

A meta-analysis on the prognosis of exosomal miRNAs in all solid tumor patients

Jiupeng Zhou, MMed^{a,*}, Hui Guo, PhD^b, Yuanli Yang, MMed^a, Yongfeng Zhang, MMed^a, Heng Liu, MMed^a

Abstract

Background: It has been reported that the encapsulated miRNAs from exosomes are potential biomarkers of tumors prognosis. Yet, the results are controversial, so it is obliged to do a meta-analysis to reach a definite conclusion.

Materials and methods: Studies were searched for published in PubMed, Embase, and Web of Science databases until April 20, 2018. A meta-analysis was conducted to appraise the role of exosomal miRNAs in prognosis of cancer patients.

Results: The different exosomal miRNAs expression was remarkably related to overall survival (OS) (hazard ratio [HR] = 2.02, 95% confidence interval [CI]: 1.84–2.21) and disease-free survival (DFS) (HR = 2.43, 95% CI: 1.86–3.17) of cancer patients. High exosomal miR-21 expression was associated with poor OS (HR = 2.59; 95% CI: 1.71–3.90) and DFS (HR = 1.84; 95% CI: 1.37–2.47). High exosomal miR-451a expression was associated with poor OS (HR = 4.81; 95% CI: 2.33–9.93) and DFS (HR = 2.64; 95% CI: 1.62–4.31). High exosomal miR-1290 expression was associated with poor OS (HR = 1.73; 95% CI: 1.29–2.33). Low exosomal miR-638 expression was associated with poor OS (HR = 2.25; 95% CI: 1.46–3.46).

Conclusion: The expression levels of exosomal miRNAs, particularly miR-21, miR-451a, miR-1290, and miR-638 could strongly predict prognosis of solid tumor patients and might be a potential target for tumor treatment.

Abbreviations: CI = confidence interval, DFS = disease-free survival, HR = hazard ratio, NOS = Newcastle–Ottawa quality assessment scale, OS = overall survival, TTR = time to tumor recurrence.

Keywords: exosomal miRNAs, meta-analysis, prognosis, tumor

1. Introduction

MiRNAs are a short (20–24nt) noncoding RNA, which regulate gene expression at the post-transcriptional level in molecular mechanisms by affecting the stability and translation of mRNA. In addition, miRNAs target protein-coding mRNA at the posttranscriptional level by directly dividing mRNA or inhibiting protein synthesis. It has been found that the imbalance of miRNA is related to the occurrence and progression of cancer, implying that miRNAs are possible to be as molecular biomarker for diagnosis and prognosis prediction of cancer.^[1,2] Exosomes are nanoscale vesicles (40–100 nm), which originate from the membrane cavity of multiple vesicles and release with cell membrane.^[3] As we all know, exosomes contain tissue-specific signals composed of proteins and selective packaged RNA, such as miRNA, which can transfer these components to other cells.^[4]

Medicine (2019) 98:16(e15335)

Received: 7 November 2018 / Received in final form: 5 March 2019 / Accepted: 26 March 2019

http://dx.doi.org/10.1097/MD.00000000015335

from exosomes are potential biomarkers of tumors prognosis, including lung cancer,^[5–7] hepatocellular carcinoma,^[8–10] hepatoblastoma,^[11,12] glioma,^[13] colorectal cancer,^[14–20] ovarian cancer,^[21] prostate cancer,^[22] kidney cancer,^[23] and pancreatic ductal adenocarcinoma.^[24,25] However, the results of some researches were disputable. Some studies demonstrated that the high expression of exosome-delivered miRNAs in human peripheral blood might be associated with the poor prognosis of tumor patients,^[17,22] others found that the high expression of exosomal miRNAs was distinctly not related to the unfavorable prognosis of tumor patients.^[6,21] Even more, some studies reported that the high expression of exosome-delivered miRNAs might be associated with the favorable prognosis of tumor patients.^[12,16] Therefore, the objective of this meta-analysis is to explore the prognosis of exosomal miRNAs in all solid tumor patients and refrain from the possible deviations.

2. Materials and methods

Ethical approval was not necessary in meta-analysis.

2.1. Literature search strategy

In order to gain the potential qualified research, systematic network document search was aimed at multiple websites database, including Embase, PubMed, and Web of Science until April 20, 2018, and the search keywords were as follows: "exosomal miR", "exosomes", "cancer", "tumor", "prognosis", and "survival". The relevant systematic reviews and references cited in the searched articles were also filted to avoid leaving out any potentially usable researches. Besides, other related articles were also available by examining the reference list by hand.

Editor: Jianxun Ding.

The authors declare that they have no conflict of interest.

^a Xi'an Chest Hospital, Xi'an, ^b The First Affiliated Hospital of Xi'an Jiaotong University, Shaanxi Province, China.

^{*}Correspondence: Jiupeng Zhou, Xi'an Chest Hospital, Xi'an 710000, Shaanxi Province, China (e-mail: 44996323@qq.com).

Copyright © 2019 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

2.2. Selected and removed criteria

The selected criteria were as follows:

- 1) the expression level of exosomal miRNAs in primary cancerous serum exosomes was measured;
- 2) dichotomous model was appraised by quantitative polymerase chain reaction (qPCR);
- 3) proven diagnosis of tumor by histopathology;
- 4) hazard ratio (HR) and 95% confidence interval (CI) between exosomal miRNAs expression and overall survival (OS) and/ or DFS/time to tumor recurrence (TTR) could be picked up directly or calculated indirectly in the study.

The removed criteria were as follows:

- 1) the letters, experiments, or articles that researched animal models;
- 2) repeated research publications;
- 3) the expression of miRNAs was detected in plasma or tumor tissue.

2.3. Date extraction and quality evaluation

Two investigators (JP zhou and YL yang) independently extracted the information and data from all eligible studies through cross-check. The data and information were got together from every study using a purpose-designed form: the author, the year of publication, the country, the type of cancer, the overall number of patients, and the standard for high exosomal miRNAs expression. The survival results of both original and adjusted data were OS and/or DFS/TTR. The disagreements between the 2 investigators were settled by means of discussion until an agreement was reached with the third investigators. Quality evaluation was based on the Newcastle–Ottawa quality assessment scale (NOS). The NOS scores varied from 0 to 9. Six points or more were deemed as high quality.

2.4. Statistical analysis

The current meta-analysis was carried out using the RevMan5.3 software and Stata SE13.0 software. The prognostic effect of exosomal miRNAs expression was assessed via both adjusted and unadjusted HRs and their 95% CIs of OS and/or DFS/TTR from the primary studies. According to the standard of every study, the expression of exosomal miRNAs was divided into high and low levels. Reported HR and 95% CI were immediately collected from the researches. If HR and 95% CI were not specific in the studies, the means of Tierney et al^[26] and Parmar et al^[27] were recommended to evaluate HRs and 95% CIs. The heterogeneity among the enrolled studies was performed by I² metric and Q statistic, the P value for the Q test <.05 and the I²value >50% were taken into account to be indexes of serious heterogeneity. The random-effects model was selected for the researches with a remarkable heterogeneity ($P \leq .05$, $I^2 \geq 50\%$). Or else, the fixed-effects model was applied (P > .05, $I^2 < 50\%$). Publication bias was appraised by Egger test and a funnel plot, and P < .05 demonstrated remarkable bias. The P value < .05 was deemed statistically significant.

3. Results

3.1. Studies identification and characteristics of eligible studies

After the initial search algorithm, a total of 396 articles were retrieved. Unrelated articles were excluded from headlines and

abstracts, and 73 articles were further evaluated in the full text. Those articles, which were review article or case and did not provid survival data to extract, dichotomous variables and valuable data, were excluded. Finally, this meta-analysis contained 21 articles including a total number of 2971 patients (Fig. 1). The average sample number of patients every study was 141.2 (range:30–326). In the 21 studies, 11 of them came from the People's Republic of China, 8 from Japan and 2 from USA In this meta-analysis, 9 different types of cancers were contained, which were 3 lung cancer, 2 hepatoblastoma, 3 hepatocellular carcinoma, 1 glioma, 7 colorectal cancer, 1 ovarian cancer, 1 prostate cancer, 1 kidney cancer, and 2 pancreatic adenocarcinoma respectively (Table 1).

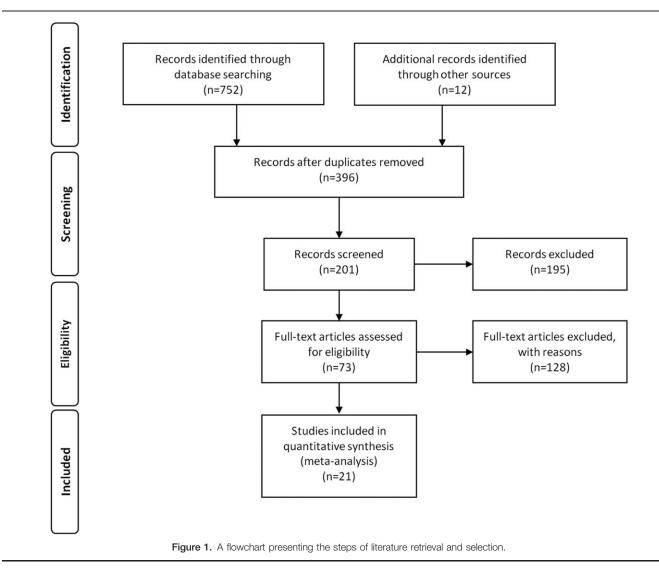
3.2. Association between exosomal miRNAs expression and prognosis

A fixed-effects model was employed to estimate the combined HR with 95% CI for the outcomes of OS in virtue of restricted heterogeneity ($I^2 = 21\%$, P = .13). Pooled HR value (95% CI) of OS associated with different exosomal miRNAs expression was 2.02 (1.84–2.21) in all solid tumor patients (Fig. 2). In addition, the random-effects model was carried out for significant heterogeneity in the studies with DFS ($I^2 = 59\%$, P = .001). The pooled HR value (95% CI) of DFS associated with different exosomal miRNAs expression was 2.43 (1.86-3.17) (Fig. 3) in all solid tumor patients. Poor prognosis was associated with the upregulation of 22 exosomal miRNAs (miR-21, miR-4257, miR-375, miR-23b-3p, miR-21-5p, miR-19a-3p, miR-194-5p, miR-1290, miR-10b-5p, let-7g-5p, miR-451a, miR-665, miR-301a, miR-19a, miR-4772-3p, miR-6803-5p, miR-203, miR-373 miR-200a, miR-200b, miR-200c, miR-1246) and with downregulation of 11 exosomal miRNAs (miR-34s, miR-125b, miR-638, miR-6869-5p, miR-190b, miR-26a-1-3p, miR-145-3p, miR-200a-3p, let-7i-5p, miR-9-5p, miR-615-3p). Five exosomal miRNAs (miR-21, miR-451a, miR-375, miR-1290 and miR-638) were evaluated by at the least 2 studies. A subgroup meta-analysis was carried out in the relevant studies (Figs. 4 and 5).

The association of exosomal miR-21 with survival outcomes was reported in 4 studies. Two reported OS and 3 reported DFS. The meta-analysis displayed that the high exosomal miR-21 expression was distinctly related to poor OS (a fixed-effect model, HR = 2.59; 95% CI: 1.71–3.90; P <.00001; $I^2 = 0\%$, P =.35). The analysis indicated a pooled HR = 1.84 (95% CI: 1.37–2.47, P <.00001), demonstrating a poor DFS of high exosomal miR-21 expression ($I^2 = 48\%$, P =.14).

Three studies discussed the relationship between exosomal miR-451a and the result of survival, of which 2 contained OS and 2 contained DFS. The results displayed that the high exosomal miR-451a expression was distinctly related to poor OS (a fixed-effect model, HR=4.81; 95% CI: 2.33–9.93; P<.00001; I^2 = 0%, P=.40). It was demonstrated that the high exosomal miR-451a expression distinctly correlated with poor DFS (a fixed-effect model, HR=2.64; 95% CI: 1.62–4.31; P<.00001; I^2 = 0%, P=.84).

Two studies expounded the part of exosomal miR-1290 in OS of tumor patients. Then, a meta-analysis was conducted on the relation of exosomal miR-1290 expression and OS. This demonstrated that elevatory exosomal miR-1290 expression correlated with poor OS (a fixed-effect model, HR = 1.73; 95% CI: 1.29–2.33; P < .001; $I^2 = 0$, P = .71).



Two studies investigated the association between exosomal miR-375 expression and OS of tumor patients. A meta-analysis was executed on the relationship of exosomal miR-375 expression and OS. The results suggested that abnormal exosomal miR-375 expression was not related to OS (a random-effect model, HR=1.72; 95% CI: 0.72–4.06; P=.23; $I^2=80\%$, P=.02).

Two studies included OS for exosomal miR-638 in tumor patients. When a meta-analysis was performed on the relationship of exosomal miR-638 expression and OS, the results indicated that a lower expression of exosomal miR-638 could predict shorter OS (a fixed-effect model, HR=2.25; 95% CI: 1.46–3.46; P < .001; $I^2 = 0$, P = .37).

3.3. Publication bias

Egger test was used to evaluate the publication bias. Egger test demonstrated significant publication bias for OS and DFS in all solid tumor patients (P = .004, P = .014) (Table 2, Fig. 6). The bias indicated the presence of a language bias, a potential publication bias, an exaggerated estimates by a flawed methodologic design in smaller studies, and a deficiency of publication of small trials with contrary results.

4. Discussion and Conclusions

Over the past few years, expression of miRNAs has been assessed in several studies in order to select potential diagnostic and/or prognostic biomarkers in solid tumors. However, the clinical significance of plasma/serum miRNA as a diagnostic biomarker is still disputable.^[28–31] The instability of miRNA in plasm/serum may be 1 reason for this controversy. Recent most studies examined the role of exosomal miRNAs in various tumors as a biomarker.

In this study, we systematically analyzed 2971 tumor samples from 21 appropriate articles and reported, for the first time, a group of 33 exosomal miRNAs related to prognosis of all solid tumors. This study aimed to identify the relationship between exosomal miRNAs and solid tumors prognosis, which could be further confirmed in future studies and ultimately assessed before the treatment to improve the treatment of solid tumors.

This research discovered that different exosomal miRNAs expression was associated with OS and DFS in all solid tumor patients, and the pooled HR value (95% CI) was respectively 2.02 (1.84–2.21) and 2.43 (1.86–3.17). The upregulation of 22 exosomal miRNAs (miR-21, miR-4257, miR-375, miR-23b-3p, miR-21–5p, miR-19a-3p, miR-194–5p, miR-1290, miR-10b-5p,

Table 1

Author, yr	Country	miRNAs investigated	Dysregulation	Number	Cancer type	Detection method	Criterion of high expression	Quality stars, NOS
Hitoshi Dejima 2017	Japan	miR–21	Upregulation	195	LC	RT-qPCR	the average of expression levels	7
		miR–4257	Upregulation	195	LC	RT-qPCR	he average of expression levels	
Qingyun Liu 2017	China	miR-375	Upregulation	196	LC	RT-qPCR	Not reported	8
		miR-23b-3p	Upregulation	196	LC	RT-qPCR	Not reported	
		miR-21-5p	Upregulation	196	LC	RT-qPCR	Not reported	
		miR-19a-3p	Upregulation	196	LC	RT-qPCR	Not reported	
		miR-194-5p	Upregulation	196	LC	RT-qPCR	Not reported	
		miR-1290	Upregulation	196	LC	RT-qPCR	Not reported	
		miR-10b-5p	Upregulation	196	LC	RT-aPCR	Not reported	
		let-7g-5p	Upregulation	196	LC	RT-qPCR	Not reported	
Rie Kanaoka 2017	Japan	miR-451a	Upregulation	285	LC	RT-aPCR	the average of expression levels	9
Wanbo Liu 2016	China	miR–21	Upregulation	32	HB	RT-qPCR	2.0-fold	8
Chenwei Jiao 2016	China	miR-34s	Downregulation	63	HB	RT-aPCR	0.5-fold	8
Weifeng Liu 2017	China	miR- 125b	Downregulation	128	HCC	RT-qPCR	the average of expression levels	9
Min Shi 2017	China	miR-638	Downregulation	126	HCC	RT-qPCR	Not reported	8
Zhen Qu 2017	China	miR-665	Upregulation	30	HCC	RT-qPCR	5-fold	9
Fengming Lan 2017	China	miR-301a	Upregulation	60	GM	RT-qPCR	the average of expression levels	8
T Matsumura 2015	Japan	miR-19a	Upregulation	227	CRC	RT-qPCR	the average of expression levels	8
Chang Liu 2016	Japan	miR-4772-3p	Upregulation	84	CRC	RT-qPCR	the 60th percentile of the Δ CT value	8
Shushan Yan 2017	China	miR-638	Downregulation	192	CRC	RT-aPCR	Not reported	8
Mitsuo Tsukamoto 2017	Japan	miR-21	Upregulation	326	CRC	RT-qPCR	the average of expression levels	9
Shushan Yan 2017	China	miR-6803-5p	Upregulation	168	CRC	RT-qPCR	Not reported	8
Shushan Yan 2017	China	miR-6869-5p	Downregulation	142	CRC	RT-aPCR	Not reported	8
Yuki Takano 2017	Japan	miR-203	Upregulation	240	CRC	RT-qPCR	the average of expression levels	9
Xiaodan Meng 2016	China	miR-373	Upregulation	163	OC	RT-qPCR	the average of expression levels	9
Adduart Mong 2010	China	miR-200a	Upregulation	163	00	RT-aPCR	the average of expression levels	0
	China	miR-200b	Upregulation	163	00	RT-aPCR	the average of expression levels	
	China	miR-2000	Upregulation	163	00	RT-qPCR	the average of expression levels	
XiaoyiHuang 2015	USA	miR-1290	Upregulation	123	PCa	RT-qPCR	the average of expression levels	7
Aldoyii ludiig 2010	UUA	miR-1246	Upregulation	123	PCa	RT-aPCR	the average of expression levels	1
		miR-375	Upregulation	123	PCa	RT-qPCR	the average of expression levels	
Meijun Du 2017	USA	miR-190b	Downregulation	109	KC	RT-aPCR	Not reported	8
Weijuli Du 2017	UUA	miR-26a-1-3p	Downregulation	109	KC	RT-qPCR	Not reported	0
		miR-145-3p	Downregulation	109	KC	RT-aPCR		
		miR-200a-3p	Downregulation	109	KC	RT-qPCR		
		let-7i-5p	Downregulation	109	KC KC	RT-qPCR RT-qPCR		
		1	Downregulation	109	KC KC	RT-qPCR RT-qPCR		
		miR-9-5p	0	109	KC KC			
Kupibika Takabasi 0010	longe	miR-615-3p	Downregulation			RT-qPCR	the querose of overcesies levels	0
Kunihiko Takahasi 2018	Japan	miR-451a	Upregulation	50	PC	RT-qPCR	the average of expression levels	9
Takuma Goto 2018	Japan	miR- 21	Upregulation	32	PC	RT-qPCR	the average of expression levels	8

CRC=colorectal cancer, GM=glioma, HB=hepatoblastoma, HCC=hepatocellular carcinoma, KC=kidney cancer, LC=lung cancer, OC=ovarian cancer, PC=pancreatic ductal adenocarcinoma, PCa=prostate cancer.

let-7g-5p, miR-451a, miR-665, miR-301a, miR-19a, miR-4772– 3p, miR-6803–5p, miR-203, miR-373 miR-200a, miR-200b, miR-200c, miR-1246) and downregulation of 11 exosomal miRNAs (miR-34s, miR-125b, miR-638, miR-6869–5p, miR-190b, miR-26a-1–3p, miR-145–3p, miR-200a-3p, let-7i-5p, miR-9–5p, miR-615–3p) correlated with poor prognosis.

We conducted the meta-analysis on these 5 exosomal miRNAs (miR-21, miR-451a, miR-375, miR-1290, and miR-638) to determine a pooled conclusion because the 5 exosomal miRNAs were identified by at least 2 studies. The study found that high exosomal miR-21 expression was related to poor OS and DFS. High exosomal miR-451a expression was related to poor OS and DFS. High exosomal miR-375 expression was not related to poor OS. High exosomal miR-1290 expression was related to poor OS. Low exosomal miR-638 expression was related to poor OS.

Aside from the above mentioned exosomal miRNAs, this study also made a systematic study of the relationship between exosomal miRNAs and lung cancer, hepatocellular carcinoma, colorectal cancer, and pancreatic ductal adenocarcinoma prognosis. Figures 4 and 5 summarize the relationship between prognosis of the tumors and exosomal miRNAs.

It is deserved to be mentioned that exosomal miR-21, a wellknown miRNA studied in different cancer types. A meta-analysis shows that exosomal miR-21 has a powerful potential to be served as a general biomarker to diagnose cancers.^[32] Zhou et al^[33] collected 63 published studies and discovered that the increase of miR-21 expression indicated the deterioration of OS in cancers. The mechanism of miR-21 affecting the metastasis and unfavorable prognosis of tumor patients has been reported. MiR-21 can induce tumor by inhibiting the negative regulation of the RAS/MEK/ERK pathway and apoptosis.^[34] It is well known that over expression of miR-21 downregulates the expression of PTEN, PDCD4, and TPM1 and promotes cell proliferation and cancer progression.^[34] In addition, overexpressed miR-21

Chudu or Cubaroun	logillagerd Patial	er.	Waight	Hazard Ratio IV. Fixed, 95% C	Hazard Ratio
Study or Subgroup	log[Hazard Ratio]				
Chang LiumiR-4772-3pUp		0.72		6.17 [1.51, 25.31]	
Chenwei JiaomiR-34sDown		0.14	10.9%	1.86 [1.41, 2.45]	
Fengming LanmiR-301aUp		0.29	2.5%	4.39 [2.49, 7.76]	
Kunihiko miR-451aUp		0.61	0.6%	3.19 [0.97, 10.54]	
Meijun Dulet-7i-5pDown	0.54	0.2	5.4%	1.72 [1.16, 2.54]	
Meijun DumiR-145-3pDown		0.19	5.9%	1.90 [1.31, 2.75]	
Meijun DumiR-190bDown		0.21	4.9%	2.16 [1.43, 3.26]	
Meijun DumiR-200a-3pDown	0.65	0.21	4.9%	1.92 [1.27, 2.89]	
Meijun DumiR-26a-1-3pDown	0.76	0.21	4.9%	2.14 [1.42, 3.23]	
Meijun DumiR-615-3pDown	0.62	0.3	2.4%	1.86 [1.03, 3.35]	
Meijun DumiR-9-5pDown	0.6	0.23	4.1%	1.82 [1.16, 2.86]	
Min ShimiR-638Down	1.02	0.32	2.1%	2.77 [1.48, 5.19]	
Mitsuo miR-21Up	0.82	0.3	2.4%	2.27 [1.26, 4.09]	
Qingyun Liulet-7g-5pUp	0.25	0.28	2.7%	1.28 [0.74, 2.22]	
Qingyun LiumiR-10b-5pUp	0.8	0.32	2.1%	2.23 [1.19, 4.17]	
Qingyun LiumiR-1290Up	0.45	0.3	2.4%	1.57 [0.87, 2.82]	
Qingyun LiumiR-194-5pUp	0.1	0.34	1.9%	1.11 [0.57, 2.15]	
Qingyun LiumiR-19a-3pUp	0.47	0.28	2.7%	1.60 [0.92, 2.77]	
Qingyun LiumiR-21-5pUp	0.75	0.26	3.2%	2.12 [1.27, 3.52]	
Qingyun LiumiR-23b-3pUp	0.88	0.26	3.2%	2.41 [1.45, 4.01]	
Qingyun LiumiR-375Up	0.1	0.27	2.9%	1.11 [0.65, 1.88]	
Rie KanaokamiR-451aUp	1.8	0.46	1.0%	6.05 [2.46, 14.90]	
T MatsumuramiR-19aUp	0.91	0.45	1.1%	2.48 [1.03, 6.00]	
Takuma GotomiR- 21Up	1.4	0.55	0.7%	4.06 [1.38, 11.92]	
Weifeng LiumiR-125bdown	0.12	0.36	1.7%	1.13 [0.56, 2.28]	
Xiaodan MengmiR-200aUp	0.53	0.38	1.5%	1.70 [0.81, 3.58]	
Xiaodan MengmiR-200bUp	1.03	0.46	1.0%	2.80 [1.14, 6.90]	
Xiaodan MengmiR-200cUp		0.44	1.1%	2.51 [1.06, 5.94]	
Xiaodan MengmiR-373Up		0.42	1.2%	2.89 [1.27, 6.57]	A DECEMBER OF A
XiaoyiHuangmiR-1246Up		0.63	0.5%	8.00 [2.33, 27.52]	
XiaoyiHuangmiR-1290Up		0.17	7.4%	1.79 [1.28, 2.49]	
XiaoyiHuangmiR-375Up		0.29	2.5%	2.69 [1.52, 4.75]	
YanmiR-638down	0.63	0.3	2.4%	1.88 [1.04, 3.38]	
YanmiR-6803-5pUp	1.08	0.4	1.3%	2.94 [1.34, 6.45]	
YanmiR-6869-5pdown		0.39	1.4%	2.32 [1.08, 4.97]	
Yuki TakanomiR-203Up		0.29	2.5%	2.27 [1.29, 4.01]	
Zhen QumiR-665Up		0.91	0.3%	3.74 [0.63, 22.28]	
Total (95% CI)			100.0%	2.02 [1.84, 2.21]	•
Heterogeneity: Chi ² = 45.44, df		4.07			

Figure 2. A forest plot for the association between the exosomal miRNA expression levels with OS. OS=overall survival.

Study or Subgroup	log[Hazard Ratio]	SE	Weight	Hazard Ratio IV, Random, 95% CI	3		lazard Ratio Random, 95%	CI	
Chang LiumiR-4772-3pUp	1.7	0.4	5.7%	5.47 [2.50, 11.99]				-	
HitoshiDejimamiR4257Up	2.09	0.69	2.9%	8.08 [2.09, 31.26]					-
HitoshiDejimamiR-21up	1.34	0.66	3.1%	3.82 [1.05, 13.92]					
Kunihiko miR-451aUp	1.05	0.45	5.1%	2.86 [1.18, 6.90]				-	
Mitsuo miR-21Up	0.85	0.23	8.6%	2.34 [1.49, 3.67]					
Rie KanaokamiR-451aUp	0.94	0.3	7.3%	2.56 [1.42, 4.61]					
MatsumuramiR-19aUp	0.91	0.45	5.1%	2.48 [1.03, 6.00]			-	191	
Vanbo LiumiR-21Up	0.36	0.2	9.1%	1.43 [0.97, 2.12]			-		
Veifeng LiumiR-125bdown	1.96	0.36	6.3%	7.10 [3.51, 14.38]			-		
Kiaodan MengmiR-200aUp	0.1	0.29	7.5%	1.11 [0.63, 1.95]			-		
Kiaodan MengmiR-200bUp	0.47	0.29	7.5%	1.60 [0.91, 2.82]					
Kiaodan MengmiR-200cUp	0.53	0.38	6.0%	1.70 [0.81, 3.58]			—		
Kiaodan MengmiR-373Up	0.47	0.3	7.3%	1.60 [0.89, 2.88]					
anmiR-638down	0.53	0.35	6.5%	1.70 [0.86, 3.37]					
anmiR-6803-5pUp	1.18	0.38	6.0%	3.25 [1.55, 6.85]					
ruki TakanomiR-203Up	1.27	0.4	5.7%	3.56 [1.63, 7.80]				_	
Total (95% CI)			100.0%	2.43 [1.86, 3.17]			•		
Heterogeneity: Tau ² = 0.16; C	Chi ² = 36.85, df = 15 (P = 0.	001); l ² = {	59%		-	_	1	100
est for overall effect: Z = 6.5	50 (P < 0.00001)				0.01	0.1 Better	DFS Worse [10 DFS	100

Figure 3. A forest plot for the association between the exosomal miRNA expression levels with DFS. DFS=disease-free survival.

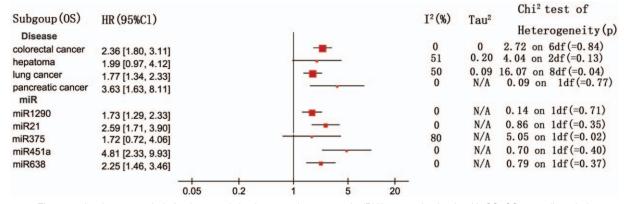


Figure 4. A subgroup analysis for the association between the exosomal miRNA expression levels with OS. OS = overall survival.

Subgoup (DFS)	HR (95C1)								I ² (%)	Tau ²	Chi ² test of Heterogeneity(p)
Disease											
colorectal cancer	2.77 [2.03, 3.79]					-	-		17	0.03	6.01 on 1df(=0.31)
lung cancer	3.16 [1.93, 5.16]					_	-		18	N/A	2.43 on 2df (=0.32)
miR											
miR21	1.84 [1.37, 2.47]					-			48	N/A	3.87 on 2df (=0.14)
miR451a	2.64 [1.62. 4.31]					-			0	N/A	0.04 on 1df (=0.84)
	-				-						
		0.1	0.2	0.5	1	2	5	10			

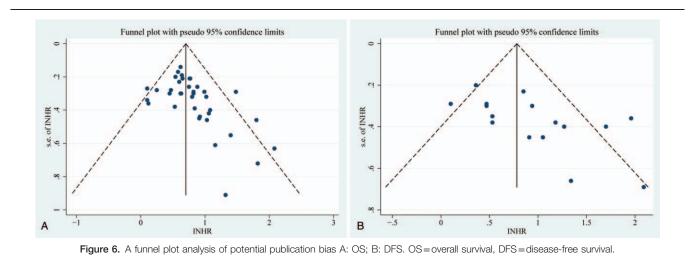
Table 2 The publication bias test including literatures.									
	Coef	95%CI	Р						
OS	1.427	0.493-2.360	.004						
DFS	3.057	0.708-5.405	.014						

DFS = disease-free survival, OS = overall survival.

enhances the phenotype of cancer stem cells and promotes the invasion, migration, and tumorigenesis in hepatocellular carcinoma.^[35] MiR-451a plays a crucial role in the development of human cancers. Kim et al^[36] found that miR-451a regulated

gliomatous cell proliferation and migration by means of the LKB1-AMPK signaling pathway. Wang et al^[37] reported that miR-451a remarkablely inhibited the proliferation of non-small cell lung cancer (NSCLC) cells in vitro, which was partly owing to the downregulation of ras-related protein. Su et al^[38] showed that overexpressed miR-451a was related to cell multiplication, migration, and apoptosis in renal cell carcinoma.

Although results of this meta-analysis were supported by powerful proof, some limitations were worth noting. In evaluating the association between exosomal miRNAs and DFS, the heterogeneity detection indicated markedly heterogeneity. The heterogeneity might be caused by different types of tumor



and different cut points of high expression of exosomal miRNAs. Moreover, some studies of small sample might also contribute to the formation of heterogeneity. Also, there might be different proportion of advanced tumors in different research centers, and could also be a cause of heterogeneity. In addition, in order to ensure the effectiveness of exosomes, the electron microscopy is usually needed. In the included studies, 6 (5, 7, 8, 10, 17, 25) provided electron microscopic pictures, $3^{[6,14,15]}$ used electron microscopy without electron microscopy. These contributed to the formation of heterogeneity in evaluating the association between exosomal miRNAs and the survival of tumor.

Second, in Egger test, it also showed that there was a significant *P* value for OS and DFS in all solid tumor patients. This meant that there was publication bias. Prejudice mainly due to the inclination of positive publication and ignore negative results. Publication bias could only increase the unreliability. In addition, the summary of the meeting was excluded, which might lead to publishing bias.

Third, although most data were directly available in research, some studies only offered survival curves, which led to possible deviations between estimated and actual statistical data. In order to reduce the deviation as much as possible, detailed steps had been taken.

At last, population primarily came from East Asia and did not well represent the population all over the world.

Generally speaking, this meta-analysis suggested that several exosomal miRNAs were associated with poor prognosis in all solid tumor patients and served as a promising biomarker to predict survival outcomes, which might be a potential target for tumor treatment. Such molecules should be combined with other clinical and molecular biomarkers to evaluate the optimal treatment option for solid tumor patients. So, larger scale, multicentre and higher quality studies are needed to verify our results.

Author contributions

Jiupeng Zhou and Yuanli Yang made contributions to conception and design, publication search, quality evaluation, data collection, statistics and manuscript writin; Yongfeng Zhang and Heng Liu made contributions to statistics and editors, and Hui Guo contributed to conception, design, statistics, and editing. **Conceptualization:** Jiupeng Zhou, Hui Guo.

Data curation: Jiupeng Zhou, Hui Guo, Yuanli Yang, Yongfeng Zhang, Heng Liu.

Software: Jiupeng Zhou.

Writing – original draft: Jiupeng Zhou.

Writing - review & editing: Jiupeng Zhou.

References

- Yi R, Li Y, Wang FL, et al. MicroRNAs as diagnostic and prognostic biomarkers in colorectal cancer. World J Gastrointest Oncol 2016;8:330–40.
- [2] Deng D, Liu Z, Du Y. Epigenetic alterations as cancer diagnostic, prognostic, and predictive biomarkers. Adv Genet 2010;71:125–76.
- [3] Peinado H, Lavotshkin AM, MateiI S, et al. Melanoma exosomes educate bone marrow progenitor cells toward a prometastatic phenotype through MET. Nat Med 2012;18:883–91.
- [4] Tovar-Camargo OA, Toden S, Goel A. Exosomal microRNA biomarkers: emerging frontiers in colorectal and other human cancers. Expert Rev Mol Diagn 2016;16:553–67.

- [5] Dejima H, Iinuma H, Kanaoka R, et al. Exosomal microRNA in plasma as a non-invasive biomarker for the recurrence of non-small cell lung cancer. Oncol Lett 2017;13:1256–63.
- [6] Qingyun Liu ZY, Shuai Y, Weijia X, et al. Circulating exosomal microRNAs as prognostic biomarkers for non-small-cell lung cancer. Oncotarget 2017;8:13048–58.
- [7] Kanaoka R, Iinuma H, Dejima H, et al. Usefulness of plasma exosomal microRNA-451a as a noninvasive biomarker for early prediction of recurrence and prognosis of non-small cell lung cancer. Oncology 2018;94:311–23.
- [8] Wang H, Hou L, Li A, et al. Expression of serum exosomal microRNA-21 in human hepatocellular carcinoma. BioMed Res Int 2014;1–5.
- [9] Shi M, Jiang Y, Yang L, et al. Decreased levels of serum exosomal miR-638 predict poor prognosis in hepatocellular carcinoma. J Cell Biochem 2017;119:4711–6.
- [10] Zhen Qu JW, Junyi Wu, Anlai Ji, et al. Exosomal miR-665 as a novel minimally invasive biomarker for hepatocellular carcinoma diagnosis and prognosis. Oncotarget 2017;8:80666–78.
- [11] Liu W, Chen S, Liu B. Diagnostic and prognostic values of serum exosomal microRNA-21 in children with hepatoblastoma: a Chinese population-based study. Pediatr Surg Int 2016;32:1059–65.
- [12] Jiao C, Jiao X, Zhu A, et al. Exosomal miR-34s panel as potential novel diagnostic and prognostic biomarker in patients with hepatoblastoma. J Pediatr Surg 2017;52:618–24.
- [13] Lan F, Qing Q, Pan Q, et al. Serum exosomal miR-301a as a potential diagnostic and prognostic biomarker for human glioma. Cell Oncol 2018;41:25–33.
- [14] Matsumura T, Sugimachi K, Iinuma H, et al. Exosomal microRNA in serum is a novel biomarker of recurrence in human colorectal cancer. Br J Cancer 2015;113:275–81.
- [15] Liu C, Eng C, Lu Y, et al. Serum exosomal miR-4772-3p is a predictor of tumor recurrence in stage II and III colon cancer. Oncotarget 2016;7: 76250–60.
- [16] Shushan Yan GD, Xiaoyu Z, Chengwen J, et al. Downregulation of circulating exosomal miR-638 predicts poor prognosis in colon cancer patients. Oncotarget 2017;8:72220–6.
- [17] Tsukamoto M, Iinuma H, Yagi T, et al. Circulating exosomal microRNA-21 as a biomarker in each tumor stage of colorectal cancer. Oncology 2017;92:360–70.
- [18] Yan S, Jiang Y, Liang C, et al. Exosomal miR-6803-5p as potential diagnostic and prognostic marker in colorectal cancer. J Cell Biochem 2017;119:4113–9.
- [19] Yan S, Wang Z, Duan Q, et al. MicroRNA-6869-5p acts as a tumor suppressor via targeting TLR4/NF-kappaB signaling pathway in colorectal cancer. J Cell Physiol 2017;233:6660–8.
- [20] Yuki Takano TM, Hisae I, Rui Y, et al. Circulating exosomal microRNA-203 is associated with metastasis possibly via inducing tumor-associated macrophages in colorectal cancer. Oncotarget 2017;8:78598–613.
- [21] Xiaodan Meng VM, Karin ML, Fabian T, et al. Diagnostic and prognostic relevance of circulating exosomal miR-373, miR-200a, miR-200b and miR-200c in patients with epithelial ovarian cancer. Oncotarget 2016;7:16923–35.
- [22] Huang X, Yuan T, Liang M, et al. Exosomal miR-1290 and miR-375 as prognostic markers in castration-resistant prostate cancer. Eur Urol 2015;67:33–41.
- [23] Meijun Du KVG, Yijun T, Michael, et al. Plasma exosomal miRNAs-based prognosis in metastatic kidney cancer. Oncotarget 2017;8:63703–14.
- [24] Goto T, Fujiya M, Konishi H, et al. An elevated expression of serum exosomal microRNA-191, -21, -451a of pancreatic neoplasm is considered to be efficient diagnostic marker. BMC Cancer 2018; 18:116.
- [25] Takahasi K, Iinuma H, Wada K, et al. Usefulness of exosomeencapsulated microRNA-451a as a minimally invasive biomarker for prediction of recurrence and prognosis in pancreatic ductal adenocarcinoma. J Hepatobiliary Pancreat Sci 2018;25:155–61.
- [26] Tierney JF, Stewart LA, Ghersi D, et al. Practical methods for incorporating summary time-to-event data into meta-analysis. Trials 2007;8:16.
- [27] Parmar MK, Torri V, Stewart L. Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. Stat Med 1998;17:2815–34.
- [28] Almeida AL, Bernardes MV, Feitosa MR, et al. Serological under expression of microRNA-21, microRNA-34a and microRNA-126 in colorectal cancer. Acta Cir Bras 2016;31(suppl 1):13–8.

- [29] Montagnana M, Benati M, Danese E, et al. Plasma expression levels of circulating miR-21 are not useful for diagnosing and monitoring colorectal cancer. Clin Lab 2016;62:967–70.
- [30] Guo R, Gu J, Zhang Z, et al. MiR-451 promotes cell proliferation and metastasis in pancreatic cancer through targeting CAB39. Biomed Res Int 2017;2017:2381482.
- [31] Bandres E, Bitarte N, Arias F, et al. microRNA-451 regulates macrophage migration inhibitory factor production and proliferation of gastrointestinal cancer cells. Clin Cancer Res 2009;15: 2281–90.
- [32] Shi J. Considering exosomal miR-21 as a biomarker for cancer. J Clin Med 2016;5:pii:E42.
- [33] Zhou X, Wang X, Huang Z, et al. Prognostic value of miR-21 in various cancers: an updating meta-analysis. PLoS One 2014;9:e102413.

- [34] Inamura K, Ishikawa Y. MicroRNA in lung cancer: novel biomarkers and potential tools for treatment. J Clin Med 2016;5:pii:E36.
- [35] Jiang J, Yang P, Guo Z, et al. Overexpression of microRNA-21 strengthens stem cell-like characteristics in a hepatocellular carcinoma cell line. World J Surg Oncol 2016;14:278.
- [36] Kim Y, Roh S, Lawler S, et al. miR451 and AMPK mutual antagonism in glioma cell migration and proliferation: a mathematical model. PLoS One 2011;6:e28293.
- [37] Wang R, Wang ZX, Yang JS, et al. MicroRNA-451 functions as a tumor suppressor in human non-small cell lung cancer by targeting ras-related protein 14 (RAB14). Oncogene 2011;30:2644–58.
- [38] Su Z, Ni L, Yu W, et al. MicroRNA-451a is associated with cell proliferation, migration and apoptosis in renal cell carcinoma. Mol Med Rep 2015;11:2248–54.