LAB/IN VITRO RESEARCH

e-ISSN 1643-3750 © Med Sci Monit, 2017; 23: 1286-1294 DOI: 10.12659/MSM.903248

Received	1: 2017.01.09	-	High Expression of Ang	iogenic Factor with	
Accepted	1: 2017.02.01		G-Patch and FHA Domai	in1 (AGGF1) Predicts Poor	
Published	1: 2017.03.14		Prognosis in Gastric Car	ncer	
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Source of support:		f support:	Departmental sources		
Background:		ground:	Angiogenic factor with G-patch and FHA domain1 (AGGF1 or VG5Q) is a newly identified human angiogenic factor. The aim of this study was to explore AGGF1 expression level in gastric cancer and detect its correlation with the prognosis.		
Material/Methods:		Iethods:	Immunohistochemistry was performed to detect AGGF1 level in gastric cancer and its adjacent noncancerous samples of 198 cases, and the relationships among the expression levels of AGGF1, vascular endothelial growth factor (VEGF) and prognosis were analyzed		
Results: Conclusions:		Results: :lusions:	Expression of AGGF1 in gastric cancer samples was significantly higher than that in adjacent noncancerous samples ($P<0.001$). The overall survival rate (OS) of patients with high AGGF1 expression was significantly lower than that of patients with low AGGF1 expression ($P=0.000$). The Cox model analysis demonstrated that expression of AGGF1 was an independent biomarker for prediction of patients' survival in gastric cancer. High expression of AGGF1 predicts poor prognosis in gastric cancer patients. AGGF1 can be used as an independent factor to predict postoperative survival of patients with gastric cancer.		
MeSH Keywords:		ywords:	Angiogenesis Inducing Agents • Stomach Neoplasms • Vascular Endothelial Growth Factor A		
	Full-t	ext PDF:	http://www.medscimonit.com/abstract/index/idArt/903248		



MEDICAL SCIENCE MONITOR

Background

Gastric cancer (GC) is one of the most aggressively malignant tumors of the digestive tract. Most patients have been in advanced stage at diagnosis and the effectiveness of surgery is limited. Invasion and metastasis is the main cause of death in patients with gastric cancer. Among the potential promoting factors, tumor angiogenesis plays an important role [1,2]. Tumor angiogenesis is the basis of tumor growth and metastasis. Therefore, it an important focus in the study of angiogenesis in gastric cancer and the search for new potential therapeutic targets.

Angiogenic factor with G-patch and FHA domain1 (AGGF1 or VG5Q), as a newly identified human angiogenic factor, was first reported by Tian et al. [3] in 2004. The gene is highly expressed in vascular endothelial cells and the encoded protein has a strong angiogenesis ability *in vitro*. Recent studies have found that AGGF1 is expressed in some types of malignant tumors and is closely related to tumor angiogenesis [4–7]. Obviously, persistent angiogenesis, as one of the main signs of tumor, is closely related to the growth, invasion, metastasis, and recurrence of gastric cancer [1,2], but the expression level of AGGF1 and its prognostic value in patients with gastric cancer have not been reported.

Therefore, in the present study, the protein expression levels of AGGF1 and vascular endothelial growth factor (VEGF) were examined by immunohistochemistry in GC and corresponding noncancerous samples. Next, Kaplan-Meier curves and log rank test were applied to analyze the survival rate. Lastly, Cox regression method was used to explore the prognostic value of AGGF1 in gastric cancer.

Material and Methods

Patient and clinicopathologic data

We selected specimens from 198 cases of gastric cancer (GC), along with the corresponding noncancerous tissues, from patients diagnosed at the Anhui Provincial Hospital of Anhui Medical University (Hefei, China) between 2007 and 2011. Detailed pathological and clinical data (including age, sex, tumor size, Borrmann type, degree of differentiation, histological type, metastasis of lymph node, invasion depth, and TNM staging) were obtained from each patient's medical records. The samples were obtained from 58 female and 140 male patients with an average age of 56±13 years old (range, 26–82 years). None of the patients had received radiotherapy or chemotherapy before surgery. The specimens were fixed in formalin and embedded in paraffin for pathological analysis and confirmation of the diagnosis. Complete clinical follow-up data was obtained from the gastric cancer database of our hospital. The study was approved by the Anhui Medical University Human Research Ethics Committee. Written informed consent was obtained from each patient.

Immunohistochemical study

Immunohistochemistry for AGGF1 and VEGF (both antibody concentrations were 1: 500) was performed on each cancerous and corresponding noncancerous tissue. The samples (4- μ m thick) were cut onto salinized glass slides consecutively. Two-step immunohistochemistry was used to detect these proteins expression according to the manufacturer's instructions.

Every section was scored on the basis of the stained tumor cells fraction and staining intensity. The proportion was classified as $0 (\leq 1\%)$, 1 (2% to 25%), 2 (26% to 50%), 2 (51% to 75%), and 4 (\geq 76%). The staining intensity was scored as 0 (no staining), 1 (weak), 2 (moderate), and 3 (strong). The expression result was calculated according to the formula: percentage score multiplied by intensity score. Total scores (0–12) were categorized as low (score 0–3) or high (score 4–12).

Statistical analysis

SPSS 20.0 software (SPSS, Inc., Chicago, IL, USA) was used for all statistical analyses. Chi-square test and Spearman correlation test were used to analyze the immunohistochemical results. Kaplan-Meier and log rank test were applied to analyze the survival rates of patients. Cox regression method was used to determine the prognostic value. A *P*-value less than 0.05 was considered to indicate statistical significance.

Results

AGGF1 expression in cancerous and noncancerous gastric tissues

In total, 198 paired cancerous and noncancerous tissue samples were analyzed by immunohistochemistry for AGGF1 expression. The AGGF1 immunoreactivity was mainly observed in the cytoplasm of neoplastic cells. High expression of AGGF1 was found in most cancer samples (132/198) and in fewer non-cancerous samples (48/198). The expression level of AGGF1 in gastric cancer was dramatically higher than that in noncancerous samples (P<0.001). Representative GC samples with different AGGF1 expression patterns are shown in Figure 1.



Figure 1. Positive (A) and negative (B) expression of AGGF1 in gastric cancer and corresponding noncancerous tissues by immunohistochemistry, respectively (200× magnification).

Correlation of AGGF1 with clinicopathological factors and VEGF

As shown in Table 1, the expression of AGGF1 was remarkably associated with lymph node metastasis (P=0.022), invasion depth (P=0.006), and TNM stage (P<0.001). Additionally, we also found there was a significantly positive correlation between VEGF and AGGF1 expression in gastric cancer samples (P=0.017, Figure 2).

Correlation of AGGF1 with patients' prognosis

Kaplan-Meier method was plotted to compare the OS and DFS according to AGGF1 expression patterns. Patients with high-expression tumors showed a more unfavorable prognosis than those with low-expression tumors (Figure 3). Univariate survival analysis (Tables 2, 3) revealed AGGF1 expression was remarkably associated with OS (P<0.001) and DFS (P<0.001), in addition to lymph node metastasis (P<0.001 for OS, P<0.001 for DFS), invasion depth (P=0.001 for OS, P<0.001 for DFS), and TNM stage (P<0.001 for OS, P<0.001 for OS, P=0.002 for DFS), invasion depth (P=0.024 for OS, P=0.024 for DFS), TNM stage (P<0.001 for OS, P<0.001 for OS, P<0.

Discussion

Since the gene was first reported, AGGF1 and its physiological functions were further revealed, especially in the cardiovascular

system. Chen et al. [8] explored the function of AGGF1 in the angiogenesis of zebrafish and found that AGGF1 regulated the formation of blood vessels and the differentiation of veins. Lu et al. [9] administered the angiogenic therapy in a mouse hindlimb ischemia model by using AGGF1 gene, which improved blood supply to the ischemic area. Another study found that AGGF1 inhibits vascular inflammatory response and improves endothelial function [10]. Based on these findings, we speculate that AGGF1 plays an important role in the growth, metastasis, and invasion of gastric cancer. We found the expression level of AGGF1 protein was significantly higher in gastric cancer tissue than that in the corresponding noncancerous tissue. Similar to our results, a recent study found that hepatocellular carcinoma also displays overexpression of AGGF1 [7]. Furthermore, patients with high AGGF1 expression had dramatically lower DFS and OS than those with low AGGF1 expression. Additionally, high AGGF1 expression in patients with gastric cancer was closely related to poor prognosis, as demonstrated by univariate and multivariate analyses.

Tumor angiogenesis plays a pivotal role in the progression and development of gastric cancer. Overexpression of VEGF is associated with unfavorable prognosis and aggressive behavior of tumors [11]. Moreover, several studies have demonstrated that increased VEGF expression and microvessel density (MVD) are strongly related to worse prognosis in gastric cancer patients [12–15]. Therefore, to explore the role of AGGF1 in angiogenesis of gastric cancer, we explored the relationships between VEGF and AGGF1 expression levels in GC tissues. We also found a significantly positive relationship between AGGF1 and VEGF expressions in gastric cancer tissues,

variables	Total	AGGF1 expression				
- Variables	Total	Low (n=66)	High (n=132)	χ²	<i>P</i> value	
Gender						
Male	140	44	96	0.780	0.377	
Female	58	22	36			
Age at surgery (yeas)						
≤60	94	35	59	1.225	0.268	
>60	104	31	73			
Size of primary tumor (cm)						
≤5	101	34	67	0.010	0.920	
>5	97	32	65			
Borrmann type						
I+ II type	67	23	44	0.045	0.832	
III+IV type	131	43	88			
Degree of differentiation						
Well/moderate	85	30	55	0.258	0.612	
Poor and not	113	36	77			
Histological type						
Adenocarcinoma	167	57	110	0.306	0.580	
Others	31	9	22			
Depth of invasion						
T1	8	5	3	12.388	0.006	
T2	24	14	10			
T3	62	20	42			
T4	94	27	77			
Lymph node metastasis						
NO	44	20	24	9.602	0.022	
N1	47	20	27			
N2	56	16	40			
N3	51	10	41			
TNM stage						
I	13	9	4	18.044	0.000	
	57	29	34			
III	119	25	83			
IV	9	3	11			
VEGF expression						
Low	76	33	43	5.648	0.017	
High	122	33	89			

Table 1. Correlations between AGGF1 protein expressions and clinicopathological factors in patients with gastric cancer.



Figure 2. High expression levels of AGGF1 (A) and VEGF (B) in gastric cancer (200× magnification).



Figure 3. Kaplan-Meier analysis of overall survival (OS) and disease-free survival (DFS) curves of patients with gastric cancer based on AGGF1 expression as positive or negative. (A) OS curve of patients with gastric cancer based on AGGF1 expression; (B) DFS curve of patients with gastric cancer based on AGGF1 expression.

suggesting that AGGF1, probably cooperating with VEGF, is involved in tumor angiogenesis of gastric cancer. The potential underlying mechanisms may be that AGGF1 induces the expression of VEGF through β -catenin-dependent signaling [4].

However, some limitations should be acknowledged in this study. Firstly, it was a retrospective study with relatively small samples. Secondly, we only used immunohistochemical method to examine the protein expression levels of AGGF1 and VEGF in gastric cancer tissues, and the gene expression level was not assessed. Lastly, the exact underlying mechanisms in the participation of AGGF1 in angiogenesis of gastric cancer need to be further explored.

Conclusions

In summary, our preliminary results show that AGGF1 protein is overexpressed in gastric cancer tissues and it can be used as an independent parameter to evaluate and predict the postoperative survival time of gastric cancer patients. The potential mechanism is probably related to the promotion of tumor angiogenesis. In future, targeting AGGF1 for the inhibition of angiogenesis may be a new therapeutic strategy for gastric cancer patients.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Table 2. Univariate analysis of the correlation between	clinicopathological parameters and overall survival time of patients with gastric
cancer.	

Variables	Mean survival time (m)	95% CI	Log-rank test	<i>P</i> value
Gender				
Male	45.013	40.729–49.296	0.344	0.557
Female	42.637	35.896–49.379		
Age at surgery (yeas)				
≤60	45.585	40.366–50.803	0.377	0.539
>60	43.358	38.348–48.367		
Size of primary tumor (cm)				
≤5	43.387	38.485–48.289	0.236	0.627
>5	45.577	40.253–50.901		
Borrmann type				
l+ ll type	46.035	39.715–52.354	0.396	0.529
III+IV type	43.472	39.063–47.881		
Degree of differentiation				
Well/moderate	48.339	42.980–53.697	3.494	0.062
Poor and not	41.237	36.406–46.067		
Histological type				
Adenocarcinoma	43.013	39.097–46.930	2.770	0.096
Others	50.453	41.753–59.153		
Depth of invasion				
T1	71.000	69.303–72.697	16.372	0.001
T2	60.055	51.567–68.544		
Т3	42.322	35.986–48.657		
T4	39.659	34.737–44.580		
Lymph node metastasis				
NO	52.465	45.378–59.551	21.639	0.000
N1	53.543	46.738-60.348		
N2	40.204	33.820–46.588		
N3	33.154	26.253-40.056		
TNM stage				
I	71.200	69.798–72.602	50.264	0.000
ll	49.310	43.488–55.132		
III	41.309	36.395–46.223		
IV	16.698	11.017–22.379		
AGGF1 expression				
Low	57.777	53.817–62.737	22.538	0.000
High	37.830	33.433-42.227		

 Table 3. Univariate analysis of the correlation between clinicopathological parameters and disease free survival time of patients with gastric cancer.

Variables	Mean survival time (m)	95% CI	Log-rank test	P value
Gender				
Male	42.455	37.772–47.138	0.124	0.725
Female	40.506	33.253–47.758		
Age at surgery (yeas)				
≤60	43.372	37.678–49.067	0.328	0.567
>60	40.841	35.408–46.274		
Size of primary tumor (cm)				
≤5	40.844	35.509–46.179	.327	.567
>5	43.252	37.460–49.044		
Borrmann type				
l+ ll type	43.836	37.014–50.657	0.458	0.499
III+IV type	41.056	36.244–45.869		
Degree of differentiation				
Well/moderate	45.897	40.024–51.769	2.837	0.092
Poor and not	41.237	33.779–44.274		
Histological type				
Adenocarcinoma	40.579	36.340-44.818	2.430	0.119
Others	48.141	38.375-57.907		
Depth of invasion				
T1	69.250	64.582–73.918	16.505	0.001
T2	59.471	50.651-68.290		
Т3	40.118	33.162-47.074		
T4	36.727	31.346-42.108		
Lymph node metastasis				
NO	50.134	42.355-57.914	19.960	0.000
N1	51.141	43.721–58.561		
N2	37.224	30.221-44.228		
N3	30.373	22.813-37.934		
TNM stage				
I	70.429	67.577–73.280	44.100	0.000
II	47.130	40.785–53.474		
III	38.787	33.389–44.186		
IV	12.800	7.380–18.220		
AGGF1 expression				
Low	56.509	51.135–61.882	23.489	0.000
High	34.898	30.098–39.699		

Table 4. Multivariate analysis of the correlation between clinicopathological parameters and overall survival time of patients with gastric cancer.

Covariates	HR	95% CI for HR	P value
Gender (male vs. female)	0.817	0.527-1.267	0.367
Age (≥60 <i>vs</i> . <60 cm)	1.052	0.704–1.573	0.803
Tumor size (≥5 <i>vs</i> . <5 cm)	1.031	0.679–1.566	0.886
Borrmann type (type I, II vs. III, IV)	1.131	0.734–1.743	0.578
Degree of differentiation	0.877	0.584–1.318	0.528
Histological type	1.539	0.822–2.882	0.178
Depth of invasion (T3, T4 vs. T1, T2)	0.341	0.135–0.865	0.024
Lymph node metastasis	0.311	0.157–0.615	0.001
TNM stage (stage I vs. II vs. III vs. IV)	0.161	0.079–0.331	0.000
AGGF1 expression (low vs. high)	0.354	0.213–0.586	0.000

Table 5. Multivariate analysis of the correlation between clinicopathological parameters and disease free survival time of patients with gastric cancer.

Covariates	HR	95% CI for HR	P value
Gender (male vs. female)	0.895	0.579–1.382	0.616
Age (≥60 <i>vs</i> . <60 cm)	1.030	0.688–1.543	0.886
Tumor size (≥5 <i>vs</i> . <5 cm)	1.045	0.689–1.586	0.836
Borrmann type (type I, II vs. III, IV)	1.102	0.715–1.696	0.660
Degree of differentiation	0.909	0.605–1.364	0.644
Histological type	1.483	0.792–2.778	0.218
Depth of invasion (T3, T4 vs. T1, T2)	0.347	0.138-0.869	0.024
Lymph node metastasis	0.334	0.169–0.658	0.002
TNM stage (stage I vs. II vs. III vs. IV)	0.196	0.096–0.401	0.000
AGGF1 expression (low vs. high)	0.366	0.222-0.604	0.000

References:

- 1. Jia S, Cai J: Update on biomarkers in development of anti-angiogenic drugs in gastric cancer. Anticancer Res, 2016; 36: 1111–18
- Brzozowa M, Michalski M, Harabin-Słowińska M, Wojnicz R: The role of tumour microenvironment in gastric cancer angiogenesis. Prz Gastroenterol, 2014; 9: 325–28
- Tian XL, Kadaba R, You SA et al: Identification of an angiogenic factor that when mutated causes susceptibility to Klippel-Trenaunay syndrome. Nature, 2004; 427: 640–45
- Major MB, Roberts BS, Berndt JD et al: New regulators of Wnt/beta-catenin signaling revealed by integrative molecular screening. Sci Signal, 2008; 1: ra12
- Xu Y, Zhou M, Wang J et al: Role of microRNA-27a in down-regulation of angiogenic factor AGGF1 under hypoxia associated with high-grade bladder urothelial carcinoma. Biochim Biophys Acta, 2014; 1842: 712–25
- Røe OD, Anderssen E, Sandeck H et al: Malignant pleural mesothelioma: Genome-wide expression patterns reflecting general resistance mechanisms and a proposal of novel targets. Lung Cancer, 2010; 67: 57–68
- 7. Wang W, Li GY, Zhu JY et al: Overexpression of AGGF1 is correlated with angiogenesis and poor prognosis of hepatocellular carcinoma. Med Oncol, 2015; 32: 131
- Chen D, Li L, Tu X et al: Functional characterization of Klippel-Trenaunay syndrome gene AGGF1 identifies a novel angiogenic signaling pathway for specification of vein differentiation and angiogenesis during embryogenesis. Hum Mol Genet, 2013; 22: 963–76
- 9. Lu Q, Yao Y, Yao Y et al: Angiogenic factor AGGF1 promotes therapeutic angiogenesis in a mouse limb ischemia model. PLoS One, 2012; 7: e46998

- Hu Y, Li L, Seidelmann SB et al: Identification of association of common AGGF1 variants with susceptibility for Klippel-Trenaunay syndrome using the structure association program. Ann Hum Genet, 2008; 72: 636–43
- 11. Kaya M, Wada T, Akatsuka T et al: Vascular endothelial growth factor expression in untreated osteosarcoma is predictive of pulmonary metastasis and poor prognosis. Clin Cancer Res, 2000; 6: 572–77
- Hsu JT, Chen TD, Chuang HC et al: Vascular endothelial growth factor expression is an independent poor prognostic factor for human epidermal growth factor receptor 2 positive gastric cancer. J Surg Res, 2017; 208: 40–50
- Chang Y, Niu W, Lian PL et al: Endocan-expressing microvessel density as a prognostic factor for survival in human gastric cancer. World J Gastroenterol, 2016; 22: 5422–29
- 14. Chen S, Zhang X, Peng J et al: VEGF promotes gastric cancer development by upregulating CRMP4. Oncotarget, 2016; 7: 17074–86
- 15. Zhao DQ, Chen J, Wu YF et al: Correlation between vascular endothelial growth factor and somatostatin receptor with progression and prognosis in gastric cancer. Hepatogastroenterology, 2014; 61: 1154–58