

ORIGINAL ARTICLE

Impact of *TP53* mutation status on systemic treatment outcome in *ALK*-rearranged non-small-cell lung cancer

A. Kron^{1,2,3†}, C. Alidousty^{1,3,4†}, M. Scheffler^{1,2,3}, S. Merkelbach-Bruse^{1,3,4}, D. Seidel^{3,5}, R. Riedel^{1,2,3}, M. A. Ihle^{1,3,4}, S. Michels^{1,2,3}, L. Nogova^{1,2,3}, J. Fassunke^{1,3,4}, C. Heydt^{1,3,4}, F. Kron^{1,3}, F. Ueckeroth^{1,3,4}, M. Serke^{1,6}, S. Krüger^{1,7}, C. Grohe^{1,8}, D. Koschel^{1,9}, J. Benedikter^{1,10}, B. Kaminsky^{1,11}, B. Schaaf^{1,12}, J. Braess^{1,13}, M. Sebastian^{1,14}, K.-O. Kambartel^{1,15}, R. Thomas^{1,16}, T. Zander^{1,3}, A. M. Schultheis^{1,3,4‡}, R. Büttner^{1,3,4‡} & J. Wolf^{1,2,3*‡}

¹Network Genomic Medicine, Cologne; ²Lung Cancer Group Cologne, Department I of Internal Medicine, University Hospital of Cologne, Cologne; ³Center for Integrated Oncology Köln Bonn, Cologne; ⁴Institute of Pathology, University Hospital of Cologne, Cologne; ⁵CECAD Cluster of Excellence, University of Cologne, Cologne; ⁶Department of Pneumology, Lungenklinik Hemer des Deutschen Gemeinschafts-Diakonieverbandes GmbH, Hemer, ⁷Department of Pneumology, Florence Nightingale Hospital, Düsseldorf; ⁸Department of Pneumology, Evangelische Lungenklinik Berlin (Paul Gerhardt Diakonie), Berlin; ⁹Department of Pneumology, Facktrankenhaus Coswig, Coswig; ¹⁰Department of Pneumology, Klinikum Bogenhausen, Munich; ¹¹Department of Pneumology, Krankenhaus Bethanien, Solingen; ¹²Lung Cancer Center, Klinikum Dortmund GmbH, Dortmund; ¹³Department of Oncology and Hematology, Krankenhaus Barmherzige Brueder, Regensburg; ¹⁴Department of Oncology and Hematology, University Hospital Frankfurt (Johannes-Wolfgang Goethe Institute), Frankfurt am Main; ¹⁵Department of Pneumology, Bethanien Hospital Moers-Lungenzentrum, Moers; ¹⁶Cologne Center for Genomics, University Hospital of Cologne, Cologne, Germany

*Correspondence to: Prof. Jürgen Wolf, Lung Cancer Group Cologne, Department I of Internal Medicine, Center for Integrated Oncology, University Hospital of Cologne, Kerpener Str. 62, D-50937 Cologne, Germany. Tel: +49-221-478-89050; Fax: +49-221-478-89051; E-mail: juergen.wolf@uk-koeln.de [†]Both authors contributed equally to this work.

[†]These authors share senior authorship.

Data presented in part at the European Society of Medical Oncology Annual Meeting 2017, Madrid, Spain (abstract #3757) and the German Society of Hematology/Oncology Annual Meeting 2017, Stuttgart, Germany (abstract A-908-0026-00596).

Background: We analyzed whether co-occurring mutations influence the outcome of systemic therapy in *ALK*-rearranged non-small-cell lung cancer (NSCLC).

Patients and methods: *ALK*-rearranged stage IIIB/IV NSCLC patients were analyzed with next-generation sequencing and fluorescence *in situ* hybridization analyses on a centralized diagnostic platform. Median progression-free survival (PFS) and overall survival (OS) were determined in the total cohort and in treatment-related sub-cohorts. Cox regression analyses were carried out to exclude confounders.

Results: Among 216 patients with *ALK*-rearranged NSCLC, the frequency of pathogenic *TP53* mutations was 23.8%, while other co-occurring mutations were rare events. In *ALK/TP53* co-mutated patients, median PFS and OS were significantly lower compared with *TP53* wildtype patients [PFS 3.9 months (95% CI: 2.4–5.6) versus 10.3 months (95% CI: 8.6–12.0), P < 0.001; OS 15.0 months (95% CI: 5.0–24.9) versus 50.0 months (95% CI: 2.2.9–77.1), P = 0.002]. This difference was confirmed in all treatment-related subgroups including chemotherapy only [PFS first-line chemotherapy 2.6 months (95% CI: 1.3–4.1) versus 6.2 months (95% CI: 1.8–10.5), P = 0.021; OS 2.0 months (95% CI: 2.9–7.2) versus 9.0 months (95% CI: 6.1–11.9), P = 0.035], crizotinib plus chemotherapy [PFS crizotinib 5.0 months (95% CI: 2.9–7.2) versus 14.0 months (95% CI: 8.0–20.1), P < 0.001; OS 17.0 months (95% CI: 6.7–27.3) versus not reached, P = 0.049] and crizotinib followed by next-generation ALK-inhibitor [PFS next-generation inhibitor 5.4 months (95% CI: 0.1–10.7) versus 9.9 months (95% CI: 6.4–13.5), P = 0.039; OS 7.0 months versus 50.0 months (95% CI: not reached), P = 0.001].

Conclusions: In *ALK*-rearranged NSCLC co-occurring *TP53* mutations predict an unfavorable outcome of systemic therapy. Our observations encourage future research to understand the underlying molecular mechanisms and to improve treatment outcome of the *ALK*/*TP53* co-mutated subgroup.

Key words: ALK-rearranged NSCLC, sequential ALK-inhibitor therapy, TP53 mutation status

© The Author(s) 2018. Published by Oxford University Press on behalf of the European Society for Medical Oncology.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

Introduction

ALK-positive non-small-cell lung cancer (NSCLC) is characterized by *ALK* gene rearrangements and an association with acinar histology, younger age and never-smoking status [1]. *ALK* rearrangements lead to constitutive activation of the encoded tyrosine kinase and downstream transforming signaling pathways [2]. Crizotinib, the first approved ALK-inhibitor, is superior to chemotherapy regarding overall response rate, progression-free survival (PFS), toxicity profile [3, 4] and overall survival (OS) [5– 7]. Next-generation inhibitors with activity against *ALK* resistance mutations are in clinical evaluation and partly already approved [8–11]. An impressive OS was reported for sequential ALK-inhibitor therapy ranging from 45 to 89.6 months [12–14].

There are considerable differences in the clinical course of *ALK*-positive NSCLC patients treated with chemotherapy or ALK inhibitors [3, 4, 15, 16]. Genetic heterogeneity of *ALK*-positive tumors could explain this observation. We have molecularly analyzed 216 *ALK*-positive patients with advanced disease and hypothesized that co-occurring mutations might underlie these differences.

Patients and methods

Patients and samples

The study was carried out within the Network Genomic Medicine [17], which offers centralized molecular diagnostics at the University Hospital of Cologne for patients with lung cancer from 300 participating partners. The study was conducted in concordance with local ethical guidelines. Patients were treated with crizotinib, ceritinib, alectinib or brigatinib according to national guidelines or within clinical trials [PROFILE1005 (NCT00932451); PROFILE1007 (NCT00932893); CLDK378X2101 (NCT-1283516); ASCEND-5 (NCT01828112); ALTA (AP26113) (NCT02094573); ACCALIA (NCT01801111)].

Fluorescence in situ hybridization

ALK, *RET* and *ROS1* rearrangements were diagnosed using break-apart fluorescence *in situ* hybridization (FISH) [17]. *MET* and *ERBB2* were tested for amplification as reported [18]. Details are described in supplementary Table S1, available at *Annals of Oncology* online.

Next-generation sequencing

Samples were analyzed with either a validated gene panel using AmpliSeq chemistry (Thermofisher, LUN3) comprising 102 amplicons of 14 different genes or a validated gene panel using GeneRead chemistry (Qiagen, LUN4), comprising 17 genes [19]. Details are described in supplementary Table S6, available at *Annals of Oncology* online. *ALK* variants were determined using the Archer[®] FusionPlex[®] Lung Kit and Archer Molecular Barcode (MBC) Adapters (both for Illumina) according to the manufacturer's instructions.

Programmed death-ligand 1 immunohistochemistry

Programmed death-ligand 1 (PD-L1) immunohistochemistry was carried out on the Leica Bond platform using primary antibody clone 28-8 (Abcam, Cambridge, UK). Interpretation was done according to the Dako PD-L1 22C3 pharmDx guidelines, results were reported based on an integrated proportion score [20, 21].

Data collection

The Network Genomic Medicine database covers molecular diagnostics and basic demographic and clinical data. For treatment outcome medical records were reviewed. PFS was determined based on RECIST v1.1. Time of death was determined either via medical records or requests to local registry offices. OS was defined as the time from first diagnosis of stage IIIB/IV until death. For subjects alive at completion of this analysis, time to death was censored at the time of last contact.

Statistical analyses

Statistical analyses were carried out using IBM SPSS software 24 (IBM, Armonk, NY). Chi-squared and two-sided Fischer's exact tests were used for analyzing qualitative variable characteristics in different groups. The Kaplan–Meier estimator was used to calculate OS and PFS. Two-sided log-rank tests were applied to compare differences between treatment groups. Cox proportional hazards model was used to adjust for potential confounders. *P* values <0.05 were considered statistically significant.

Results

Patient characteristics

Between January 2011 and December 2016, 423 *ALK*-positive patients were identified using FISH. From 289 patients with written informed consent, 53 had no stage IIIB/IV and 20 were lost to follow up. About 216 patients were eligible (Figure 1A). Median age, distribution of sex and histology are in line with earlier reports (Table 1) [3, 4]. Median follow-up was 34 months.

From 147 (68%) patients' tumors were analyzed by next-generation sequencing [LUN3 panel: 90 patients (61%); LUN4 panel: 57 patients (39%)]. Fifty patients (23%) were tested by additional single gene sequencing. Thirty-four (17%) of 197 patients were tested for PD-L1 expression, 135 (69%) received further FISH analyses. In 34 of 216 *ALK*-positive patients (16%) distribution of *ALK* variants was assessed by RNA sequencing (Figure 1A).

For 175 patients (81%) follow-up data for OS were available including 7 patients (3.2%) treated with best supportive care. Thus, 168 patients (77.8%) were subdivided (Figure 1B) into cohort A including 42 patients (19.4%) treated with chemotherapy only, cohort B including 71 patients (33%) with crizotinib and chemotherapy, cohort C including 18 patients (8.3%) with first-line crizotinib and cohort D including 37 patients (17.1%) with ceritinib after crizotinib with or without chemotherapy. Supplementary Figure S2, available at *Annals of Oncology* online shows treatment sequences in cohort D.

From 41 patients (19%, cohort Z) no complete therapy data until death or final follow-up were available including 5 patients treated with alectinib and 2 with brigatinib.

Co-occurring mutations, PD-L1 status and *ALK* variants

Mutations in *TP53* were the most frequent co-occurring mutations with 23.8% (34/143) of the tested patients. Among 36 *TP53* mutations 34 were classified as nonfunctional [22], 2 were of unknown functional significance (supplementary Table S5, available at *Annals of Oncology* online). All other co-alterations occurred rarely with frequencies between 0.6% for *BRAF* (1/171), 0.6% for *KRAS* (1/174) and 3.6% (4/112) for low-level *MET* amplification (Figure 2A and

Annals of Oncology

Original article



Figure 1. (A) Flowsheet of molecular diagnostics. NGS, next-generation sequencing; FISH, fluorescence *in situ* hybridization; IHC, immunohistochemistry. (B) Allocation of patients to cohorts for evaluation of treatment-related OS. BSC, best supportive care; PFS, progression-free survival; OS, overall survival.

supplementary Table S4, available at *Annals of Oncology* online). Four patients showed more than 1 co-occurring alteration (supplementary Figure S1, available at *Annals of Oncology* online).

PD-L1 expression of tumor cells was assessed in 34 patients (supplementary Table S4, available at *Annals of Oncology* online). In eight patients (23.5%) the PD-L1 score [21] was 5, i.e. more than 50% of the tumor cells expressed PD-L1. PD-L1 positivity was significantly correlated with *TP53* mutations [*TP53* wildtype (wt): 41% PD-L1 positive/*TP53* mutated: 90% PD-L1 positive; P = 0.009]. No significant difference was observed for high PD-L1 positivity (score 5) between wt and mutated *TP53* (Figure 2B).

In 18 of 34 patients (53%) *ALK* variant 1 was found, in 14 patients (41%) variant 3a/b and in 2 patients (6%) variant 2 (supplementary Table S8, available at *Annals of Oncology* online).

PFS dependent on therapy and TP53 mutations

PFS was assessed in 157 patients with first-line chemotherapy (140 patients cohorts A–D plus 17 patients cohort Z, see Figure 1B), thereof 149 patients with platinum-based chemotherapy (109 resp. 103 PD at data cutoff), for crizotinib after chemotherapy in 112 patients (cohorts B–D and partly Z; 73 PD at data cutoff), for

Annals of Oncology

	n	%
Sex	216	
Male	111	51.4
Female	105	48.6
Age at diagnosis (years)		
Mean	58.09	
Standard deviation	14.52	
Median	58 (19–89)	
Histology		
AD	210	97.2
Adenosquamous	3	1.4
w/o differentiation	3	1.4
Smoking history		
Never	86	46.3
Former	62	33.3
Current	38	20.4
n/a	30	
ECOG performance status		
0	63	43.4
1	63	43.4
2	16	11.1
3	3	2.1
n/a	71	
Tumor stage at diagnosis		
l	4	1.9
	9	4.2
IIIA	14	6.5
IIIB	23	10.6
IV	166	76.8

w/o, without differentiation; n/a, not available; AD, adenocarcinoma.

crizotinib first line in 24 patients (cohorts C and D and partly Z; 12 PD at data cutoff) and for ceritinib after crizotinib with or without chemotherapy in 43 patients (cohorts D and partly Z; 28 PD at data cutoff). PFS of next-generation ALK inhibitors was calculated in 50 patients combined for ceritinib, alectinib (5 patients, 4 PD at data cutoff) and brigatinib (2 patients, 1 PD at data cutoff) because of the small patient number. PFS for first-line chemotherapy [5.4 months (95% CI: 3.7–7.1)] and for the subgroup of first-line platinum-based chemotherapy [5.5 months (95% CI: 3.8–7.2)] was inferior to PFS for first-line crizotinib [12.3 months (95% CI: 0.0–34.9); P = 0.001] and for crizotinib after chemotherapy [9.4 months (95% CI: 6.4–12.4); P < 0.001]. PFS for next-generation ALK inhibitors as sequential therapy after crizotinib was 7.0 months [(95% CI: 5.4–8.6); P = 0.449].

TP53 mutations were a negative prognostic factor for PFS regardless of systemic therapy. Median PFS with first-line chemotherapy was 2.6 months (95% CI: 1.3–4.1) with mutated *TP53* (n=27) and 6.2 months (95% CI: 1.8–10.5) with *TP53* wt (n=75) (P=0.021). For crizotinib first-line median PFS was 5.5 months only (95% CI: 0.0–10.9) with mutated *TP53* (n=3) versus 29.9 months (95% CI: 0.0–63.9) with *TP53* wt (n=15) (P=0.007). Similarly, for crizotinib after chemotherapy median PFS was 5.0 months (95% CI: 2.3–7.8) with mutated *TP53*

Original article



*TP53 mutation status not available for 2 cases

Figure 2. (A) Frequencies of co-occurring genetic aberrations in *ALK*-positive NSCLC patients. Results of NGS, single gene sequencing and FISH analysis in 197 *ALK* FISH-positive patients. (B) Correlation between PD-L1 positivity (expression score) and *TP53* mutation status in 34 *ALK*-positive patients.

(n = 19) versus 14.0 months (95% CI: 9.5–18.6) with *TP53* wt (n = 56) (P = 0.004). Regardless of treatment line, *TP53* mutation status segregated the median PFS of crizotinib-treated patients in an unfavorable *TP53*-mutated group [n = 22; 5.0 months (95% CI: 2.9–7.2)] and a favorable *TP53* wt group (n = 71; 14.0 months (95% CI: 8.0–20.1); P < 0.001]. Also, median PFS with next-generation ALK inhibitors after crizotinib was worse in patients with mutated *TP53* [n = 11; 5.4 months (95% CI: 0.1–10.7)] compared with *TP53* wt [n = 22, 9.9 months (95% CI: 0.1–10.7)] compared with *TP53* wt [n = 22, 9.9 months (95% CI: 0.4–13.5); P = 0.039]. In total, PFS of *TP53* co-mutated patients was 3.9 months $[n = 60 \quad (95\% \text{ CI: } 2.4–5.6)]$ and 10.3 months in *TP53* wt patients $[n = 168 \quad (95\% \text{ CI: } 8.6–12.0)]$ regardless of treatment (P < 0.001) (Figure 3A and supplementary Table S2, available at *Annals of Oncology* online).

The *ALK* variant 3a/b subgroup (cohorts A–D, n = 20) showed a nonsignificant trend toward better PFS with 11.9 months (95% CI: 0.9–23.1) versus variant 1 (n = 31) with 7.9 months (95% CI: 1.6–14.4) (P = 0.285). *TP53* mutations were negative predictive in both variant subgroups (n = 30; P = 0.001) with a strong trend in variant 1 [2.6 month (95% CI: 0.0–10.9) versus 15.9 months (95% CI: 1.4–30.6); P = 0.068] and reaching statistical significance in variant 3a/b (P = 0.022). Cox regression suggested a negative impact of *TP53* mutations on PFS regardless of ALK

Original article



Figure 3. (A) PFS with different systemic treatments dependent on *TP53* mutation status. Kaplan–Meier blots for the total cohort (n = 228), for chemotherapy (n = 102), for crizotinib (n = 93) and for next-generation ALK inhibitors (n = 33). (B) OS in the treatment-related cohorts dependent on *TP53* mutation status. Kaplan–Meier blots for the total cohort (n = 143), for chemotherapy (n = 22), for crizotinib (n = 63) and for ceritinib (n = 24).

variants (n = 30; P = 0.002) (supplementary Tables S8 and S9, available at *Annals of Oncology* online).

OS dependent on therapy and TP53 mutations

OS was assessed for 168 patients in cohorts A–D (Figure 1B). Median OS with chemotherapy only (cohort A, n = 42, 31 events at data cutoff) was with 9.0 months (95% CI: 5.0–12.9) inferior to all other cohorts treated with ALK inhibitors: cohort B (n = 71, 32 events) 31.0 months (95% CI: 0.4–61.6); P < 0.001, cohort C (n = 18, 2 events) median not reached (P = 0.001), cohort D (n = 37, 20 events) 45.0 months (95% CI: 32.3–57.7); P < 0.001. OS of patients treated with crizotinib starting from the first dose (n = 89; cohorts B + C) was 17.0 months (95% CI: 10.6–23.9).

TP53 mutations were a strong negative predictor for median OS in all cohorts. Median OS of mutated *TP53* patients (n=34) was 15.0 months (95% CI: 5.0–24.9) compared with 50.0 months (95% CI: 22.9–77.1) for *TP53* wt patients (n=109) (P=0.002). With chemotherapy only (cohort A), the median OS in *TP53*-mutated patients (n=7) was 2.0 months (95% CI: 0.0–4.6) compared with 9.0 months (95% CI: 6.1–11.9) in *TP53* wt patients (n=15) (P=0.035). For crizotinib-treated patients (cohorts B + C), OS for *TP53*-mutated patients (n=13) was 17.0 months (95% CI: 6.7–27.3) compared with *TP53* wt patients (n=50) for whom the median OS was not reached (P=0.049). Also for patients treated with ceritinib after crizotinib (cohort D), a striking difference in OS was observed with 7.0 months only (95% CI: not reached) (P=0.001) for *TP53* wt patients (n=20).

Within the *TP53*-mutated patient cohort, median OS with chemotherapy only (n=7) was 2.0 months (95% CI: 0.0–4.6) and thus inferior to ALK-inhibitor treatment (n=15) with 17.0 months (95% CI: 4.6–29.4) (P=0.025). For *TP53* wt patients treated with chemotherapy only (n=15), the median OS was 9.0 months (95% CI: 6.1–11.9) compared with a median OS of 50.0 months (95% CI: 22.3–77.7) (P < 0.001) for *TP53* wt patients treated with ALK inhibitors with or without chemotherapy (n=54) (Figure 3B and supplementary Table S3, available at *Annals of Oncology* online).

In univariate analysis including age, sex, smoking history, current smoker status, Eastern Cooperative Oncology Group (ECOG) performance status, number of brain metastases, number of treatment lines before crizotinib or ceritinib and *TP53* mutation status only current smoker status and *TP53* mutations were significant negative prognostic factors for OS (P=0.016 and P=0.002, respectively). In multivariate Cox regression analysis only *TP53* mutation remained an independent negative prognostic factor (P=0.004) (supplementary Table S7, available at *Annals of Oncology* online).

Patients with *ALK* variant 3a/b (n = 14) had a nonsignificant better OS of 50 months (95% CI: 0.0–108.9) compared with variant 1 (n = 18) with 29.0 months (95% CI: 9.4–48.6) (P = 0.815). *TP53* mutations were prognostic negative in both variant subgroups reaching statistical significance in variant 1 (P = 0.032) (supplementary Table S8, available at *Annals of Oncology* online).

Discussion

We show that in *ALK*-positive NSCLC *TP53* mutations separate roughly one quarter of patients with a substantially worse

outcome. PFS and OS were inferior compared with *TP53* wt patients treated with chemotherapy and ALK inhibitors.

TP53 alterations may damage tumor suppressor functions as loss of function mutations or trigger inhibition of apoptosis and genomic instability as gain of function mutations [23]. Thus, a negative prognostic impact of *TP53* mutations in cancer has been postulated and preclinical observations support this hypothesis [24]. While in unselected NSCLC such a negative prognostic impact has not been proven unequivocally [25–29], it has been reported in numerous reports for *EGFR*-mutated NSCLC treated with *EGFR* inhibitors. These results, however, only partly reached statistical significance [30–34]. In *ALK*-positive lung cancer, *TP53* mutations so far have not been described as significant negative prognostic factors.

The outcome in our treatment-related subgroups independently of *TP53* status confirmed what has been described in clinical trials [3, 4, 16] and registry analyses [6, 7, 12–14, 35]: superiority of ALK inhibitors over chemotherapy in terms of PFS and superiority of sequential ALK-inhibitor therapy compared with crizotinib monotherapy in terms of OS. Our results additionally show that *TP53* mutations represent the by far most frequent cooccurring mutations in *ALK*-positive NSCLC. By comparison, we found other co-mutations with a frequency of below 4% only; among them rarely those with actionable mutations like *BRAFV600*, high-level *MET* amplification or activating *KRAS* mutation.

Most important, our results suggest that about one-fourth of *ALK*-positive patients do not substantially benefit from recent progress of targeted therapy. As a limitation, concerning the use of next-generation ALK inhibitors statistically valid OS data could be assessed only for ceritinib. Future studies will have to prove, whether our findings can be confirmed for other next-generation ALK inhibitors.

In many cancer types, *TP53* mutations were shown to be associated with higher genetic instability [24]. Accordingly, we could recently show that early *TP53* mutations can lead to chromosomal instability in *ALK*-positive NSCLC [36]. It is tempting to speculate that a higher mutational burden might lead to a better efficacy of immune checkpoint inhibitors. Of note, the proportion of patients with PD-L1 positive tumor cells is enriched in our *TP53*-mutated group, although the number of patients is rather small.

Recently, in post ALK-inhibitor treatment biopsies it was shown that the type of *ALK* variant influences the development of ALK-inhibitor resistance mutations. In particular, *EML4-ALK* variant 3 was correlated with the development of *ALK G1202R* resistance mutation and a better PFS under treatment with the third-generation ALK-inhibitor lorlatinib, but not with first- and second-generation ALK inhibitors [37]. Similarly, in our pretreatment biopsies we saw a nearly equal distribution between *ALK* variants 1 and 3a/b and no significant influence of first- and second-generation ALK inhibitors on PFS. *TP53* mutations were negative prognostic in terms of PFS and OS in both variant subgroups. Based on the low patient number, which limits our conclusions, significance was only partly reached. It remains to be elucidated whether *TP53* mutation status and *ALK* variant status are independent prognostic factors.

In summary, we here describe *TP53* mutations as the first pretreatment biomarker in *ALK*-positive NSCLC identifying

Original article

patients with a substantially worse outcome from therapy. In future clinical trials stratification of this patient subgroup should be considered and new treatment strategies investigated to improve the outcome of *ALK/TP53* co-mutated patients.

Funding

None declared.

Disclosure

AK: Advisory Role for BMS, AbbVie, Novartis; CA: Research Funding from Roche; MS: Honoraria, Advisory Role and Travel Expenses from Novartis, BMS, Takeda, Boehringer Ingelheim; SM-B: Honoraria and Advisory Role from AstraZeneca, BMS, Novartis; DS: BB Biotech AG; RR: Advisory Role and Travel Expenses from Boehringer Ingelheim, Lilly; SM: Honoraria, Advisory Role and Research Funding from Novartis, Pfizer, Roche, Boehringer Ingelheim; LN: Honoraria, Advisory Role, Research Funding and Travel Expenses from Pfizer, Celgene, Novartis, Roche, BMS, Boehringer Ingelheim, MSD; JF: Honoraria from BMS and AstraZeneca; FK: Honoraria and Advisory Role from Takeda, AbbVie, BMS, MSD, Roche, Novartis, Celgene; MS: Honoraria, Advisory Role and Travel Expenses from Pfizer, BMS, Roche, MSD, Lilly, Celgene, AstraZeneca; SK: Honoraria, Advisory Role and Travel Expenses from Roche, Novartis, Chugai, AstraZeneca, Boehringer Ingelheim; DK: Advisory Role and Travel Expenses from Roche, Boehringer Ingelheim, Novartis, Grifols; JB: Honoraria, Advisory Role and Travel Expenses from Boehringer Ingelheim, BMS, AstraZeneca, Novartis, MSD, Pfizer, Roche, Chugai; BS: Honoraria from Boehringer Ingelheim, BMS, Roche; JB: Honoraria from Amgen, Janssen; MS: Honoraria and Advisory Role from Pfizer, Roche, Novartis, Takeda; K-OK: Advisory Role and Research Funding from MSD, BMS, Roche, Pfizer, Boehringer Ingelheim; RT: Advisory Role and Research Funding from Roche, J&J, Novartis, AstraZeneca, Bayer, New Oncology AG, Clovis, Boehringer Ingelheim, Merck, MSD, Lilly, Sanofi-Aventis, Daiichi-Sankyo, Puma; TZ: Honoraria and Advisory Role from Roche, Novartis, Lilly, Pfizer, Merck; AMS: Honoraria, Advisory Role, Research Funding and Travel Expenses from BMS, Roche; RB: Honoraria from Pfizer, Novartis; JW: Advisory Role, Research Funding and Travel Expenses from AbbVie, AstraZeneca, BMS, Boehringer Ingelheim, Chugai, Ignyta, Lilly, MSD, Novartis, Pfizer, Roche. All remaining authors have declared no conflicts of interest.

References

- 1. Inamura K, Takeuchi K, Togashi Y et al. EML4-ALK lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. Mod Pathol 2009; 22(4): 508-515.
- 2. Wellstein A, Toretsky JA. Hunting ALK to feed targeted cancer therapy. Nat Med 2011; 17(3): 290–291.
- Shaw AT, Kim D-W, Nakagawa K et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. N Engl J Med 2013; 368(25): 2385–2394.

- Solomon BJ, Mok T, Kim D-W et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. N Engl J Med 2014; 371(23): 2167–2177.
- Clinical Lung Cancer Genome Project (CLCGP); Network Genomic Medicine (NGM). A genomics-based classification of human lung tumors. Sci Transl Med 2013; 5(209): 209ra153.
- 6. Barlesi F, Mazieres J, Merlio JP et al. Routine molecular profiling of patients with advanced non-small-cell lung cancer: results of a 1-year nationwide programme of the French Cooperative Thoracic Intergroup (IFCT). Lancet 2016; 387(10026): 1415–1426.
- Kris MG, Johnson BE, Berry LD et al. Using multiplexed assays of oncogenic drivers in lung cancers to select targeted drugs. JAMA 2014; 311(19): 1998–2006.
- 8. Gadgeel SM. Sequencing of ALK inhibitors in ALK+ non-small cell lung cancer. Curr Treat Options Oncol 2017; 18(6): 36.
- 9. Lin JJ, Riely GJ, Shaw AT. Targeting ALK: precision medicine takes on drug resistance. Cancer Discov 2017; 7(2): 137–155.
- 10. Markham A. Brigatinib: first global approval. Drugs 2017; 77(10): 1131–1135.
- Mok TSK, Crino L, Felip E et al. The accelerated path of ceritinib: translating pre-clinical development into clinical efficacy. Cancer Treat Rev 2017; 55: 181–189.
- Duruisseaux M, Besse B, Cadranel J et al. Overall survival with crizotinib and next-generation ALK inhibitors in ALK-positive non-small-cell lung cancer (IFCT-1302 CLINALK): a French nationwide cohort retrospective study. Oncotarget 2017; 8(13): 21903–21917.
- 13. Gainor JF, Tan DS, De Pas T et al. Progression-free and overall survival in ALK-positive NSCLC patients treated with sequential crizotinib and ceritinib. Clin Cancer Res 2015; 21(12): 2745–2752.
- 14. Ito K, Hataji O, Fujimoto H et al. Sequential use of ALK inhibitors: an optional approach. J Thorac Oncol 2016; 11(12): e153–e154.
- Peters S, Camidge DR, Shaw AT et al. Alectinib versus crizotinib in untreated ALK-positive non-small-cell lung cancer. N Engl J Med 2017; 377(9): 829–838.
- Soria JC, Tan DSW, Chiari R et al. First-line ceritinib versus platinumbased chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. Lancet 2017; 389(10072): 917–929.
- Heydt C, Kostenko A, Merkelbach-Bruse S et al. ALK evaluation in the world of multiplex testing: Network Genomic Medicine (NGM): the Cologne model for implementing personalised oncology. Ann Oncol 2016; 27(suppl_3): iii25–iii34.
- Schildhaus HU, Heukamp LC, Merkelbach-Bruse S et al. Definition of a fluorescence in-situ hybridization score identifies high- and low-level FGFR1 amplification types in squamous cell lung cancer. Mod Pathol 2012; 25(11): 1473–1480.
- 19. Koitzsch U, Heydt C, Attig H et al. Use of the GeneReader NGS System in a clinical pathology laboratory: a comparative study. J Clin Pathol 2017; 70(8): 725–728.
- Scheel AH, Baenfer G, Baretton G et al. Interlaboratory concordance of PD-L1 immunohistochemistry for non-small-cell lung cancer. Histopathology 2018; 72(3): 449–459.
- Scheel AH, Dietel M, Heukamp LC et al. Harmonized PD-L1 immunohistochemistry for pulmonary squamous-cell and adenocarcinomas. Mod Pathol 2016; 29(10): 1165–1172.
- 22. Kato S, Han SY, Liu W et al. Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. Proc Natl Acad Sci USA 2003; 100(14): 8424–8429.
- 23. Brosh R, Rotter V. When mutants gain new powers: news from the mutant p53 field. Nat Rev Cancer 2009; 9(10): 701–713.
- 24. Hainaut P, Pfeifer GP. Somatic TP53 mutations in the era of genome sequencing. Cold Spring Harb Perspect Med 2016; 6.
- Ahrendt SA, Hu Y, Buta M et al. p53 mutations and survival in stage I non-small-cell lung cancer: results of a prospective study. J Natl Cancer Inst 2003; 95(13): 961–970.
- 26. Govindan R, Weber J. TP53 mutations and lung cancer: not all mutations are created equal. Clin Cancer Res 2014; 20(17): 4419–4421.

Annals of Oncology

- 27. Kosaka T, Yatabe Y, Onozato R et al. Prognostic implication of EGFR, KRAS, and TP53 gene mutations in a large cohort of Japanese patients with surgically treated lung adenocarcinoma. J Thorac Oncol 2009; 4(1): 22–29.
- Rosell R, Monzo M, Pifarre A et al. Molecular staging of non-small cell lung cancer according to K-ras genotypes. Clin Cancer Res 1996; 2(6): 1083–1086.
- 29. Tsao MS, Aviel-Ronen S, Ding K et al. Prognostic and predictive importance of p53 and RAS for adjuvant chemotherapy in non small-cell lung cancer. J Clin Oncol 2007; 25(33): 5240–5247.
- 30. Canale M, Petracci E, Delmonte A et al. Impact of TP53 mutations on outcome in EGFR-mutated patients treated with first-line tyrosine kinase inhibitors. Clin Cancer Res 2017; 23(9): 2195–2202.
- Labbe C, Cabanero M, Korpanty GJ et al. Prognostic and predictive effects of TP53 co-mutation in patients with EGFR-mutated non-small cell lung cancer (NSCLC). Lung Cancer 2017; 111: 23–29.
- Yu HA, Jordan E, Ni A et al. Concurrent genetic alterations identified by next-generation sequencing in pre-treatment, metastatic EGFR-mutant lung cancers. J Clin Oncol 2016; 34: 9053–9053.

- 33. Aisner DL, Sholl LM, Berry LD et al. The impact of smoking and TP53 mutations in lung adenocarcinoma patients with targetable mutations— The Lung Cancer Mutation Consortium (LCMC2). Clin Cancer Res 2018; 24(5): 1038–1047.
- 34. VanderLaan PA, Rangachari D, Mockus SM et al. Mutations in TP53, PIK3CA, PTEN and other genes in EGFR mutated lung cancers: correlation with clinical outcomes. Lung Cancer 2017; 106: 17–21.
- 35. Kostenko A, Michels SYF, Fassunke J et al. Survival following implementation of next-generation sequencing in routine diagnostics of advanced lung cancer: results of the German Network Genomic Medicine. J Clin Oncol 2016; 34: 9085–9085.
- Alidousty C, Baar T, Martelotto LG et al. Genetic instability and recurrent MYC amplification in ALK-translocated NSCLC: a central role of TP53 mutations. J Pathol 2018; 246: 67–76.
- Lin JJ, Zhu VW, Yoda S et al. Impact of EML4-ALK variant on resistance mechanisms and clinical outcomes in ALK-positive lung cancer. J Clin Oncol 2018; 36: 1199–1206.