

Research Article

Identification and Validation of QTLs for Macronutrient Contents in Brown and Milled Rice Using Two Backcross Populations between *Oryza sativa* and *O. rufipogon*

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Received 3 February 2021; Revised 14 May 2021; Accepted 31 May 2021; Published 12 June 2021

Academic Editor: Yong Feng

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Mineral malnutrition as a prevalent public health issue can be alleviated by increasing the intake of dietary minerals from major staple crops, such as rice. Identification of the gene responsible for mineral contents in rice would help breed cultivars enriched with minerals through marker-assisted selection. Two segregating populations of backcross inbred lines (BIL) were employed to map quantitative trait loci (QTLs) for macronutrient contents in brown and milled rice, BC₁F₅, and BC₂F_{4,5} derived from an interspecific cross of Xieqingzao B (*Oryza sativa*) and Dongxiang wild rice (*O. rufipogon*). Phenotyping the populations was conducted in multiple locations and years, and up to 169 DNA markers were used for the genotyping. A total of 17 QTLs for P, K, Na, Ca, and Mg contents in brown and milled rice distributed on eight regions were identified in the BC₁F₅ population, which is explained to range from 5.98% to 56.80% of phenotypic variances. Two regions controlling *qCa1.1* and *qCa4.1* were validated, and seven new QTLs for Ca and Mg contents were identified in the BC₂F_{4,5} population. 18 of 24 QTLs were clustered across seven chromosomal regions, indicating that different mineral accumulation might be involved in common regulatory pathways. Of 24 QTLs identified in two populations, 16 having favorable alleles were derived from *O. rufipogon* and 10 were novel. These results will not only help understand the molecular mechanism of macronutrient accumulation in rice but also provide candidate QTLs for further gene cloning and grain nutrient improvement through QTL pyramiding.

1. Introduction

Mineral nutrients are vital for human health, and more than 25 minerals are required by humans. These minerals can be obtained directly from the daily diet in an appropriate amount [1], and the important minerals are phosphorus (P), sodium (Na), potassium (K), magnesium (Mg), calcium (Ca), iron (Fe), zinc (Zn), copper (Cu), and manganese (Mn). Macroelements, such as P, K, Na, Mg, and Ca, play a

pivotal role in the metabolic functions of fluid balance, neurotransmission, blood pressure regulation, and human's immune system [2, 3]. Although the micronutrient Fe, Zn, and I deficiencies are very common worldwide, macronutrient Mg and Ca deficiencies are serious in some countries [4, 5] and spread across more countries recently [6, 7]. The deficiencies might be partly due to high dietary dependence on a few major staple cereals lacking adequate essential minerals.

Rice (*Oryza sativa* L.) is one of the most important staple foods for over half of the global population. Rice grains store macronutrients (P, K, Mg, and Ca) as well as micronutrients (Fe, Zn, Cu, and Mn) for seed germination and seedling growth [8–10]. Rice grain could be a source of energy and mineral nutrients for rice seedling and human development. Mineral deficiencies may be improved by enhancing grain mineral nutrients. However, most of modern high-yielding rice cultivars do not have sufficient essential minerals to meet the daily dietary requirement, especially in most preferred consumption of polished rice [11]. It is a growing demand to enhance essential mineral contents in rice grain through breeding-based biofortification approaches. Wide variations have been observed for grain mineral contents in diverse rice genotypes [12], which provide a desirable potential to breed rice varieties enriched with essential nutrient. Understanding the genetic basis of mineral accumulation in rice is imperative and will greatly facilitate the enhancement of mineral nutrient in the rice breeding program [13].

Quantitative trait locus (QTL) mapping is a powerful and desirable approach to explore the chromosomal regions controlling quantitative traits for the marker-assisted selection (MAS) strategy in rice. Up to now, a larger number of QTLs associated with mineral contents in rice have been identified using various mapping populations derived from biparental inter- or intra-subspecific and interspecific crosses [5, 11, 13–16]. Using the RILs from the intra-subspecific cross Zhenshan 97/Miyang 46, Yu et al. [13] identified 20 QTLs for P, K, Mg, Ca, Zn, Mn, and Cu contents in milled rice and Wang et al. [5] identified 51 and 61 QTLs for the contents of 13 and 15 minerals including P, K, Na, Mg, and Ca in brown rice and straw, respectively. Garcia-Oliveira et al. [15] identified 31 QTLs for P, K, Mg, Ca, Fe, Zn, Mn, and Cu contents in brown rice using introgression lines (ILs) from an interspecific cross of cultivar “Teqing” and Yunnan wild rice (*O. rufipogon*). Using both RILs and backcross introgression lines (BILs) from an inter-subspecific cross Lemont/Teqing, Zhang et al. [16] reported 134 QTLs for the contents of 16 minerals including P, K, Mg, and Ca in brown rice, which were scattered over 39 genomic regions in 12 rice chromosomes. Descalota-Empleo et al. [11] mapped 50 QTLs for 13 mineral contents in milled rice, including P, K, Na, Mg, and Ca, using two sets of doubled haploid (DH) lines from two inter-subspecific crosses IR64/IR69428 and BR29/IR75862. In addition, Du et al. [14] identified 23 and 9 QTLs for seven mineral contents including P, K, Mg, and Ca in brown rice in the Lingshui and Hangzhou ecological environments, respectively. Of them, only two QTLs for the Mg content were simultaneously identified across two environments, implying that the environment impacts the detection of QTL for grain mineral contents.

Wild species have a high diversity of desirable genes for agronomic traits, and their allelic variation is an important gene pool for genetic improvement [17]. It has been proved that wild rice has much higher contents of minerals than cultivated rice [18, 19], which makes wild rice valuable for biofortification of rice cultivars with enhanced mineral contents.

Dongxiang wild rice (*O. rufipogon* Griff., DWR) is a common wild rice in the northernmost habitat in China and even the world (latitude: 28°14' N), which harbors various valuable genes for improving many traits in rice, i.e., biotic and abiotic stresses [20]. A set of backcross inbred lines (BILs) at BC₁F₅ derived from an interspecific cross cultivar of Xieqingzao B and an accession of DWR were employed to identify QTLs for planthopper resistance [21], cold tolerance [20], and heavy metal content [22]. In the present study, we conducted QTL mapping for the contents of major macronutrients in brown and milled rice using the BILs and then validated the primarily identified QTLs for Mg and Ca contents using the BILs with increasing genetic homogeneity in the maternal XB background. The main objective was to identify candidate QTLs for macronutrient contents in rice for further fine mapping and cloning. The identified QTLs and favorable alleles from *O. rufipogon* could be used for developing rice varieties with enriched mineral nutrients.

2. Materials and Methods

2.1. Plant Materials. Two mapping populations derived from an interspecific cross between a cultivar of Xieqingzao B (XB) (*Oryza sativa* L.) and an accession of Dongxiang wild rice (*O. rufipogon* Griff.) were used in the present study. The first population, consisting of 202 BC₁F₅ BILs previously reported by Chen et al. [23], was used to the initial map QTL for the P, K, Na, Mg, and Ca contents in the present study. XB is an *indica* maintainer line of dwarf-abortive cytoplasmic male sterile line Xieqingzao A, bred by the Rice Research Institute, Anhui Academy of Agricultural Sciences, in the 1980s.

The second population, consisting of 132 BC₂F_{4.5} BILs, was used to verify QTL for Mg and Ca contents detected in the BC₁F₅ population and identify new QTL in a more homogenous background. This population was developed from a plant in the BC₁F₅ population as described and illustrated in Figure 1. The BC₁F₅ BIL plant A58 was backcrossed with recurrent parent XB and selfed to generate 415 BC₂F₂ plants.

After one generation of selfing, the BC₂F_{2.3} lines were assayed with 186 polymorphic simple sequence repeat (SSR) markers distributed across 12 chromosomes. The 186 polymorphic SSR markers included 108 SSRs in the original XB/XB//DWR map [23, 24], 78 other SSRs were selected from the Gramene database (<http://www.gramene.org/>). Of them, 52 SSRs were heterozygous at these loci in the population. Hereafter, these 50 SSRs were used to genotype 415 BC₂F_{2.3} lines. Based on genotypic and phenotypic data, five BC₂F₃ plants from four BC₂F_{2.3} family lines were selected to generate the BC₂F₄ population. Further selection with these markers and agronomic traits resulted in 132 plants from the five BC₂F₄ populations (72 plants for each population), and they were selfed to generate 132 BC₂F_{4.5} lines. The resulting BC₂F_{4.5} lines were genetically more homogenous in the XB background than their parental BC₁F₅ and, thus, were used to verify the QTLs detected in the BC₁F₅ population and further identify new QTLs.

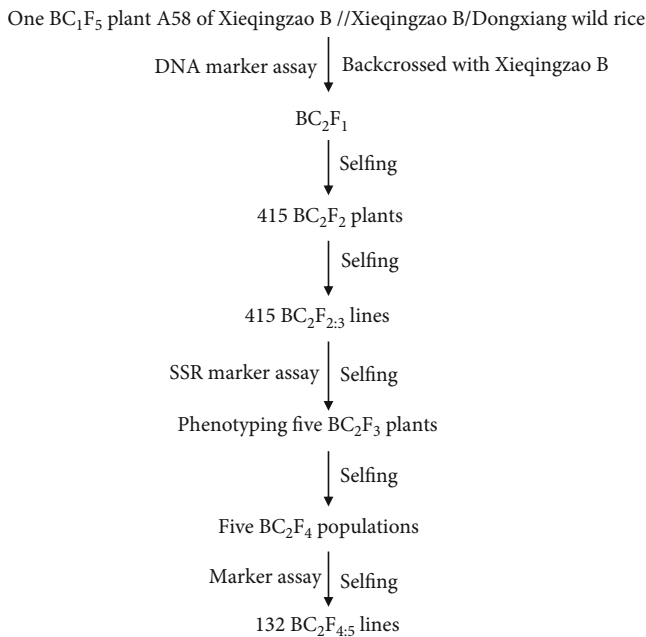


FIGURE 1: Development of the BC₂F_{4:5} population.

2.2. Field Experiments. Rice populations were grown in the experimental fields of the China National Rice Research Institute located in both Lingshui, Hainan province (latitude: 18° 30' N, longitude: 110° 02' E), and Hangzhou, Zhejiang province (latitude: 30° 04' N, longitude: 119° 55' E). The 202 BC₁F₅ lines and the recurrent parent XB were grown in Lingshui, Hainan province, in the 2011 winter season (November 2011 to April 2012) (designated as LS11) and Hangzhou, Zhejiang province, in the 2012 summer season (May to October) (designated as HZ12), respectively. The physicochemical properties of the paddy soil in the LS11 and HZ12 trails were listed in Supplementary Table 1. The 132 BC₂F_{4:5} lines and the parent XB were grown in Hangzhou, Zhejiang province, in the 2016 summer season (May to October) (designated as HZ16) and 2017 summer season (May to October) (designated as HZ17), respectively. In all the field trials, each BIL line consisted of 24 plants in 2 rows, 12 plants per row by transplanting at a spacing specification of 16.7 cm × 26.7 cm between plants. There were no replicates in the BC₁F₅ population. For the BC₂F_{4:5} population, the experiments were conducted under a randomized complete block design with two replicates. Field management and weed control followed the regular agricultural practices. Rice grains of each BIL line were obtained by bulk harvesting the center 10 plants at maturity and threshing manually.

2.3. Sample Preparation. Sample grains were stored and air dried at room temperature for three months. For the LS11 and HZ16 trials, 20 g of the grains from each line were dehulled on a Satake sheller (Satake Corporation, Hiroshima, Japan), ground into flour using a Cyclotec 1093 sample grinder (FOSS Tecator, Hoganas, Sweden), and screened through a 0.18 mm mesh sieve. For the HZ12 and HZ17 trials, 30 g of the grains from each line were dehulled and

divided in half. One-half of the dehulled rice samples were ground into flour using a Cyclotec 1093 sample grinder and screened through a 0.18 mm mesh; the remaining half were milled for one minute firstly in a Kett Pearlest polisher (Kett Electric Laboratory, Tokyo, Japan) and then grounded using a Cyclotec 1093 sample grinder and screened through a 0.18 mm mesh sieve.

2.4. Measurement of the P, K, Na, Mg, and Ca Contents. Approximately 0.5 g flour of each sample was added into a 50 mL polypropylene tube and digested with 8.0 mL 68–70% nitric acid (HNO₃) and 2.0 mL 70% perchloric acid (HClO₃) using a digital block digestion system (model ED54, Labtech Inc., Beijing, China). The digestion procedure was as follows: the digestion tube was heated at 80°C for 0.5 hr, 150°C for 2 hrs, and 180°C until 0.5 mL solution remained. After cooling at room temperature, the final digested residues were diluted to 25 mL with double-deionized water. The contents of P, K, Na, Mg, and Ca in the digested solutions were simultaneously determined using an inductively coupled plasma atomic emission spectrometer (IRIS Intrepid II XSP, Thermo Electron Corporation, Milford, MA, USA) according to the manufacturer's instructions. The certified standards (National Institute Center of Standards in China) in flour samples (GBW10010, CRM Rice) were employed to calibrate the contents of five macronutrients. All the measurements were performed in duplication and the average value over two duplications was used for data analysis.

2.5. DNA Marker Analysis. Total genomic DNA was extracted following the protocol described by Chen et al. [23]. PCR amplification was performed according to Chen et al. [25]. The PCR products were visualized on 2.5% agarose gel stained with GelRed (Biotium, Hayward, CA, USA) or 6% nondenaturing polyacrylamide gel using silver staining.

2.6. Map Construction and Data Analysis. Linkage map of the BC₁F₅ population was constructed using simple sequence repeat (SSR) and restriction fragment length polymorphism (RFLP) markers previously by Chen et al. [23] and updated by Huang et al. [24]. The updated map consisted of 149 DNA markers including 108 SSR markers and 41 RFLPs and spanned 1306.4 cM. The linkage map for the BC₂F_{4:5} population consisted of 186 SSRs which were constructed using MapMaker/Exp 3.0 [26]. The map distance between genetic markers was determined using the Kosambi function and presented in centimorgan.

Statistics were performed for mean, standard deviation (SD), coefficient of variation (CV), skewness, and kurtosis of the phenotypic traits in all trials using the Command DSum of the software Windows QTL Cartographer 2.5 [27]. Pearson's correlation coefficients between the traits were calculated using IBM SPSS Statistics 19. QTL analysis was conducted using the composite interval mapping (CIM) approach of the Windows QTL Cartographer 2.5 [27]. The CIM were performed using the default parameters with backward and forward regressions with a probability threshold of 0.01. The LOD threshold larger than 2.5 was

used to claim a putative QTL. The QTLs detected in this study were designated according to the nomenclature reported by McCouch and CGSNL [28].

3. Results

3.1. Performance of K, P, Na, Mg, and Ca Contents in Brown and Milled Rice. The contents of the five macroelements in brown and milled rice collected from two mapping populations in each trial were presented in Table 1. In all the trials, the contents of the five macronutrients were continuously distributed with the absolute value of skewness and kurtosis less than 1, except for P, K, Na, and Mg contents in milled rice for the HZ12, a typical pattern of quantitative inheritance (Table 1).

For brown rice measured in the BC₁F₅ population in both LS11 and HZ12 trials, the average contents of P, K, and Ca in brown rice were higher in the HZ12 trial than in the LS11 trial, but similar for Na and Mg. For brown rice from the BC₂F_{4.5} population in both HZ16 and HZ17 trials, the average contents of Mg and Ca were higher in the HZ16 trial than in the HZ17 trial (Table 1). These results indicated that the contents of macronutrient in rice were affected by the growing environments, especially the soil (Supplementary Table 1), but the magnitude of influence varied among elements. Pearson correlation analysis indicated that a significantly negative correlation ($r = -0.45$, $P < 0.01$) was only observed for Na between the LS11 and HZ12 trials and no significant correlation was observed for the remaining four macronutrients.

For both the brown and milled rice in the HZ12 and HZ17 trials, the contents of macronutrients in milled rice were lower than those in brown rice. In the HZ12 trial, the average contents of five macronutrients of the BC₁F₅ population in milled rice showed a decrease of 57.14% for P, 57.82% for K, 58.14% for Na, 65.28% for Mg, and 57.10% for Ca, when compared with those in brown rice. Similarly, the reductions of 64.15% for P, 63.50% for K, 8.74% for Na, 75.54% for Mg, and 31.24% for Ca, were observed in the milled rice from the brown rice for parental XB. Pearson correlation analysis indicated that a significantly positive correlation was observed for Na, P, and Ca between brown and milled rice, with the correlation coefficients of 0.65 ($P < 0.01$), 0.17 ($P < 0.05$), and 0.16 ($P < 0.05$), respectively, and no significant relationship was observed for the remaining two macronutrients. In the HZ17 trial, the average contents of Mg and Ca in milled rice of the BC₂F_{4.5} population decreased by 88.12% and 51.99%, respectively, compared to those in brown rice. Likewise for XB, the decreases in the milled rice from the brown rice were 84.14% and 49.29%, respectively, for Mg and Ca contents. The results showed that these macroelements were largely contained in rice bran rather than endosperm. In other words, most of the macroelements in rice are removed during the milling process, which is previously reported [8, 9]. The average contents of Mg and Ca in brown and milled rice of the BC₂F_{4.5} population in both HZ16 and HZ17 trials were lower than those of the BC₁F₅ population in the HZ12 trial. Furthermore, the differences between the average contents of Mg and Ca

in brown and milled rice of the BC₂F_{4.5} population and XB were smaller than those of the BC₁F₅ population and XB. This explained that the BC₂F_{4.5} population was more homogenous in the XB background than the BC₁F₅ population.

Correlations between each pair of the macronutrients within a rice type and a trial were presented in Table 2. One significantly negative correlation was observed for Na-Ca in the LS11 trial, while six significantly positive correlations were found in the LS11 trial ranging from 0.75 to 0.98. For the HZ12 trial, there were six significantly positive correlations ranging from 0.34 to 0.97 in either the brown or milled rice, with an odd for Na-Ca, negative (-0.13) in the brown rice but positive (0.41) in the milled rice. All other correlations were either insignificant or low. Overall, the correlations of P-K, P-Mg, and K-Mg were high for all the types of rice samples and trials.

3.2. QTLs Detected in the BC₁F₅ Population. In total, 20 QTLs for the five macronutrient contents were claimed from both the LS11 and HZ12 trials in both the brown and rice samples, eight in brown rice showing in the LS11 trial only, seven in brown rice showing in the HZ12 trials only, and five in the milled rice showing in the HZ12 trials (Table 3). The identified QTLs were distributed on eight chromosomes of 1-4, 6-7, 9, and 12 (Figure 2). The proportion of phenotypic variance explained by a single QTL (R^2) ranged from 5.70 to 56.80%. The *qNal* for Na was found on chromosome 1 flanked by RM315 and RG236 in both brown and milled rice from the HZ12 trial, which was considered as a duplicate QTL. Therefore, the total number of the identified QTLs decreased to 17. Fourteen of these QTLs formed five clusters (Figure 2).

The largest cluster on chromosome 9 consisted of four QTLs, followed by two clusters of three QTLs on chromosomes 4 and 12. In the RM316-RZ698 region on chromosome 9, the XB allele increased P, K, Mg, and Ca contents in brown rice by 276.52 mg/kg, 148.38 mg/kg, 127.30 mg/kg, and 9.38 mg/kg, respectively (Table 3). In the QTL regions in the vicinity of RM142 on chromosome 4, the DWR allele increased P, Mg, and Ca contents in brown rice by 276.47 mg/kg, 101.80 mg/kg, and 22.94 mg/kg, respectively. In the RM101-RG463 region on chromosome 12, the DWR allele increased P, K, and Mg contents in milled rice by 546.40 mg/kg, 312.64 mg/kg, and 199.10 mg/kg, respectively. All three QTLs, *qP12*, *qK12*, and *qMg12*, had their corresponding highest R^2 value of 53.17%, 33.07%, and 56.80%, respectively.

The other two clusters on chromosomes 1 and 3 each included two QTLs. In the RM315-RG236 region on chromosome 1, the DWR allele increased Ca and Na contents in brown rice and Na content in milled rice by 12.84 mg/kg, 3.35 mg/kg, and 3.48 mg/kg, respectively. It is noted that only one QTL exhibited significant effects on the same macronutrient content in both brown and milled rice. The RG482-RZ519 region on chromosome 3 showed significant effects on P and Mg contents in brown rice, with the DWR allele increasing the two traits by 401.37 mg/kg and 172.90 mg/kg, respectively.

TABLE 1: Phenotypic performance of macronutrient contents in brown and milled rice in two sets of BIL populations derived from interspecific cross of Xieqingzao B/Dongxiang wild rice.

Population	Element	Trial*	Tissue	Mean (mg/kg)	SD	CV	Range	Skewness	Kurtosis	XB (mg/kg)	
BC ₁ F ₅	P	LS11	Brown rice	3785.6	803.6	0.14	2096.0–5868.0	0.34	-0.39	3550.0	
		HZ12	Brown rice	3956.8	642.9	0.11	2112.0–5868.0	0.49	0.36	2848.0	
			Milled rice	1695.8	439.0	0.13	988.0–3426.0	1.59	2.63	1021.0	
	K	LS11	Brown rice	2109.9	476.6	0.14	1089.0–3384.0	0.25	-0.37	2102.0	
		HZ12	Brown rice	2475.9	388.9	0.11	1507.0–3568.0	0.20	-0.005	1810.0	
			Milled rice	1044.3	286.6	0.13	522.0–2214.0	1.28	2.29	661.0	
	Mg	LS11	Brown rice	1442.4	361.3	0.15	696–2416.0	0.39	-0.29	1461.0	
		HZ12	Brown rice	1339.4	251.9	0.11	732.0–2218.0	0.67	1.01	1068.0	
			Milled rice	465.0	163.1	0.15	214.0–1062.0	1.45	1.94	261.0	
	Ca	LS11	Brown rice	133.0	29.1	0.14	61.7–208.6	0.29	-0.42	121.9	
		HZ12	Brown rice	197.4	42.9	0.14	114.4–313.4	0.37	-0.36	179.1	
			Milled rice	84.7	33.7	0.17	3.3–201.8	0.18	0.31	123.2	
	Na	LS11	Brown rice	18.75	9.23	0.22	3.45–41.72	0.21	-1.14	22.12	
		HZ12	Brown rice	14.36	6.41	0.18	0.08–36.22	0.54	0.10	10.30	
			Milled rice	6.01	4.44	0.17	0.06–25.96	1.10	1.64	9.40	
	BC ₂ F _{4:5}	Mg	HZ16	Brown rice	1383.9	90.6	0.07	1207.3–1638.5	0.61	0.17	1257.0
		Ca		Brown rice	148.2	18.3	0.12	115.2–201.4	0.54	-0.35	134.6
		Mg	HZ17	Brown rice	1339.5	31.5	0.02	1275.5–1397.6	-0.18	-0.81	1332.3
			Milled rice	159.1	46.6	0.29	89.7–246.7	0.31	-1.36	211.2	
Ca		HZ17	Brown rice	137.7	12.1	0.09	117.4–159.7	0.05	-1.13	141.8	
			Milled rice	66.1	7.4	0.11	54.0–80.7	0.26	-0.95	71.9	

*LS11: 2011 winter season in Lingshui; HZ12: 2012 summer season in Hangzhou; HZ16: 2016 summer season in Hangzhou; HZ17: 2017 summer season in Hangzhou.

TABLE 2: Correlation coefficients among five macroelement contents in the LS11 and HZ12 trials.

Trial	Tissue	Element	K	Mg	Ca	Na
LS11	Brown rice	P	0.94**	0.98**	0.78**	-0.11
		K		0.93**	0.76**	-0.12
		Mg			0.75**	-0.12
		Ca				0.31**
HZ12	Brown rice	P	0.90**	0.95**	0.42**	0.05
		K		0.86**	0.52**	-0.08
		Mg			0.36**	0.12
	Milled rice	Ca				-0.13
		P	0.91**	0.97**	0.34**	-0.01
		K		0.92**	0.43**	0.05
	Mg			0.38**	0.03	
	Ca				0.41**	

“*” and “**” represent significance levels of $P < 0.05$ and 0.01 , respectively.

In the remaining three regions, one QTL for the K content and two QTLs for the Ca content in each region were detected in the LS11 and HZ12 trials, respectively. They were *qCa1.1* located in the RM5359–RG173 region on chromosome 1, *qK2* in RM250–RM213 on chromosome 2, and *qCa7* in RM214–RG678 on chromosome 7.

3.3. Validation of Two QTL Regions in the BC₂F_{4:5} Population. Of 11 segregating regions in the BC₂F_{4:5} population, QTLs were distributed across 5 regions on chromosomes 1, 4, 6, 9, and 11 (Figure 3; Table 4). Four QTLs for the Mg content and six for the Ca content were identified in both HZ16 and HZ17 trials, including *qCa1.1* and *qCa4.1*, and the others in only one of the trials (Figure 3 and Table 4). The *qCa1.1* for both brown and milled rice Ca contents on chromosome 1 was discovered in the HZ17 trial, making the total number of the identified QTLs down to 9. The R^2 value for brown rice ranged from 13.50% to 17.20% and 19.60% to 19.90% in the HZ16 and HZ17 trials, respectively, and the R^2 value for the milled rice in the HZ17 trial was 17.70% to 63.39%.

Four and three QTLs for the Ca content in both the HZ16 and HZ17 trials (Figure 3 and Table 4) explained 13.5% to 17.2% and 19.60% to 53.38% of the phenotypic variation, respectively. Of them, two segregating QTL regions of the BC₂F_{4:5} population (Figure 3) were covered or neighbored by the QTLs for the Ca content in the BC₁F₅ population, including *qCa1.1* in the interval RM5359–RG173 on chromosome 1 and *qCa4.1* in the interval RG499–RM142 on chromosome 4. The DWR alleles of the *qCa1.1* consistently decreased the brown rice Ca content, while the alleles in *qCa4.1* increased the brown rice Ca content in both the HZ16 and HZ17 trials (Tables 3 and 4). Additionally, the *qCa1.1* showed significant effects on the Ca content in milled

TABLE 3: QTLs for macroelement contents in brown and milled rice detected in the BC₁F₅ BIL population.

Trial	Tissue	Element	QTL	Interval	LOD	A*	R ² %	Previous reports
LS11	Brown rice	P	<i>qP3</i>	RG482-RZ519	3.59	401.37	11.36	
			<i>qP9</i>	RM316-RZ698	3.04	-276.52	8.73	
		K	<i>qK2</i>	RM250-RM213	2.53	175.05	7.30	
			<i>qK9</i>	RM316-RZ698	2.49	-148.38	7.03	[16]
		Mg	<i>qMg3</i>	RG482-RZ519	3.45	172.90	10.30	[5, 15]
			<i>qMg9</i>	RM316-RZ698	3.45	-127.30	9.10	[5, 32]
		Ca	<i>qCa9</i>	RM316-RZ698	2.60	-9.38	7.47	[15, 35]
HZ12	Brown rice	P	<i>qP4</i>	RM142-RM273	2.66	276.47	7.67	
		Mg	<i>qMg4</i>	RG449-RM142	2.52	101.80	6.70	
		Ca	<i>qCa1.1</i>	RM5359-RG173	3.47	-14.58	6.94	[34]
			<i>qCa1.2</i>	RM315-RG236	3.02	12.84	5.98	[35, 36]
		<i>qCa4.1</i>	RG449-RM142	5.24	22.94	13.00		
	Milled rice	Na	<i>qNa1</i>	RM214-RG678	3.02	-19.89	6.46	
		P	<i>qP12</i>	RM315-RG236	2.59	3.35	10.59	[37]
		K	<i>qK12</i>	RM101-RG463	5.06	546.40	53.17	[15, 16, 32]
		Mg	<i>qMg12</i>	RM101-RG463	3.02	312.64	33.07	
		Na	<i>qNa1</i>	RM101-RG463	4.48	199.10	56.80	[15]
Na	<i>qNa1</i>	RM315-RG236	2.87	3.48	22.37	[37]		

A* indicates an additive effect of replacing a Xieqingzao B allele by a Dongxiang wild rice allele; R² indicates the proportion of phenotypic variance explained by the QTL effect.

rice in the HZ17 trial. *qCa1.1* and *qCa4.1* were consistent across both HZ16 and HZ17 trials, respectively.

The other three QTLs identified in the BC₂F_{4.5} population were not detected in the BC₁F₅ population. One of them, *qCa6.1* in the region of RM588–RM204 on chromosome 6, showed significant effects on the brown rice Ca content in the HZ16 trial, and its DWR allele increased the Ca content by 10.11 mg/kg (Table 4). In the neighboring region of *qCa6.1* between RM276 and RG64 in the BC₁F₅ population, *qCa6* for the milled rice Ca content was identified, in which favorable alleles were derived from DWR. At the *qCa4.2* and *qCa11*, dominant XB alleles increased the milled and brown rice Ca contents by 2.98 mg/kg and 5.17 mg/kg, respectively, with R² of 53.38% and 13.50%.

One and three QTLs were detected for both the brown and milled rice Mg contents in the HZ16 and HZ17 trials, respectively, with R² from 15.50% to 73.39%. The QTL *qMg1* had the XB allele with a partial dominant effect that increased the Mg content in brown rice by 41.76 mg/kg. The remaining three QTLs, *qMg4.1*, *qMg9.1*, and *qMg11* had the enhancing alleles from DWR and showed the over-dominant, partially dominant, and additive effects, respectively.

4. Discussion

In general, the mineral nutrient content is very low in rice, even though it serves as a major staple food for half the world's population. Therefore, a rice-based diet is associated with mineral deficiency in rice-consuming people. A slight increment of the mineral content in rice should effectively alleviate erupting dietary deficiency worldwide [29]. The

increment relies on breeding rice varieties with enhanced mineral nutrient contents using MAS [30]. Identification of QTLs controlling mineral contents in rice is the prerequisite for pyramiding of QTL for mineral biofortification with MAS.

4.1. Genetic and Environmental Effects on Macronutrient Content QTL Detection. Using various mapping populations from the same cross phenotype across environments could help eliminate genetic and environmental noise for dissecting consistent QTL. In the present study, we used a same interspecific cross between Xieqingzao B (*Oryza sativa* L.) and Dongxiang wild rice (*O. rufipogon* Griff.) to generate two populations of BC₁F₅ and BC₂F_{4.5} BILs for QTL analysis across two environments and two seasons, respectively. As a result, 17 QTLs for macronutrient contents in brown and milled rice were initially identified in BC₁F₅ BILs, and then, 9 QTLs for Ca and Mg contents were in BC₂F_{4.5} BILs in which the background should be more homogeneous than that of the BC₁F₅ BILs. In addition, more QTLs with higher statistical significance and effectiveness were resulted from the BC₂F_{4.5} BILs than from the BC₁F₅ BILs. The higher significance and effectiveness might be due to a high genetic homogeneity of 72.04% at 186 marker loci in the XB genetic background, indicating that BC₂F_{4.5} BILs are more appropriate to detect QTL with smaller effect than BC₁F₅ BILs. The result is in accord with the findings reported by Zhang et al. [16].

Mineral contents in rice were influenced by multiple factors [14, 22], especially the soil. The climate in Lingshui, Hainan, is a typical tropical monsoonal, with sufficient sunshine and abundant rainfall, while that in Hangzhou, Zhejiang, is a

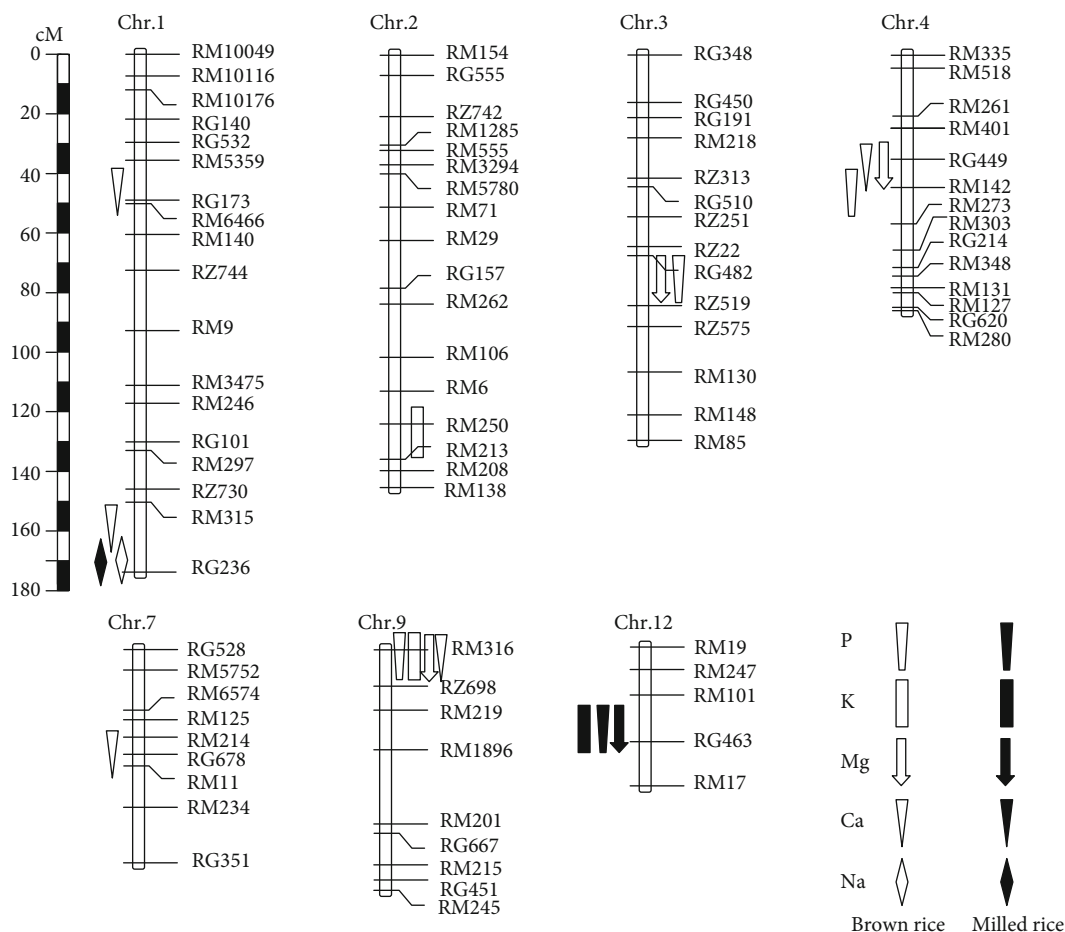


FIGURE 2: Chromosomal positions of the QTLs conferring P, K, Na, Mg, and Ca contents in brown and milled rice in the BC₁F₅ XB²/Dongxiang wild rice. QTLs on the right and left hand sides of the chromosomes indicate that they were detected in the LS11 and HZ12 trials, respectively.

TABLE 4: QTLs for Mg and Ca contents in brown and milled rice detected in the BC₂F_{4.5} population.

Trial	Tissue	Element	QTL	Interval	LOD	A *	D	R ²	Previous reports
HZ16	Brown rice	Mg	<i>qMg1</i>	RM10300-RM10345	4.86	-41.76	-18.81	15.50	[11]
		Ca	<i>qCa1.1</i>	RM10300-RM10345	8.73	-9.00	-4.36	15.10	[34]
			<i>qCa4.1</i>	RM273-RM303	3.12	17.88	-4.93	13.60	
			<i>qCa6.1</i>	RM588-RM204	9.62	10.11	9.66	17.20	[5, 13, 37]
			<i>qCa11</i>	RM21-RM206	7.27	-5.17	10.56	13.50	[14]
HZ17	Brown rice	Ca	<i>qCa1.1</i>	RM10300-RM10345	2.62	-11.93	7.34	19.90	[34]
			<i>qCa4.1</i>	RM273-RM303	2.53	6.61	8.60	19.60	
	Milled rice	Mg	<i>qMg4.1</i>	RM348-RM131	3.38	33.59	-81.32	17.90	
			<i>qMg9.1</i>	RM1896-RM24152	2.91	65.49	-19.67	17.70	
		Ca	<i>qMg11</i>	RM254-RM7163	14.56	45.72	-7.32	63.39	[11, 33]
	Ca	<i>qCa1.1</i>	RM10300-RM10345	2.54	-6.75	5.68	18.77	[34]	
		<i>qCa4.2</i>	RM348-RM131	2.88	-2.98	13.58	53.38	[15, 16]	

A* indicates an additive effect of replacing a Xieqingzao B allele by a Dongxiang wild rice allele; R² indicates the proportion of phenotypic variance explained by the QTL effect.

humid subtropical. The physicochemical properties of the soil (Supplementary Table 1) were obviously different at two distinct ecological locations. Accordingly, the contents

of macronutrients in brown rice were different between LS11 and HZ12 trials. Taking Ca content in brown rice as an example, the values in the HZ12 trial ranged from 114.4

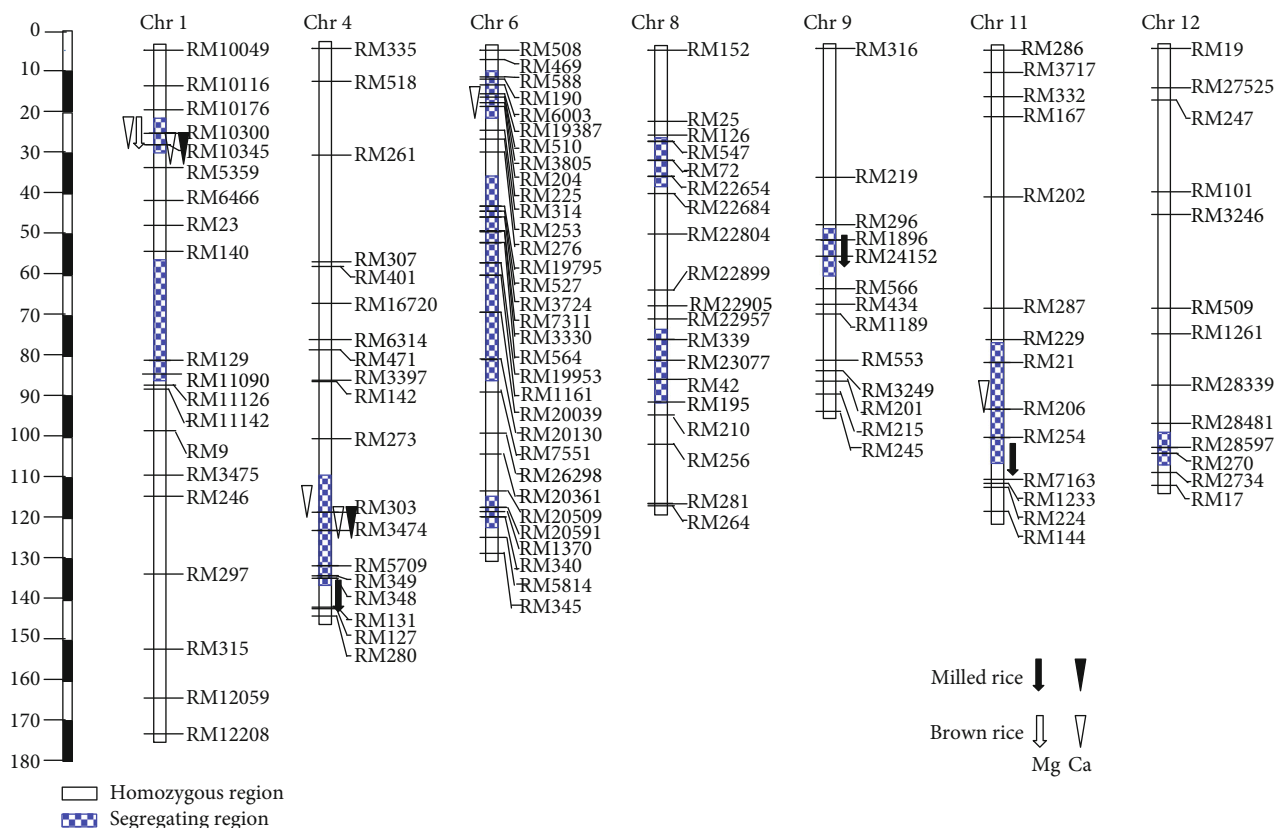


FIGURE 3: Chromosomal positions of the QTLs conferring Mg and Ca contents in brown and milled rice in the $BC_2F_{4.5}$ population of Xieqingzao B/Dongxiang wild rice. QTLs on the right and left hand sides of the chromosomes indicate their origination from the HZ16 and HZ17 trials, respectively.

to 313.4 mg/kg^{-1} with an average content of 197.4 mg/kg^{-1} , which were higher than the values ranging from 61.7 to 208.6 mg/kg^{-1} with a mean value of 133.0 mg/kg^{-1} in the LS11 trial. Consistent with previous studies [14, 22, 31], 17 QTLs for macronutrient contents in BC_1F_5 BILs were all environment specific. These results suggested that different environmental factors, especially the soil, affected the accumulations of the macronutrients as well as QTL detection. However, it is notable that two QTLs, *qCa1.1* and *qCa4.1*, were validated with constant allelic directions by identification across populations and seasons. These environment-specific and/or stable QTLs across environments with consistent allelic effects are of great importance, which provided good candidates for rice biofortification breeding and further gene cloning.

4.2. QTL Cluster for Macronutrient Contents. QTL clusters for multi-macronutrient contents in this study are in line with previous reports [5, 11, 14–16]. In total, 14 and 4 of the QTLs in the BC_1F_5 and $BC_2F_{4.5}$ populations were clustered over five and two genomic regions distributed on six chromosomes, respectively. Among them, six regions were also reported in previous studies and the remaining regions were novel. Three genomic regions of eight QTL clusters were especially important. One super QTL cluster on chromosome 9 contained *qP9*, *qK9*, *qCa9*, and *qMg9*, of which

qK9 was also identified for the K content in brown rice reported previously [16]. The second QTL cluster *qP12/qK12/qMg12* was within the interval RM101–RG463 on chromosome 12. Three QTLs in this region, *qP12*, *qK12* and *qMg12*, all had the largest R^2 and favorable alleles from *O. rufipogon*, of which *qP12* and *qMg12* were reported in previous studies [15, 16, 32]. The remaining QTL cluster *qCa1.1/qMg1.1* was adjacent to the interval in the vicinity of RM5359 on chromosome 1, of which *qCa1.1* was across two mapping populations and three trials, providing a good candidate for enhancing the Ca content in both the brown and milled rice. In addition, further fine mapping and cloning are required to confirm the allelic relationships of these QTL clusters.

The significant correlations of macronutrient contents with each other were observed, and those correlations were the same in both the brown and milled rice samples. Notably, the highly positive correlations were found for each pair of K, P, Mg, and Ca contents in the brown and milled rice samples in both HZ12 and LS11 trials. Their high correlations have been previously reported [5, 16]. Accordingly, two QTL clusters for K-P-Mg contents in brown and milled rice were collocated on chromosomes 9 and 12, respectively. These QTL clusters demonstrate that the pleiotropic gene or tightly linked genes controlling different macronutrient contents were located on the same chromosomal region, probably

because these different macronutrients are involved in the common regulatory pathways. Once the synergistic associations for these macronutrients are confirmed, the improvement of the macronutrient contents could be very effective because a single effort in rice breeding practices could result in an enhancement of the multi-macronutrient content simultaneously.

4.3. Comparison of QTLs for Macronutrient Contents between Present and Previous Studies. Based on the physical position of flanking markers, we compared the similarity for the QTLs identified in the present study with earlier reports for the same macronutrient within a physical distance of 1 Mb. Of 24 QTLs, 14 were coincident with previous studies and 10 were novel QTLs. *qP12* and *qK9* accorded with a QTL related to the P content [15, 16, 32] and K content [16], respectively, in previous reports. The *qMg1* coincided with *qMg1.1* for the Mg content in milled rice [11], *qMg3* with *qMg3-1* [15] and *qMg3.2* [5], *qMg9* with *qMg9.1* for brown rice [5] and *qMg9* for milled rice [33], *qMg11* with *qMg11.1* [11] and *qMg11* [33], and *qMg12* with *qMg12-1* [15]. *qCa1.1* and *qCa1.2* accorded with *qCa1.1* for the Ca content in milled rice [34] and a QTL for brown rice [35, 36], respectively, *qCa4.2* with a QTL for the Ca content in brown rice [15, 16], *qCa6.1* with *qCa6.2* and *qCa6* for brown rice [5, 37] and *qCa6* for milled rice [13], *qCa9* with a QTL for the Ca content in brown rice in earlier studies [15, 36], *qCa11* with a QTL for brown rice reported by Du et al. [14], and *qNa1* with *qNa1* for the Na content [37]. These QTLs across diverse genetic backgrounds and environments are considered as consensus QTLs. They are worthy of exploiting the full potential for biofortifying rice with enriched macronutrient in breeding practices, while effort should be focused on the identification of molecular markers tightly linked to the target QTLs for MAS breeding. Moreover, these consensus QTLs in various studies are also good candidates for verification of allelic effects, fine mapping, and cloning.

A large number of genes involved in mineral absorption and translocation have been isolated and characterized in rice. As expected, some potential genes related to macronutrient uptake, transportation, and homeostasis in rice were within the chromosomal regions of the QTLs in present study. These QTL regions harbor the following potential genes, such as *OsPAP10c* for P, *OsHAK2* for Na, *OsCNGC4*, *OsCNGC5*, *OsKCO3*, and *OsCNGC17* for K; *OsMGT3* for Mg; and *OsEFCAX1*, *OsCam1-3*, *OsCAX3*, *OsCCX5*, and *OsCML6* for Ca. Interestingly, *OsEFCAX1* (LOC_Os01g11414) and *OsCam1-3* (LOC_Os01g16240) for Ca are present within the *qCa1.1/qMg1* regions consistently across multiexperiments of the present study (Figures 2 and 3), which are involved in calcium-mediated signaling and homeostasis and expressed in all tissues including seeds [38, 39]. It would be important to generate near-isogenic lines differing in alleles at these gene loci on the XB genetic background for investigation of the relationships with the QTLs identified in the present study.

4.4. Utilization of Dongxiang Wild Rice for Rice Biofortification Breeding. To satisfy the growing demand of food and dietary nutrition security in the rice-consuming population, more and more attention has been focused on enhancing the contents of essential nutrients while decreasing the concentrations of toxic metals in rice grain. However, the complex interactions between toxic elements and essential nutrients add difficulty to this global effort [40]. Previously, we have used the same BC₁F₅ BILs to map QTL for grain yield with contributing components [24]. In the previous and present studies, we have reported eight QTLs for yield components and six QTLs for macronutrient contents which distribute over four chromosomal regions or clusters on chromosomes 4, 6, 7, and 12. Two major clusters on chromosomes 4 and 12 holding four QTLs are *qTGW-12* and *qMg12/qP12/qK12* and *qSF-4/qNFGP-4/qGYD-4* and *qCa4.2*. These two clusters demonstrate that the DWR alleles have synergistic effects for yield and macronutrient traits in the QTL regions on chromosomes 4 and 12. Many other reports have revealed the synergistic effects, as well [31, 34, 41]. The remaining two clusters on chromosomes 6 and 7 harbor three QTLs, *qNFGP-6/qSF-6*, *qCa6.1*, and *qNFSP-7/qTFGP-7/qCa7*. However, the DWR alleles in the clustered QTLs on chromosomes 6 and 7 showed the opposite allele effects on the macronutrient content and yield components. These results demonstrate that the *O. rufipogon* alleles in favor of some traits may be unfavorable to the others. These deleterious associations between grain mineral nutrients and yield traits have been reported previously [5, 11, 16, 42]. Interruption of deleterious linkages should be done before those desirable genes could be transferred from *O. rufipogon* into *O. sativa* in cultivar improvement.

Up to now, much efforts have been made to exploit favorable alleles from wild rice species for grain quality enhancement [43], grain yield, and resistance improvement [44]. Wild rice is rich in some essential nutrients in comparison of cultivated rice [18, 19]. Dongxiang wild rice is the progenitor of cultivated rice, having numerous favorable alleles [21, 45]. These alleles are widely utilized in genomic and breeding studies for improvement of important agronomic traits. Notably, 11 of 17 (64.70%) and 5 of 9 (55.55%) QTLs in the BC₁F₅ and BC₂F_{4,5} BILs have favorable alleles from *O. rufipogon* for enrichment of macronutrient contents. Similarly, previous studies showed that three AA-genome wild rice species, *O. rufipogon*, *O. nivara*, and *O. meridionalis*, have favorable alleles of 83.87% (26 of 31 QTLs), 83.33% (25 of 30 QTLs), and 100% (4 of 4 QTLs), respectively, for gain mineral nutrient contents [15, 46, 47]. These results suggest that wild relatives of rice cultivar may greatly contribute to improving grain mineral nutrient contents in cultivated species of *O. sativa*. They have precious potentials for the biofortification of cultivated rice with enhanced grain mineral contents. Introgression of favorable alleles from *O. rufipogon* may introduce some unfavorable allele into elite rice cultivars because of their tight linkage. It becomes important to interrupt these types of linkage for breeding rice varieties for integrating grain yield with mineral traits.

5. Conclusions

A total of 24 QTLs for the contents of five macronutrients in rice were detected using two BIL populations derived from the same interspecific cross. Dongxiang wild rice contributed 65.38% (16 out of 24) favorable QTL alleles for the improvement of the grain macronutrient content in future breeding. Two QTLs *qCa1.1* and *qCa4.1* with consistent allelic directions across different populations and/or trials should be taken into account in grain nutrient biofortification through QTL pyramiding.

Data Availability

All the relevant data have been incorporated into the original research manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Biao-lin Hu, Yan-shou Wu, and Jian-hua Yin designed the research. Ting Wu, Xia Li, De-run Huang, and Feng-lin Huang performed the research. Biao-lin Hu and Jian-hua Yin supervised the data analysis and interpretation. Ting Wu and Feng-lin Huang analyzed the data. Biao-lin Hu and Yan-shou Wu wrote the paper. Jian-hua Yin and Biao-lin Hu critically revised the paper for publication.

Acknowledgments

This study was supported by grants from the Natural Science Foundation of Jiangxi Province (no. 20171ACB21071), National Natural Science Foundation of China (no. 31760378), the Natural Science Foundation of Jiangxi Province (no. 2020BABL205018), and the Open Fund of the State Key Laboratory of Rice Biology (no. 20200201).

Supplementary Materials

Supplementary Table 1 The physicochemical characteristics of the soil in the experimental fields in LS11 and HZ12. (*Supplementary Materials*)

References

- [1] P. J. White and P. H. Brown, "Plant nutrition for sustainable development and global health," *Annals of Botany*, vol. 105, no. 7, pp. 1073–1080, 2010.
- [2] D. P. Chaudhary, R. Sharma, and D. D. Bansal, "Implications of magnesium deficiency in type 2 diabetes: a review," *Biological Trace Element Research*, vol. 134, no. 2, pp. 119–129, 2010.
- [3] A. J. Stein, "Global impacts of human mineral malnutrition," *Plant Soil*, vol. 335, no. 1-2, pp. 133–154, 2010.
- [4] D. B. Kumssa, E. J. M. Joy, E. L. Ander et al., "Dietary calcium and zinc deficiency risks are decreasing but remain prevalent—Dietary calcium and zinc deficiency risks are decreasing but remain prevalent," *Scientific Reports*, vol. 5, no. 1, article 10974, 2015.
- [5] C. C. Wang, Z. Tang, J. Y. Zhuang, Z. Tang, X. Y. Huang, and F. J. Zhao, "Genetic mapping of ionic quantitative trait loci in rice grain and straw reveals *OsMOT1;1* as the putative causal gene for a molybdenum QTL *qMo8*," *Molecular Genetics and Genomics*, vol. 295, no. 2, pp. 391–407, 2020.
- [6] A. Mahender, A. Anandan, S. K. Pradhan, and E. Pandit, "Rice grain nutritional traits and their enhancement using relevant genes and QTLs through advanced approaches," *SpringerPlus*, vol. 5, no. 1, p. 2086, 2016.
- [7] N. Rawat, K. Neelam, V. K. Tiwari, and H. S. Dhaliwal, "Biofortification of cereals to overcome hidden hunger," *Plant Breeding*, vol. 132, no. 5, pp. 437–445, 2013.
- [8] L. L. Lu, S. K. Tian, H. B. Liao et al., "Analysis of metal element distributions in rice (*Oryza sativa* L.) seeds and relocation during germination based on X-ray Fluorescence imaging of Zn, Fe, K, Ca, and Mn," *PLoS One*, vol. 8, no. 2, article e57360, 2013.
- [9] T. Iwai, M. Takahashi, K. Oda, Y. Terada, and K. T. Yoshida, "Dynamic changes in the distribution of minerals in relation to phytic acid accumulation during rice seed development," *Plant Physiology*, vol. 160, no. 4, pp. 2007–2014, 2012.
- [10] J. Pariasca-Tanaka, E. Vandamme, A. Mori et al., "Does reducing seed-P concentrations affect seedling vigor and grain yield of rice?," *Plant and Soil*, vol. 392, no. 1-2, pp. 253–266, 2015.
- [11] G. I. Descalsota-Empleo, A. Amparado, M. A. Inabangan-Asilo et al., "Genetic mapping of QTL for agronomic traits and grain mineral elements in rice," *Crop Journal*, vol. 7, no. 4, pp. 560–572, 2019.
- [12] S. R. M. Pinson, L. Tarpley, W. G. Yan et al., "Worldwide genetic diversity for mineral element concentrations in rice grain," *Crop Science*, vol. 55, no. 1, pp. 294–311, 2015.
- [13] Y. H. Yu, Y. F. Shao, J. Liu et al., "Mapping of quantitative trait loci for contents of macro- and microelements in milled rice (*Oryza sativa* L.)," *Journal of Agricultural and Food Chemistry*, vol. 63, no. 35, pp. 7813–7818, 2015.
- [14] J. Du, D. Zeng, B. Wang, Q. Qian, S. Zheng, and H. Q. Ling, "Environmental effects on mineral accumulation in rice grains and identification of ecological specific QTLs," *Environmental Geochemistry and Health*, vol. 35, no. 2, pp. 161–170, 2013.
- [15] A. L. Garcia-Oliveira, L. B. Tan, Y. C. Fu, and C. Q. Sun, "Genetic identification of quantitative trait loci for contents of mineral nutrients in rice grain," *Journal of Integrative Plant Biology*, vol. 51, no. 1, pp. 84–92, 2009.
- [16] M. Zhang, S. R. M. Pinson, L. Tarpley et al., "Mapping and validation of quantitative trait loci associated with concentrations of 16 elements in unmilled rice grain," *Theoretical and Applied Genetics*, vol. 127, no. 1, pp. 137–165, 2014.
- [17] S. D. Tanksley and S. R. McCouch, "Seed banks and molecular maps: unlocking genetic potential from the wild," *Science*, vol. 277, no. 5329, pp. 1063–1066, 1997.
- [18] Z. Q. Cheng, X. Q. Huang, Y. Z. Zhang et al., "Diversity in the content of some nutritional components in husked seeds of three wild rice species and rice varieties in Yunnan province of China," *Journal of Integrative Plant Biology*, vol. 47, no. 10, pp. 1260–1270, 2005.
- [19] S. L. Jiang, C. H. Shi, and J. G. Wu, "Studies on mineral nutrition and safety of wild rice (*Oryza* L.)," *International Journal of Food Sciences and Nutrition*, vol. 60, pp. 139–147, 2009.
- [20] B. L. Hu, T. Wu, X. Li, Y. S. Wu, and J. H. Yin, "QTL mapping for cold tolerance at early seedling stage in Dongxiang wild rice (*Oryza rufipogon* Griff.) under severe cold stress,"

- International Journal of Agriculture and Biology*, vol. 22, pp. 1017–1022, 2019.
- [21] J. Chen, D. R. Huang, L. Wang, G. J. Liu, and J. Y. Zhuang, "Identification of quantitative trait loci for resistance to white-backed planthopper, *Sogatella furcifera*, from an interspecific cross *Oryza sativa* × *O. rufipogon*," *Breeding Science*, vol. 60, no. 2, pp. 153–159, 2010.
- [22] D. R. Huang, Y. Y. Fan, B. L. Hu, Y. Q. Xiao, D. Z. Chen, and J. Y. Zhuang, "Assessment and genetic analysis of heavy metal content in rice grain using an *Oryza sativa* × *O. rufipogon* backcross inbred line population," *Journal of the Science of Food and Agriculture*, vol. 98, no. 4, pp. 1339–1345, 2018.
- [23] J. Chen, B. Hu, D. Z. Chen, G. J. Liu, K. L. Zheng, and J. Y. Zhuang, "Development of chromosomal segment substitution lines from a backcross recombinant inbred population of interspecific rice cross," *Rice Science*, vol. 13, no. 1, pp. 15–21, 2006.
- [24] D. R. Huang, J. Chen, L. J. Hou, Y. Y. Fan, and J. Y. Zhuang, "Identification of QTLs for yield traits in the BC₂F₅ population of Xieqingzao B//Xieqingzao B/Dongxiang wild rice," *Journal of Agricultural Biotechnology*, vol. 16, no. 6, pp. 977–982, 2008.
- [25] X. Chen, S. Temnykh, Y. Xu, Y. G. Cho, and S. R. McCouch, "Development of a microsatellite framework map providing genome-wide coverage in rice (*Oryza sativa* L.)," *Theoretical and Applied Genetics*, vol. 95, no. 4, pp. 553–567, 1997.
- [26] E. Lander, P. Green, J. Abrahamson et al., "MAPMAKER: an interactive computer package for constructing primary genetic linkage maps of experimental and natural populations," *Genomics*, vol. 1, no. 2, pp. 174–181, 1987.
- [27] S. C. Wang, C. J. Basten, and Z. B. Zeng, *Windows QTL cartographer 2.5*, Department of Statistics, North Carolina State University, Raleigh, NC, USA, 2012, <http://statgen.ncsu.edu/qtlcart/WQTLCart.htm>.
- [28] S. R. McCouch and Committee on Gene Symbolization, Nomenclature and Linkage, Rice Genetics Cooperative, "Gene nomenclature system for rice," *Rice*, vol. 1, no. 1, pp. 72–84, 2008.
- [29] G. B. Gregorio, D. Senadhira, T. Htut, and R. D. Graham, "Breeding for trace mineral density in rice," *Food and Nutrition Bulletin*, vol. 21, no. 4, pp. 382–386, 2000.
- [30] G. I. L. Descalsota, B. P. M. Swamy, H. Zaw et al., "Genome-Wide association mapping in a rice MAGIC plus population detects QTLs and genes useful for biofortification," *Frontier in the Plant Science*, vol. 9, p. 1347, 2018.
- [31] Q. Xu, T. Q. Zheng, X. Hu et al., "Examining two sets of introgression lines in rice (*Oryza sativa* L.) reveals favorable alleles that improve grain Zn and Fe concentrations," *PLoS One*, vol. 10, no. 7, article e0131846, 2015.
- [32] C. R. J. Stangoulis, B. L. Huynh, M. R. Welch, E. Y. Choi, and R. D. Graham, "Quantitative trait loci for phytate in rice grain and their relationship with grain micronutrient content," *Euphytica*, vol. 154, no. 3, pp. 289–294, 2007.
- [33] G. J. Norton, C. M. Deacon, L. Z. Xiong, S. Y. Huang, A. A. Meharg, and A. H. Price, "Genetic mapping of the rice ionome in leaves and grain: identification of QTLs for 17 elements including arsenic, cadmium, iron and selenium," *Plant and Soil*, vol. 329, no. 1-2, pp. 139–153, 2010.
- [34] B. P. M. Swamy, G. I. L. Descalsota, C. T. Nha et al., "Identification of genomic regions associated with agronomic and biofortification traits in DH populations of rice," *PLoS One*, vol. 13, no. 8, article e0201756, 2018.
- [35] Z. H. Sun, F. H. Xiao, Y. W. Zeng et al., "QTL analysis of calcium content in brown rice *Japonica*," *Acta Botanica Boreali-Occidentalia Sinica*, vol. 30, no. 3, pp. 481–486, 2010.
- [36] M. Yang, K. Lu, F. J. Zhao et al., "Genome-Wide association studies reveal the genetic basis of ionic variation in rice," *Plant Cell*, vol. 30, no. 11, pp. 2720–2740, 2018.
- [37] Y. Huang, C. Tong, F. F. Xu, Y. L. Chen, C. Y. Zhang, and J. S. Bao, "Variation in mineral elements in grains of 20 brown rice accessions in two environments," *Food Chemistry*, vol. 192, pp. 873–878, 2016.
- [38] B. Boonburapong and T. Buaboocha, "Genome-wide identification and analyses of the rice calmodulin and related potential calcium sensor proteins," *BMC Plant Biology*, vol. 7, no. 1, p. 4, 2007.
- [39] A. Singh, P. Kanwar, A. K. Yadav et al., "Genome-wide expression and functional analysis of calcium transport elements during abiotic stress and development in rice," *The FEBS Journal*, vol. 281, no. 3, pp. 894–915, 2014.
- [40] M. A. Khaliq, B. James, Y. H. Chen et al., "Uptake, translocation, and accumulation of cd and its interaction with mineral nutrients (Fe, Zn, Ni, Ca, Mg) in upland rice," *Chemosphere*, vol. 215, pp. 916–924, 2019.
- [41] S. Ishikawa, T. Abe, M. Kuramata et al., "A major quantitative trait locus for increasing cadmium-specific concentration in rice grain is located on the short arm of chromosome 7," *Journal of Experimental Botany*, vol. 61, no. 3, pp. 923–934, 2010.
- [42] A. Anandan, G. Rajiv, R. Eswaran, and M. Prakash, "Genotypic variation and relationships between quality traits and trace elements in traditional and QTL analysis for grain iron and zinc concentrations in two *O. nivara* derived backcross populations improved rice (*Oryza sativa* L.) genotypes," *Journal of Food Science*, vol. 76, no. 4, pp. 122–130, 2011.
- [43] C. Q. Zhang, J. H. Zhu, S. J. Chen et al., "Wx^{lv} the Ancestral Allele of Rice *Waxy* Gene," *Molecular Plant*, vol. 12, no. 8, pp. 1157–1166, 2019.
- [44] D. S. Brar and G. S. Khush, "Wild relatives of rice: a valuable genetic resource for genomics and breeding research," in *The wild Oryza genomes, Compendium of Plant Genomes*, T. Mondal and R. Henry, Eds., pp. 1–25, Springer, Cham, 2018.
- [45] X. X. Dong, X. Y. Wang, L. S. Zhang et al., "Identification and characterization of OsEBS, a gene involved in enhanced plant biomass and spikelet number in rice," *Plant Biotechnology Journal*, vol. 11, no. 9, pp. 1044–1057, 2013.
- [46] B. P. M. Swamy, K. Kaladhar, K. Anuradha, A. K. Batchu, T. Longvah, and N. Sarla, "QTL analysis for grain iron and zinc concentrations in two *O. nivara* derived backcross populations," *Rice Science*, vol. 25, no. 4, pp. 197–207, 2018.
- [47] R. Ishikawa, M. Iwata, K. Taniko et al., "Detection of quantitative trait loci controlling grain zinc concentration using Australian wild rice, *Oryza meridionalis*, a potential genetic resource for biofortification of rice," *PLoS One*, vol. 12, no. 10, article e0187224, 2017.