urology*, vol. 7, no. 5, pp. 245–257, 2010.
- [13] S. M. Reamon-Buettner, V. Mutschler, and J. Borlak, "The next innovation cycle in toxicogenomics: environmental epigenetics," *Mutation Research*, vol. 659, no. 1-2, pp. 158– 165, 2008.
- [14] Q. Yang, C. Qiu, J. Yang, Q. Wu, and Q. Cui, "miREnvironment database: providing a bridge for microRNAs, environmental factors and phenotypes," *Bioinformatics*, vol. 27, pp. 3329–3330, 2011.
- [15] R. T. Lima, S. Busacca, G. M. Almeida, G. Gaudino, D. A. Fennell, and M. H. Vasconcelos, "MicroRNA regulation of core apoptosis pathways in cancer," *European Journal of Cancer*, vol. 47, no. 2, pp. 163–174, 2011.
- [16] E. Turrini, S. Haenisch, S. Laechelt, T. Diewock, O. Bruhn, and I. Cascorbi, "MicroRNA profiling in K-562 cells under imatinib treatment: influence of miR-212 and miR-328 on

ABCG2 expression," *Pharmacogenet Genomics*, vol. 22, pp. 198–205, 2012.

- [17] M. Avissar-Whiting, K. R. Veiga, K. M. Uhl et al., "Bisphenol A exposure leads to specific microRNA alterations in placental cells," *Reproductive Toxicology*, vol. 29, no. 4, pp. 401–406, 2010.
- [18] J. H. Kang, F. Kondo, and Y. Katayama, "Human exposure to bisphenol A," *Toxicology*, vol. 226, no. 2-3, pp. 79–89, 2006.
- [19] Z. Lin and E. K. Flemington, "MiRNAs in the pathogenesis of oncogenic human viruses," *Cancer Letters*, vol. 305, no. 2, pp. 186–199, 2011.
- [20] A. Eulalio, L. Schulte, and J. Vogel, "The mammalian microRNA response to bacterial infections," *RNA Biology*, vol. 9, no. 6, pp. 742–775, 2012.
- [21] Y. Ladeiro, G. Couchy, C. Balabaud et al., "MicroRNA profiling in hepatocellular tumors is associated with clinical features and oncogene/tumor suppressor gene mutations," *Hepatology*, vol. 47, no. 6, pp. 1955–1963, 2008.
- [22] R. C. Miranda, "MicroRNAs and fetal brain development: implications for ethanol teratology during the second trimester period of neurogenesis," *Frontiers in Genetics*, vol. 3, article 77, 2012.
- [23] J. D. Tapocik, M. Solomon, M. Flanigan et al., "Coordinated dysregulation of mRNAs and microRNAs in the rat medial 4 prefrontal cortex following a history of alcohol dependence," *The Pharmacogenomics Journal*. In press.
- [24] M. A. Maccani and V. S. Knopik, "Cigarette smoke exposureassociated alterations to non-coding RNA," *Frontiers in Genetics*, vol. 3, article 53, 2012.
- [25] A. Izzotti, P. Larghero, C. Cartiglia et al., "Modulation of microRNA expression by budesonide, phenethyl isothiocyanate and cigarette smoke in mouse liver and lung," *Carcinogenesis*, vol. 31, no. 5, pp. 894–901, 2010.
- [26] M. A. Maccani, M. Avissar-Whiting, C. E. Banister, B. McGonnigal, J. F. Padbury, and C. J. Marsit, "Maternal cigarette smoking during pregnancy is associated with downregulation of miR-16, miR-21 and miR-146a in the placenta," *Epigenetics*, vol. 5, no. 7, pp. 583–589, 2010.
- [27] A. Alisi, L. Da Sacco, G. Bruscalupi et al., "Mirnome analysis reveals novel molecular determinants in the pathogenesis of diet-induced nonalcoholic fatty liver disease," *Laboratory Investigation*, vol. 91, no. 2, pp. 283–293, 2011.
- [28] L. A. Davidson, N. Wang, M. S. Shah, J. R. Lupton, I. Ivanov, and R. S. Chapkin, "n-3 Polyunsaturated fatty acids modulate carcinogen-directed non-coding microRNA signatures in rat colon," *Carcinogenesis*, vol. 30, no. 12, pp. 2077–2084, 2009.
- [29] C. J. Davis, S. G. Bohnet, J. M. Meyerson, and J. M. Krueger, "Sleep loss changes microRNA levels in the brain: a possible mechanism for state-dependent translational regulation," *Neuroscience Letters*, vol. 422, no. 1, pp. 68–73, 2007.
- [30] S. Nielsen, C. Scheele, C. Yfanti et al., "Muscle specific microRNAs are regulated by endurance exercise in human skeletal muscle," *Journal of Physiology*, vol. 588, no. 20, pp. 4029–4037, 2010.
- [31] M. J. Drummond, J. J. McCarthy, C. S. Fry, K. A. Esser, and B. B. Rasmussen, "Aging differentially affects human skeletal muscle microRNA expression at rest and after an anabolic stimulus of resistance exercise and essential amino acids," *American Journal of Physiology*, vol. 295, no. 6, pp. E1333– E1340, 2008.
- [32] Y. Gidron, M. De Zwaan, K. Quint, and M. Ocker, "Influence of stress and health-behaviour on miRNA expression," *Molecular Medicine Reports*, vol. 3, no. 3, pp. 455–457, 2010.

- [33] C. J. Marsit, K. Eddy, and K. T. Kelsey, "MicroRNA responses to cellular stress," *Cancer Research*, vol. 66, no. 22, pp. 10843– 10848, 2006.
- [34] O. M. Niemoeller, M. Niyazi, S. Corradini et al., "MicroRNA expression profiles in human cancer cells after ionizing radiation," *Radiation Oncology*, vol. 6, no. 1, article 29, 2011.
- [35] R. Garzon, F. Pichiorri, T. Palumbo et al., "MicroRNA gene expression during retinoic acid-induced differentiation of human acute promyelocytic leukemia," *Oncogene*, vol. 26, no. 28, pp. 4148–4157, 2007.
- [36] S. Das, N. Foley, K. Bryan et al., "MicroRNA mediates DNA demethylation events triggered by retinoic acid during neuroblastoma cell differentiation," *Cancer Research*, vol. 70, no. 20, pp. 7874–7881, 2010.
- [37] X. M. Wu, X. Q. Shao, X. X. Meng et al., "Genome-wide analysis of microRNA and mRNA expression signatures in hydroxycamptothecin-resistant gastric cancer cells," *Acta Pharmacologica Sinica*, vol. 32, no. 2, pp. 259–269, 2011.
- [38] J. L. Tong, C. P. Zhang, F. Nie et al., "MicroRNA 506 regulates expression of PPAR alpha in hydroxycamptothecin-resistant human colon cancer cells," *FEBS Letters*, vol. 585, pp. 3560– 3568, 2011.
- [39] E. San José-Enériz, J. Román-Gómez, A. Jiménez-Velasco et al., "MicroRNA expression profiling in Imatinib-resistant Chronic Myeloid Leukemia patients without clinically significant ABL1-mutations," *Molecular Cancer*, vol. 8, article 69, 2009.
- [40] E. I. Zimmerman, C. M. Dollins, M. Crawford et al., "Lyn kinase-dependent regulation of miR181 and myeloid cell leukemia-1 expression: implications for drug resistance in myelogenous leukemia," *Molecular Pharmacology*, vol. 78, no. 5, pp. 811–817, 2010.
- [41] Y. Yu, L. Yang, M. Zhao et al., "Targeting microRNA-30a-mediated autophagy enhances imatinib activity against human chronic myeloid leukemia cells," *Leukemia*, vol. 26, no. 8, pp. 1752–1760, 2012.
- [42] M. Y. Shah, X. Pan, L. N. Fix, M. A. Farwell, and B. Zhang, "5-fluorouracil drug alters the microrna expression profiles in MCF-7 breast cancer cells," *Journal of Cellular Physiology*, vol. 226, no. 7, pp. 1868–1878, 2011.
- [43] L. Rossi, E. Bonmassar, and I. Faraoni, "Modification of miR gene expression pattern in human colon cancer cells following exposure to 5-fluorouracil in vitro," *Pharmacological Research*, vol. 56, no. 3, pp. 248–253, 2007.
- [44] T. Boren, Y. Xiong, A. Hakam et al., "MicroRNAs and their target messenger RNAs associated with ovarian cancer response to chemotherapy," *Gynecologic Oncology*, vol. 113, no. 2, pp. 249–255, 2009.
- [45] J. Hodzic, E. Giovannetti, B. D. Calvo, A. D. Adema, and G. J. Peters, "Regulation of deoxycytidine kinase expression and sensitivity to gemcitabine by micro-RNA 330 and promoter methylation in cancer cells," *Nucleosides, Nucleotides and Nucleic Acids*, vol. 30, pp. 1214–1222, 2011.
- [46] Q. S. Xia, Y. Ishigaki, L. Sun, R. Chen, and Y. Motoo, "Effect of anti-cancer drugs on the expression of BIC/miR-155 in human pancreatic cancer PANC-1 cells," *Zhonghua Yi Xue Za Zhi*, vol. 90, no. 2, pp. 123–127, 2010.
- [47] S. Samakoglu, D. S. Deevi, H. Li et al., "Preclinical rationale for combining an EGFR antibody with cisplatin/gemcitabine for the treatment of NSCLC," *Cancer Genomics Proteomics*, vol. 9, pp. 77–92, 2012.
- [48] A. Sorrentino, C. G. Liu, A. Addario, C. Peschle, G. Scambia, and C. Ferlini, "Role of microRNAs in drug-resistant ovarian

cancer cells," *Gynecologic Oncology*, vol. 111, no. 3, pp. 478–486, 2008.

- [49] W. Zhu, X. Shan, T. Wang, Y. Shu, and P. Liu, "MiR-181b modulates multidrug resistance by targeting BCL2 in human cancer cell lines," *International Journal of Cancer*, vol. 127, no. 11, pp. 2520–2529, 2010.
- [50] L. Guo, Y. Liu, Y. Bai, Y. Sun, F. Xiao, and Y. Guo, "Gene expression profiling of drug-resistant small cell lung cancer cells by combining microRNA and cDNA expression analysis," *European Journal of Cancer*, vol. 46, no. 9, pp. 1692– 1702, 2010.
- [51] R. Hummel, D. I. Watson, C. Smith et al., "Mir-148a improves response to chemotherapy in sensitive and resistant oesophageal adenocarcinoma and squamous cell carcinoma cells," *Journal of Gastrointestinal Surgery*, vol. 15, no. 3, pp. 429–438, 2011.
- [52] Y. Huang, A. Chuang, H. Hao et al., "Phospho-ΔNp63α is a key regulator of the cisplatin-induced microRNAome in cancer cells," *Cell Death and Differentiation*, vol. 18, no. 7, pp. 1220–1230, 2011.
- [53] E. Moussay, V. Palissot, L. Vallar et al., "Determination of genes and microRNAs involved in the resistance to fludarabine in vivo in chronic lymphocytic leukemia," *Molecular Cancer*, vol. 9, article 115, 2010.
- [54] R. Zhou, P. Yuan, Y. Wang et al., "Evidence for selective microRNAs and their effectors as common long-term targets for the actions of mood stabilizers," *Neuropsychopharmacol*ogy, vol. 34, no. 6, pp. 1395–1405, 2009.
- [55] H. Chen, N. Wang, M. Burmeister, and M. G. McInnis, "MicroRNA expression changes in lymphoblastoid cell lines in response to lithium treatment," *International Journal of Neuropsychopharmacology*, vol. 12, no. 7, pp. 975–981, 2009.
- [56] H. Rong, T. B. Liu, K. J. Yang et al., "MicroRNA-134 plasma levels before and after treatment for bipolar mania," *Journal* of Psychiatric Research, vol. 45, no. 1, pp. 92–95, 2011.
- [57] W. P. Tsang and T. T. Kwok, "Epigallocatechin gallate upregulation of miR-16 and induction of apoptosis in human cancer cells," *Journal of Nutritional Biochemistry*, vol. 21, no. 2, pp. 140–146, 2010.
- [58] S. Ali, A. Ahmad, S. Banerjee et al., "Gemcitabine sensitivity can be induced in pancreatic cancer cells through modulation of miR-200 and miR-21 expression by curcumin or its analogue CDF," *Cancer Research*, vol. 70, no. 9, pp. 3606– 3617, 2010.
- [59] C. D. Davis and S. A. Ross, "Evidence for dietary regulation of microRNA expression in cancer cells," *Nutrition Reviews*, vol. 66, no. 8, pp. 477–482, 2008.
- [60] J. Yang, Y. Cao, J. Sun, and Y. Zhang, "Curcumin reduces the expression of Bcl-2 by upregulating miR-15a and miR-16 in MCF-7 cells," *Medical Oncology*, vol. 27, no. 4, pp. 1114– 1118, 2010.
- [61] Y. Li, X. Zhu, J. Gu et al., "Anti-miR-21 oligonucleotide enhances chemosensitivity of leukemic HL60 cells to arabinosylcytosine by inducing apoptosis," *Hematology*, vol. 15, no. 4, pp. 215–221, 2010.
- [62] J. Ji, J. Shi, A. Budhu et al., "MicroRNA expression, survival, and response to interferon in liver cancer," *New England Journal of Medicine*, vol. 361, no. 15, pp. 1437–1447, 2009.
- [63] C. A. McClung and E. J. Nestler, "Neuroplasticity mediated by altered gene expression," *Neuropsychopharmacology*, vol. 33, no. 1, pp. 3–17, 2008.
- [64] A. Z. Pietrzykowski, R. M. Friesen, G. E. Martin et al., "Posttranscriptional regulation of BK channel splice variant

stability by miR-9 underlies neuroadaptation to alcohol," *Neuron*, vol. 59, no. 2, pp. 274–287, 2008.

- [65] V. Chandrasekar and J. L. Dreyer, "microRNAs miR-124, let-7d and miR-181a regulate cocaine-induced plasticity," *Molecular and Cellular Neuroscience*, vol. 42, no. 4, pp. 350– 362, 2009.
- [66] V. Chandrasekar and J. L. Dreyer, "Regulation of MiR-124, Let-7d, and MiR-181a in the accumbens affects the expression, extinction, and reinstatement of cocaine-induced conditioned place preference," *Neuropsychopharmacology*, vol. 36, no. 6, pp. 1149–1164, 2011.
- [67] A. Wanet, A. Tacheny, T. Arnould, and P. Renard, "miR-212/132 expression and functions: within and beyond the neuronal compartment," *Nucleic Acids Research*, vol. 40, pp. 4742–4753, 2012.
- [68] J. A. Hollander, H. I. Im, A. L. Amelio et al., "Striatal microRNA controls cocaine intake through CREB signalling," *Nature*, vol. 466, no. 7303, pp. 197–202, 2010.
- [69] H. I. Im, J. A. Hollander, P. Bali, and P. J. Kenny, "MeCP2 controls BDNF expression and cocaine intake through homeostatic interactions with microRNA-212," *Nature Neuroscience*, vol. 13, no. 9, pp. 1120–1127, 2010.
- [70] S. L. Lin, H. Kim, and S. Y. Ying, "Intron-mediated RNA interference and microRNA (miRNA)," *Frontiers in Bioscience*, vol. 13, no. 6, pp. 2216–2230, 2008.
- [71] D. Bhaumik, G. K. Scott, S. Schokrpur, C. K. Patil, J. Campisi, and C. C. Benz, "Expression of microRNA-146 suppresses NF-κB activity with reduction of metastatic potential in breast cancer cells," *Oncogene*, vol. 27, no. 42, pp. 5643–5647, 2008.
- [72] M. J. Jardim, R. C. Fry, I. Jaspers, L. Dailey, and D. Diaz-Sanchez, "Disruption of MicroRNA expression in human airway cells by diesel exhaust particles is linked to tumorigenesis-associated pathways," *Environmental Health Perspectives*, vol. 117, no. 11, pp. 1745–1751, 2009.
- [73] V. Bollati, B. Marinelli, P. Apostoli et al., "Exposure to metalrich particulate matter modifies the expression of candidate MicroRNAs in peripheral blood leukocytes," *Environmental Health Perspectives*, vol. 118, no. 6, pp. 763–768, 2010.
- [74] X. Wang, L. Ye, Y. Zhou, M. Q. Liu, D. J. Zhou, and W. Z. Ho, "Inhibition of anti-HIV microRNA expression: a mechanism for opioid-mediated enhancement of HIV infection of monocytes," *American Journal of Pathology*, vol. 178, no. 1, pp. 41–47, 2011.
- [75] G. Sun, H. Li, X. Wu et al. et al., "Interplay between HIV-1 infection and host microRNAs," *Nucleic Acids Research*, vol. 40, pp. 2181–2196, 2012.
- [76] M. T. Bolisetty, G. Dy, W. Tam, and K. L. Beemon, "Reticuloendotheliosis virus strain T induces miR-155, which targets JARID2 and promotes cell survival," *Journal of Virology*, vol. 83, no. 23, pp. 12009–12017, 2009.
- [77] Y. J. Ma, J. Yang, X. L. Fan et al. et al., "Cellular microRNA let-7c inhibits M1 protein expression of the H1N1 influenza A virus in infected human lung epithelial cells," *Journal of Cellular and Molecular Medicine*, vol. 16, no. 10, pp. 2539– 2546, 2012.
- [78] N. Motsch, T. Pfuhl, J. Mrazek, S. Barth, and F. A. Grässer, "Epstein-Barr virus-encoded latent membrane protein 1 (LMP1) induces the expression of the cellular microRNA miR-146a," *RNA Biology*, vol. 4, no. 3, pp. 131–137, 2007.
- [79] J. H. Kim, W. S. Kim, and C. Park, "Epstein-Barr virus latent membrane protein-1 protects B-cell lymphoma from rituximab-induced apoptosis through miR-155-mediated

Akt activation and up-regulation of Mcl-1," Leukemia & Lymphoma, vol. 53, no. 8, pp. 1586–1591, 2012.

- [80] M. Tomita, Y. Tanaka, and N. Mori, "MicroRNA miR-146a is induced by HTLV-1 tax and increases the growth of HTLV-1infected T-cells," *International Journal of Cancer*, vol. 130, pp. 2300–2309, 2012.
- [81] K. Pichler, G. Schneider, and R. Grassmann, "MicroRNA miR-146a and further oncogenesis-related cellular microR-NAs are dysregulated in HTLV-1-transformed T lymphocytes," *Retrovirology*, vol. 5, article 100, 2008.
- [82] K. V. Mohan, K. Devadas, S. Sainath Rao, I. Hewlett, and C. Atreya, "Identification of XMRV infection-associated microRNAs in four cell types in culture," *PLoS ONE*, vol. 7, Article ID e32853, 2012.
- [83] A. Dittmer and K. Forstemann, "Murine cytomegalovirus infection of cultured mouse cells induces expression of miR-7a," *Journal of General Virology*, vol. 93, pp. 1537–1547, 2012.
- [84] D. Santhakumar, T. Forster, N. N. Laqtom et al., "Combined agonist-antagonist genome-wide functional screening identifies broadly active antiviral microRNAs," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 31, pp. 13830–13835, 2010.
- [85] Y. Zhang, Y. Jia, R. Zheng et al., "Plasma microRNA-122 as a biomarker for viral-, alcohol-, and chemical-related hepatic diseases," *Clinical Chemistry*, vol. 56, no. 12, pp. 1830–1838, 2010.
- [86] G. L. Zhang, Y. X. Li, S. Q. Zheng, M. Liu, X. Li, and H. Tang, "Suppression of hepatitis B virus replication by microRNA-199a-3p and microRNA-210," *Antiviral Research*, vol. 88, no. 2, pp. 169–175, 2010.
- [87] R. E. Lanford, E. S. Hildebrandt-Eriksen, A. Petri et al., "Therapeutic silencing of microRNA-122 in primates with chronic hepatitis C virus infection," *Science*, vol. 327, no. 5962, pp. 198–201, 2010.
- [88] C. Braconi, N. Valeri, P. Gasparini et al., "Hepatitis C virus proteins modulate microRNA expression and chemosensitivity in malignant hepatocytes," *Clinical Cancer Research*, vol. 16, no. 3, pp. 957–966, 2010.
- [89] M. P. Myklebust, O. Bruland, O. Fluge, A. Skarstein, L. Balteskard, and O. Dahl, "MicroRNA-15b is induced with E2F-controlled genes in HPV-related cancer," *British Journal* of Cancer, vol. 105, pp. 1719–1725, 2011.
- [90] C. L. Jopling, M. Yi, A. M. Lancaster, S. M. Lemon, and P. Sarnow, "Molecular biology: modulation of hepatitis C virus RNA abundance by a liver-specific MicroRNA," *Science*, vol. 309, no. 5740, pp. 1577–1581, 2005.
- [91] J. I. Henke, D. Goergen, J. Zheng et al., "microRNA-122 stimulates translation of hepatitis C virus RNA," *EMBO Journal*, vol. 27, no. 24, pp. 3300–3310, 2008.
- [92] L. Qiu, H. Fan, W. Jin et al., "MiR-122-induced downregulation of HO-1 negatively affects miR-122-mediated suppression of HBV," *Biochemical and Biophysical Research Communications*, vol. 398, no. 4, pp. 771–777, 2010.
- [93] F. El-Assaad, C. Hempel, V. Combes et al., "Differential microRNA expression in experimental cerebral and noncerebral malaria," *Infection and Immunity*, vol. 79, no. 6, pp. 2379–2384, 2011.
- [94] D. Delić, M. Dkhil, S. Al-Quraishy, and F. Wunderlich, "Hepatic miRNA expression reprogrammed by Plasmodium chabaudi malaria," *Parasitology Research*, vol. 108, no. 5, pp. 1111–1121, 2011.

- [95] M. A. Maccani and C. J. Marsit, "Exposure and fetal growthassociated miRNA alterations in the human placenta," *Clinical Epigenetics*, vol. 2, pp. 401–404, 2011.
- [96] S. Xi, M. Yang, Y. Tao et al., "Cigarette smoke induces C/EBP-β-mediated activation of mir-31 in normal human respiratory epithelia and lung cancer cells," *PLoS ONE*, vol. 5, no. 10, Article ID e13764, 2010.
- [97] F. Schembri, S. Sridhar, C. Perdomo et al., "MicroRNAs as modulators of smoking-induced gene expression changes in human airway epithelium," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 7, pp. 2319–2324, 2009.
- [98] A. Izzotti, G. A. Calin, P. Arrigo, V. E. Steele, C. M. Croce, and S. De Flora, "Downregulation of microRNA expression in the lungs of rats exposed to cigarette smoke," *The FASEB Journal*, vol. 23, no. 3, pp. 806–812, 2009.
- [99] P. Kaur, A. Armugam, and K. Jeyaseelan, "MicroRNAs in neurotoxicity," *Journal of Toxicology*, vol. 2012, Article ID 870150, 15 pages, 2012.
- [100] P. Sathyan, H. B. Golden, and R. C. Miranda, "Competing interactions between micro-RNAs determine neural progenitor survival and proliferation after ethanol exposure: evidence from an ex vivo model of the fetal cerebral cortical neuroepithelium," *Journal of Neuroscience*, vol. 27, no. 32, pp. 8546–8557, 2007.
- [101] S. Balaraman, U. H. Winzer-Serhan, and R. C. Miranda, "Opposing actions of ethanol and nicotine on microRNAs are mediated by nicotinic acetylcholine receptors in fetal cerebral cortical-derived neural progenitor cells," *Alcoholism: Clinical and Experimental Research*, vol. 36, no. 10, pp. 1669– 1677, 2012.
- [102] T. L. Tal, J. A. Franzosa, S. C. Tilton et al., "MicroR-NAs control neurobehavioral development and function in zebrafish," *The FASEB Journal*, vol. 26, pp. 1452–1461, 2012.
- [103] S. Bala and G. Szabo, "MicroRNA signature in alcoholic liver disease," *International Journal of Hepatology*, vol. 2012, Article ID 498232, 6 pages, 2012.
- [104] H. Yin, M. Hu, R. Zhang, Z. Shen, L. Flatow, and M. You, "MicroRNA-217 promotes ethanol-induced fat accumulation in hepatocytes by down-regulating SIRT1," *The Journal* of *Biological Chemistry*, vol. 287, pp. 9817–9826, 2012.
- [105] K. R. Diller, "Stress protein expression kinetics," Annual Review of Biomedical Engineering, vol. 8, pp. 403–424, 2006.
- [106] B. A. Miller, "The role of TRP channels in oxidative stressinduced cell death," *Journal of Membrane Biology*, vol. 209, no. 1, pp. 31–41, 2006.
- [107] J. Dresios, A. Aschrafi, G. C. Owens, P. W. Vanderklish, G. M. Edelman, and V. P. Mauro, "Cold stress-induced protein Rbm3 binds 60S ribosomal subunits, alter microRNA levels, and enhances global protein synthesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 6, pp. 1865–1870, 2005.
- [108] G. J. Wilmink, C. L. Roth, B. L. Ibey et al., "Identification of microRNAs associated with hyperthermia-induced cellular stress response," *Cell Stress and Chaperones*, vol. 15, no. 6, pp. 1027–1038, 2010.
- [109] R. Kulshreshtha, R. V. Davuluri, G. A. Calin, and M. Ivan, "A microRNA component of the hypoxic response," *Cell Death* and Differentiation, vol. 15, no. 4, pp. 667–671, 2008.
- [110] K. Saito, E. Kondo, and M. Matsushita, "MicroRNA 130 family regulates the hypoxia response signal through the Pbody protein DDX6," *Nucleic Acids Research*, vol. 39, pp. 6086–6099, 2011.

- [111] Y. C. Chan, S. Khanna, S. Roy, and C. K. Sen, "MiR-200b targets Ets-1 and is down-regulated by hypoxia to induce angiogenic response of endothelial cells," *Journal of Biological Chemistry*, vol. 286, no. 3, pp. 2047–2056, 2011.
- [112] R. Kulshreshtha, M. Ferracin, S. E. Wojcik et al., "A microRNA signature of hypoxia," *Molecular and Cellular Biology*, vol. 27, no. 5, pp. 1859–1867, 2007.
- [113] Z. Hua, Q. Lv, W. Ye et al., "Mirna-directed regulation of VEGF and other angiogenic under hypoxia," *PLoS ONE*, vol. 1, no. 1, article e116, 2006.
- [114] R. B. Donker, J. F. Mouillet, D. M. Nelson, and Y. Sadovsky, "The expression of Argonaute2 and related microRNA biogenesis proteins in normal and hypoxic trophoblasts," *Molecular Human Reproduction*, vol. 13, no. 4, pp. 273–279, 2007.
- [115] R. Jiang, F. B. Hu, E. L. Giovannucci et al., "Joint association of alcohol and folate intake with risk of major chronic disease in women," *American Journal of Epidemiology*, vol. 158, no. 8, pp. 760–771, 2003.
- [116] S. J. James, I. P. Pogribny, M. Pogribna, B. J. Miller, S. Jernigan, and S. Melnyk, "Mechanisms of DNA damage, DNA hypomethylation, and tumor progression in the folate/methyl-deficient rat model of hepatocarcinogenesis," *Journal of Nutrition*, vol. 133, pp. 3740S–3747S, 2003.
- [117] H. R. Guo, "Cancer risks associated with arsenic in drinking water," *Environmental Health Perspectives*, vol. 115, no. 7, pp. A339–A340, 2007.
- [118] H. Chen, S. F. Li, J. Liu, B. A. Diwan, J. C. Barrett, and M. P. Waalkes, "Chronic inorganic arsenic exposure induces hepatic global and individual gene hypomethylation: implications for arsenic hepatocarcinogenesis," *Carcinogenesis*, vol. 25, no. 9, pp. 1779–1786, 2004.
- [119] S. Bhana and D. R. Lloyd, "The role of p53 in DNA damage-mediated cytotoxicity overrides its ability to regulate nucleotide excision repair in human fibroblasts," *Mutagene*sis, vol. 23, no. 1, pp. 43–50, 2008.
- [120] P. Pitsikas, D. Lee, and A. J. Rainbow, "Reduced host cell reactivation of oxidative DNA damage in human cells deficient in the mismatch repair gene hMSH2," *Mutagenesis*, vol. 22, no. 3, pp. 235–243, 2007.
- [121] L. Barbour, L. G. Ball, K. Zhang, and W. Xiao, "DNA damage checkpoints are involved in postreplication repair," *Genetics*, vol. 174, no. 4, pp. 1789–1800, 2006.
- [122] J. Pothof, N. S. Verkaik, W. Van Ijcken et al., "MicroRNAmediated gene silencing modulates the UV-induced DNAdamage response," *EMBO Journal*, vol. 28, no. 14, pp. 2090– 2099, 2009.
- [123] S. Shin, H. J. Cha, E. M. Lee et al., "Alteration of miRNA profiles by ionizing radiation in A549 human non-small cell lung cancer cells," *International Journal of Oncology*, vol. 35, no. 1, pp. 81–86, 2009.
- [124] H. J. Cha, K. M. Seong, S. Bae et al., "Identification of specific microRNAs responding to low and high dose *y*-irradiation in the human lymphoblast line IM9," *Oncology Reports*, vol. 22, no. 4, pp. 863–868, 2009.
- [125] C. Qiu, G. Chen, and Q. Cui, "Towards the understanding of microRNA and environmental factor interactions and their relationships to human diseases," *Scientific Reports*, vol. 2, article 318, 2012.