



Impact of Normoglycemia in Reducing Microvascular Complications in Patients with Type 2 Diabetes: A Follow-Up Study

Fukashi Ishibashi¹ and Mitra Tavakoli^{2*}

¹ Ishibashi Clinic, Hiroshima, Japan, ² University of Exeter Medical School, Exeter, United Kingdom

Aims: Hyperglycemia is associated with an increased risk of microvascular complications in patients with type 2 diabetes. The aim of the present study was to investigate whether the reduction of the levels of HbA1c by tight glycemic control (GC) decreases the rate of microvascular complications and improves the neurological measures in patients with type 2 diabetes.

Methods: Detailed clinical and neurological examinations including corneal confocal microscopy (CCM) were performed in 141 Japanese patients with type 2 diabetes and 60 age-matched control subjects at baseline and follow-up with GC for 4 years. Patients were stratified according to the mean HbA1c level during follow-up into good (HbA1c < 53.0 mmol/mol, mean; 47.5 mmol/mol), fair (53.0 mmol/mol/mol \leq HbA1c < 58.5 mmol/mol, mean; 55.6 mmol/mol), and poor (HbA1c \geq 58.5 mmol/mol/mol, mean; 68.9 mmol/mol) GC groups with similar HbA1c levels at baseline (84.5–88.2 mmol/mol).

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> *Correspondence: Mitra Tavakoli m.tavakoli@exeter.ac.uk

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Ishibashi F and Tavakoli M (2018) Impact of Normoglycemia in Reducing Microvascular Complications in Patients with Type 2 Diabetes: A Follow-Up Study. Front. Endocrinol. 9:52. doi: 10.3389/fendo.2018.00052 **Results:** At baseline, CCM revealed significant nerve fiber damage in all patients compared to that in controls. The interval changes in most corneal nerve fiber (CNF) parameters and neurophysiological functions were significantly related with the mean HbA1c levels during follow-up. Interestingly, the baseline HbA1c level did not impact on neurological functions at follow-up. Interval changes in neuropathy outcomes were associated with mean clinical factors during follow-up and hypoglycemic strategies. Good GC improved all nerve functions, including CNF branch density and bead, but not the length and main fiber density. Fair GC deteriorated some nerve functions. Poor GC compromised all neuropathy outcomes. Irrespective of GC levels, retinopathy increased after follow-up period, while nephropathy decreased.

Conclusion: This study showed that tight GC was beneficial just for nephropathy among microvascular complications. Despite strict GC, the retinopathy progressed in patients with type 2 diabetes. Glucose control did not improve neurophysiological and corneal nerve measurements unless near-normoglycemia was reached.

Keywords: microvascular complications, neuropathy outcome, corneal nerve fibers, glycemic control, nearnormoglycemia, type 2 diabetes, treatment strategy

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Abbreviations: BF, beading frequency; BS, bead size; CCM, corneal confocal microscopy; CNBD, corneal nerve branch density; CNF, corneal nerve fiber; CNFD, corneal nerve fiber density; CNFL, corneal nerve fiber length; DPN, diabetic peripheral neuropathy; ETDRS, Early Treatment Diabetic Retinopathy Study; MCV, motor nerve conduction velocity; NCV, nerve conduction velocity; NDS, neuropathy disability score; SCV, sensory nerve conduction velocity; SNF, small nerve fiber.

INTRODUCTION

Hyperglycemia is associated with an increased risk of microvascular complications in patients with type 2 diabetes mellitus (T2DM). Several large clinical trials concluded that intensive glycemic control (GC) in type 2 diabetes is associated with a reduction in microvascular complications, in particular nephropathy and vascular outcomes (1–3).

It has been shown that intensive GC has an equivocal efficacy for diabetic peripheral neuropathy (DPN) in T2DM (4). The earliest nerve fibers to undergo damage and subsequent repair are small nerve fibers (SNFs) (5, 6). Therefore, the functional and morphological measures for SNF neuropathy are essential for estimating the efficacy of intervention for DPN.

Small nerve fibers can be assessed objectively by quantifying intraepidermal nerve fiber density in skin biopsies; however, this is an invasive procedure that requires expert laboratory assessment and has considerable variability even among control (7). The use of corneal confocal microscopy (CCM) for rapid, noninvasive clinical assessment of corneal nerve fibers (CNFs) has grown significantly in recent years (8). It has proven to be particularly useful as a diagnostic marker for the detection of diabetic neuropathy (9–11) and a range of other peripheral neuropathies (12–14).

Due to potential severe hypoglycemia (3), euglycemia using insulin-providing strategy in T2DM patients may be difficult. The benefit of strict GC on neurophysiological functions, CNF parameters, and other microangiopathies markers in patients with poorly controlled type 2 diabetes is unclear and needs to be investigated.

The aim of the present study is to establish whether the tight control of HbA1c improves the CNF measures and neurophysiological functions along with the occurrence of nephropathy and retinopathy in patients with poorly controlled type 2 diabetes by strict GC for 4 years.

SUBJECTS AND METHODS

Subjects

At the baseline, 141 Japanese patients with poorly controlled type 2 diabetes (HbA1c > 58.5 mmol/mol) who underwent GC for 3-5 years and 60 healthy control subjects without diabetes (HbA1c < 38.8 mmol/mol) were enrolled between June 2011 and July 2012. The patients were followed up for 4 years in average (March 2015 to November 2016) at Ishibashi Clinic, Hiroshima, Japan. The patients with diabetes were divided into the following three subgroups according to baseline HbA1c levels: subgroup 1; 58.5 mmol/mol < HbA1c \leq 74.9 mmol/mol, subgroup 2; 74.9 mmol/mol < HbA1c ≤ 91.3 mmol/mol, and subgroup 3; HbA1c > 91.3 mmol/mol, and also according to the mean HbA1c level during follow-up [good (HbA1c < 53.0 mmol/mol), fair (53.0 mmol/mol \leq HbA1c < 58.5 mmol/mol), and poor (HbA1c \geq 58.5 mmol/mol) control subgroups]. The patients assigned to the insulin-sensitizing or insulin-providing strategy were treated with biguanides or pioglitazone, or with sulfonylureas or insulin, respectively (15).

The HbA1c for patients was measured monthly during the follow-up period. And hence, the mean HbA1c levels during

follow-up are representative, and the linear regression provided changes in neuropathy outcomes by reduction in the mean HbA1c level per year of follow-up.

The exclusion criteria were any kind of other neuropathy apart from diabetes, vitamin B deficiency, and severe nonproliferative or proliferative diabetic retinopathy defined by Early Treatment Diabetic Retinopathy Study (ETDRS) (16), any type of corneal disease and history of refractive surgery, using contact lenses and significant media opacity at baseline and end point. Written informed consent was obtained from all subjects based on the Declaration of Helsinki. The ethics committee of Ishibashi Clinic approved the protocol of this study.

Corneal Confocal Microscopy

All study subjects were examined using a Heidelberg Retina Tomograph (III) in vivo CCM (Rostock Corneal module, Heidelberg Engineering, Heidelberg, Germany) (17). Patients with T2DM underwent CCM examination at baseline and end point. Control subjects underwent CCM examination once at the start of the study. A minimum of six high-quality images per subject from the sub-basal nerve plexus and Bowman's layer from the center of cornea was selected and quantified for the following CNF parameters: (1) corneal nerve fiber density (CNFD), the total number of major nerve fibers/mm² of corneal tissue; (2) corneal nerve fiber length (CNFL), the total length of all nerve fibers (mm/mm²); (3) corneal nerve branch density (CNBD), the number of branches emanating from all major nerve trunks/ mm²; (4) beading frequency (BF) (/0.1 mm); and (5) bead size (BS) (µm²) determined after enlarging five times and smoothing the original image of CCM using S-Spline Max algorithm (PhotoZoomPro4, Gungle Inc., Tokyo, Japan). The pixel numbers of 120 beads were counted using Photoshop Elements 8.0 (Adobe Systems Inc., San Jose, CA, USA) and averaged (18). Except for BS, all measurements were performed using ImageJ (Texelcraft, Tokyo, Japan).

Assessment of Neuropathy and Neurophysiological Examinations

All healthy control subjects and patients with T2DM underwent detailed neurophysiological examinations at baseline and final follow-up visits. The neurological deficits were assessed using the modified neuropathy disability score (NDS) (19), which includes evaluations of vibration perception, pin prick, temperature perception, and ankle reflexes. The classification for the evaluation of neuropathy was based on the Toronto consensus (20) which considered combination of symptoms (numbness or reduced ability to feel pain or temperature changes, a tingling or a burning sensation, sharp pains, increased sensitivity to touch, and muscle weakness), signs (a symmetric decrease in distal sensation and decreased or the absence of ankle reflexes), and electrophysiological tests. Hence, the patients with NDS >2 and sensory nerve conduction velocity (SCV) of sural nerve (SN) <42 m/s were labeled with neuropathy based on the Toronto criteria.

Electrophysiology and nerve conduction velocity (NCV) studies were performed using an electromyography instrument (Neuropak S1, NIHON KOHDEN, Tokyo, Japan). The motor nerve conduction velocity (MCV) of median nerve (MN) and

sensory (SCV, ulnar, and SN) NCVs and their action potential amplitudes were determined. Skin temperature was maintained above 32°C.

The vibration perception threshold (VPT) was measured at the left medial malleolus using a biothesiometer (Biomedical Instruments, Newbury, OH, USA). The warm and cold perception thresholds (PTs) at the dorsum of the foot were determined using a thermal stimulator (Intercross-200, Intercross Co., Tokyo, Japan). To assess cardiac autonomic neuropathy (CAN), the coefficient of variation of R–R intervals (CV_{R-R}) was calculated from the R–R intervals of 200 samples on electrocardiogram.

Clinical and Laboratory Data

The BMI, blood pressure, and HbA1c levels were measured monthly during the terms of study (48 months in average). The measured HbA1c levels were converted to National Glycohemoglobin Standardization Program units (21) and subsequently to International Federation of Clinical Chemistry values. The serum lipid levels [low-density lipoprotein (LDL) cholesterol (C), high-density lipoprotein (HDL)-C, and triglycerides], and urinary creatinine and albumin levels were assessed every 3 months. The albumin-to-creatinine ratio (ACR) >30 mg/ gCr twice a year was labeled as nephropathy (22).

At baseline and end point, bilateral retinal fundus images were captured (the field of assessment: 45°) and graded according to the ETDRS scale: no apparent retinopathy, 0; mild nonproliferative diabetic retinopathy, 1; moderate nonproliferative diabetic retinopathy, 2 (16).

Statistical Analysis

Statistical analyses were performed using SPSS ver. 19 (SPSS, Chicago, IL, USA). A p-value of <0.05 was considered to be statistically significant.

The post hoc analysis of sample power revealed that by using a one-sided ANOVA (significance of 0.05) and Kruskal-Wallis test for CNF measures and neurophysiological tests, the present study population provided statistical power ranging from 0.71 to 0.99. All values are presented as mean ± standard error of the mean (SEM). All data sets were tested for normality using the Shapiro-Wilk test. For normally distributed variables, comparisons between controls and patients with diabetes (baseline, end point, and mean value during follow-up) or between controls and each diabetic subgroups divided by the baseline or the mean HbA1c levels were performed using oneway ANOVA for continuous variables and the χ^2 -test for categorical variables followed by Bonferroni correction. For non-normally distributed variables, the Kruskal-Wallis test was applied followed by the Mann-Whitney U-test and Bonferroni correction for continuous variables and the McNemar test for categorical variables. The differences between baseline and end point were assessed using the paired t-test and Wilcoxon's signed rank test for normally and non-normally distributed continuous variables, respectively. Correlations between the neuropathy outcome measures and the mean clinical factors during follow-up or treatment strategies were assessed using Spearman's rank correlation coefficient or multiple regression analysis. The sensitivity, specificity, positive-predictive value

(PPV), and negative-predictive value (NPV) of CNF measures and neurophysiological tests in differentiating between control subjects and patients with or without neuropathy were assessed at baseline and end point using receiver-operating characteristic analysis.

RESULTS

Clinical, Neurophysiological, and CNF Measure Data of Controls and Patients with Type 2 Diabetes and Their Subgroups Based on Baseline HbA1c Levels At Baseline

The HbA1c levels were clearly different in three subgroups based on baseline results. The retinopathy was presented in 21.3% of total patients and between 18.2 and 23.4% in subgroups based on ETDRS scale (16). A total of 37.6% patients and 34.1-42.6% of subgroups had nephropathy based on the nephropathy definition by American Diabetes Association (22). The neuropathy was found to be 17.7% in total patients and 14.0-21.3% in subgroups based on Toronto criteria (20). Most neurophysiological tests (NDS, MCV and amplitude of MN, SCV and amplitude of ulnar nerve, SN amplitude, VPT, CV_{R-R}, warm and cold PTs, and most CNF measures) were impaired in patients or subgroups compared to those in controls (Table 1). Figure 1 shows representative corneal sub-basal nerve plexuses in a control subject (Figure 1A) and a patient with type 2 diabetes (Figure 1B). Compared with the control, the diabetic patient had reduced CNFD and increased tortuosity.

The BS in patient with type 2 diabetes (**Figure 1D**) is larger than that in control subject (**Figure 1C**).

At End Point

In all patients, the HbA1c level improved by average 30.6 mmol/ mol by tight GC. HbA1c levels in all subgroups were similar (53.0– 56.4 mmol/mol). In all patients and in particular subgroups 2 and 3, the retinopathy significantly increased and progressed; however, the nephropathy significantly decreased. The incidence of neuropathy did not significantly change in all subgroups. Overall, the GC was not beneficial for improving the neurophysiological functions and CNF parameters, and some neurophysiological functions rather deteriorated after GC (**Table 1**). Because GC did not improve the functions of large fibers assessed by NCVs and VPT, or neither of SNFs (CNF and temperature perceptions) in all patients, the differential impact by GC on large and SNFs was not obtained (**Table 1**).

After follow-up among the patient subgroups, despite strict GC and improvement in mean HbA1c by around 30.6 mmol/ mol, the cumulative incidence of neuropathy increased from 17.7% at baseline to 21.3% (p = 0.383) and retinopathy increased from 21.3 to 35.5% (p < 0.001); however, the cumulative incidence of nephropathy reduced from 37.6 to 22.0% (p < 0.001) (**Table 1**). Neuropathy decreased insignificantly in subgroup 3 (those with the poorest GC at baseline which suffered from severe neuropathy).

| | Control T2DM-Total Patients with type 2 diabetes stratified based on HbA1c level at base | | | | | | | evel at baseline | |
|--|--|----------------------------|------------------------------------|----------------------------|---------------------------|---|--------------------------|---------------------------------|-----------------------------|
| | Control subjects | T2DM | T2DM | Subgr | oup 1 | Subgro | oup 2 | Subgrou | p 3 |
| | | | | 58.5 < HbA1c m | \leq 74.9 mmol/ | 74.9 < HbA1c mo | ≤ 91.3 mmol/ ol | HbA1c > 91.3 mol | 3 mmol/ |
| | | Baseline | End point | Baseline | End point | Baseline | End point | Baseline | End point |
| Number (M/F) | 60 (40/20) | 141 (98/43) | 141 (98/43) | 50 (37/13) | 50 (37/13) | 44 (30/14) | 44 (30/14) | 47 (31/16) | 47 (31/16) |
| Age (years) | 53.1 ± 0.9 | 53.2 ± 0.7 | 57.1 ± 0.7 | 53.8 ± 1.0 | 57.7 ± 1.0 | 53.6 ± 1.2 | 57.7 ± 1.3 | 52.2 ± 1.5 | 55.9 ± 1.5 |
| Follow-up period (years) | | | 4.0 ± 0.1 | | 4.0 ± 0.2 | | 4.1 ± 0.2 | | 3.7 ± 0.1 |
| Duration of diabetes (years) | | 8.9 ± 0.6 | 12.9 ± 0.7 | 11.7 ± 1.1 | 15.7 ± 1.1 | 9.6 ± 1.2 | 13.6 ± 1.2 | $5.2 \pm 0.8^{*,\dagger}$ | $8.9\pm0.8^{\star,\dagger}$ |
| Smoking (%) | 26.7 | 31.9 | 24.8 | 38.0 | 30.0 | 29.5 | 20.5 | 27.7 | 23.4 |
| Alcohol consumption (%) | 40.0 | 36.9 | 37.6 | 52.0 | 50.0 | 40.9 | 40.9 | 17.0 | 21.3 |
| Body mass index (kg/m²) | 22.9 + 0.5 | $26.3 + 0.4^{\ddagger}$ | 26.4 + 0.4 | 26.2 + 0.6 [‡] | 26.4 + 0.7 | 26.7 + 0.7 [‡] | 26.6 + 0.8 | 26.0 + 0.7 [‡] | 26.2 + 0.7 |
| Systolic blood pressure (mmHa) | 132 ± 1.8 | $148 \pm 1.7^{\ddagger}$ | $139 \pm 0.8^{\circ}$ | 144 + 1 9 [‡] | 138 + 1.3" | $150 \pm 3.1^{\ddagger}$ | 140 ± 1.2^{1} | $150 \pm 3.7^{\ddagger}$ | 139 ± 1.6^{1} |
| Diastolic blood pressure (mmHg) | 77.8 ± 0.9 | $88.4 \pm 0.7^{\ddagger}$ | $80.3 \pm 0.6^{\circ}$ | 867 ± 0.9 [‡] | 80 2 ± 1 2 [§] | 89 1 + 1 2 [‡] | 80.9 ± 0.8§ | $90.0 \pm 1.7^{\ddagger}$ | 80.2 + 1.1§ |
| HbA1c (mmol/mol) | 36.8 ± 0.4 | 85.8 ± 1.8‡ | 55 2 ± 0 9 ^{‡.§} | $66.4 \pm 0.7^{\ddagger}$ | 56.3 ± 1.1 [‡] § | 82.6 ± 0.8 ^{±,*} | $56.4 \pm 2.0^{\pm 9}$ | $110 \pm 2.4^{\pm *^{\dagger}}$ | 53 0 ± 1 7 [‡] § |
| DL cholesterol (mmol/L) | 3.23 ± 0.09 | 3 66 ± 0 00# | 3.33 ± 0.08 | 3.40 ± 0.14 | 3.30 ± 0.14 | $3.78 \pm 0.15^{\#}$ | 3.48 ± 0.12 | 3.81 ± 0.17 | 3.21 ± 0.13 |
| HDL cholesterol (mmol/L) | 1.74 ± 0.06 | 1.47 ± 0.03 | 1.42 ± 0.00^{-1} | 1.40 ± 0.14 | 1.41 ± 0.06 | 1 44 + 0.05** | 1.40 ± 0.12 | 1.52 ± 0.07 | $1.42 \pm 0.07^{\circ}$ |
| | 1.74 ± 0.00 | $1.47 \pm 0.04^{\circ}$ | 1.43 ± 0.04 | 1.43 ± 0.03 | 1.41 ± 0.00 | 1.44 ± 0.00 | 1.44 ± 0.00 | 1.00 ± 0.07 | 1.42 ± 0.07 |
| Inglycendes (mmol/L) | 1.53 ± 0.14 | $2.45 \pm 0.16^{\circ}$ | 1.90 ± 0.12" | $2.17 \pm 0.20^{\circ}$ | 2.17 ± 0.22 | $2.40 \pm 0.30^{\circ}$ | 1.79 ± 0.15 | 2.79 ± 0.41 | 1.69 ± 0.24 |
| eGFR (mL/min) | 81.2 ± 1.9 | 87.7 ± 1.7" | /3.0±1.5° | 80.7 ± 2.2 | $71.1 \pm 2.3^{\circ}$ | 87.7 ± 2.8 | /b./ ± 2./* | $95.0 \pm 3.5^{-0.01}$ | $73.4 \pm 2.7^{\circ}$ |
| Albumin-creatinine ratio (mg/gCr) | 7.9 ± 1.2 | 94.9 ± 31.2‡ | 55.4 ± 16.6 [§] | 31.9 ± 5.9‡ | 41.7 ± 19.2 | 57.4 ± 19.4‡ | 33.2 ± 18.3 [¶] | 197 ± 90.3‡ | 90.8 ± 41.9§ |
| ETDRS retinopathy scale | 0 | 0.40 ± 0.06 | 0.62 ± 0.08§ | 0.48 ± 0.12 | 0.54 ± 0.13 | 0.34 ± 0.10 | 0.66 ± 0.13^{9} | 0.36 ± 0.11 | 0.66 ± 0.13¶ |
| Neurophysiological tests | | | | | | | | | |
| Neuropathy disability score $(0-10)$ | 0 | 42 ± 02 | 41 ± 02 | 37 ± 0.3 | 37 ± 0.3 | 46 ± 03 | 46 ± 0.3 | 44 + 04 | 40 ± 04 |
| MCV of modian nonvo (m/s) | 581,044 | 52.2 L 0.41 [‡] | 527 L 0.42 | 53.3 L 0.72 [‡] | 52.5 ± 0.76 | 51.5 ± 0.60 [‡] | 52.4 ± 0.71 | 52.1 ± 0.75‡ | 52 1 ± 0.74 |
| Amplitude of modian nonvo (m) | 7 08 1 0 21 | 52.0 ± 0.41 | 5 76 ± 0.42 | 6.00 ± 0.72 | 52.0 ± 0.70 | 51.0 ± 0.00 | 52.4 ± 0.71 | 6.12 ± 0.73 | 6.10 ± 0.74 |
| SOV of upper parts (m/s) | 7.90 ± 0.51 | $5.91 \pm 0.22^{\circ}$ | 5.70 ± 0.21 | $0.00 \pm 0.00^{\circ}$ | 5.12 ± 0.31 | $5.09 \pm 0.00^{\circ}$ | 5.50 ± 0.55 | 0.12 ± 0.09 | 0.19 ± 0.39 |
| | 03.9 ± 0.01 | $39.0 \pm 0.37^{+}$ | 39.0 ± 0.42 | $00.2 \pm 0.09^{+}$ | 39.3 ± 0.04 | $56.0 \pm 0.09^{+}$ | 30.4 ± 0.03 | $36.3 \pm 0.09^{+}$ | 39.2 ± 0.72 |
| Amplitude of ulnar herve (μv) | 30.5 ± 2.0 | 15.4 ± 0.53+ | 14.7 ± 0.51 | 16.4 ± 0.82+ | 15.9 ± 0.86 | 15.3 ± 0.99 ⁺ | 14.4 ± 0.91 | 14.3 ± 0.93+ | 13.7 ± 0.89 |
| SCV of sural nerve (m/s) | 46.9 ± 0.53 | 46.2 ± 0.39 | $45.3 \pm 0.43^{\circ}$ | 46.4 ± 0.65 | 45.2 ± 0.68 | 46.0 ± 0.72 | 44.8 ± 0.84 | 46.2 ± 0.67 | 45.9 ± 0.74 |
| Amplitude of sural nerve (μ V) | 14.5 ± 0.84 | $8.90 \pm 0.25^{+}$ | $8.26 \pm 0.28^{\circ}$ | $10.0 \pm 0.39^{**}$ | 9.35 ± 0.48 | $8.23 \pm 0.39^{+}$ | 7.58 ± 0.46 | $8.33 \pm 0.47^{+.++}$ | $7.74 \pm 0.48^{++}$ |
| VPT (µ/120 c/s) | 2.06 ± 0.24 | 3.73 ± 0.16 [‡] | 3.74 ± 0.19 | 3.51 ± 0.26 [‡] | 3.53 ± 0.25 | 3.97 ± 0.37 [‡] | 4.11 ± 0.48 | 3.74 ± 0.20 [∓] | 3.60 ± 0.24 |
| CV _{R-R} (%) | 3.90 ± 0.12 | 3.24 ± 0.10 [‡] | 3.04 ± 0.11 [¶] | 3.28 ± 0.15 [#] | 2.97 ± 0.16 | 3.18 ± 0.20 [#] | 2.79 ± 0.20 | 3.28 ± 0.18 [#] | 3.35 ± 0.19 |
| Warm perception threshold (W/m ²) | -512 ± 11.8 | -580 ± 11.0‡ | -596 ± 12.9 | -579 ± 21.3 | -597 ± 22.2 | -591 ± 18.4# | -600 ± 20.3 | -571 ± 17.1 | -572 ± 19.6 |
| Cold perception threshold (W/m²) | 478 ± 11.1 | $536 \pm 7.5^{\ddagger}$ | 545 ± 8.2" | 538 ± 13.9# | 557 ± 15.2 | 549 ± 13.1# | 558 ± 14.2 | 536 ± 7.5 | 519 ± 12.3 |
| Corneal nerve fiber measures | | | | | | | | | |
| Density (no/mm²) | 32.2 ± 0.74 | $17.6 \pm 0.5^{\ddagger}$ | 16.8 ± 0.5 | $18.5 \pm 0.87^{\ddagger}$ | 17.7 ± 0.79 | $17.1 \pm 0.90^{\ddagger}$ | 15.2 ± 0.88 | $17.1 \pm 0.94^{\ddagger}$ | 17.4 ± 0.98 |
| Length (mm/mm ²) | 15.4 ± 0.30 | $9.08 \pm 0.23^{\ddagger}$ | 8 83 + 0 22 | $9.62 \pm 0.41^{\ddagger}$ | 9.17 ± 0.37 | $8.79 \pm 0.39^{\ddagger}$ | 8.34 ± 0.37 | $8.77 \pm 0.40^{\ddagger}$ | 8.93 ± 0.42 |
| Branch density (no/mm²) | 13.2 ± 0.67 | 122 ± 0.40 | 122 ± 0.50 | 11.7 ± 0.79 | 9.90 ± 0.73 | $13.6 \pm 0.75^{\dagger\dagger}$ | $13.6 \pm 0.83^{++}$ | 11.6 ± 0.75 | 13.2 ± 0.90 |
| Brading frequency $(no/0.1 \text{ mm})$ | 10.2 ± 0.07 23.1 ± 0.30 | 10.7 ± 0.20‡ | 12.2 ± 0.00 19.6 ± 0.20 | $20.0 \pm 0.32^{\ddagger}$ | 19.50 ± 0.70 | 10.0 ± 0.70 $10.4 \pm 0.43^{\ddagger}$ | 10.0 ± 0.00 | $19.7 \pm 0.38^{\ddagger}$ | 20.3 ± 0.33 |
| Bead size (µm ²) | 7.78 ± 0.14 | $10.4 \pm 0.07^{\ddagger}$ | 10.2 ± 0.10 | $9.94 \pm 0.11^{\ddagger}$ | 10.2 ± 0.15 | $10.7 \pm 0.08^{\pm,*}$ | 10.4 ± 0.18 | $10.5 \pm 0.11^{\pm,*}$ | 10.1 ± 0.17 |
| | | | | | | | | | |
| Insulin providing (%) | | 55.3 | 66 7 | 66.0 | 76.0 | 56.8 | 70 5 | 42.6 | 53 2 ^{±‡} |
| Insulin consitizing (%) | | 44.0 | 75.0% | 66.0 | 80.0 | 42 Ott | 70.0 | 42.U 01.2* | 70.25 |
| $\frac{1}{2} \sum_{i=1}^{n} \frac{1}{2} \sum_{i=1}^{n} \frac{1}$ | | 44.0 | 10.20 | 00.0 | 0U.U 70.0 ⁸ | 40.2** | 12.1 " EC 06 | 21.0 | 12.0° E1 18 |
| | | 19.1 | 00.33 | 2ð.U | 1 2.Us | 10.9 | 20.83 | 12.8 | 51.1ª |
| | | | | | | | | | (Continued) |

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| | Control | T2DM- | - Total | Ра | tients with type 2 (| diabetes stratifiec | I based on HbA1c | evel at baseline |
|--------------------------------|------------------|----------|-------------------|-------------------|-----------------------|---------------------|--------------------|------------------------|
| | Control subjects | T2DM | T2DM | Subg | roup 1 | Subgi | roup 2 | Subgroup 3 |
| | | | | 58.5 < HbA1c π | : ≤ 74.9 mmol/ Iol | 74.9 < HbA1c m | ≤ 91.3 mmol/ ol | HbA1c > 91.3 mn mol |
| | | Baseline | End point | Baseline | End point | Baseline | End point | Baseline |
| Presence of microvascular comp | lications | | | | | | | |
| Presence of nephropathy (%) | 0 | 37.6 | 22.0 [§] | 36.0 | 24.0 | 34.1 | 15.9" | 42.6 |
| Presence of retinopathy (%) | 0 | 21.3 | 35.5 [§] | 22.0 | 28.0 | 18.2 | 40.91 | 23.4 |
| Presence of neuropathy (%) | 0 | 17.7 | 21.3 | 14.0 | 16.0 | 18.2 | 31.8 | 21.3 |

Data are the mean \pm standard error of the mean in the control subjects, at baseline, and end point in total patients with type 2 diabetes and their subgroups divided by baseline HbA1c levels.

p < 0.001 compared with subgroup 1, p < 0.01 compared with subgroup 2, p < 0.001 compared with control subjects, s > 0.001 compared with baseline, p < 0.05 compared with baseline, p < 0.05 compared with baseline. p < 0.05 compared with control subjects.

 $^{\dagger t}p < 0.01$ compared with subgroup 1, $^{\pm }p < 0.05$ compared with subgroup *p < 0.01 compared with control subjects,</p>

estimated glomerular filtration rate; ETDRS, Early Treatment Diabetic Retinopathy Study; HDL, high-density lipoprotein; LDL, low-density lipoprotein; DPP-4, dipeptidyl peptidase-4; eGFR, CV_{R-B} coefficient of variation of R-R interval;

sensory nerve conduction velocity; VPT, vibration perception threshold. motor nerve conduction velocity; SCV, WCV,

Clinical, Neurophysiological, and CNF Measure Data in the Subgroups of Patients with Type 2 Diabetes Stratified by the Mean HbA1c Levels during Follow-Up

Because the baseline HbA1c levels did not impact on neuropathy outcomes, we stratified patients by the mean HbA1c levels during follow-up.

At Baseline

The duration of diabetes in good GC subgroup was significantly shorter than others. The percentage of each original subgroup stratified by baseline HbA1c levels in good, fair, and poor control patients was not significantly different. The percentage of subgroup 3 in good control patients seemed to be larger than those in other two groups. The HbA1c levels in all subgroups were similar. Diabetic retinopathy was found more under fair and poor GC than under good GC. There was no difference in the prevalence of nephropathy and neuropathy, and the results of all neuropathy outcome measures between subgroups. Insulin-providing and insulin-sensitizing agents were more frequently prescribed under fair and poor GC than under good GC (Table 2). There was no difference in smoking habit (28.6-34.1%) and alcohol intake (31.8-39.6%) in all subgroups.

At End Point and Mean Levels

The HbA1c levels at end point and the mean levels (Figure 2A) were clearly different among subgroups. The decrease in HbA1c levels from baseline in the good GC subgroup was significantly larger than those in other GC subgroups. The retinopathy increased under poor GC. The nephropathy and neuropathy decreased under good GC. Good GC significantly improved all neurophysiological tests, while fair GC worsened some neurophysiological tests. Under poor GC, all neurophysiological functions deteriorated. Many neurophysiological functions under fair and poor GC were inferior to those under good GC (Table 2, Figure 3).

Even under good GC, CNFD (p = 0.750) and CNFL (p = 0.069) did not improve, while they were significantly higher than those under poor GC (p = 0.009 and 0.008, respectively). CNBD, BF, and BS were improved under good GC, and fair GC decreased BS. The poor GC worsened all CNF measures (p = <0.001-0.027) (Table 2; Figures 2B-F).

Under fair and poor GC, insulin-providing agents were more frequently prescribed compared with baseline. Insulin-sensitizing agents were more frequently prescribed than at baseline under good and fair GC. Dipeptidyl peptidase-4 (DPP-4) inhibitor was prescribed more frequently than at baseline in all subgroups and was prescribed less frequently under good GC than others (Table 2).

Associations between Changes in **Neuropathy Outcomes and Clinical Factors in All Patients**

The alterations in CNF parameters by GC were closely associated with the mean HbA1c level during follow-up. The duration of diabetes and the HDL-C level were negatively and positively associated with CNBD, respectively, but not other CNF

End point

mmol/

25.5" 38.3"

17.0

Presence of neuropathy (%)

TABLE 1 | Continued



measures. The mean HbA1c level had significant correlations with improvement in neurological dysfunctions in total patients. However, the HbA1c level at baseline did not significantly relate with CNF measures or neurophysiological tests (standard β : -0.139-0.05, p = 0.059-0.960). Except for CNFD (standard β : -159, p = 0.060) and ulnar nerve amplitude (standard β : -0.142, p = 0.098), changes in all other neuropathy outcome measures by GC significantly associated with the magnitude of reduction in HbA1c levels during follow-up (standard β : -0.197-0.368, $p = \langle 0.001 - 0.020 \rangle$. The duration of diabetes might worsen the MN MCV. The LDL-C level was positively associated with the VPT. The insulin-providing strategy deteriorated the nerve functions. The DPP-4 inhibitor impaired the SN SCV and amplitude (Table 3). The insulin-sensitizing strategy had no association with nerve functions. The mean SBP and DBP (standard β : -0.005 to -0.159, p = 0.962 - 0.194) had no significant association with interval changes in any neuropathy outcomes. The interval changes in HbA1c levels were correlated with the interval changes in CNF measures and neurophysiological tests (Figure 4).

One important finding in this study is that 10 mmol/mol reduction in HbA1c levels per year significantly improved CNF measures and neurophysiological dysfunctions (**Table 3**).

In 116 patients without neuropathy at baseline, 14 patients developed neuropathy after follow-up. When the clinical factors at baseline and their mean levels during follow-up were compared between patients with or without the development of neuropathy, the mean HbA1c levels (p < 0.001) during follow-up and the duration of diabetes (p = 0.004) were the most significant factors. None of patients in good GC subgroup developed neuropathy. Among the neuropathy outcome measures at baseline, the MN MCV (p = 0.039) and the SN SCV (p < 0.001) in patients developing neuropathy were slower than those in patients without neuropathy.

The Differential Abilities of CNF Measures and Neurophysiological Tests between Controls and Patients without Neuropathy and between Patients with or without Neuropathy

At baseline and end point, CNFD and CNFL had the excellent ability to differentiate between controls and patients without

neuropathy and better than CNBD (**Table 4**; **Figure 5A**). CNF measures lost differential ability between patients with or without neuropathy. The neurophysiological tests possessed modest differential ability between controls and patients without neuropathy, and the MN MCV, SN amplitude, and VPT were better than warm PT at baseline (**Figure 5B**). The neurophysiological test still possessed the modest differential ability between patients with or without neuropathy. They had good PPV between patients with or without neuropathy and good NPV between patients with or without neuropathy. The area under curve of most neuropathy outcome measures at baseline and end point appeared to be similar between controls and patients without neuropathy and between patients with or without neuropathy and patients without neuropathy and between patients with or without neuropathy and patients without neuropathy and between patients with or without neuropathy (**Table 4**).

DISCUSSION

The strict GC is the only strategy for slowing DPN progression in patients with type 2 diabetes. Thus, identifying potentially modifiable risk factors for neuropathy is crucial (23). In type 1 diabetes, cardiovascular risk factors are associated with the neuropathy incidence (23). It is also critical to identify the study cohort with mild to moderate DPN, because severe DPN may be less amenable to intervention (24). The current cohort had mild neuropathy at baseline (NDS: 4.2 ± 0.2). As the SNFs are most likely to show objective responses to interventions (25), SNF measures of DPN are essential to evaluate the efficacy of GC in clinical studies.

The corneal sub-basal nerve plexus is predominantly composed of SNFs (26), and CCM can quantify SNF pathology in diabetic neuropathy (27, 28). A recent longitudinal study in type 1 diabetes without neuropathy revealed that GC, HDL-C, and age impact on CNF measures (29). However, the risk factors associated with the altered CNF measures in type 2 diabetes have never been studied. Because in patients with long-standing type 1 diabetes baseline HbA1c levels predict the DPN development (30), we stratified the patients by the baseline HbA1c levels. HbA1c levels in subgroups decreased to 53.0-56.4 mmol/mol for 4 years. There was no difference in the improvement of neuropathy outcome measures between subgroups. However, the incidence of neuropathy decreased insignificantly in subgroup with the poorest control of HbA1c at baseline. The baseline HbA1c levels were not associated with the interval changes in neuropathy outcomes. The magnitude of reduction in HbA1c levels from baseline had a significant relationship with the changes in many neuropathy outcome measures, but the significance levels were less marked compared with the association with the mean HbA1c levels during followup. Then, we divided the patients into three subgroups according to the mean HbA1c level during follow-up. After follow-up, good GC reduced and poor GC increased the incidence of neuropathy. Under good GC, some CNF measures and all neurophysiological functions improved. The significantly larger decrease in HbA1c levels from baseline in good control subgroup due to relatively more inclusion of subgroup-3 members (patients with the highest HbA1c level at baseline) may contribute to the improvement in neuropathy outcomes. The fair control reduced BS but worsened some neurophysiological functions. This might be due to insufficient GC level in this subgroup. Because the mean HbA1c levels

TABLE 2 | Clinical characteristics, neurophysiological tests, corneal nerve fiber measures, and hypoglycemic strategies at baseline, at end point, and mean values in the diabetic subgroups divided by mean HbA1c levels during follow-up period.

| | | iod | | | | | | | |
|---|-----------------|-------------------------|-----------------|-------------------|--------------------------------------|------------------|-------------------------|-----------------------------|----------------------------|
| | | Good control | | | Fair control | | I | Poor control | |
| | HbA1 | c < 53.0 mmol/m | ol | 53.0 ≤ H | lbA1c < 58.5 mmol | /mol | HbA1c | ≥ 58.5 mmol/mo | I |
| | Baseline | End point | Mean | Baseline | End point | Mean | Baseline | End point | Mean |
| Number (M/F) | 48 (33/15) | 48 (33/15) | | 49 (35/14) | 49 (35/14) | | 44 (30/14) | 44 (30/14) | |
| Subgroups 1/2/3 (%) | 27.1/29.2/43.7 | | | 42.8/28.6/28.6 | | | 36.4/36.4/27.2 | | |
| Age (years) | 51.9 ± 1.4 | 55.7 ± 1.4 | 53.8 ± 1.4 | $56.1 \pm 1.1^*$ | $60.3 \pm 1.1^*$ | 58.2 ± 1.1* | 51.3 ± 1.2 ⁺ | 55.1 ± 1.2‡ | 53.4 ± 1.3† |
| Follow-up period (years) | | 3.8 ± 0.1 | | | 4.2 ± 0.2 | | | 4.0 ± 0.1 | |
| Duration of diabetes (years) | 4.2 ± 0.6 | 7.9 ± 0.6 | 6.1 ± 0.6 | $9.6 \pm 1.2^*$ | $13.8 \pm 1.2^*$ | $11.7 \pm 1.2^*$ | 13.1 ± 1.1§ | $17.0 \pm 1.1^{\circ}$ | 15.1 ± 1.1 [§] |
| Body mass index (kg/m ²) | 26.3 ± 0.7 | 25.3 ± 0.6 | 24.9 ± 0.6 | 26.2 ± 0.7 | 26.6 ± 0.8 | 26.1 ± 0.8 | 26.3 ± 0.7 | 27.3 ± 0.8" | 26.9 ± 0.7 |
| Systolic blood pressure (mmHg) | 148 ± 3.2 | 137 ± 1.1¶ | 137 ± 1.1 | 151 ± 2.9 | 140 ± 1.4" | 140 ± 1.1 | $143 \pm 2.5^{+}$ | 139 ± 1.6 | 141 ± 1.1 |
| Diastolic blood pressure (mmHg) | 89.4 ± 1.4 | 79.8 ± 1.0 [#] | 81.7 ± 0.9 | 89.6 ± 1.4 | $80.0 \pm 1.1^{\#}$ | 82.1 ± 0.8 | 85.9 ± 1.0 | 81.6 ± 0.9" | 83.5 ± 0.6 |
| HbA1c (mmol/mol) | 88.2 ± 2.7 | $47.4 \pm 0.7^{\#}$ | 47.5 ± 0.5 | 84.5 ± 3.2 | $53.9 \pm 0.5^{\text{\#},\text{\$}}$ | 55.6 ± 0.2§ | 85.2 ± 3.3 | 64.4 ± 2.2 ^{#,§,‡} | 68.9 ± 1.7 ^{§,**} |
| Decrease in HbA1c from baseline (mmol/mol) | | -40.8 ± 3.0 | | | $-30.6 \pm 3.4^{*}$ | | | -20.8 ± 3.9§ | |
| LDL cholesterol (mmol/L) | 3.81 ± 0.15 | 3.28 ± 0.12# | 3.27 ± 0.10 | 3.68 ± 0.17 | 3.16 ± 0.14" | 3.19 ± 0.11 | 3.44 ± 0.14 | 3.56 ± 0.13 | 3.50 ± 0.11 |
| HDL cholesterol (mmol/L) | 1.37 ± 0.06 | 1.40 ± 0.08 | 1.40 ± 0.05 | 1.49 ± 0.06 | 1.43 ± 0.06 | 1.45 ± 0.06 | 1.55 ± 0.07 | 1.44 ± 0.06 | 1.47 ± 0.06 |
| Triglycerides (mmol/L) | 2.43 ± 0.25 | 2.00 ± 0.24 | 1.81 ± 0.13 | 2.42 ± 0.25 | 1.89 ± 0.20 | 1.76 ± 0.12 | 2.50 ± 0.43 | 1.98 ± 0.19 | 1.96 ± 0.23 |
| Presence of nephropathy (%) | 27.1 | 10.4 [¶] | | 38.8 | 26.5 | | 47.7 | 29.5 | |
| Presence of retinopathy (%) | 4.2 | 10.4 | | 24.5* | 34.7 | | 36.4§ | 63.6 ^{#,§} | |
| Presence of neuropathy (%) | 18.8 | 6.3 [¶] | | 16.3 | 18.4 | | 18.2 | 40.9",§,† | |
| Neurophysiological tests | | | | | | | | | |
| MCV of median nerve (m/s) | 53.3 ± 0.51 | 55.9 ± 0.51# | | 51.6 ± 0.79 | 51.4 ± 0.78§ | | 52.1 ± 0.78 | 50.5 ± 0.62 ^{§,"} | |
| Amplitude of median nerve (mV) | 5.92 ± 0.38 | 6.41 ± 0.37¶ | | 5.95 ± 0.35 | 5.68 ± 0.37 | | 5.86 ± 0.41 | 5.14 ± 0.38*," | |
| SCV of ulnar nerve (m/s) | 59.5 ± 0.55 | 61.7 ± 0.60# | | 58.5 ± 0.64 | $58.4 \pm 0.66^{++}$ | | 59.0 ± 0.72 | 56.8 ± 0.74 ^{§,#} | |
| Amplitude of ulnar nerve (μ V) | 16.5 ± 0.98 | 17.9 ± 0.91# | | 14.6 ± 0.90 | 13.9 ± 0.81* | | 15.0 ± 0.84 | 12.1 ± 0.74 ^{§,#} | |
| SCV of sural nerve (m/s) | 46.3 ± 0.74 | 47.6 ± 0.72# | | 46.4 ± 0.70 | 45.0 ± 0.74# | | 45.9 ± 0.56 | 43.1 ± 0.63#,§ | |
| Amplitude of sural nerve (μV) | 8.82 ± 0.39 | 9.43 ± 0.44 | | 9.04 ± 0.48 | 8.40 ± 0.51# | | 8.82 ± 0.44 | 6.82 ± 0.44 ^{#,§} | |
| VPT (µ/120 c/s) | 3.60 + 0.24 | 2.85 + 0.22# | | 3.80 + 0.24 | 3.82 + 0.25* | | 3.80 + 0.36 | 4.62 + 0.46 ^{#,§} | |
| CV _{B-R} (%) | 3.24 + 0.16 | 3.53 + 0.18# | | 3.23 ± 0.21 | 3.04 + 0.21",†† | | 3.27 ± 0.15 | 2.52 + 0.13 ^{#,§} | |
| Warm perception threshold (W/m ²) | -564 + 14.8 | $-520 + 13.2^{\#}$ | | -593 + 23.0 | -618 + 24.0 ^{#,††} | | -582 + 18.3 | -633 + 19.7 ^{#,§} | |
| Cold perception threshold (W/m²) | 532 ± 12.8 | 497 ± 13.3 [#] | | 544 ± 14.1 | 557 ± 13.4",†† | | 532 ± 11.7 | 583 ± 12.9 ^{#,§} | |
| Corneal nerve fiber measures | | | | | | | | | |
| Density (no/mm²) | 17.9 ± 0.9 | 18.2 ± 0.7 | | 18.3 ± 1.0 | 17.6 ± 0.9 | | 16.4 ± 0.8 | 14.5 ± 1.0 ^{++,} " | |
| Length (mm/mm²) | 9.01 ± 0.34 | 9.44 ± 0.33 | | 9.31 ± 0.46 | 9.20 ± 0.40 | | 8.90 ± 0.39 | 7.75 ± 0.40 ^{++,#} | |
| Branch density (no/mm ²) | 11.4 + 0.6 | 13.6 + 0.9# | | 12.0 + 0.9 | 12.3 ± 0.9 | | 13.4 + 0.8 | $10.5 + 0.7^{\#}$ | |
| Beading frequency (no/0.1 mm) | 19.6 + 0.3 | 20.9 + 0.3 | | 19.8 + 0.4 | 19.7 ± 0.2 | | 19.7 + 0.3 | 18.1 + 0.3 ^{#,§,‡} | |
| Bead size (µm²) | 10.4 ± 0.1 | $9.4 \pm 0.1^{\#}$ | | 10.5 ± 0.1 | 10.2 ± 0.1^{1} | | 10.3 ± 0.1 | 11.2 ± 0.2 ^{#,§,†} | |
| Hypoglycemic strategies | | | | | | | | | |
| Insulin providing (%) | 22.9 | 8.3 | | 63.3 [§] | 93.9 ^{§,#} | | 81.8§ | 100 ^{§,"} | |
| Insulin sensitizing (%) | 18.8 | 77.1# | | 49.0 [§] | 71.4¶ | | 65.9§ | 77.3 | |
| DPP-4 inhibitor (%) | 8.3 | 39.6# | | 26.5 | 65.3*,# | | 22.7 | 77.3 ^{§,#} | |

Data are the mean ± standard error of the mean at baseline, at end point, and mean values during follow-up in the subgroups of patients with type 2 diabetes divided by the mean HbA1c levels during follow-up period.

*p < 0.05 compared with good control subgroup, *p < 0.05 compared with fair control subgroup, *p < 0.01 compared with fair control subgroup, *p < 0.01 compared with good control subgroup, *p < 0.01 compared with baseline, *p < 0.05 compared with baseline, *p < 0.001 compared with fair control subgroup, *p < 0.01 compared with good control subgroup.

CV_{R-R}, coefficient of variation of R-R interval; DPP-4, dipeptidyl peptidase-4; HDL, high-densityl ipoprotein; LDL, low-densityl ipoprotein; MCV, motor nerve conduction velocity; SCV, sensory nerve conduction velocity; VPT, vibration perception threshold.

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bead size (F) between control subjects, baseline, and end point among subgroups stratified by the mean HbA1c levels during follow-up. Hatched column; control subjects, open column: at baseline, solid column: mean level (A) or at end point (B–F). *p < 0.001 compared with good control subgroup, *p < 0.01 compared with good control subgroup, *p < 0.01 compared with baseline, *p < 0.021 compared with baseline,

during follow-up in fair control subgroup were $55.6 \pm 0.2 \text{ mmol}/\text{mol}$, this GC level was similar or inferior to those in clinical trials which are not beneficial for the amelioration of DPN (1, 31). Poor GC deteriorated most neuropathy outcome measures. These results indicated that the near-normoglycemia (HbA1c: 47.5 mmol/mol) achieved by GC is beneficial for neuropathy, while the mean HbA1c level higher than 53.0 mmol/mol in fair control subgroup and at end point HbA1c levels in total and subgroups stratified by the baseline HbA1c are not effective to prevent the deterioration of neuropathy outcomes.

In the Kumamoto study (31), the tight GC (HbA1c: 54.1 mmol/ mol) prevented NCV but not CAN decline in type 2 diabetes. In the United Kingdom Prospective Diabetes Study (UKPDS) (1), the intensive (HbA1c: 53.0 mmol/mol) and conventional GC (HbA1c: 62.8 mmol/mol) had a similar effect on DPN and CAN. In the ACCORD (Action to Control Cardiovascular Risk in Diabetes) trial (3), the intensive treatment (HbA1c: 45.4 mmol/ mol) prevented loss of ankle jerk and light-touch sensation, but resulted in an increased total and CVD-related mortality and severe hypoglycemia. Furthermore, reliable neurophysiological tests were not included. These randomized trials could not establish the optimum GC level for preventing the deterioration of neuropathy outcomes in type 2 diabetes and did not evaluate the SNF morphology.

When patients with recent onset type 1 diabetes were followed up under near-normoglycemia (HbA1c: 47.5 mmol/ mol) for 24 years, the decline in nerve functions was completely prevented (32). Although the HbA1c levels of <53.0 mmol/mol is considered to be a reasonable treatment goal (33), more stringent HbA1c goals (\leq 47.5 mmol/mol) are suggested for selected patients if achievable without hypoglycemia (32). Therefore, the near-normoglycemia is prerequisite for preventing neurophysiological deterioration. These findings were compatible with our study showing that all neurophysiological tests were improved under near-normoglycemia (mean HbA1c: 47.5 mmol/mol).

In patients with type 1 diabetes receiving simultaneous pancreas-kidney transplantation, euglycemia improved CNF measures within 12 months after transplantation (34). In type 1 diabetes, CNFD and CNFL, and BF alter related to the annual mean HbA1c level for previous 7–10 years and 1 year before CCM examination, respectively (17). The CNF measures may improve with different metabolic memory after the establishment of GC.



An improved HbA1c level was associated with repair in CNFD but not in CNBD in a small mixed cohort of type 1 and type 2 diabetes (35), indicating the different sensitivities to GC among CNF measures. Because the above study included a small number of type 1 diabetes-dominant patients, their results were different from our study.

As BF and BS showed better association with the mean HbA1c level than other CNF measures, the beads may be more responsible to strict GC than other CNF measures. The beads visualized by CCM are composed of the accumulated mitochondria, glycogen particles, and vesicles (26), and mitochondria play a pivotal role in nerve conduction *via* their adenosine triphosphate production in nerve fibers. We already reported that the hyperglycemia-induced expansion of beads in CNFs might be a predictor of slow NCV in patients with type 2 diabetes (18). However, further investigations are required to evaluate the pathophysiological role of altered beading structure in developing DPN. The near-normoglycemia did not improve CNFD and CNFL, but prevented further declines under persistent hyperglycemia.

As the GC strategy influences DPN (15), we examined the impact of treatment strategies on the changes in neuropathy outcomes by GC. The infrequency of insulin-providing agents and DPP-4 inhibitors in the good GC subgroup than in others may contribute to the tight GC without hypoglycemia. As the insulin-providing strategy was detrimental to most neurophysiological

functions, the impact of GC on the neuropathy outcomes should be assessed with careful attention to treatment strategies.

In good GC subgroup, BMI significantly decreased after GC, and hence, good response to lifestyle intervention may improve the neuropathy outcomes (6). Among the components of metabolic syndrome affecting DPN (36), dyslipidemia had a marginal impact on neuropathy outcome measures. Smoking and alcohol consumption, which may influence on DPN (36), had no impact on DPN.

Because the performance and area under curve of most CNF parameters at baseline and end point for differentiating between controls and patients with or without neuropathy were satisfactory, the assessment of neuropathy outcome measures at baseline may predict the subsequent changes especially in those with more severe damage and those that are the subjects that may further benefit from GC. Because CNFD and CNFL excellently differentiated between controls and patients without neuropathy, these are the best measures for detecting the early DPN. Neurophysiological tests had modest sensitivity and specificity. Before and after developing mild neuropathy, high PPV and NPV were found, respectively, suggesting that the neurophysiological tests contribute to the sensitive diagnosis in the early stage and reliable diagnosis after developing neuropathy.

In the current study, with optimal GC, nephropathy decreased, while the retinopathy increased. The initial extreme

TABLE 3 | Correlation between interval changes by glycemic control in measures of corneal nerve fibers or nerve functions and mean clinical factors during follow-up or hypoglycemic strategies, and the slope of the linear regression of the interval changes in neuropathy outcome measures by 10 mmol/mol HbA1c reduction per year in patients with type 2 diabetes.

| Interval changes in | | | Mean clinica | l factors d | uring follow-up | o period | | | H | ypoglycen | nic strategy | | | |
|--------------------------------|------------------------|-----------------|--------------|-------------|-----------------|----------|-----------------|-------|-------------------|-----------|------------------|-------|---|--------|
| neuropathy outcome measures | Duratior diabetes m | n of ellitus | HbA1c | | LDL cholesterol | | HDL cholesterol | | Insulin providing | | DPP-4 inhibitor | | Linear regression by 10 mmol/mol annual HbA1c reduction | |
| | Standard β | р | Standard β | р | Standard β | p | Standard β | p | Standard β | р | Standard β | р | Slope | р |
| Corneal nerve fiber | | | | | | | | | | | | | | |
| Density | -0.063 | 0.530 | -0.267 | 0.023 | -0.069 | 0.459 | 0.075 | 0.379 | 0.039 | 0.723 | 0.035 | 0.677 | 0.096 | 0.015 |
| Length | -0.077 | 0.442 | -0.260 | 0.025 | -0.036 | 0.697 | 0.037 | 0.661 | -0.050 | 0.648 | 0.068 | 0.418 | 0.049 | 0.002 |
| Branch density | -0.206 | 0.023 | -0.242 | 0.021 | -0.127 | 0.128 | 0.264 | 0.001 | -0.076 | 0.442 | -0.131 | 0.088 | 0.163 | <0.001 |
| Beading frequency | -0.147 | 0.120 | -0.316 | 0.004 | -0.170 | 0.052 | -0.033 | 0.679 | -0.009 | 0.932 | -0.017 | 0.828 | 0.070 | 0.002 |
| Bead size | 0.147 | 0.070 | 0.399 | <0.001 | -0.006 | 0.931 | -0.063 | 0.354 | 0.180 | 0.043 | 0.151 | 0.029 | -0.045 | <0.001 |
| Nerve functions | | | | | | | | | | | | | | |
| MCV of median nerve | -0.240 | 0.010 | -0.238 | 0.027 | -0.070 | 0.407 | -0.012 | 0.877 | -0.116 | 0.251 | -0.021 | 0.791 | 0.136 | <0.001 |
| Amplitude of ulnar nerve | 0.112 | 0.208 | -0.376 | <0.001 | -0.033 | 0.681 | 0.102 | 0.173 | -0.290 | 0.003 | -0.022 | 0.774 | 0.056 | 0.007 |
| SCV of sural nerve | -0.034 | 0.649 | -0.425 | <0.001 | 0.068 | 0.330 | 0.013 | 0.835 | -0.296 | <0.001 | -0.181 | 0.005 | 0.101 | <0.001 |
| Amplitude of sural nerve | -0.095 | 0.234 | -0.371 | <0.001 | -0.109 | 0.140 | 0.068 | 0.310 | -0.220 | 0.013 | -0.171 | 0.012 | 0.038 | 0.001 |
| Vibration perception threshold | 0.149 | 0.060 | 0.398 | <0.001 | 0.213 | 0.004 | -0.074 | 0.263 | 0.175 | 0.044 | 0.004 | 0.953 | -0.033 | <0.001 |
| Warm perception threshold | -0.084 | 0.338 | -0.254 | 0.014 | 0.015 | 0.857 | 0.094 | 0.205 | -0.322 | 0.001 | -0.003 | 0.960 | 1.91 | 0.001 |
| Cold perception threshold | 0.101 | 0.231 | 0.357 | <0.001 | -0.024 | 0.760 | -0.042 | 0.556 | 0.215 | 0.022 | 0.087 | 0.227 | -2.17 | <0.001 |

Bold p value reveals p < 0.05.

DPP-4, dipeptidyl peptidase-4; HDL, high-density lipoprotein; LDL, low-density lipoprotein; MCV, motor nerve conduction velocity; SCV, sensory nerve conduction velocity.



hyperglycemia paired with rapid and substantial HbA1c reduction might develop the retinopathy (37). The differences from previous studies in the impact of GC on microangiopathy may originate from the differences in patient characteristics (race, age, BMI, duration of diabetes, and HbA1c levels) at baseline (1-3, 31). Perhaps, based on these results, we can control the neuropathy and nephropathy, but not retinopathy. The duration of diabetes in good control subgroup was shorter than that in others. This may contribute to the difference in metabolic memory. Several epidemiological studies (38) suggest that an early intensive GC can reduce the risk of diabetic microangiopathy. The duration of diabetes at baseline was 8.9 years in the current study, and this is an important factor. The emergence of metabolic memory suggests the need for an early aggressive treatment aiming to normalize the metabolic control for minimizing diabetic complications (38).

TABLE 4 | Differential abilities of corneal nerve fiber (CNF) measures and neurophysiological tests between control subjects and diabetic patients without neuropathy, or between diabetic patients with or without neuropathy at the baseline and end point.

| | | Control s | ubjects vs. | diabetic pati at baselin | ents without e | Diabet | ic patients | atients with vs. without neuropathy at baseline | | | | | | | |
|--------------------------------------|-----------|--------------------|-----------------|-----------------------------|-------------------|------------|-------------|---|-----------------|-------------|-------------|------------|------------|--|--|
| | | AUC 95%CI | <i>p</i> -Value | Sensitivity | Specificity | PPV (%) | NPV (%) | AUC 95% CI | <i>p</i> -Value | Sensitivity | Specificity | PPV (%) | NPV (%) | | |
| CNF measures | | | | | | | | | | | | | | | |
| CNF density | Baseline | 0.96 0.93–0.98 | <0.001 | 0.86 | 0.93 | 96.1 | 77.8 | 0.61 0.49–0.73 | 0.061 | 0.60 | 0.66 | 27.3 | 88.4 | | |
| | End point | 0.97 0.95–0.99 | <0.001 | 0.92 | 0.93 | 96.4 | 86.1 | 0.51 0.37–0.64 | 0.916 | 0.48 | 0.66 | 23.1 | 85.4 | | |
| CNF length | Baseline | 0.96 0.93–0.98 | <0.001 | 0.94 | 0.87 | 93.2 | 88.2 | 0.62 0.50–0.74 | 0.051 | 0.72 | 0.59 | 27.3 | 90.7 | | |
| | End point | 0.97 0.95–0.99 | <0.001 | 0.93 | 0.92 | 95.6 | 87.3 | 0.59 0.47–0.71 | 0.160 | 0.52 | 0.66 | 24.5 | 86.4 | | |
| Corneal nerve branch density | Baseline | 0.55 0.47–0.64 | 0.241 | 0.40 | 0.78 | 78.0 | 40.2 | 0.52 0.39–0.65 | 0.806 | 0.44 | 0.66 | 21.6 | 84.4 | | |
| | End point | 0.59 0.50–0.67 | 0.048 | 0.44 | 0.78 | 79.7 | 42.0 | 0.54 0.41–0.67 | 0.525 | 0.68 | 0.50 | 22.7 | 87.9 | | |
| Neurophysiological t | ests | | | | | | | | | | | | | | |
| Motor nerve conduction velocity of | Baseline | 0.84 0.78–0.90 | <0.001 | 0.72 | 0.85 | 90.2 | 60.8 | 0.65 0.51–0.78 | 0.031 | 0.60 | 0.76 | 34.9 | 89.8 | | |
| median nerve (MN) | End point | 0.82 0.75–0.88 | <0.001 | 0.70 | 0.85 | 90.0 | 59.3 | 0.58 0.45–0.71 | 0.209 | 0.56 | 0.63 | 24.6 | 86.9 | | |
| Amplitude of MN | Baseline | 0.70 0.62–0.78 | <0.001 | 0.55 | 0.68 | 77.1 | 44.1 | 0.62 0.50–0.75 | 0.059 | 0.60 | 0.66 | 27.8 | 88.5 | | |
| | End point | 0.73 0.65–0.80 | <0.001 | 0.57 | 0.78 | 83.5 | 48.4 | 0.55 0.44–0.67 | 0.374 | 0.56 | 0.60 | 23.0 | 86.3 | | |
| Sensory nerve conduction velocity of | Baseline | 0.53 0.44–0.63 | 0.511 | 0.72 | 0.40 | 70.0 | 42.8 | 1.00 1.00–1.00 | <0.001 | 1.00 | 0.98 | 92.7 | 100 | | |
| sural nerve (SN) | End point | 0.53 0.45–0.62 | 0.443 | 0.54 | 0.60 | 72.4 | 40.4 | 0.91* 0.85–0.96 | <0.001 | 0.84 | 0.83 | 51.3 | 96.0 | | |
| Amplitude of SN | Baseline | 0.79 0.71–0.87 | <0.001 | 0.76 | 0.73 | 84.6 | 61.1 | 0.68 0.56–0.80 | 0.004 | 0.56 | 0.78 | 35.0 | 89.1 | | |
| | End point | 0.81* 0.74–0.89 | <0.001 | 0.81 | 0.75 | 86.2 | 67.1 | 0.68 0.56–0.79 | 0.004 | 0.64 | 0.66 | 28.6 | 89.4 | | |
| Vibration perception threshold (PT) | Baseline | 0.80 0.72–0.88 | <0.001 | 0.81 | 0.73 | 85.4 | 66.6 | 0.55 0.42–0.68 | 0.446 | 0.60 | 0.58 | 23.5 | 87.0 | | |
| | End point | 0.79 0.71–0.87 | <0.001 | 0.78 | 0.75 | 85.7 | 63.4 | 0.51 0.39–0.64 | 0.829 | 0.56 | 0.53 | 20.3 | 84.7 | | |
| Warm PT | Baseline | 0.64 0.56–0.73 | <0.001 | 0.66 | 0.55 | 73.8 | 45.2 | 0.50 0.38–0.62 | 0.989 | 0.52 | 0.52 | 18.8 | 83.3 | | |
| | End point | 0.64 0.55–0.72 | 0.002 | 0.53 | 0.70 | 77.2 | 43.3 | 0.52 0.40–0.64 | 0.704 | 0.64 | 0.47 | 20.5 | 85.7 | | |
| Cold PT | Baseline | 0.65 0.57–0.74 | <0.001 | 0.65 | 0.57 | 74.3 | 45.4 | 0.53 0.39–0.66 | 0.700 | 0.56 | 0.63 | 24.6 | 86.9 | | |
| | End point | 0.67 0.59–0.75 | <0.001 | 0.67 | 0.57 | 75.0 | 47.2 | 0.53 0.40–0.65 | 0.691 | 0.56 | 0.56 | 21.5 | 85.5 | | |

 $^{*}p < 0.01$ compared with baseline.

Bold p value reveals p < 0.05.

AUC, area under curve; CI, confidence interval; NPV, negative predictive value; PPV, positive predictive value.

Strengths and Limitations

The present study measured the main modifiable risk factors including HbA1c level, blood pressure, and BMI monthly, and the lipid profile, estimated glomerular filtration rate, and ACR every 3 months. Therefore, the mean clinical factors impacting

on DPN are representative, and the contribution of HbA1c levels attained by GC and other risk factors on neuropathy outcome measures and other microangiopathies was reliably evaluated. We estimated the morphological and functional measures of SNFs. The altered CNFs were found in all patients with type 2



perception threshold; SN, sural nerve; VPT, vibration perception threshold.

diabetes at baseline, and near-normoglycemia improved some CNF parameters and all neurophysiological tests. The present study had some limitations. First, we assessed neuropathy outcome measures only at baseline and end point. Ideally, these examinations should be performed annually for assessing the trends of their changes. Second, the development of DPN depends on the metabolic memory (4). Our follow-up period might be too short to reflect the metabolic memory on changes in neuropathy outcome measures. Therefore, we could not detect the impact of baseline HbA1c levels on neuropathy outcomes. Third, this was a retrospective cohort study with a relatively small number of participants. Future prospective studies using a larger cohort are required to consolidate the present results. Finally, the process of selecting CCM images can be more robust to avoid bias. In the current study, based on established protocol and to avoid overlapping of images, we selected at least six high-quality images per subject (39). However, the averaging of parameter values based on multiple CCM images does not necessarily result in good approximations of the respective reference values of the whole image area, indicating the potential for the inevitable local bias when selecting CCM images in the central corneal area (40). Furthermore, cornea is an avascular tissue. In patients with type 2 diabetes, CNFL and CNBD were inversely correlated with tear levels of insulin-like growth factor-binding protein 3 which sequestrates insulin-like growth factor 1 (41). The corneal epithelium-derived neurotrophic factors promote corneal nerve regeneration in mice (42). We acknowledged that there exists a controversy of whether the response of CNF to diabetes or GC is same as that in peripheral sensory nerves in extremities.

In conclusion, the HbA1c level closer to 47.5 mmol/mol achieved mainly with insulin-sensitizing agents and lifestyle

modification would be a safe glycemic goal for improving the outcome measures of DPN in patients with poorly controlled type 2 diabetes with mild to moderate neuropathy. The near-normoglycemia is effective for preventing the development of neuropathy and nephropathy, but not retinopathy. However, achieving this is not feasible in many patients.

ETHICS STATEMENT

Written informed consent was obtained from all subjects based on the Declaration of Helsinki. The ethics committee of Ishibashi Clinic approved the protocol of this study.

AUTHOR CONTRIBUTIONS

FI designed the study, researched data, and wrote the entire manuscript. MT advised on the statistical analysis, interpreted the results, contributed to writing, and reviewed and revised the whole manuscript. FI and MT are the guarantors of this work and, as such, had full access to all data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis and interpretation.

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REFERENCES

- 1. UK Prospective Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* (1998) 352(9131):837–53. doi:10.1016/S0140-6736(98)07019-6
- ADVANCE Collaborative Group, Patel A, MacMahon S, Chalmers J, Neal B, Billot L, et al. Intensive blood glucose control and vascular outcomes in patients with type 2 diabetes. N Engl J Med (2008) 358(24):2560–72. doi:10.1056/ NEJMoa0802987
- Ismail-Beigi F, Craven T, Banerji MA, Basile J, Calles J, Cohen RM, et al. Effect of intensive treatment of hyperglycaemia on microvascular outcomes in type 2 diabetes: an analysis of the ACCORD randomised trial. *Lancet* (2010) 376(9739):419–30. doi:10.1016/S0140-6736(10)60576-4
- Ang L, Jaiswal M, Martin C, Pop-Busui R. Glucose control and diabetic neuropathy: lessons from recent large clinical trials. *Curr Diab Rep* (2014) 14(9):528. doi:10.1007/s11892-014-0528-7
- Sumner CJ, Sheth S, Griffin JW, Cornblath DR, Polydefkis M. The spectrum of neuropathy in diabetes and impaired glucose tolerance. *Neurology* (2003) 60(1):108–11. doi:10.1212/WNL.60.1.108
- Smith AG, Russell J, Feldman EL, Goldstein J, Peltier A, Smith S, et al. Lifestyle intervention for pre-diabetic neuropathy. *Diabetes Care* (2006) 29(6):1294–9. doi:10.2337/dc06-0224
- Smith AG, Howard JR, Kroll R, Ramachandran P, Hauer P, Singleton JR, et al. The reliability of skin biopsy with measurement of intraepidermal nerve fiber density. *J Neurol Sci* (2005) 228(1):65–9. doi:10.1016/j.jns.2004.09.032
- Tavakoli M, Petropoulos IN, Malik RA. Corneal confocal microscopy to assess diabetic neuropathy: an eye on the foot. J Diabetes Sci Technol (2013) 7(5):1179–89. doi:10.1177/193229681300700509
- Tavakoli M, Quattrini C, Abbott C, Kallinikos P, Marshall A, Finnigan J, et al. Corneal confocal microscopy. *Diabetes Care* (2010) 33(8):1792–7. doi:10.2337/dc10-0253
- Ziegler D, Papanas N, Zhivov A, Allgeier S, Winter K, Ziegler I, et al. Early detection of nerve fiber loss by corneal confocal microscopy and skin biopsy in recently diagnosed type 2 diabetes. *Diabetes* (2014) 63(7):2454–63. doi:10.2337/db13-1819
- Wu T, Ahmed A, Bril V, Orszag A, Ng E, Nwe P, et al. Variables associated with corneal confocal microscopy parameters in healthy volunteers: implications for diabetic neuropathy screening. *Diabet Med* (2012) 29(9):e297–303. doi:10.1111/j.1464-5491.2012.03678.x
- Tavakoli M, Marshall A, Banka S, Petropoulos IN, Fadavi H, Kingston H, et al. Corneal confocal microscopy detects small-fiber neuropathy in Charcot– Marie–Tooth disease type 1A patients. *Muscle Nerve* (2012) 46(5):698–704. doi:10.1002/mus.23377
- Tavakoli M, Marshall A, Thompson L, Kenny M, Waldek S, Efron N, et al. Corneal confocal microscopy: a novel noninvasive means to diagnose neuropathy in patients with Fabry disease. *Muscle Nerve* (2009) 40(6):976–84. doi:10.1002/mus.21383
- Asghar O, Petropoulos IN, Alam U, Jones W, Jeziorska M, Marshall A, et al. Corneal confocal microscopy detects neuropathy in subjects with impaired glucose tolerance. *Diabetes Care* (2014) 37(9):2643–6. doi:10.2337/ dc14-0279
- Pop-Busui R, Lu J, Brooks MM, Albert S, Althouse AD, Escobedo J, et al. Impact of glycemic control strategies on the progression of diabetic peripheral neuropathy in the bypass angioplasty revascularization investigation 2 diabetes (BARI 2D) cohort. *Diabetes Care* (2013) 36(10):3208–15. doi:10.2337/ dc13-0012
- Wilkinson CP, Ferris FL III, Klein RE, Lee PP, Agardh CD, Davis M, et al. Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. *Ophthalmology* (2003) 110(9):1677–82. doi:10.1016/S0161-6420(03)00475-5
- Ishibashi F, Okino M, Ishibashi M, Kawasaki A, Endo N, Kosaka A, et al. Corneal nerve fiber pathology in Japanese type 1 diabetic patients and its correlation with antecedent glycemic control and blood pressure. J Diabetes Investig (2012) 3(2):191–8. doi:10.1111/j.2040-1124.2011.00157.x
- Ishibashi F, Kojima R, Taniguchi M, Kosaka A, Uetake H, Tavakoli M. The expanded bead size of corneal C-nerve fibers visualized by corneal confocal microscopy is associated with slow conduction velocity of the peripheral nerves

in patients with type 2 diabetes mellitus. *J Diabetes Res* (2016) 2016:3653459. doi:10.1155/2016/3653459

- Young MJ, Boulton AJ, MacLeod AF, Williams DR, Sonksen PH. A multicentre study of the prevalence of diabetic peripheral neuropathy in the United Kingdom hospital clinic population. *Diabetologia* (1993) 36(2):150–4. doi:10.1007/BF00400697
- Tesfaye S, Boulton AJM, Dyck PJ, Freeman R, Horowitz M, Kempler P, et al. Diabetic neuropathies: update on definitions, diagnostic criteria, estimation of severity, and treatments. *Diabetes Care* (2010) 33(10):2285–93. doi:10.2337/ dc10-1303
- Kashiwagi A, Kasuga M, Araki E, Oka Y, Hanafusa T, Ito H, et al. International clinical harmonization of glycated hemoglobin in Japan: from Japan Diabetes Society to National Glycohemoglobin Standardization Program values. *J Diabetes Investig* (2012) 3(1):39–40. doi:10.1111/j.2040-1124.2012.00207.x
- Molitch ME, DeFronzo RA, Franz MJ, Keane WF, Mogensen CE, Parving HH, et al. Nephropathy in diabetes. *Diabetes Care* (2004) 27(Suppl 1):S79–83. doi:10.2337/diacare.27.2007.S79
- Tesfaye S, Chaturvedi N, Eaton SEM, Ward JD, Manes C, Ionescu-Tirgoviste C, et al. Vascular risk factors and diabetic neuropathy. *N Engl J Med* (2005) 352(4):341–50. doi:10.1056/NEJMoa032782
- Brown MJ, Bird SJ, Watling S, Kaleta H, Hayes L, Eckert S, et al. Natural progression of diabetic peripheral neuropathy in the Zenarestat study population. *Diabetes Care* (2004) 27(5):1153–9. doi:10.2337/diacare.27.5.1153
- Malik RA, Veves A, Tesfaye S, Smith G, Cameron N, Zochodne D, et al. Small fibre neuropathy: role in the diagnosis of diabetic sensorimotor polyneuropathy. *Diabetes Metab Res Rev* (2011) 27(7):678–84. doi:10.1002/dmrr.1222
- Müller LJ, Marfurt CF, Kruse F, Tervo TMT. Corneal nerves: structure, contents and function. *Exp Eye Res* (2003) 76(5):521–42. doi:10.1016/ S0014-4835(03)00050-2
- Quattrini C, Tavakoli M, Jeziorska M, Kallinikos P, Tesfaye S, Finnigan J, et al. Surrogate markers of small fiber damage in human diabetic neuropathy. *Diabetes* (2007) 56(8):2148–54. doi:10.2337/db07-0285
- Chen X, Graham J, Dabbah MA, Petropoulos IN, Ponirakis G, Asghar O, et al. Small nerve fiber quantification in the diagnosis of diabetic sensorimotor polyneuropathy: comparing corneal confocal microscopy with intraepidermal nerve fiber density. *Diabetes Care* (2015) 38(6):1138–44. doi:10.2337/ dc14-2422
- Dehghani C, Pritchard N, Edwards K, Russell AW, Malik RA, Efron N. Risk factors associated with corneal nerve alteration in type 1 diabetes in the absence of neuropathy: a longitudinal *in vivo* corneal confocal microscopy study. *Cornea* (2016) 35(6):847–52. doi:10.1097/ICO.000000000000060
- Hyllienmark L, Ludvigsson J, Alstrand N, Cooray G, Jonsson B, Wahlberg-Topp J. Early electrophysiological abnormalities and clinical neuropathy. *Diabetes Care* (2013) 36(10):3187–94. doi:10.2337/dc12-2226
- 31. Ohkubo Y, Kishikawa H, Araki E, Miyata T, Isami S, Motoyoshi S, et al. Intensive insulin therapy prevents the progression of diabetic microvascular complications in Japanese patients with non-insulin-dependent diabetes mellitus: a randomized prospective 6-year study. *Diabetes Res Clin Pract* (1995) 28(2):103–17. doi:10.1016/0168-8227(95)01064-K
- 32. Ziegler D, Behler M, Schroers-Teuber M, Roden M. Near-normoglycaemia and development of neuropathy: a 24-year prospective study from diagnosis of type 1 diabetes. *BMJ Open* (2015) 5:e006559. doi:10.1136/bmjopen-2014-006559
- American Diabetes Association. Standards of medical care in diabetes-2014. Diabetes Care (2014) 37(Suppl 1):S14–80. doi:10.2337/dc14-S014
- Tavakoli M, Mitu-Pretorian M, Petropoulos IN, Fadavi H, Asghar O, Alam U, et al. Corneal confocal microscopy detects early nerve regeneration in diabetic neuropathy after simultaneous pancreas and kidney transplantation. *Diabetes* (2013) 62(1):254–60. doi:10.2337/db12-0574
- Tavakoli M, Kallinikos P, Iqbal A, Herbert A, Fadavi H, Efron N, et al. Corneal confocal microscopy detects improvement in corneal nerve morphology with an improvement in risk factors for diabetic neuropathy. *Diabet Med* (2011) 28(10):1261–7. doi:10.1111/j.1464-5491.2011.03372.x
- Grisold A, Callaghan BC, Feldman EL. Mediators of diabetic neuropathy: is hyperglycemia the only culprit? *Curr Opin Endocrinol Diabetes Obes* (2017) 24(2):103–11. doi:10.1097/MED.00000000000320
- Shurter A, Genter P, Ouyang D, Ipp E. Euglycemic progression: worsening of diabetic retinopathy in poorly controlled type 2 diabetes in minorities. *Diabetes Res Clin Pract* (2013) 100(3):362–7. doi:10.1016/j.diabres.2013.03.018

- Ceriello A. The emerging challenge in diabetes: the "metabolic memory". Vascul Pharmacol (2012) 57(5-6):133-8. doi:10.1016/j.vph.2012.05.005
- Kalteniece A, Ferdousi M, Adam S, Schofield J, Azmi S, Petropoulos I, et al. Corneal confocal microscopy is a rapid reproducible ophthalmic technique for quantifying corneal nerve abnormalities. *PLoS One* (2017) 12(8):e0183040. doi:10.1371/journal.pone.0183040
- Winter K, Scheibe P, Köhler B, Allgeier S, Guthoff RF, Stachs O. Local variability of parameters for characterization of the corneal subbasal nerve plexus. *Curr Eye Res* (2016) 41(2):186–98. doi:10.3109/02713683.2015. 1010686
- 41. Stuard WL, Titone R, Robertson DM. Tear levels of insulin-like growth factor binding protein 3 correlate with subbasal nerve plexus changes in patients with type 2 diabetes mellitus. *Invest Ophthalmol Vis Sci* (2017) 58(11):6105–12. doi:10.1167/iovs.17-22425
- Di G, Qi X, Zhao X, Zhang S, Danielson P, Zhou Q. Corneal epithelium-derived neurotrophic factors promote nerve regeneration. *Invest Ophthalmol Vis Sci* (2017) 58(11):4695–702. doi:10.1167/iovs.16-21372

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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