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Sub-acute toxicity evaluation of aqueous leaf extract from *Passiflora edulis Sims f. edulis* (Gulupa) in Wistar rats

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ABSTRACT

The genus Passiflora (Passifloraceae) comprises about 500 species. The Passiflora edulis stands out because of its economic and medicinal importance. It is widely planted in tropical and subtropical regions worldwide, especially in South America, the Caribbean, South Africa, and Asia. The aqueous extract of Passiflora edulis Sims f. edulis (Gulupa) leaves is used in traditional medicine for its soothing and tranquilizing effects on the central nervous system. Therefore, evaluating its safety for human use is a fundamental requirement to continue the development of new therapies within the framework of regulatory, preclinical, and clinical guidelines. Here, the sub-acute toxicity study was conducted following the Organization for Economic Cooperation and Development (OECD) guideline 407 for 28 days in Wistar albino rats. The study showed that 1000 mg/kg/day of the aqueous extract in 10 adult Wistar rats (five males and five females) was well tolerated. The hematological results are at normal levels. However, monocytopenia and eosinopenia were observed with a significant difference (P < 0.05) for both male and female rats treated with the aqueous extract of Passiflora edulis. The results show that liver and kidney function profiles were conserved. However, an increase in ALT is observed with significant differences between male and female rats treated with the extract compared to the controls. Study findings were limited to non-adverse histopathological results of a slightly increased incidence of focal periportal lymphocytic infiltrate in the liver and focal corticomedullary nephrocalcinosis in the kidney compared to control. Therefore, the aqueous extract of Passiflora edulis has a good safety profile in oral administration, was well tolerated, and did not cause any lethality or adverse effects in the sub-acute toxicity study in male and female rats. The NOAEL (no observed adverse effect level) for the 28-day subacute toxicity study was considered to be 1000 mg/kg.

1. Introduction

Traditional and complementary medicine (TCM) is used by at least 80% of the Member States across all WHO (World Health Organization) regions, particularly Eastern Mediterranean, Southeast Asia, and Western Pacific regions reported using TCM [1–3]. Likewise, the growth in the number of Member States developing regulations and national policies for TCM has been reported [1]. It is also informed that this treatment has been gradually integrated into treating chronic diseases such as diabetes [4] and complex diseases like cancer [5,6]. The increased use of plants in phytotherapy has meant that new and diverse therapeutic resources have become available, along with conserving the traditional

knowledge of the countries where they are used [7].

In general, the population perceives treatments with medicinal plants as safe due to their origin [2,8]; however, the use of these treatments can affect different organs [9] as well as cause interactions with conventional drugs are possible, leading to changes in the efficacy of treatments or toxic manifestations [10]. In this regard, around 80,000 exposures to plants, and homeopathic supplements were reported for 2020 in the United States [11,12]. Most of these exposures are of minimal toxicity largely because they involve pediatric ingestions of low quantity [12].

Passiflora edulis Sims f. edulis, known as gulupa in Colombia or purple passion fruit worldwide, is a plant native to South America, specifically

* Correspondence to: Carrera 7 No. 43–82, Edificio Jesús Emilio Ramírez, Lab 304A, Bogotá C.P.110211, Colombia. *E-mail address: albarra@javeriana.edu.co* (S.L. Albarracín).

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Received 23 August 2023; Received in revised form 17 October 2023; Accepted 23 October 2023 Available online 24 October 2023 2214-7500/© 2023 Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/). Brazil. However, since the 19th century, it has been widely distributed in Asia, the Caribbean, Africa, India, and Australia [13]. Among the different varieties of *Passiflora edulis*, the yellow passion fruit, *Passiflora edulis*, are the main varieties, and the purple passion fruit, *Passiflora edulis*, are the main varieties, and their secondary metabolites are of great interest in the pharmaceutical and food industry [14,15].

The genus *Passiflora spp.* has been associated with anti-inflammatory and antioxidant activity due to the presence of flavonoids [16–18]. *Passiflora edulis* has been used as a sedative, diuretic, anthelmintic, antidiarrheal, stimulant, and in the treatment of hypertension, menopausal symptoms, infant colic, and for the treatment of symptoms of alcoholism, anxiety, migraine, nervousness, and insomnia [19,20].

Previous reports on evaluating the oral toxicity of Passiflora edulis extracts did not show toxicity signs in hematological and biochemical parameters [21]. For instance, a sub-acute toxicity study of Passiflora nepalensis (Passifloraceae) in male and female rats at the doses of 40, 80, 160, and 320 mg/kg for fourteen consecutive days did not induce any short-term toxicity [22]. Another sub-acute study in Wistar albino rats was conducted by oral administration of 100, 200, 300, and 400 mg/kg of aqueous leaf extract Passiflora edulis. The subacute study indicated that the extract was safe for bone marrow function and was neither hepatotoxic nor nephrotoxic [23]. In an acute toxicity study of Passiflora alata in mice, deaths were not observed for a single dose up to 4800 mg/kg. However, mice acutely treated with extract 150, 300, and 600 mg/kg presented DNA damage determined by comet assay in peripheral blood cells 3 h after treatment. In another sub-acute study, rats treated with aqueous extract from Passiflora alata at 300 mg/kg dose for 14 days did not present biochemical, hematological, or histopathological significant alterations compared to the control group. However, these rats showed irritability and did not show weight gain [24]. An acute oral toxicity study was performed in mice to determine the safety profile following preclinical requirements to demonstrate the absence of effects or alterations induced by the aqueous extract obtained from Passiflora edulis f. edulis at 2000 mg/kg dose [25]. The results showed no abnormal change in mice behavioral patterns and hematologic parameters, including RBC, Hb, WBC, MCV, MCH, platelets, neutrophils, and lymphocytes. In these studies, regardless of the dosing frequency, it is necessary to determine the safety profile following preclinical requirements [26] to develop new therapeutic alternatives such as polymolecular drugs or isolated molecules.

2. Materials and methods

2.1. Animals and ethical approval

Wistar albino rats of 9 weeks of age were maintained in accordance with recommendations regarding housing, temperature, and humidity in the Guide for the Care and Use of Laboratory Animals at the Unit of Comparative Biology accredited facilities. All experimental procedures were approved by the Pontificia Universidad Javeriana Animal Care and Use Committee. The protocols carried out were endorsed by the Institutional Committee for the Care and Use of Laboratory Animals of the Pontificia Universidad Javeriana (CICUA), FUA No. 057–18. All rats were housed with free access to a standard pellet diet and water *ad libitum*. They were kept at a constant temperature of 22 ± 1 °C, the relative humidity of 55 ± 5 %, and 12/12 h light/dark cycle.

2.2. Preparation of aqueous extract

Leaves of *Passiflora edulis* Sims f *edulis* were collected from the departments of Nariño and Boyacá, Colombia. The taxonomic determination of the species was carried out by the biologist Nestor García in the herbarium of the Pontificia Universidad Javeriana. The leaves were dried in a circulating air oven at a controlled temperature of 35°C and crushed with a blade mill. The dried and ground material was subjected to an extraction procedure by infusion with boiling water for 10 min, in

a ratio (1:10 P/V) of solvent plant material, emulating the traditional use. Subsequently, this extract was subjected to lyophilization. Preparing the aqueous extract of *Passiflora edulis* under these conditions corresponds to what is done in traditional medicine as an infusion of the leaves. Ultra Performance Liquid Chromatography coupled with Photodiode Array Detection (UPLC-PDA) was in performed Chromatographic profiling. Three batches of the *Passiflora edulis* extract were prepared. The results of the UPLC-DAD analysis show the chromatographic profile's maintenance between the different batches of the extract. The above allowed us to infer that the chromatographic fingerprints are maintained regardless of the standardized extraction process in each batch. The above allows us to assume that the compounds present in the three batches are conserved in structure and retention times.

2.3. Toxicity assessment

The study was conducted following the recommendations of the Organization for Economic Cooperation and Development (OECD).

2.4. Sub-acute oral toxicity study

The sub-acute toxicity study was conducted following the Organization for Economic Cooperation and Development (OECD) guideline 407 for 28 days in Wistar albino rats [27]. Twenty Wistar rats of nine weeks old were divided into two groups of ten animals each (five males and five females). One group was treated with a dose of 1000 mg/kg/day with the aqueous extract for 28 days and the other corresponded to the control (vehicle group) received 0.5 ml of physiological saline. As per the OECD guideline, lower dose groups were not included as no effects were expected at 1000 mg/kg/day. The extract or vehicle was administered by oral gavage. The animals were observed daily at least twice a day for 28 days, and weight was recorded twice weekly. Mortality, motor activity, righting reflex, corneal reflex, pineal reflex, the tendency to escape, pallor, piloerection, salivation, and diarrhea, among others, were recorded.

All animals were sacrificed at the end of the study, for which they were anesthetized with isoflurane on day 29 and after fasting overnight. Blood was drawn by intracardiac puncture, and the samples were collected in two tubes (one with EDTA) for the biochemical tests: Urea (BUN), alanine aminotransferase (ALT), gamma-glutamyl transferase (GGT), alkaline phosphatase, creatinine, total protein, and bilirubin. Another sample was collected with heparin for hematological analysis (erythrocyte count, total and differential leukocyte count, platelet count, hemoglobin, hematocrit). On the other hand, a complete necropsy was conducted on all animals by a veterinary pathologist assisted by a team of trained individuals. The brain, liver, kidneys, heart, adrenals, testes, uterus, ovaries, and spleen were trimmed and weighed immediately. For histopathology, liver and kidney samples were taken and preserved in 10 % buffered formaldehyde. Hematoxylin-eosin (H&E) staining was used for tissue processing.

2.5. Statistical analysis

Data were analyzed using GraphPad Prism v 8.0 (GraphPad Software, La Jolla, California, United States, www.graphpad.com). Data are expressed as mean \pm standard deviation of the mean (SD) unless otherwise specified. Blood parameters were processed by factorial analysis of variance (ANOVA). The differences were considered statistically significant when p < 0.05.

3. Results

3.1. Chemical profile of the Passiflora edulis extract

The chromatographic profiles by UPLC-PDA show between minutes

4 and 10 (wavelength 254 nm) peaks corresponding to flavonoids with the typical UV absorption spectra of these molecules (Fig. 1). Among those, peaks 3, 5, 6, 7, and 9 show UV absorption of luteolin derivatives (λ_{max} = 260,270, 350 nm), while peaks 4 and 8 presented UV absorption related to apigenin derivatives (λ_{max} = 270, 340 nm). Some of those flavonoids have already been reported for the genus *Passiflora*, such as vitexin 2-O-rhamnoside, luteolin 7-O-glucoside, vicenin-2, 6,8-di-Cglucosyl chrysin and spinosyn [20,24]. The chromatographic profiles of the three batches (L1, L2, and L3) manufactured for the aqueous extract of *Passiflora edulis* are shown in Fig. 2, where batch-to-batch consistency is observed at 254 and 350 nm, which ensures that the results obtained can be extrapolated to subsequent batches for further preclinical and clinical development.

Using the analysis by Time-of-Flight Mass Spectrometry (UHPLC-MS-QTOF), the retention times (RT), molecular ion, and some fragmentation patterns that are presented in Table 1 were obtained. According to the molecular mass of the compounds, the results show the presence of molecules between 300 and 600 g/mol, according to the literature, could be associated with the presence of flavonoid compounds, and characteristically those glycosylated as vitexin, isovitexin, vicenin, chrysin [28–33].

4. Sub-acute study toxicity for 28 days

4.1. Survival and clinical observations

Here, the administration of 1000 mg/Kg/day of the aqueous extract did not generate deaths during the experimental period. There were no clinical findings or observations related to the alteration of the animals' well-being related to the consumption of the extract. As expected, no weight loss was evident in the animals during the 29 days of the study (Fig. 3). However, differences in weight gain were observed between males and females, which can be explained by gender characteristics. There were no treatment-related differences in body weight or body weight gain in male or female rats (Table 2). However, male rats had higher body weights from the start of the study and greater weight gains compared to females. Male rats receiving the extract had lower body weights at the beginning of day 1 of the test and greater body weight gains at the end of the study (32.22 %). However, these differences in body weight compared to control did not reach statistical significance when analyzed using a one-way ANOVA. These differences were interpreted as normal biological variability since these values correspond to the increase in body weight, similar to control males. Furthermore, the weight and body weight gain of female rats receiving the extract were like those of the control group. There were no treatment-related effects on terminal body weights or any organ weights in female and male rats receiving extract compared to control. There were no statistically identified differences in terminal body weights or organ weights in either rat males or females receiving extract compared to controls (Table 3). There were no treatment-related macroscopic pathological or histopathological observations. All pathological observations were interpreted as spontaneous alterations not associated with exposure to the extract.

4.2. Hematological parameters

Table 4 shows the hematological parameters of the animals in the study and shows the behavior of the variables analyzed by gender and by treatment. The results do not show important hematological alterations since it is evident that most of these parameters are within the normal ranges reported for Wistar and Sprague Dawley [34,35]. rats. However, a small monocytopenia is observed with a significant difference (P < 0, 05) in monocytes for both male and female rats treated with the aqueous extract of *Passiflora edulis*. Additionally, eosinopenia was observed with differences in eosinophil counts and percentages between the control and experimental groups for both sexes. No significant changes in total neutrophils were observed. However, no changes associated with loss of animal well-being or weight loss were observed during the study. These results suggest that the aqueous extract of *Passiflora edulis* is not toxic but induces a slight leukocytopenia with a decrease in monocyte and eosinophil counts.

4.3. Biochemical parameters

Regarding the blood biochemistry analysis, Table 5 shows the summary of the liver function profile represented by ALT, GGT, FA, conjugated and unconjugated bilirubin. The data do not show alterations and these parameters are within the normal ranges reported for Wistar and Sprague Dawley rats [34,35]. However, an increase in ALT is observed with significant differences between male and female rats treated with the extract compared to the control. These data must be correlated with the histological findings to establish whether the extract generates a hepatotoxic effect. Table 6 shows the summary of the renal function profile represented by urea, creatinine, BUN, and plasma proteins. These parameters did not show changes or significant differences between the groups.

4.4. Histopathological examination

Histopathological examinations of liver sections in rats treated with *Passiflora edulis* extract are shown in Fig. 4. Microscopic observation of liver sections from the controls and the animals treated with the extract

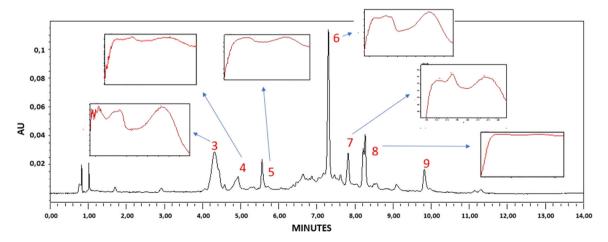


Fig. 1. UPLC chromatographic profile and DAD spectra of the aqueous *Passiflora edulis* extract at 254 nm FE: Phenomenex Kinetex® C18 column ($100 \times 2.1 \text{ mm}$; 2.6 µm) at temperature: 26 °C. FM: Gradient of acetonitrile and formic acid 0.1%.

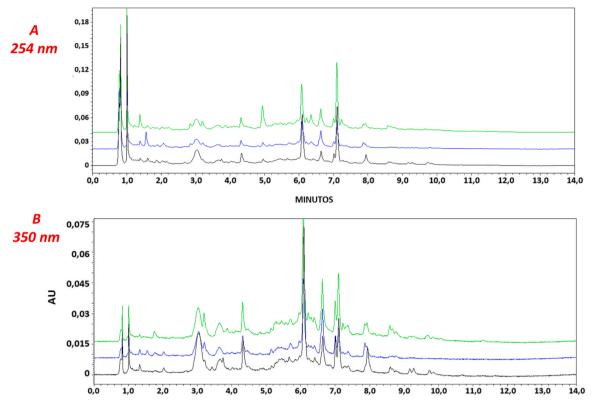


Fig. 2. UPLC chromatographic profile and DAD spectra of the different batches from *Passiflora edulis* extract: A) 254 nm and B) 350 nm. Black: L1; Blue: L2 and Green: L3. FE: Phenomenex Kinetex® C18 column (100 × 2.1 mm; 2.6 µm) at temperature: 26 °C. FM: Gradient of acetonitrile and formic acid at 0.1%.

Table 1

Analysis by Time-of-Flight Mass Spectrometry (UHPLC-MS-QTOF). The table shows the retention times (RT), molecular ion and some fragmentation patterns can be correlated with could be associated with the presence of flavonoid compounds, and characteristically those glycosylated as vitexin, isovitexin, vicenin, chrysin.

Retention time	[M-H]	Fragments	Identification	UV max abs. nm
2,45	431,19126	-	Vitexin / isovitexin - Glucoside.	266,346
3,13	593.15009	433.20749 340.10273 298.80642	Vicenin (apigenin glucoside)	265, 346
5,86	723.50054	713.47185 575.13924 497.33320 298.80640	Chrysin - glucoside	258, 342
6,86	607.16544	561.15963 395.07593	Spinosin	266, 346
6,97	697.41488	445,25465	Cyclopassifloside	
9,00	697.41501	681.41279 680.40880 679.40.366 639.37441 395.07603 327.21722	glycoside	

showed healthy liver cells without fatty accumulation, preserved cytoplasm, prominent nucleus, and intact central vein (Fig. 4A-4D). However, the histopathological study revealed the presence of a slight periportal mono-morphonuclear infiltrate (lymphocyte-macrophage) in the liver (Figs. 4E and 4F), with a slight increase in hepatocyte binucleation. The above results suggest that male and female rats administered the extract at 1000 mg/kg dose showed typical liver architecture without necrosis.

The results of histopathological examination of kidney sections from

control animals and those treated with Passiflora edulis extract are shown in Fig. 5. Kidneys from vehicle-administered rats demonstrated intact glomeruli and tubules (Fig. 5A-5D). Rats administered with 1000 mg/kg extract showed normal architectural structures similar to the control group. However, the histological study showed a slight generalized increase of intraglomerular mesangium (Fig. 5E and F) compared with the control, and a slight focal increase of cortico-medullary tubular nephron-calcifications was observed in both the control and the experimental groups (Fig. 6A and B).

5. Discussion

The therapeutic properties of Passiflora's infusion (aqueous extract) include traditional medicine such as anxiolytic, sedative, antispasmodic, neuroprotective, anti-inflammatory, and antioxidant [19,20,25,26]. The range of daily dosage varies from 30 mg to 120 mg of total flavonoids expressed as vitexin in an herbal medicine made from Passiflora [36]. In clinical studies that involved the use of Passiflora to treat anxiety 10-100 mg, only one of the studies was conducted with monopreparations of Passiflora. This difficult report doses to use in humans [37]. These preparations can be used acutely or chronically, depending on the pharmacological intention or traditional knowledge. For instance, the therapeutic activity for the central nervous system could involve a more prolonged treatment than its use as an anti-inflammatory. Regardless of the dosing frequency, it is necessary to determine the safety profile following nonclinical requirements [27] to advance in developing new therapeutic alternatives either as polymolecular drugs or as isolated molecules. The oral administration of the aqueous of Passiflora edulis Sims f edulis (300 mg/kg) showed an antidepressant activity [38]. On the other hand, the aqueous extract of Passiflora edulis at 50, 100, and 150 mg/kg showed anxiolytic-like effects in rats' elevated plus-maze and inhibitory avoidance tests. In addition, the extract did not disrupt the rat memory process in habituation to an open-field test [39].

Regarding the chemical profile of the Passiflora edulis extract, the

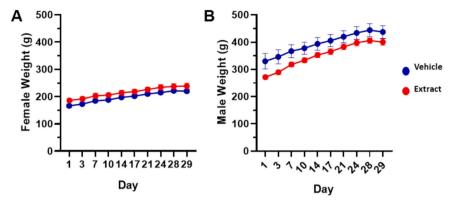


Fig. 3. Body weight of male and female rats. Values represent the mean \pm SD (n = 10). No significant differences were observed.

Table 2

Body weight gains summary (g) and weight gains (g) for male and female rats during the study. The table shows weight monitoring on different days and long-term weight gain at 7, 14, 21, and 29 of the subacute study.

Group		Days on Test								
		1	7	Gain	14	Gain	21	Gain	29	Gain
Male	Vehicle	329.8	366.64	36.84	393.38	63.58 ± 19	419.74	89.94	437.30	107.50
	n = 5	\pm 63.55	\pm 53.22	\pm 13.46	\pm 51.91		\pm 49.56	\pm 26.03	\pm 52.06	\pm 26.84
	Extract	271.4	318.1	$\textbf{46.70} \pm \textbf{4.47}$	352.94	81.54	383.10	111.70	400.42	129.02
	n = 5	\pm 15.95	\pm 18.81		\pm 20.77	\pm 7.27	\pm 23.52	± 11.45	\pm 26.39	\pm 15.60
Female	Vehicle	166.4	166.4	18.18 ± 5.05	197.42	31.02	210.22	43.82 ± 7.10	220.14	53.74 ± 8.39
	n = 5	\pm 13.90	\pm 13.90		± 11.12	\pm 8.26	\pm 14.39		\pm 15.39	
	Extract	185.6	185.6	16.90	213.84	28.24	225.86	40.26 ± 7.83	238.54	52.94
	n = 5	± 12.66	\pm 12.66	\pm 11.11	\pm 19.20	\pm 8.93	\pm 19.43		\pm 23.86	± 11.75

 Table 3

 Organ weight summary (g) for male and female rats on the final day of the study.

 The table shows the weight of the main organs obtained on necropsy.

Organ	Male		Female		
	Vehicle	Extract	Vehicle	Extract	
Liver	20.05 ± 2.3	19.41 ± 2.56	10.77 ± 1.14	11.34 ± 1.80	
Kidney	3.35 ± 0.47	3.01 ± 0.28	1.73 ± 0.12	1.75 ± 0.21	
Lung	2.46 ± 0.23	1.77 ± 0.12	1.44 ± 0.12	1.24 ± 0.09	
Heart	1.62 ± 0.24	1.55 ± 0.14	0.88 ± 0.16	1.11 ± 0.20	
Brain	1.97 ± 0.11	1.86 ± 0.05	1.69 ± 0.15	1.61 ± 0.04	
Testis	6.40 ± 1.39	5.14 ± 0.55			
Uterus			1.13 ± 0.24	0.65 ± 0.12	
Pancreas	1.62 ± 0.24	1.55 ± 0.14	0.38 ± 0.08	0.44 ± 0.14	
Spleen	1.00 ± 0.14	0.90 ± 0.13	0.57 ± 0.12	0.71 ± 0.11	
Stomach	1.85 ± 0.27	1.88 ± 0.07	1.55 ± 0.20	1.35 ± 0.23	
Adrenal	$\textbf{0.08} \pm \textbf{0.02}$	0.06 ± 0.01	0.14 ± 0.01	$\textbf{0.08} \pm \textbf{0.006}$	
Bladder	$\textbf{0.13} \pm \textbf{0.01}$	$\textbf{0.12} \pm \textbf{0.03}$	$\textbf{0.06} \pm \textbf{0.01}$	$\textbf{0.34} \pm \textbf{0.39}$	

mass spectrometry data (Table 1) suggest the presence of a group of compounds of the glycosylated flavonoid type with masses between 400 and 600 g/mol. According to the literature, they may correspond to 4 flavonoids: vitexin/iso-vitexin ([M-H]-: 431.19129), vicenin ([M-H]-: 593.15009), chrysin ([M-H]-: 723.50054) and spinosyn ([M-H]-: 607.16544). These molecules of flavonoids can present cores of apigenin, chrysin, and swertisin, respectively. Results previously reported by Farag et al. [32] identified flavone glycoside conjugates as the major class of compounds in Passiflora spp., using mass spectrophotometry and liquid chromatography with diode array detection with UV maxima 270 and 325-350 nm. Additionally, as suggested by Urrego et al. [33] we also propose the presence of the cyclopassifloside compound ([M-H]-: 697.41501) in Passiflora edulis f edulis sims. Other authors have reported the absence of absorption of some compounds at wavelengths > 205 nm and detection with UHPLC-DAD-ELSD of the presence of terpene compounds without chromophore groups, possibly saponins and cyclopasifloic acid glycoside terpenoid structures in species of

Passiflora spp [31–33,40].

A previous acute oral toxicity study was performed in our lab in mice to determine the safety profile and the absence of effects or alterations induced by the *Passiflora edulis* aqueous extract at 2000 mg/kg single dose. All animals survived to the end of the test and were observed for 14 days. Our data suggest that the *Passiflora edulis* extract is well tolerated for mice. For this reason, we select the 1000 mg/kg dose for the subacute study.

This study determined the sub-acute toxicity for 28 days of an aqueous *Passiflora edulis* f *edulis* extract. This preparation, in which batch-to-batch consistency was confirmed, was based on traditional Colombian knowledge. The NOAEL for sub-acute oral 28-day toxicity was 1000 mg/kg/day in a rat model. This result suggests low toxicity seen in acute and 7-day toxicity studies with an aqueous *Passiflora edulis* f *edulis* extract at oral doses of up to 100, 200, 300, and 400 mg/kg body weight in a rat model [21]. Considering the components reported for aqueous *Passiflora edulis* f *edulis* extract [20–27], the literature reports that extracts with glycosylated flavonoids have low toxicity in oral administration [41].

Determining weight gain or relative organ weight is essential to identify possible damage from exposure to toxic substances in the extract. The weight of the damaged organ would be altered depending on the degree of toxicity and the proportion of the body weight. In this subacute toxicity study, the extract administered up to 1000 mg/kg did not cause any change in clinical signs, mortality, or morbidity. In addition, weight gain was observed in the animals throughout the 29 days of the study. These results suggest that the extract did not cause any detrimental effect on the relative weight of the organs compared to the control animals at the end of the study.

The liver is the main organ metabolizing and detoxifying drugs and environmental chemicals. For this reason, evaluating liver function is essential to determine the toxicity of drugs and plant extracts. AST, ALT, ALP, total protein, and total bilirubin concentration are liver disease's most common clinical biomarkers [27]. Serum ALT activity has been used as an essential biomarker of liver injury in preclinical studies, as

Table 4

Effect of Passiflora edulis ex	extract on hematological	parameters of control and
experimental groups. $*p < 0$	0.05.	

			- 1	- 1
	Males -	Males -	Females -	Females -
	Control	Extract	Control	Extract
	(n = 5)	(n = 5)	(n = 5)	(n = 5)
Hematocrit (%)	52.33	48.30	$\textbf{47.18} \pm \textbf{3.61}$	46.25 ± 3.66
	± 1.55	\pm 3.75		
Hemoglobin (g/	17.20	16.93	14.74 ± 0.81	15.63 ± 1.20
dL)	± 0.61	\pm 1.21		
RBCs (IU)	$\textbf{8.66} \pm \textbf{0.38}$	8.10	$\textbf{7.48} \pm \textbf{0.45}$	$\textbf{7.74} \pm \textbf{0.38}$
		\pm 0.46		
VCM	60.56	59.60	63.08 ± 3.94	59.73 ± 1.90
	\pm 3.79	\pm 2.01		
HCM	19.88	20.88	19.71 ± 0.43	20.20 ± 0.56
	± 0.28	± 0.67		
MCC	32.90	35.05	31.36 ± 1.99	33.80 ± 0.37
	\pm 1.71	± 0.35		
Leukocytes	8163 ± 666	7675	6134 ± 1398	5025 ± 960
		± 1723		
Neutrophils (%)	20.80	15.00	13.82 ± 3.70	11.75 ± 3.40
	\pm 2.36	\pm 6.22		
Eosinophils (%)	2.09	0.25	2.43	0.25
	\pm 0.74 *	\pm 0.50 *	\pm 0.77 *	\pm 0.50 *
Lymphocytes	75.76	78.50	82.11 ± 4.52	88.00 ± 3.83
(%)	\pm 3.27	\pm 9.61		
Monocytes (%)	1.50	0.50	1.13	0 *
	\pm 0.87 *	\pm 0.57 *	\pm 0.93 *	
Total	1702 ± 277	1089	871 ± 315	582 ± 188
Neutrophils		\pm 250		
(µL)				
Total	127	155	175.52	135
Eosinophils	\pm 42.45 *	\pm 70.29 *	\pm 34.50 *	\pm 27.00 *
(µL)				
Total	6175 ± 445	6053	4995 ± 1025	4429 ± 912
Lymphocytes		± 1758		
(µL)				
Total	124.7	77.00	45.00	0 *
Monocytes	\pm 77.32 *	\pm 32.37 *	\pm 52.37 *	
(µL)				
Total Platelets	949.3	1150	1130	1094
	\pm 41.88	\pm 137.4	\pm 130.1	\pm 30.18

damaged hepatocytes release ALT into the extracellular space with increased blood [27]. In this study, serum ALT increased values compared to the control groups in male and female rats. The ALT levels of 184 U/L and 328.0 U/L suggest the diagnostic criteria of liver damage, compared to the control groups of 50.25 U/L and 64.17 U/L in male and female rats, respectively. However, histopathological examination of the liver does not correlate with alterations in cellular structure, inflammation, or any other unfavorable effect.

In the present study, the control and extract-treated groups observed no significant alterations in the serum levels of GGT, FA, total protein, and total bilirubin. The present study found that administration of *Passiflora edulis* extract at a dose of 1000 mg/kg did not adversely affect the liver. Likewise, histological examination of the liver showed small changes in the tissue and induced a statistically significant increase in ALT. However, the results suggest that administration of the extract is relatively safe for the liver. These results can be correlated with several works that have reported that *Passiflora edulis* showed hepatoprotection effect in rats [42–45]. In addition, in ethanol-induced hepatic injury, the daily treatment with Fruit juice protected the liver and decreased AST and ALT activity, inflammation, and oxidative stress [44].

The results obtained from the histopathological study show in the liver the presence of a slight mono-morphonuclear infiltrate (lymphocyte-macrophage), periportal, and a slight increase in hepatocyte binucleation. The above may suggest a mild chronic proinflammatory hepatic parenchymal proinflammatory effect with associated mild intrahepatic cholangitis. This effect could be unrelated to the direct cytopathic effect of administering the aqueous extract at a dose of 1000 mg/kg/day for 28 days, given that the hepatic and cholangiositic profile evidenced between the control and experimental groups did not show any statistical difference in the hepatic biochemical biomarkers. Toxicological studies in repeated doses at 28 days [46,47] after oral administration of 1000 mg/kg/day of a glycosylated flavonoid similar to those reported for the extract of Passiflora, have also found a minimal focal and multifocal infiltrate in the liver, where the authors suggest that such histopathological liver finding may not be related to toxicological significance.

The kidneys are considered frequent targets of toxicity. Monitoring biochemical markers and histological changes is radical for determining kidney health. The biochemical results did not show significant differences in the urea, creatinine, and BUN levels. These results suggest that *Passiflora* extract does not affect the normal renal function following sub-acute exposure compared to the control group. Similarly, histological analysis showed no differences in the tissue architecture of the organ between the treatment and control groups.

The renal analysis showed histopathological results with a slight focal increase of corticomedullary tubular nephron-calcifications; a related toxicological change may not justify that because the appearance

Table 6

Effect of *Passiflora edulis* extract on kidney function profile of control and experimental groups.

Kindey Function Profile					
	Males - Control (n = 5)	Males - Extract (n = 5)	Females -Control (n = 5)	Females - Extract (n = 5)	
Urea (mg/dL)	$\begin{array}{c} 58.10 \\ \pm \ 2.71 \end{array}$	$58.12 \\ \pm 8.28$	56.68 ± 5.99	$\begin{array}{c} 69.60 \\ \pm \ 13.79 \end{array}$	
BUN (mg/dL)	$\begin{array}{c} 33.36 \\ \pm \ 11.04 \end{array}$	$\begin{array}{c} 27.16 \\ \pm \ 4.08 \end{array}$	$\textbf{30.57} \pm \textbf{5.46}$	32.52 ± 6.44	
Creatinine (mg/ dL)	$\textbf{0.73} \pm \textbf{0.05}$	0.60 ± 0.19	$\textbf{0.75} \pm \textbf{0.05}$	$\textbf{0.84} \pm \textbf{0.18}$	
Total Plasma Proteins (g/ dL)	$\textbf{8.26} \pm \textbf{0.40}$	$\textbf{6.02} \pm \textbf{0.51}$	$\textbf{7.56} \pm \textbf{0.36}$	$\textbf{7.56} \pm \textbf{0.94}$	
Kidney Biopsia	Corticomedullary tubular nephrocalcifications				

Table 5

Effect of Passiflora edulis extract on liver function profile of control and experimental groups.

Liver Function Profile				
	Males - Control ($n = 5$)	Males - Extract ($n = 5$)	Females -Control $(n = 5)$	Females - Extract (n = 5)
ALT (U/L)	50.25 ± 4.19 *	184.4 ± 271.9 *	64.17 ± 10.15 *	328.0 ± 325.0 *
GGT (U/L)	2.88 ± 2.17	1.00 ± 1.23	2.64 ± 2.16	3.20 ± 3.11
AF (IU/L)	367.6 ± 93.74	277.5 ± 10.47	246.8 ± 35.31	$\textbf{288.7} \pm \textbf{68.89}$
Total Bilirubin (mg/dL)	0.076 ± 0.070	0.075 ± 0.095	0.022 ± 0.016	0
Indirect Bilirubin (mg/ dL)	0.046 ± 0.034	0.048 ± 0.055	$\textbf{0.012} \pm \textbf{0.010}$	0
Direct Bilirubin (mg/ dL)	$\textbf{0.074} \pm \textbf{0.073}$	0.050 ± 0.057	$\textbf{0.023} \pm \textbf{0.016}$	0
Liver Biopsia	Portal triads without changes.	Slight periportal mono-morphonuclear infiltrate	Portal triads without changes.	Slight periportal mono-morphonuclear infiltrate

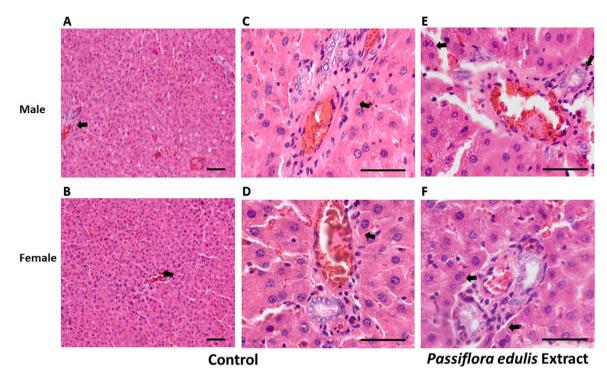


Fig. 4. Hepatic histological study of the animals in sub-acute toxicity. Samples were stained with hematoxylin and eosin (H&E). A. and B. are 10X images, and arrowheads show unchanged portal triads. Bar 10 µm. C. and D. are 40X images of male and female rat samples respectively from the control group. E and F are 40X images of male and female rat samples respectively from the experimental group. Arrowheads show mono-morphonuclear infiltrate and hepatocyte binucleation. Bar 100 µm.

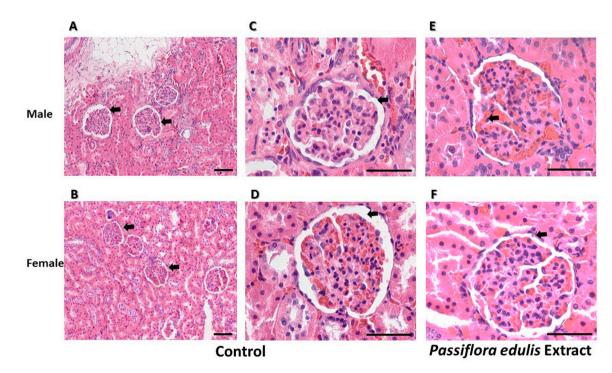
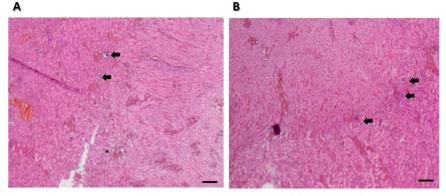


Fig. 5. Renal histological study of the animals in sub-acute. Samples were stained with hematoxylin and eosin (H&E). A. and B. are 10X images (Bar 10 µm). C. and D. are 40X images of male and female rat samples respectively from the control group. Arrowheads show unchanged cortico-medullary glomeruli. E and F are 40X images of male and female rat samples respectively from the experimental group. Arrowheads show cortico-medullary glomeruli with increased mesangium. Bar 100 µm.

of spontaneous nephrocalcinosis has been reported for the experimental animal model [48] together with the fact that no signs of loss of animal welfare were evidenced. These corticomedullary tubular nephron-calcifications have been related to different factors, such as diet changes [49], which can significantly reduce urinary pH [50]. The renal histopathological study showed a slight generalized increase of the intraglomerular mesangium; it should be noted that this change did not modify the renal filtration function since the creatinine result between



Control

Passiflora edulis Extract

Fig. 6. Changes in the renal histological study of the animals in sub-acute. Samples were stained with hematoxylin and eosin (H&E). A. and B. are 10X representative images from female rat samples, respectively, of the control and experimental group. Arrowheads show tubular nephron-calcification (Bar 10 µm). The same finds were observed in the rat male samples.

the control and experimental groups with the administration of 1000 mg/Kg/day of the aqueous extract was similar.

The hematopoietic system is very vulnerable to toxic substances; therefore, it is essential to determine the physiological and pathological states. In this study, subacute administration of *Passiflora edulis* extract caused significant changes in the leukocyte profile of rats treated with the extract compared to the control.

However, it should be noted that the histopathological scenarios reported do not express direct cytopathological findings, or at least in a toxicological magnitude associated with the experiment, which explains the results of the hematological and biochemical analyses (Table 2, Table 3). However, hematology does show a significant difference in the decrease in monocyte and eosinophil counts, which the high variability of the data can explain. On the other hand, the significant variability of the reference values reported for the species in the literature and the experimental ones is given by the high standard deviations reported for the evaluated parameters, on average around 50% (Table 1).

Finally, the aqueous extract of *Passiflora edulis f edulis* had a good safety profile in oral administration for the examined parameters, was well tolerated, and did not cause any lethality or adverse effects in the sub-acute toxicity study in male and female rats. The present study provides data on the chemical profile and sub-acute toxicity of the aqueous extract *Passiflora edulis*, commonly used in traditional medicine. However, other studies should be performed to evaluate the extract's safety, such as acute, subacute, and chronic toxicity, genotoxicity, and carcinogenicity, to understand better the plant's safety profile and its potential use in traditional medicine.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

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References

- Organization WH, WHO Global Report on Traditional and Complementary Medicine 2019, World Health Organization, 2019.
- [2] C.C. Falzon, A. Balabanova, Phytotherapy: an introduction to herbal medicine, Prim. Care Clin. Pract. 44 (2017) 217–227.
- [3] J.M. Riddle, Historical data as an aid in pharmaceutical prospecting and drug safety determination, J. Alter. Complement. Med. 5 (1998) 195–201.
- [4] P. Governa, G. Baini, V. Borgonetti, et al., Phytotherapy in the management of diabetes: a review, Molecules 23 (2018) 105.
- [5] P. Sharma, S.F. McClees, F. Afaq, Pomegranate for prevention and treatment of cancer: an update, Molecules 22 (2017) 177.
- [6] M.S. Baliga, J.J. Dsouza, Amla (Emblica officinalis Gaertn), a wonder berry in the treatment and prevention of cancer, Eur. J. Cancer Prev. 20 (2011) 225–239.
- [7] G.D. Antonio, C.D. Tesser, R.O. Moretti-Pires, Fitoterapia na atenção primária à saúde, Rev. Saude Publica 48 (2014) 541–553.
- [8] L. Cuzzolin, S. Zaffani, G. Benoni, Safety implications regarding use of phytomedicines, Eur. J. Clin. Pharmacol. 62 (2006) 37–42.
- [9] R. Capasso, A.A. Izzo, L. Pinto, et al., Phytotherapy and quality of herbal medicines, Fitoterapia 71 (2000) S58–S65.
- [10] E.M. Williamson, Drug interactions between herbal and prescription medicines, Drug Saf. 26 (2003) 1075–1092.
- [11] D.D. Gummin, J.B. Mowry, M.C. Beuhler, et al., 2020 Annual report of the American association of poison control centers' national poison data system (NPDS): 38th annual report, Clin. Toxicol. 59 (2021) 1282–1501.
- [12] B. Froberg, D. Ibrahim, R.B. Furbee, Plant poisoning, Emerg. Med. Clin. North Am. 25 (2007) 375–433.
- [13] Faleiro F.G., Morera M.P., Costa A.M., et al 2020. Pasifloras: especies cultivadas en el mundo.
- [14] X. He, F. Luan, Y. Yang, et al., Passiflora edulis: An insight into current researches on phytochemistry and pharmacology, Front Pharmacol. 11 (2020), 617.
- [15] I.L. Gadioli, M. da Cunha, S.B. de, M.V.O. de Carvalho, et al., A systematic review on phenolic compounds in Passiflora plants: exploring biodiversity for food, nutrition, and popular medicine, Crit. Rev. Food Sci. Nutr. 58 (2018) 785–807.
- [16] F.C. De Santana, F.B. Shinagawa, E. da S. Araujo, et al., Chemical composition and antioxidant capacity of Brazilian Passiflora seed oils, J. Food Sci. 80 (2015) C2647–C2654.
- [17] M.P. Argentieri, M. Levi, F. Guzzo, P. Avato, Phytochemical analysis of Passiflora loefgrenii Vitta, a rich source of luteolin-derived flavonoids with antioxidant properties, J. Pharm. Pharmacol. 67 (2015) 1603–1612.
- [18] S.P. Ingale, S.B. Kasture, Antioxidant and antiparkinsonian activity of Passiflora incarnata leaves, Orient Pharm. Exp. Med. 14 (2014) 231–236.
- [19] G.S. Taïwe, V. Kuete, Passiflora edulis. Medicinal Spices and Vegetables from Africa, Elsevier, 2017, pp. 513–526.

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- [20] N. Urrego, P. Sepúlveda, M. Aragón, et al., Flavonoids and saponins from Passiflora edulis f. edulis leaves (purple passion fruit) and its potential anti-inflammatory activity, J. Pharm. Pharmacol. 73 (2021) 1530–1538.
- [21] K. Devaki, U. Beulah, G. Akila, V.K. Gopalakrishnan, Effect of aqueous extract of Passiflora edulis on biochemical and hematological parameters of Wistar albino rats, Toxicol. Int. 19 (2012) 63.
- [22] S.S. Patel, S. Verma, G. Nayak, A.K. Singhai, N. Ganesh, Acute and sub-acute toxicity studies of Passiflora nepalensis in rats, Rev. Bras. De. Farmacogn. 21 (2011) 730–736.
- [23] K. Devaki, U. Beulah, G. Akila, V.K. Gopalakrishnan, Effect of aqueous extract of passiflora edulis on biochemical and hematological parameters of Wistar albino rats, Toxicol. Int. 19 (2012) 63–67.
- [24] J.M. Boeira, R. Fenner, A.H. Betti, G. Provensi, Le.A. Lacerda, P.R. Barbosa, et al., Toxicity and genotoxicity evaluation of Passiflora alata Curtis (Passifloraceae), J. Ethnopharmacol. 128 (2010) 526–532.
- [25] C.R. Anurangi, S. Shamina, Preliminary phytochemical screening and acute & subacutetoxicity study on different concentrations of unripen fruit peel flour of Passiflora edulis in male albino rats, World J. Pharm. Pharm. Sci. 7 (2018) 828–834, https://doi.org/10.20959/wjpps20183-11113.
- [26] 2022. Test No. 425: Acute Oral Toxicity: Up-and-Down Procedure. OECD.
- [27] 2008. Test No. 407: Repeated Dose 28-day Oral Toxicity Study in Rodents. OECD.
 [28] A.S. Ayres, L.L.S. de Araújo, T.C. Soares, et al., Comparative central effects of the aqueous leaf extract of two populations of Passiflora edulis, Rev. Bras. Farm. 25
- (2015) 499–505.
 [29] M. Coleta, M.T. Batista, M.G. Campos, et al., Neuropharmacological evaluation of the putative anxiolytic effects of Passiflora edulis Sims, its sub-fractions and flavonoid constituents, Phyther. Res. Int. J. Devoted Pharm. Toxicol. Eval. Nat. Prod. Deriv. 20 (2006) 1067–1073.
- [30] L. Pineli, L. de, O. de, J. da S.Q. Rodrigues, A.M. Costa, et al., Antioxidants and sensory properties of the infusions of wild passiflora from Brazilian savannah: potential as functional beverages, J. Sci. Food Agric. 95 (2015) 1500–1506.
- [31] G. Costa, A. Gazola, S. Zucolotto, L. Castellanos, F. Ramos, F. Reginatto, et al., Chemical profiles of traditional preparations of four South American Passiflora species by chromatographic and capillary electrophoretic techniques. Revista Brasileira de, Rev. Bras. Farmacogn. 26 (2016) 4.
- [32] M.A. Farag, A. Otify, A. Porzel, C.G. Michel, A. Elsayed, L.A. Wessjohann, Comparative metabolite profiling and fingerprinting of genus Passiflora leaves using a multiplex approach of UPLC-MS and NMR analyzed by chemometric tools, Anal. Bioanal. Chem. 408 (2016) 3125–3143.
- [33] N. Urrego, P. Sepúlveda, M. Aragón, F.A. Ramos, G.M. Costa, L.F. Ospina, et al., Flavonoids and saponins from Passiflora edulis f. edulis leaves (purple passion fruit) and its potential anti-inflammatory activity, J. Pharm. Pharmacol. 73 (11) (2021) 1530–1538.
- [34] S.T. Wolford, R.A. Schroer, F.X. Gohs, et al., Reference range database for serum chemistry and hematology values in laboratory animals, J. Toxicol. Health 18 (1986) 161–188.
- [35] L.E. Lillie, N.J. Temple, L.Z. Florence, et al., Reference values for young normal Sprague-Dawley rats: weight gain, hematology and clinical chemistry, Hum. Exp. Toxicol. 15 (1993) 612–616.
- [36] Lyca R. da Fonseca, Rafaele de A. Rodrigues, Aline de S. Ramos, Jefferson D. da Cruz, José Luiz P. Ferreira, Jefferson Rocha de A. Silva, Ana Claudia F. Amaral,

Toxicology Reports 11 (2023) 396-404

Herbal medicinal products from passiflora for anxiety: an unexploited potential, Sci. World J. (2020), https://doi.org/10.1155/2020/6598434.

- [37] Lukasz Dobrek, Krystyna Głowacka, Depression and its phytopharmacotherapy a narrative review, Int J. Mol. Sci. 24 (2023) 4772, https://doi.org/10.3390/ ijms24054772.
- [38] A.S.F.S.J. Ayres, W.B. Santos, D.D. Junqueira-Ayres, G.M. Costa, F.A. Ramos, L. Castellanos, et al., Monoaminergic neurotransmission is mediating the antidepressant-like effects of Passiflora edulis Sims fo. Edulis, Neurosci. Lett. 660 (2017) 79–85, https://doi.org/10.1016/j.neulet.2017.09.010.
- [39] Xirui He, Fei Luan, Yan Yang, Ze Wang, Zefeng Zhao, Jiacheng Fang, Min Wang, Manhua Zuo, Yongsheng Li, Passiflora edulis: an insight into current researches on phytochemistry and pharmacology, Front. Pharmacol. 11 (2020), 617, https://doi. org/10.3389/fphar.2020.00617.
- [40] S.M. Zucolotto, C. Fagundes, F.H. Reginatto, F.A. Ramos, L. Castellanos, C. Duque, et al., Analysis of C-glycosyl flavonoids from South American Passiflora species by HPLC-DAD and HPLC-MS, Phytochem. Anal. 23 (2012) 232–239.
- [41] J. Ritskes-Hoitinga, A.C. Beynen, Nephrocalcinosis in the rat: a literature review, Prog. Food Nutr. Sci. 16 (1) (1992) 85–124.
- [42] S. Zibadia, R. Faridc, S. Moriguchid, Y.R. Lue, L.Y. Fooe, P.M. Tehranic, et al., Oral administration of purple passion fruit peel extract attenuates blood pressure in female spontaneously hypertensive rats and humans, Nutr. Res. 27 (2007) 408–416, https://doi.org/10.1016/j.nutres.2007.05.004.
- [43] R.V. Kavitha, J.R. Kumar, S. Balasubramanian, S.N. Swamy, D. Keerthini, N. Shambhavi, et al., Comparative evaluation of hepatoprotective effects of exotic fruits and common vegetables extracts on CCl4 induced hepatotoxicity: an in vitro study, Int. J. Pharm. Sci. Res. 7 (2016) 3388–3393, https://doi.org/10.13040/ LJPSR.0975-8232.
- [44] Y.J. Zhang, T. Zhou, F. Wang, Y. Zhou, Y. Li, J.J. Zhang, et al., The effects of syzygium samarangense, passiflora edulis and solanum muricatum on alcoholinduced liver injury, Int. J. Mol. Sci. 17 (2016) 1616, https://doi.org/10.3390/ ijms17101616.
- [45] S. Shanmugam, D. Sivaraj, B. Dos Santos Lima, P. Dos Passos Menezes, Y.M.B.G. de Carvalho, A.A. de Souza Araújo, N. Narain, M.R. Serafini, L.J. Quintans Júnior, L. Scotti, M.T. Scotti, T. Parimelazhagan, Polyphenols rich Passiflora leschenaultii leaves modulating Farnesoid X Receptor and Pregnane X Receptor against paracetamol-induced hepatotoxicity in rats, Biomed. Pharmacother. 88 (2017) 1114–1121, https://doi.org/10.1016/j.biopha.2017.01.156.
- [46] Mukinda J.T., 2005. Acute and chronic toxicity of the flavonoid-containing plant, Artemisia afra in rodents.
- [47] Y. Li, A.D. Kandhare, A.A. Mukherjee, S.L. Bodhankar, Acute and sub-chronic oral toxicity studies of hesperidin isolated from orange peel extract in Sprague Dawley rats, Regul. Toxicol. Pharmacol. 105 (2019) 77–85.
- [48] S. Dimitrov, T. Lange, J. Born, Selective mobilization of cytotoxic leukocytes by epinephrine, J. Immunol. 184 (2010) 503–511.
- [49] J. Ritskes-Hoitinga, A.C. Beynen, Nephrocalcinosis in the rat: a literature review, Prog. Food Nutr. Sci. 16 (1992) 85–124.
- [50] H. Mohamed, A. Alhaidary, A. Beynen, Nephrocalcinosis and urinary mineral concentrations in rats fed diets containing various concentrations of magnesium, J. Anim. Vet. Adv. 9 (2010) 2405–2408.