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Original article

Biochemical Profiling and Elucidation of Biological Activities of *Beta vulgaris* L. Leaves and Roots Extracts

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ABSTRACT

Research background: Red beet (*Beta vulgaris* L.) is commercially grown in Punjab and Khyber Pakhtunkhwa (KPK) regions while cultivated as vegetable in Baluchistan and Sindh regions of Pakistan. It is well known for its health-promoting role in several clinical and pathologic outcome due to abundance of betalains and other bioactive compounds. The purpose of study was to interpret bioactivity of *B. vulgaris* leaves (BvLM) and roots (BvRM) extracts for finding natural cure of neurodegenerative diseases (NDs).

Experimental approach: BvLM and BvRM extracts were evaluated for phytochemical composition, antioxidant, anti-inflammatory and anticholinesterase potential using standard protocols with modifications.

Results: Phytochemicals analysis of BvLM and BvRM extracts depicted the presence of flavonoids, phenols, alkaloids, saponins and glycosides. The IC₅₀ values for free radical scavenging activity for BvRM and BvLM showed that BvLM (DPPH: 2.20 ± 0.72 µg/mL, H₂O₂: 0.0519 ± 1.02 µg/mL) is more significant as compared to BvRM (DPPH: 2.312 ± 0.72 µg/mL, H₂O₂: 2.668 ± 0.49 µg/mL). BvLM showed significant protection against heat induced hemolysis of HRBCs and protein denaturation (2.322 ± 0.1 > 2.324 ± 0.06 µg/mL) as compared to BvRM (8.572 ± 0.2 > 50.18 ± 1.0 µg/mL). Both extracts found to exhibit strong inhibitory potential against acetylcholinesterase enzyme.

Discussion/Conclusion: Our study highlighted *B. vulgaris* extracts as rich and nutritious source of antioxidants, anti-inflammatory and cholinesterase inhibitors that could be helpful in preventing and treating neurodegenerative disorders (NDs). In consideration of multifactorial and complex etiology of NDs, BvLM and BvRM extracts would be apt candidates for development of therapeutic strategy for management of multiple cognitive disorders.

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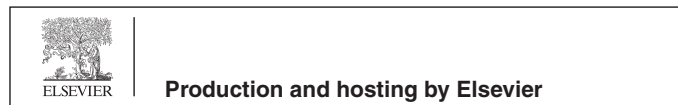
1. Introduction

Exploring enhanced disease ameliorating therapeutic strategies against cognitive disorders like Alzheimer's disease (AD) is a

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leading challenge of recent era. Cognitive impairment and neuropathological etiology is in consistent with cholinergic deficiency due to presence of neurofibrillary tangles and senile plaques besides the degeneration of cholinergic and cortical neurons in basal forebrain (Davies and Maloney, 1976; Varadarajan et al., 2000). World Health Organization (WHO) estimated approximately 71% of the dementia cases occur in developing countries (Ferri et al., 2005; Goverdhan et al., 2012; Mathew and Subramanian, 2014). According to an estimate, by 2050, prevalence of dementia is expected to rise around 46.8 million people (Association, 2015). Therefore, cognitive disorders have become primary research areas due to their huge economic, social and healthcare impacts upon society. Financial burden along with

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social stigma linked with the loss of cognitive capabilities and lead to psychological sufferings in patients (Borsje et al., 2016). Loss of acetylcholine (ACh) neurotransmitter is critically important in causing dysfunction of cholinergic neurotransmission in aging or age-related disorders, responsible for cognitive deficits (Vinutha et al., 2007). Acetylcholinesterase (AChE) is primarily responsible for ACh breakdown within cerebral cortex synapses (Bierer et al., 1995). Complex and multifaceted pathophysiology in neurodegeneration includes inflammation (Heneka et al., 2015), accretion of amyloid peptide (A β), highly phosphorylated tau protein, oxidative stress, mitochondrial dysfunction (Federico et al., 2012), neuronal apoptosis and cholinergic signaling deficit (Mufson et al., 2008). Strategies to improve cholinergic function include the cholinergic receptors activation or providing long term ACh availability to neuronal synaptic cleft via suppressing ACh hydrolysis by AChE (Palop and Mucke, 2016). Release of oxidative and inflammatory stress and ACh stimulation via inhibition of AChE seem imperative potential targets for neurodegeneration therapeutics. Commercially available AChE inhibitors for treatment of mild to moderate neurodegenerative disorders (NDs) include donepezil, tacrine, galanthamine, huperzine A, physostigmine (eserine) and rivastigmine. Synthetic drugs exhibit limited efficacy and significant side effects including gastrointestinal disturbances, hepatotoxicity, low bioavailability, short term biological activity (Chopra et al., 2011; Lee et al., 2011). These drugs also reported to temper only cholinergic symptoms, not to design for targeting specific mechanistic pathways involved in NDs pathogenesis (Lovell and Markesbery, 2007; Nunomura et al., 2001). To counteract these side effects, natural therapeutic options are considered for enhancing NDs treatment efficiency (Li et al., 2014). Natural antioxidants and anti-inflammatory bioactive components possessed by medicinal plants extracts are reported to prevent cell damage and exhibit neuroprotective effects for the management of NDs (Aguilar and Borowski, 2013; Chen et al., 2016). Animal studies for improving cognitive functions using curcumin, a natural compound, indicated the recovery in memory, improved cholinergic function and reversal of stress-induced loss in neurogenesis (Poulose et al., 2017). Another study indicated the anti-cholinesterase activity of saffron ultimately modulating the cholinergic functions (Bukhari et al., 2018).

Beta vulgaris commonly known as beetroot is an herbaceous biennial crop belonging to Chenopodiaceae family. *B. vulgaris* contains saponins, carotenoids, glycine betaine, folates, betanin, betacyanines, polyphenols and flavonoids (Lechner et al., 2010; Socaciu, 2007). *B. vulgaris* poses a variety of health benefits and aids in combating various disorders and health conditions (Clifford et al., 2015).

Current research study is an attempt to extract mixture of bioactive components from *B. vulgaris* leaves and roots and to compare their antioxidant, anti-inflammatory and anti-acetylcholinesterase activities in order to persuade their relevance for therapy of cognitive disorders. This study is to find a promising and natural treatment for neurodegenerative disorders and to what extent the plant shows inhibition activity of certain enzymes most dominantly AChE. To detect if the plant poses any anti-inflammatory and antioxidant activities and whether it is acceptable to pursue and conduct the research further in *in vivo* in order to find a better put treatment of neurodegenerative disorders.

2. Materials and methods

2.1. Ethics statement

A part of current research study involved the utilization of human blood samples. This study was approved by ethical review

board of Department of Biosciences, COMSATS University Islamabad (CUI), Islamabad, Pakistan. Moreover, blood samples from healthy volunteers who fulfilled the selection criteria were collected after a signed informed consent.

2.2. Chemicals and reagents

Chemicals such as NaCl, chloroform, concentrated sulfuric acid, ferric chloride, lead acetate etc were collected from the general laboratory of the campus. 25 g of AlbuMAX™ Lipid-Rich BSA Cat#11020021(Gibco™ United States) and 5,5'-Dithiobis(2-nitro benzoic acid) (DTNB; Ellman's Reagent) Cat# 22582 (Gibco™ United States) were purchased from Thermo Fisher Scientific. Acetylthiocholine chloride (ATCCL) Cat # A5626 (Sigma Aldrich, St. Louis, United States), Gelatin solution Cat # G1393 (Sigma Aldrich, United States), Acetylcholinesterase (AChE) from Electric eel Cat # C3389 (Sigma Aldrich, United States) were purchased from Sigma.

2.3. Study area, plant material collection and identification

Fresh Red beets (*B. vulgaris*) plant species were collected from a local vendor on Park Road Near cultivation point and transported to laboratory (Fig. 1a). The plant was identified by a taxonomist and a voucher specimen was deposited. The stems joining the roots and leaves were detached and discarded. The leaves and roots were washed with fresh water. The roots were peeled and sliced into fine and thin pieces. Both the leaves and thinly sliced roots were shade dried at room temperature until all the water content had evaporated and the sample had become fit for grinding. Once the sample had completely dried, both the roots and leaves were separately grinded into powder using a mechanical blender and placed in containers for later use.

2.4. Preparation of extracts

Soxhlet's extraction method was used for the preparation of crude leaves and roots extracts. The procedure described by De Castro and Priego-Capote (2010) and RNS Yadav (Yadav and Agarwala, 2011) was followed with slight modifications. The leaf and root powder was collected in the apparatus, separately and was treated with extraction solvents i.e. n-hexane and hydro-methanol solution. Both the solvents were heated at 67 °C, until the siphon tube of the Soxhlet's apparatus showed a colorless extract. The n-hexane and hydromethanol solution treated leaves and roots extracts were dried using rotary dryer and fan drying method respectively (Fig. 1b & c).

2.5. Percentage yield of the extracts

Percentage yield of extracts was calculated using the following formula:

$$\text{percent yield} = \left(\frac{\text{actual yield}}{\text{theoretical yield}} \right) \times 100\%$$

2.6. Qualitative phytochemical screening

Following biochemical tests were performed for the detection of bioactive compounds.

2.6.1. Tests for flavonoids

B. vulgaris hydromethanolic extracts of roots (BvRM) and leaves (BvLM) were treated with few drops of diluted sodium hydroxide. Indication of intense yellow color after the addition of few drops of

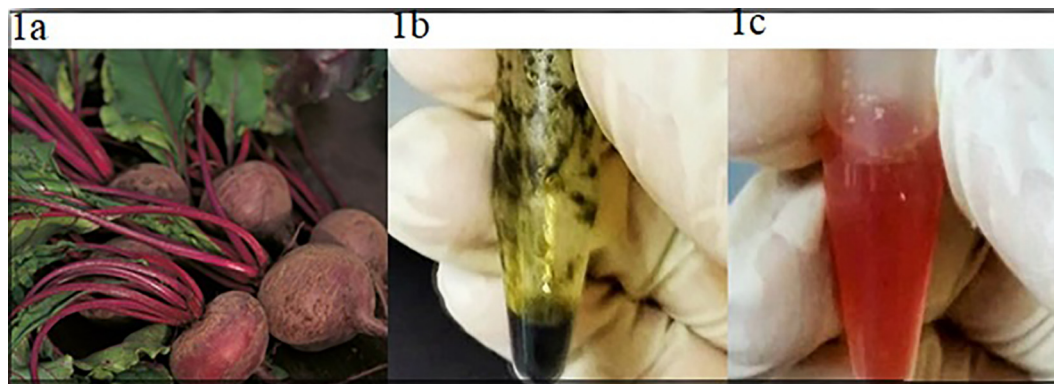


Fig. 1. Snapshot of beta vulgaris plant along (1a) with its hydromethanolic leaves (BvLM) (1b) and roots (BvRM) (1c) extracts.

dilute acid, lead to the presence of flavonoids (Shah and Hossain, 2014).

2.6.2. Tests for phenols

Ferric Chloride Test. The BvRM and BvLM extracts were dissolved and boiled separately in distilled water and filtered. Neutral ferric chloride (5% FeCl_3) solution was added to the mixture and observation of dark green coloration indicated the presence of phenolic compounds (Usman et al., 2009).

2.6.3. Lead acetate test

A bulky white precipitation was observed which ensured the presence of phenols, when BvRM and BvLM extracts were dissolved separately in distilled water to which 10% lead acetate solution (3 mL) was added (Singh and Bag, 2013).

2.6.4. Tests for alkaloids

1% HCl (2 mL) was added to BvRM and BvLM extracts separately. The mixture was heated gently followed by the addition of Mayer's and Wagner's reagents. Precipitate's turbidity indicated the presence of alkaloids (Shrestha et al., 2015).

2.6.5. Test for saponins

BvRM and BvLM extracts (1 mL) were added to 1 mL of deionized water, separately, the mixture was shaken for 30 s vigorously. The tubes were made to stand for 15 min to indicate the presence and absence of persistent frothing. If the frothing was persistent it indicated the presence of saponins (Shrestha et al., 2015).

2.6.6. Test for glycosides

Salkowski's Test. Chloroform (2 mL) was added to BvRM and BvLM extracts (1 mL). Concentrated H_2SO_4 (2 mL) was added carefully and was mixed gently. Indication of reddish brown color in chloroform lower layer lead to the presence of glycosides (Yadav and Agarwala, 2011).

2.7. Quantitative phytochemical analysis

2.7.1. Total phenolic content

Folin-Ciocalteu (FC) Reagent method was used for the determination of polyphenols in the aqueous BvRM and BvLM extracts. In 1 mL of roots and leaves extract separately, FC-reagent (0.5 mL) and 20% (w/v) solution of Na_2CO_3 was added. Mixture was incubated at room temperature for 15 min and absorbance was recorded at 650 nm along with standard (gallic acid) (Baba and Malik, 2015).

2.8. Determination of antioxidant activities of BvRM and BvLM extracts

2.8.1. DPPH (2, 2-Diphenyl 1-2 picrylhydrazyl) assay

BvRM and BvLM extracts were analyzed for their antioxidant properties. For this purpose, DPPH free radical scavenging assay was performed. Multiple concentrations (20–250 $\mu\text{g}/\text{mL}$) of BvRM and BvLM extracts were used. Briefly, reaction mixture contained 1 mL (0.3 mM) of DPPH (D9132, Sigma Aldrich, United States), 1 mL extract sample and 1 mL methanol. Sample mixture was allowed to react at room temperature and then incubated at 37 °C for 45 min in dark. Absorbance was noted at 517 nm using blank, negative control and gallic acid as positive control (Dhanasekaran, et al., 2015). The percent radical scavenging efficacy was determined using formula:

$$\% \text{Radical scavenging activity} = \frac{\text{Abs Control} - \text{Abs Sample}}{\text{Abs Control}} \times 100$$

The experiments were performed in triplicates. The half maximal inhibitory concentration (IC_{50}) values were calculated using Graphpad Prism Version 7.04.

2.8.2. Hydrogen peroxide radical (H_2O_2) scavenging assay

BvRM and BvLM extracts were evaluated for their ability to scavenge hydrogen peroxide by a method described by Ruch et al. (1989). Briefly, H_2O_2 (2 mM) solution was prepared in phosphate buffered saline (PBS, 50 mM; pH 7.4). In 0.3 mL of PBS, 0.1 mL of test sample was added followed by addition of H_2O_2 (0.6 mL) and vortexed. BvRM and BvLM in multiple concentrations (20–250 $\mu\text{g}/\text{mL}$) were used to analyze IC_{50} value. The absorbance of each concentration of tested sample along with blank and standard (gallic acid) was measured at 230 nm. Following equation was used to determine and calculate the hydrogen peroxide scavenging ability of the extracts:

$$\% \text{ Scavenged } [\text{H}_2\text{O}_2] = \frac{\text{Absorbance Control} - \text{Absorbance Sample}}{\text{Absorbance Control}} \times 100$$

2.9. Anti-inflammatory potential of extracts

2.9.1. Inhibition of protein denaturation method

BvRM and BvLM extracts were prepared in multiple serial dilutions (25–200 $\mu\text{g}/\text{mL}$). Aspirin was used as a standard drug and prepared in the concentrations from 1 to 500 $\mu\text{g}/\text{mL}$ in methanol. 1 g of Bovine Serum Albumin (BSA) was dissolved in PBS to obtain 1% of solution of BSA. The pH of the solution was adjusted to 6.3 by adding few drops of 1 N HCl. 1 mL of the extracts of different

concentrations were taken in the test tubes and 1% BSA (1 mL) was added to each tube. Similarly 1 mL of aspirin of multiple concentrations (1–500 µg/mL) was taken in different tubes and 1% BSA (1 mL) was added to them. The samples were heated in water bath at 37 °C for 20 min and then at 57 °C for 20 min. After heating the samples were allowed to cool at room temperature. The absorbance of the samples was measured at 660 nm using UV-spectrophotometer. The entire experiment was performed in triplicates and the percentage inhibition was calculated:

$$\% \text{inhibition} = \frac{\text{Absorbance of Control} - \text{Absorbance of Test Sample}}{\text{Absorbance of Control}} \times 100$$

2.9.2. Human red blood cells stabilization (HRBC) test

HRBC test was performed in two steps. The first step involved preparation of erythrocyte suspension. The suspension was prepared by collecting fresh human blood (10 mL) from a healthy volunteer who had not taken any NSAIDs for 2 weeks prior to the experiment. The blood was collected in heparinized centrifuge tubes and was centrifuged for 10 min at 3000 rpm. After centrifugation the sample was washed with equal volume of normal saline solution 2 times and centrifuged for 3 min at 3000 rpm until a clear supernatant was obtained. The supernatant was discarded and the volume of blood was measured and made to 10% (v/v) with normal saline solution. The second step of this test involved heat induced hemolysis where reaction mixture of 2 mL was prepared comprising of 1 mL of BvRM and BvLM extracts concentration ranging from (20–200 µg/mL) and 1 mL of 10% RBC suspension. Saline solution was used instead of the extracts in the case of control. Aspirin was used as a positive control. The samples were incubated for 30 min at 56 °C then centrifuged for 5 min at 2500 rpm. The absorbance of supernatant was measured at 560 nm. The test was performed in triplicates. Percentage of hemolysis of human red blood cell membrane was calculated:

$$\% \text{ hemolysis} = \left(\frac{\text{Absorbance of Test Sample}}{\text{Absorbance of Control}} \right) \times 100$$

Percentage of membrane stabilization was also calculated:

$$\% \text{ protection} = \left(100 - \left[\frac{\text{Absorbance of Test Sample}}{\text{Absorbance of Control}} \right] \right) \times 100$$

2.10. Acetylcholinesterase inhibition assay

The method described by Md. Abul Hasnat et al. (2013) was used to determine the inhibitory activity of BvRM and BvLM extracts of acetyl-cholinesterase. 20 µL of 0.2 units/mL of enzyme solution, 120 µL of 0.1 M sodium phosphate buffer and 20 µL of BvRM and BvLM extracts were mixed and incubated in microplate for 15 min at 37 °C. After 15 min the substrate (ATCCL) and DTNB were added to each well in the volume of 20 µL. Formation of colored product 5-thio-2-nitrobenzoate anion indicated the hydrolysis of ATCCL. The anion was formed because of the reaction of DTNB and thiochloine which is released because of enzyme hydrolysis. After the formation of colored product, the absorbance was measured at 410 nm after 10 min. Donepezil was used as a positive control. The experiment was performed in duplicates. AChE percentage inhibition was calculated using the equation:

$$\% \text{ Inhibition} = \left(1 - \frac{\text{Absorbance of sample}}{\text{Absorbance of control}} \right) \times 100$$

Multiple serial dilutions of BvRM, BvLM and donepezil (400 µg/mL to 12.5 µg/mL) were prepared for calculation of 50% inhibitory concentration (IC₅₀).

2.11. Statistical analysis

The statistical analysis of data was expressed in mean standard deviation of triplicate readings. IC₅₀ values were calculated using nonlinear regression in GraphPad Prism (Prism, 1994). *P* values were calculated by performing Kruskal-Wallis test (*P* < 0.05) in order to find the significance of difference between means of different extracts. GraphPad Prism 7 was used for performing statistical analysis. Microsoft Excel 2007 was also used for graphical evaluations.

3. Results

3.1. Yield calculation for extracts

The percentage yield of BvRM, BvLM, and BvLH extracts were calculated. Out of three extracts, BvRM showed the highest percentage yield (Table 1).

3.2. Estimation of qualitative analysis of phytochemicals present in *B. vulgaris* extracts

Phytochemical screening tests of BvLM and BvRM extracts depicted the presence of flavonoids, phenols, saponins, alkaloids and glycosides (Table 2).

3.3. Quantitative phytochemical analysis of extracts

3.3.1. Total phenolic content determination

To evaluate the presence of total phenolic content in the BvRM and BvLM extracts, Folin-Ciocalteu Reagent method was performed. Gallic acid being phenolic in nature was used as a standard. BvLM showed a higher value of 0.143 ± 0.00 as compared to BvRM (0.041 ± 0.003), indicating that leaves have a higher phenolic content as compared to the roots of *B. vulgaris* (Fig. 2).

3.4. Antioxidant activities of BvRM and BvLM extracts









3.4.1. DPPH (2, 2-Diphenyl 1-2 picrylhydrazyl) assay

Fig. 3a has shown the radical scavenging potential of different concentrations of *B. vulgaris* extracts. BvRM extract at 25 µg/mL concentration attained the highest scavenging activity of 81.09% as compared to BvLM (73.81%). These results illustrated the competitive antioxidative potency of BvRM and BvLM extracts with

Table 1
Table showing percentage yield of various extracts of *Beta vulgaris*.

Serial #	Extracts	Abbreviations	Percentage yield (%)
01	Roots extract in hydromethanol solvent	BvRM	54.06%
02	Leaves extract in hydromethanol solvent	BvLM	4.89%
03	Leaves extract in n-hexane solvent	BvLH	1.46%

Table 2
Table showing various phytochemicals present in the hydromethanolic extracts of *Beta vulgaris*.

Serial #	Phytochemical tests	Characteristics found	Findings	Results
01	Phenolic compounds in BvLM	Green coloration in FeCl ₃ test	+++	
02	Phenolic compounds in BvRM	Light green coloration in FeCl ₃ test	++	
03	Flavonoids in BvLM	Formation of intense yellow color which became colorless after the addition of few drops of dilute acid	+	
04	Flavonoids in BvRM	Formation of intense yellow color which became colorless after the addition of few drops of dilute acid	+++	
05	Saponins in BvLM	Mild frothing	+	
06	Saponins in BvRM	Mild frothing	+	
07	Alkaloids in BvLM	Precipitate's turbidity	++	
08	Alkaloids in BvRM	Precipitate's turbidity	++	
09	Glycosides in BvLM	Reddish brown color indicated the presence of glycosides in the lower layer of chloroform	+++	
10	Glycosides in BvRM	Reddish brown color indicated the presence of glycosides in the lower layer of chloroform	++	

+: present ++: moderately present +++: highly present.

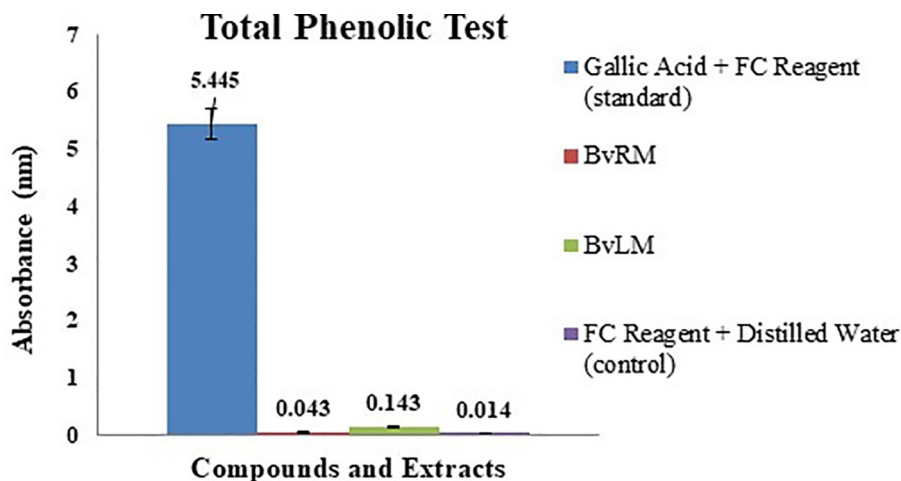


Fig. 2. Total phenolic content of BvRM and BvLM extract in terms of absorption measured along with the standard Gallic Acid.

purified standard. For the interpretation of results, 50% inhibitory concentration (IC₅₀) for each extract was calculated. The results showed that BvRM exhibit IC₅₀ value of 2.32 ± 0.72 µg/mL as compared to BvLM (2.20 ± 0.72 µg/mL) (Fig. 3b). The reason why BvLM has lower IC₅₀ value may be that it has more phenolic content as compared to the BvRM extract. Lower IC₅₀ with higher scavenging activity depicted the significant therapeutic potential of bioactive constituents found in extracts. However, we can say that both the extracts can possibly act as primary antioxidants. The

scavenging activity of these extract was found to be comparable to gallic acid (IC₅₀: 2.071 ± 0.72 µg/mL) (Fig. 3b).

3.4.2. Hydrogen peroxide radical (H₂O₂) scavenging assay

Hydrogen peroxide Radical (H₂O₂) Scavenging Assay was performed to evaluate BvRM and BvLM extracts for their antioxidant activity. Both extracts showed significant radical scavenging activity when compared to the gallic acid. Comparatively, BvLM showed higher scavenging activity than BvRM. The scavenging activity of

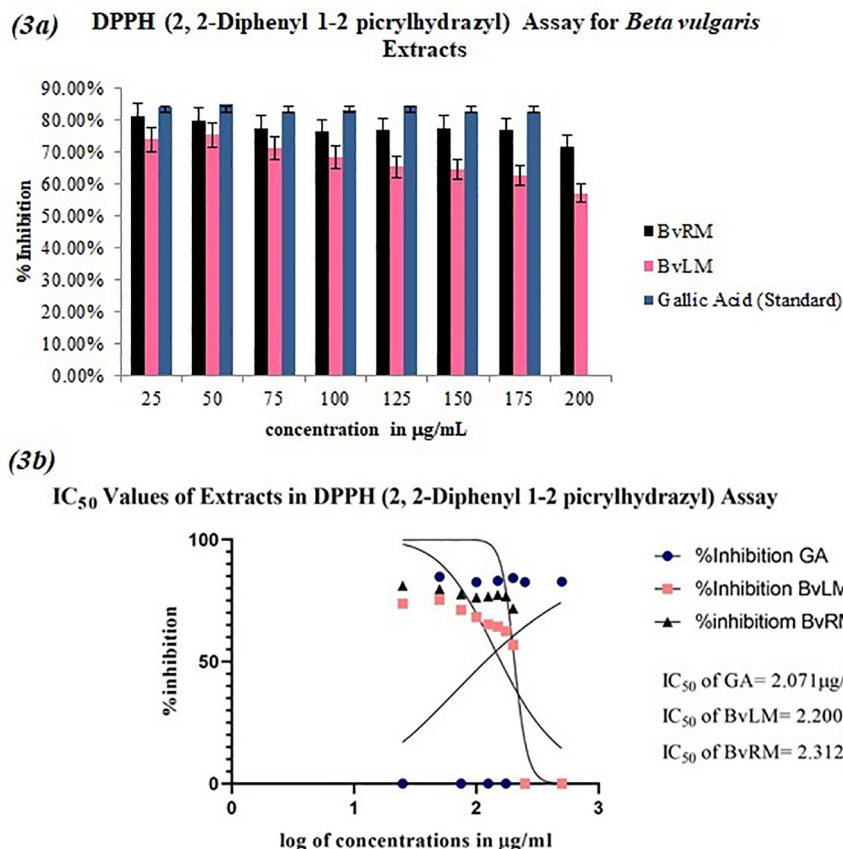


Fig. 3. (3a) A bar plot showing DPPH radical scavenging activity of the different extracts of *Beta vulgaris* in comparison to Gallic acid as standard along with their respective error amount as a percentage (3b) IC₅₀ values of the selected plant extracts for DPPH radical scavenging activity along with the standard Gallic acid.

BvLM (100 µg/mL) with percentage inhibition being found 47.60% (Fig. 4a) on the other hand BvRM showed 44.69% scavenging activity at 100 µg/mL as compared to standard Gallic acid (100 µg/mL) illustrating 51% scavenging activity.

Depicted IC₅₀ values of BvLM (0.0519 ± 1.02 µg/mL) and BvRM (2.668 ± 0.49 µg/mL) illustrated BvLM as stronger radical scavenger than BvRM. BvLM (IC₅₀: 0.0519 ± 1.02 µg/mL) was found to be more significant than the standard Gallic acid (IC₅₀: 1.367 ± 0.1 µg/mL) (Fig. 4b). The scavenging activity for H₂O₂ of plant extracts is given in the following order BvLM > BvRM (0.0519 ± 1.02 > 2.668 ± 0.49 µg/mL).

3.5. Estimation of Anti-inflammatory activity of BvRM and BvLM extracts

3.5.1. Estimation of inhibition of protein denaturation

The BvRM and BvLM extracts were analyzed for their inhibition of protein denaturation using Bovine Serum Albumin (BSA) as a protein of interest. Both extracts showed significant inhibition of protein denaturation. BvLM showed 96.45% inhibitory activity at concentration of 175 µg/mL. Both extracts displayed anti-inflammatory activity as compared to standard drug, aspirin (Fig. 5a) possibly proving the higher protein denaturation inhibitory activity. BvRM depicted relatively constant inhibition potential (~80%) at multiple dilution concentrations ranging from 25 to 200 µg/mL while BvLM showed the same inhibitory pattern with the deviation of increased protein protective potency at 150 µg/mL and 175 µg/mL concentrations. Both extracts exhibited the same but higher inhibitory potency of protein denaturation as compared to standard drug. The *P* values of both the standard and extracts were significant (*P* = 0.0063).

The IC₅₀ values of BvRM and BvLM were almost similar (2.324 ± 0.06 µg/mL and 2.322 ± 0.1 µg/mL, respectively) with BvLM showing a slightly higher IC₅₀ value than BvLM (Fig. 5b).

3.5.2. Estimation of human red blood cells (HRBCs) membrane stabilization test

The BvRM and BvLM extracts were investigated for their anti-inflammatory activity using HRBC stabilization test. Aspirin, an anti-inflammatory drug was used as standard. BvLM extract at 50 µg/mL concentration showed the highest percentage of protection (99.58%) from hemolysis (Fig. 6a). In heat induced hemolysis assay, the BvLM extract showed the minimum percentage of hemolysis at 100 µg/mL with percentage being 0.4% (Fig. 6b). Statistical analysis showed that the values were significant (*P* = 0.0185). BvRM and BvLM showed IC₅₀ values of 50.18 ± 1.0 µg/mL and 8.572 ± 0.2 µg/mL, respectively while aspirin showed IC₅₀ value of 0.341 ± 0.47 µg/mL indicating that percentage protection of hemolysis by selected plant extracts is significantly higher than standard drug (Fig. 6c).

3.6. Estimation of acetylcholinesterase inhibition activity

The inhibitors of AChE are being used widely to find targeted therapy for neurodegenerative diseases. The BvRM and BvLM extracts were also investigated for their AChE inhibition activity. Both extracts depicted significant inhibition activity along with the standard drug, donepezil. BvRM and BvLM depicted 89.8% and 93.3% inhibition of AChE in comparison to 94.2% inhibition by donepezil at 100 µg/mL (Fig. 7). Dose response analysis illustrated the low IC₅₀ values for both extracts along with donepezil depicting their high efficacy for AChE inhibition (Fig. 8).

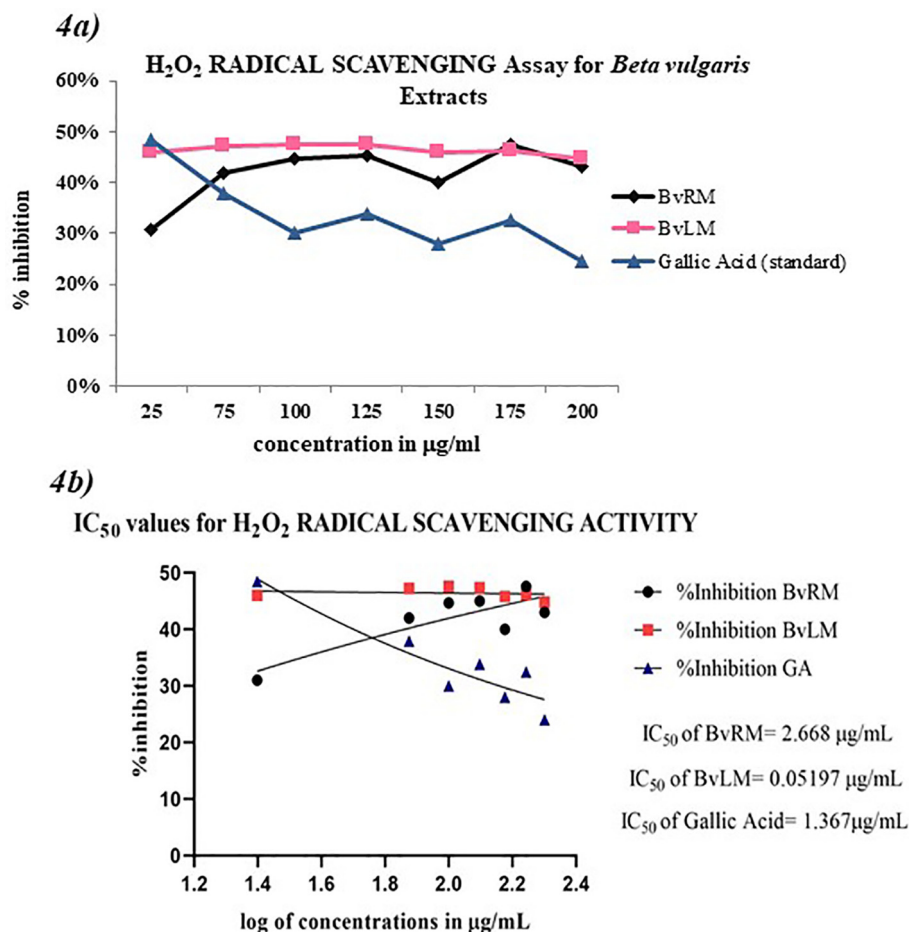


Fig. 4. (4a) A linear plot showing H₂O₂ radical scavenging activity of the different extracts of *Beta vulgaris* in comparison to Gallic acid as standard (4b) IC₅₀ values for H₂O₂ radical scavenging assay shown by different extracts of *Beta vulgaris* along with the standard Gallic acid.

4. Discussion

From botanical and health perspective, the importance of *B. vulgaris* is highly appreciated. However, proving its importance in the treatment of neurodegenerative diseases is under consideration. It is now known that beetroots do have the ability to cure dementia – a symptom shown by cognitive disorders, by providing oxygenation to the somatomotor cortex of the brain. In the current study, elaborative phytochemical investigation and pharmacognostic potential of *B. vulgaris* extracts is reported. When subjected to *in vitro* testing, BvRM and BvLM extracts showed a plethora of characteristics. Presence of multiple secondary metabolites like saponins, flavonoids, alkaloids, glycosides and phenolic compounds in preliminary phytochemical screening assays depicted extracts as rich nutritious source. Interestingly, quantitative analysis indicated the presence of phenolic compounds specifically in leaves extract (BvLM) which directly correlate with multiple bioactivities of extract. The phenolic compounds of BvLM extracts contain potential bioactive secondary metabolites (Edziri et al., 2019). However if not a lot, in comparison to the leaves, the roots also contain secondary metabolites formed by the phenolic compounds.

Free radicals are produced during various metabolic processes in body and are associated with diverse range of disorders like diabetes, neurodegenerative diseases, cancer, coronary heart disease, immunosuppression and many others (Kumar and Pandey, 2013; Kumar and Pandey, 2015). Singlet oxygen and hydrogen peroxide are considered in the category of non-free radicals whilst hydroxyl, superoxide, lipid peroxy and nitric oxide are general free radicals

(Halliwell and Gutteridge, 2015). Generally, natural body defense mechanisms like chain breaking antioxidants generation and protective antioxidant systems neutralize the destructive effect of free radicals (Jacob, 1995). Moreover devastating tissue injury occurs when rate of free radical generation exceeds the limit of natural antioxidant mechanisms. Therefore, various diseases like neurodegenerative disorders can be treated with drugs/agents (natural or synthetic) having radical scavenging potential. Natural flora is a valuable source of antioxidants which have been reported to provide protection from radicals induced damage (Kumar et al., 2013). The significant anti-radical activities of BvLM and BvRM extracts were observed using DPPH (with IC₅₀ values of 2.20 ± 0.72 and 2.32 ± 0.72 µg/mL respectively) and H₂O₂ radical scavenging assays (with IC₅₀ values of 0.0519 ± 1.02 and 2.668 ± 0.49 µg/mL respectively). The antioxidant ability of BvLM and BvRM may be correlated with the presence of phenols especially BvLM since it showed a better radical scavenging ability as compared to BvRM due to the presence of phenols in a greater amount. These results are in concordance with the study done by Irshad et al. (2012).

Inflammation is considered as a beneficial process under normal conditions. Inflammation being an immune response can also have deteriorating impact on its host, at times. The traditional treatment for inflammation involves the use of non-steroidal and anti-inflammatory drugs commonly known as NSAIDs (Non-Steroidal Anti-Inflammatory Drugs). These drugs do pose severe side effects in the case of chronic doses (Varga et al., 2017). The BvRM and BvLM extracts were evaluated for their

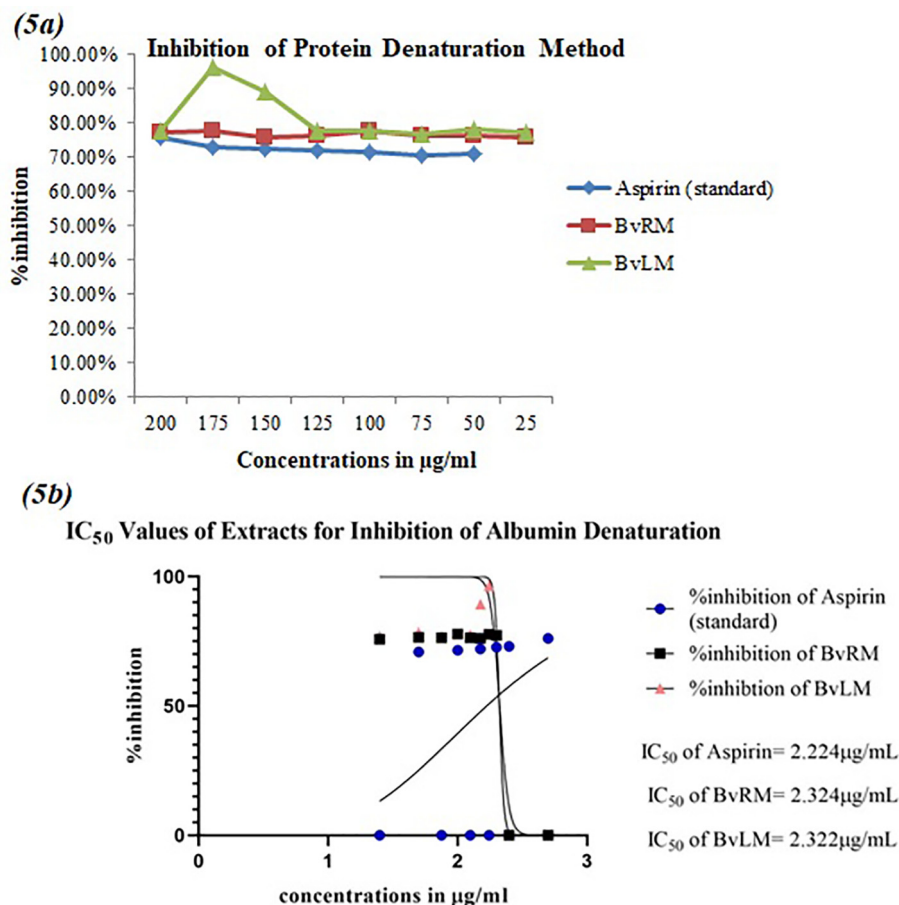


Fig. 5. (5a) A linear plot showing inhibition of protein denaturation by different extracts of *Beta vulgaris* in comparison to aspirin as a standard drug (5b) IC₅₀ values for log of concentrations of *Beta vulgaris* extracts for the inhibition of albumin denaturation along with aspirin as a standard drug.

anti-inflammatory action potential by conducting protein denaturation and HRBCs stabilization assays. Both extracts, BvLM and BvRM had the ability to inhibit protein/albumin denaturation when compared with the standard anti-inflammatory drug aspirin. Results of anti-inflammatory assays illustrated that *B. vulgaris*, commonly used as functional food, can highly be utilized for the cure of inflammation and related symptoms instead of synthetic conventional drugs which poses severe side effects. They also have the ability to boost immunity because of their anti-inflammatory properties and their richness in Vitamin C (Schlueter and Johnston, 2011). BvLM and BvRM extracts inhibited protein denaturation significantly exhibiting IC₅₀ values of 2.322 ± 0.1 and 2.324 ± 0.06 µg/mL respectively. Results from current study presented evidence for membrane stabilization and inhibition of heat induced hemolysis by extracts as additive mechanism of their anti-inflammatory potential.

In neurological disorders treatment, strong inhibitors of acetylcholinesterase enzyme (AChE) are of great importance. This enzyme is involved in catalyzing the acetylcholine (ACh), therefore having potential for treatment of dementia (Nawaz and Choudhary, 2004) in addition to many other neuronal disorders (Kandiah et al., 2017). Decrease in ACh concentration via dysfunction of multiple biochemical pathways eventually leads to NDs (Callahan et al., 2017). AChE causes the termination of signal transmission in synapses via ACh. Therefore inhibition of this metabolizing enzyme may provide effective therapy for many other neurological disorders (Xiao et al., 2017). Unfortunately, currently available drugs are significantly associated with severe side effects including hepatotoxicity and are potent to use only in mild condi-

tion of disorder (McEneny-King et al., 2017). Therefore, prime importance of current study is to explore novel remedies for NDs treatment. Various research groups including current study are scrutinizing indigenous flora for finding potential bioactive compounds for treatment of neurological disorders. Present study illustrated the very significant AChE inhibitory activity by *B. vulgaris* extracts. Results demonstrated that percentage inhibition values of BvLM and BvRM are not significantly different from in comparison to donepezil (standard drug) at the same concentration. BvLM and BvRM extracts exhibited IC₅₀ values of 4.99 µg/mL and 3.731 µg/mL in comparison to donepezil (IC₅₀: 3.691 µg/mL) against AChE. Results have shown the significant therapeutic potency of extracts for cholinesterase. Previously *Carpolobia lutea* and many other plants were analyzed for their antioxidant activity and found therapeutically effective against AChE (Nwidu et al., 2017). Although beetroots show promising properties and activities, yet further studies regarding isolation will authenticate its therapeutic potential as neuroprotective agent. In future, our research group will continue study for the exploration of pharmacological activities of purified phytochemicals of *B. vulgaris* using *in vitro* and *in silico* techniques.

5. Conclusion

The current research work is an exclusive study for finding phytochemical composition, radical scavenging, anti-inflammatory and anti-acetylcholinesterase activity of methanolic extracts of *B. vulgaris* roots and leaves part. The results of AChE inhibition and

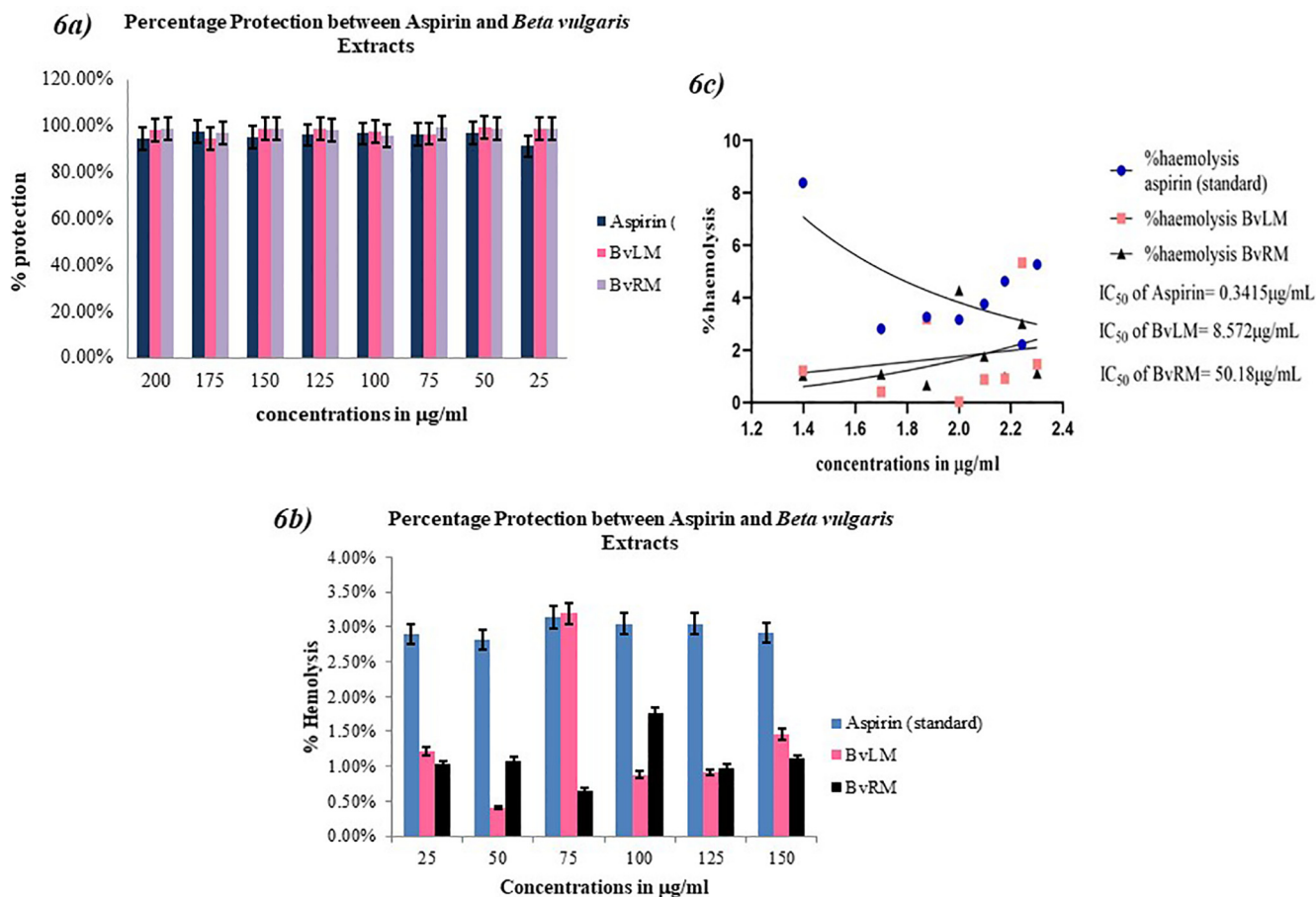


Fig. 6. (6a) A bar plot showing percentage protection of HRBCs by different extracts of *Beta vulgaris* in comparison to aspirin as standard drug along with their respective error amount as a percentage (6b) A bar plot showing percentage inhibition of hemolysis of HRBCs by different extracts of *Beta vulgaris* in comparison to aspirin as standard drug along with their respective error amount as a percentage (6c) IC₅₀ values for log of concentrations of *Beta vulgaris* extracts for the inhibition of hemolysis of HRBCs along with aspirin as a standard drug.

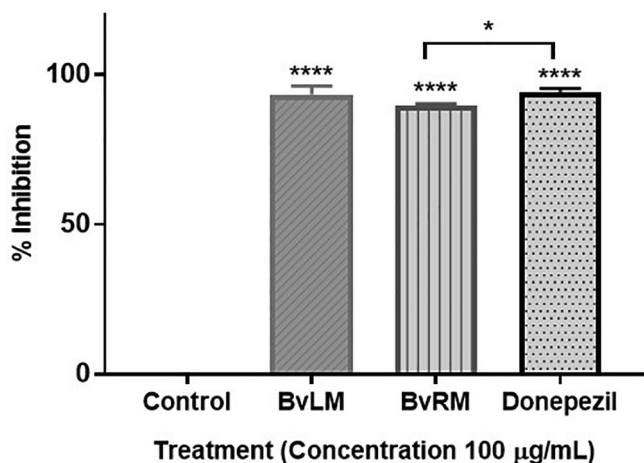


Fig. 7. A bar plot is depicting percentage inhibition of acetylcholinesterase enzyme by BvRM and BvLM extracts in comparison to standard drug, donepezil. Extracts with 100 µg/mL and 200 µg/mL concentrations were used in conjunction with standard drug.

radical scavenging assays were highly significant. Research data indicate that *B. vulgaris* is greatly enriched with antioxidants which can be implemented as potential therapeutic agents for the treatment of various neurological disorders. In the light of above

mentioned pharmacognostic activity of *B. vulgaris* extracts, further studies on bioassay guided separation of bioactive compounds and their mechanism of action is recommended.

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Availability of data and materials

The data presented in this manuscript belong to research work of Shifa Shah and has not been deposited in any repository yet. However, the materials are available to the researchers upon request.

Authors' contributions

SS carried out experimental work, data collection and literature search. SR supervised research work, helped in study design, data evaluation and drafted the final version of the manuscript. AMB and SMS helped in plant collection, provision of laboratory reagents and processing data interpretation. ZJ and AN reviewed the manuscript and statistical work. Authors approved the final manuscript for publication.

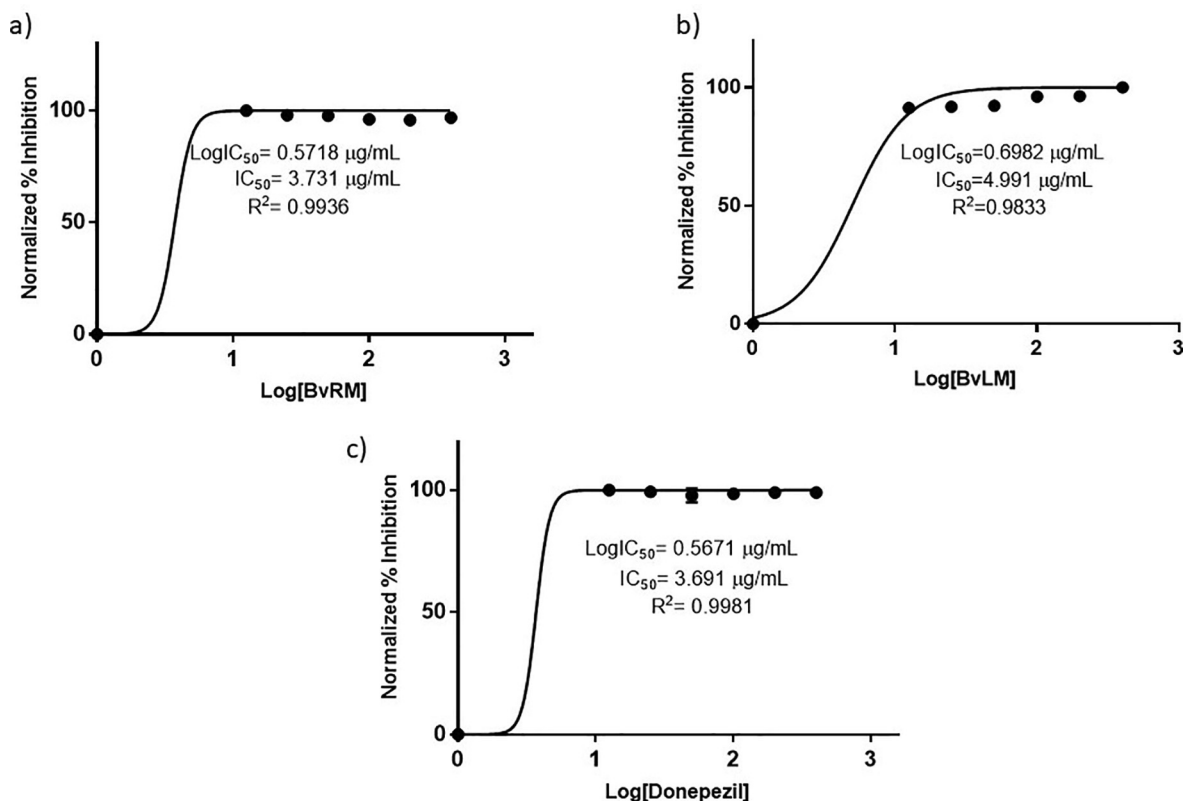


Fig. 8. IC₅₀ values calculation using dose response analysis. (a) Serial dilutions of BvRM extract were used and 3.731 µg/mL dose was found as IC₅₀. (b) Serial dilutions of BvLM extract were used and 4.991 µg/mL dose was counted as IC₅₀. (c) Serial dilutions of standard drug (donepezil) were used and 3.691 µg/mL dose was found as IC₅₀.

Declaration of Competing Interest

The authors declare that they have no competing interest.

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References

- Aguiar, S., Borowski, T., 2013. Neuropharmacological review of the nootropic herb *Bacopa monnieri*. *Rejuvenation Res.* 16, 313–326.
- Association, A.S., 2015. 2015 Alzheimer's disease facts and figures. *Alzheimers Dement* 11, 332–384.
- Baba, S.A., Malik, S.A., 2015. Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *J. Taibah. Univ. Sci.* 9, 449–454.
- Bierer, L.M., Haroutunian, V., Gabriel, S., Knott, P.J., Carlin, L.S., Purohit, D.P., Perl, D. P., Schmeidler, J., Kanof, P., Davis, K.L., 1995. Neurochemical correlates of dementia severity in Alzheimer's disease: relative importance of the cholinergic deficits. *J. Neurochem.* 64, 749–760.
- Borsje, P., Hems, M.A., Lucassen, P.L., Bor, H., Koopmans, R.T., Pot, A.M., 2016. Psychological distress in informal caregivers of patients with dementia in primary care: course and determinants. *Fam. Pract.* 33, 374–381.
- Bukhari, S.I., Manzoor, M., Dhar, M., 2018. A comprehensive review of the pharmacological potential of *Crocus sativus* and its bioactive apocarotenoids. *Biomed. Pharmacother.* 98, 733–745.
- Callahan, P.M., Bertrand, D., Bertrand, S., Plagenhoef, M.R., Terry Jr, A.V., 2017. Tropisetron sensitizes $\alpha 7$ containing nicotinic receptors to low levels of acetylcholine in vitro and improves memory-related task performance in young and aged animals. *Neuropharmacology* 117, 422–433.
- Chen, L.-E., Wu, F., Zhao, A., Ge, H., Zhan, H., 2016. Protection efficacy of the extract of *Ginkgo biloba* against the learning and memory damage of rats under repeated high sustained. *Evid. Based Complement. Alternat. Med.* 2016.

- Chopra, K., Misra, S., Kuhad, A., 2011. Current perspectives on pharmacotherapy of Alzheimer's disease. *Expert Opin. Pharmacother.* 12, 335–350.
- Clifford, T., Howatson, G., West, D.J., Stevenson, E.J., 2015. The potential benefits of red beetroot supplementation in health and disease. *Nutrients* 7, 2801–2822.
- Davies, P., Maloney, A., 1976. Selective loss of central cholinergic neurons in Alzheimer's disease. *The Lancet* 308, 1403.
- De Castro, M.L., Priego-Capote, F., 2010. Soxhlet extraction: past and present panacea. *J. Chromatogr.* 1217, 2383–2389.
- Edziri, H., Jaziri, R., Haddad, O., Anthonissen, R., Aouni, M., Mastouri, M., Verschaeve, L., 2019. Phytochemical analysis, antioxidant, anticoagulant and in vitro toxicity and genotoxicity testing of methanolic and juice extracts of *Beta vulgaris* L. S. *Afr. J. Bot.* 126, 170–175.
- Federico, A., Cardaioli, E., Da Pozzo, P., Formichi, P., Gallus, G.N., Radi, E., 2012. Mitochondria, oxidative stress and neurodegeneration. *J. Neurol. Sci.* 322, 254–262.
- Ferri, C.P., Prince, M., Brayne, C., Brodaty, H., Fratiglioni, L., Ganguli, M., Hall, K., Hasegawa, K., Hendrie, H., Huang, Y., 2005. Global prevalence of dementia: a Delphi consensus study. *The Lancet* 366, 2112–2117.
- Goverdhan, P., Sravanthi, A., Mamatha, T., 2012. Neuroprotective effects of meloxicam and selegiline in scopolamine-induced cognitive impairment and oxidative stress. *Int J Alzheimers Dis* 2012.
- Halliwell, B., Gutteridge, J.M., 2015. *Free radicals in biology and medicine*. Oxford University Press, USA.
- Hasnat, M., Pervin, M., Lim, B.O., 2013. Acetylcholinesterase inhibition and in vitro and in vivo antioxidant activities of *Ganoderma lucidum* grown on germinated brown rice. *Molecules* 18, 6663–6678.
- Heneka, M.T., Carson, M.J., El Khoury, J., Landreth, G.E., Brosseron, F., Feinstein, D.L., Jacobs, A.H., Wyss-Coray, T., Vitorica, J., Ransohoff, R.M., 2015. Neuroinflammation in Alzheimer's disease. *The Lancet Neurology* 14, 388–405.
- Irshad, M., Zafaryab, M., Singh, M., Rizvi, M., 2012. Comparative analysis of the antioxidant activity of *Cassia fistula* extracts. *Int. J. Med. Chem.* 2012.
- Jacob, R.A., 1995. The integrated antioxidant system. *Nutr. Res.* 15, 755–766.
- Kandiah, N., Pai, M.-C., Senanarong, V., Looi, I., Ampil, E., Park, K.W., Karanam, A.K., Christopher, S., 2017. Rivastigmine: the advantages of dual inhibition of acetylcholinesterase and butyrylcholinesterase and its role in subcortical vascular dementia and Parkinson's disease dementia. *Clin. Interv. Aging* 12, 697.
- Kumar, S., Pandey, A.K., 2013. Chemistry and biological activities of flavonoids: an overview. *Sci. World J.*
- Kumar, S., Pandey, A.K., 2015. Free radicals: health implications and their mitigation by herbals. *J. Adv. Med. Med. Res.* 438–457.
- Kumar, S., Mishra, A., Pandey, A.K., 2013. Antioxidant mediated protective effect of *Parthenium hysterophorus* against oxidative damage using in vitro models. *BMC Complement. Altern. Med.* 13, 120.

- Lechner, J.F., Wang, L.-S., Rocha, C.M., Larue, B., Henry, C., McIntyre, C.M., Riedl, K.M., Schwartz, S.J., Stoner, G.D., 2010. Drinking water with red beetroot food color antagonizes esophageal carcinogenesis in N-nitrosomethylbenzylamine-treated rats. *J. Med. Food* 13, 733–739.
- Lee, S.-H., Sancheti, S.A., Bafna, M.R., Sancheti, S.S., Seo, S.-Y., 2011. Acetylcholinesterase inhibitory and antioxidant properties of *Rhododendron yedoense* var. *Poukhanense* bark. *J. Med. Plants Res* 5, 248–254.
- Li, S., Chen, G., Zhang, C., Wu, M., Wu, S., Liu, Q., 2014. Research progress of natural antioxidants in foods for the treatment of diseases. *Food Sci. Hum. Well* 3, 110–116.
- Lovell, M.A., Markesbery, W.R., 2007. Oxidative DNA damage in mild cognitive impairment and late-stage Alzheimer's disease. *Nucleic Acids Res.* 35, 7497–7504.
- Mathew, M., Subramanian, S., 2014. In vitro screening for anti-cholinesterase and antioxidant activity of methanolic extracts of ayurvedic medicinal plants used for cognitive disorders. *PLoS ONE* 9.
- McEneaney-King, A., Osman, W., Edginton, A.N., Rao, P.P., 2017. Cytochrome P450 binding studies of novel tacrine derivatives: predicting the risk of hepatotoxicity. *Bioorg. Med. Chem. Lett.* 27, 2443–2449.
- Mufson, E.J., Counts, S.E., Perez, S.E., Ginsberg, S.D., 2008. Cholinergic system during the progression of Alzheimer's disease: therapeutic implications. *Expert Rev. Neurother.* 8, 1703–1718.
- Nawaz, S.A., Choudhary, M.I., 2004. New cholinesterase inhibiting bisbenzylisoquinoline alkaloids from *Cocculus pendulus*. *Chem. Pharm. Bull. (Tokyo)* 52, 802–806.
- Nunomura, A., Perry, G., Aliev, G., Hirai, K., Takeda, A., Balraj, E.K., Jones, P.K., Ghanbari, H., Wataya, T., Shimohama, S., 2001. Oxidative damage is the earliest event in Alzheimer disease. *J. Neuropathol. Exp. Neurol.* 60, 759–767.
- Nwidu, L.L., Elmorsy, E., Thornton, J., Wijamunige, B., Wijesekara, A., Tarbox, R., Warren, A., Carter, W.G., 2017. Anti-acetylcholinesterase activity and antioxidant properties of extracts and fractions of *Carpolobia lutea*. *Pharm. Biol.* 55, 1875–1883.
- Palop, J.J., Mucke, L., 2016. Network abnormalities and interneuron dysfunction in Alzheimer disease. *Nat. Rev. Neurosci.* 17, 777.
- Poulose, S.M., Miller, M.G., Scott, T., Shukitt-Hale, B., 2017. Nutritional factors affecting adult neurogenesis and cognitive function. *Adv. Nutr.* 8, 804–811.
- Prism, G., 1994. Graphpad software. San Diego, CA, USA.
- Ruch, R.J., Cheng, S.-J., Klaunig, J.E., 1989. Prevention of cytotoxicity and inhibition of intercellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogenesis* 10, 1003–1008.
- Schlueter, A.K., Johnston, C.S., 2011. Vitamin C: overview and update. *J. Evid. Based Complementary Altern. Med.* 16, 49–57.
- Shah, M.D., Hossain, M.A., 2014. Total flavonoids content and biochemical screening of the leaves of tropical endemic medicinal plant *Merremia borneensis*. *Arab. J. Chem.* 7, 1034–1038.
- Shrestha, P., Adhikari, S., Lamichhane, B., Shrestha, B.G., 2015. Phytochemical screening of the medicinal plants of Nepal. *IOSR J. Environ. Sci. Toxicol. Food Technol.* 1, 11–17.
- Singh, K.L., Bag, G., 2013. Phytochemical analysis and determination of total phenolics content in water extracts of three species of *Hedychium*. *Int. J. PharmTech. Res.* 5, 1516–1521.
- Socaciu, C., 2007. Food colorants: chemical and functional properties. CRC Press.
- Usman, H., Abdulrahman, F.I., Usman, A., 2009. Qualitative phytochemical screening and in vitro antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). *Afr. J. Tradit. Complement. Altern. Med.* 6.
- Varadarajan, S., Yatin, S., Aksanova, M., Butterfield, D.A., 2000. Alzheimer's amyloid β -peptide-associated free radical oxidative stress and neurotoxicity. *J. Struct. Biol.* 130, 184–208.
- Varga, Z., rafay ali Sabzwari, S., Vargova, V., 2017. Cardiovascular risk of nonsteroidal anti-inflammatory drugs: an under-recognized public health issue. *Cureus* 9.
- Vinutha, B., Prashanth, D., Salma, K., Sreeja, S., Pratiti, D., Padmaja, R., Radhika, S., Amit, A., Venkateshwarlu, K., Deepak, M., 2007. Screening of selected Indian medicinal plants for acetylcholinesterase inhibitory activity. *J. Ethnopharmacol.* 109, 359–363.
- Xiao, S., Wang, T., Ma, X., Qin, Y., Li, X., Zhao, Z., Liu, X., Wang, X., Xie, H., Jiang, Q., 2017. Efficacy and safety of a novel acetylcholinesterase inhibitor octahydroaminoacridine in mild-to-moderate Alzheimer's disease: a phase II multicenter randomised controlled trial. *Age Ageing* 46, 767–773.
- Yadav, R., Agarwala, M., 2011. Phytochemical analysis of some medicinal plants. *J. Phytol.*