1 Supplementary information

- 2 Lateral plate mesoderm cell-based organoid system for NK cell regeneration
- 3 from human pluripotent stem cells
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34 Supplementary Figures and Figure legends

Supplementary Fig.S1 a b 0.06 HAND1 HAND1 MSGN1 Relative expression Relative expression MSGN1 0.06 0.04 0.04 0.02 0.02 0.00 0.00 Day 0 Day 1 Day 2 Day 0 Day 1 Day 2 d С 100 **ESC** 100-**ESC iPSC** BRACHYURY⁺ cells (%) iPSC APLNR⁺ cells (%) 75 75 50-50 25 25-0

Supplementary Fig. S1 Analysis of primitive streak and lateral plate mesoderm-specific markers of hPSC-derived cells

Day 2

Day 1

Day 0

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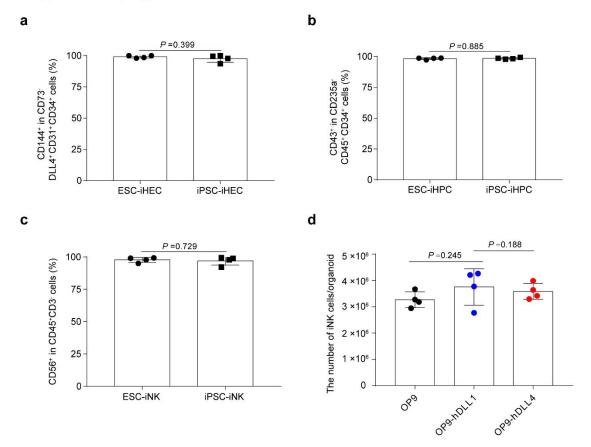
a Real-time quantitative PCR analysis of HANDI and MSGNI gene expression levels of ESC-derived cells on Day 0, Day 1, and Day 2. **b** Real-time quantitative PCR analysis of HANDI and MSGNI gene expression levels of iPSC-derived cells on Day 0, Day 1, and Day 2. **c** The proportion of BRACHYURY⁺ cells in hPSC-derived cells on Day 0, Day 1, and Day 2. Data were collected from three independent experiments. n = 3. **d** The proportion of APLNR⁺ cells in hPSC-derived cells on Day 0, Day 1, and Day 2. Data were collected from three independent experiments. n = 3. ESC: human ESC line hPSC-2; iPSC: human iPSC line hPSC-6.

Day 0

Day 1

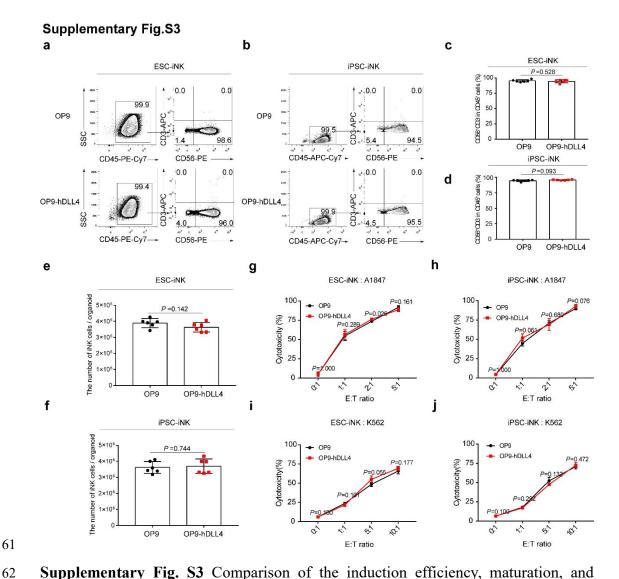
Day 2

Supplementary Fig.S2



Supplementary Fig. S2 Analysis of the efficiency of iHEC, iHPC, and iNK cell regeneration from hPSCs

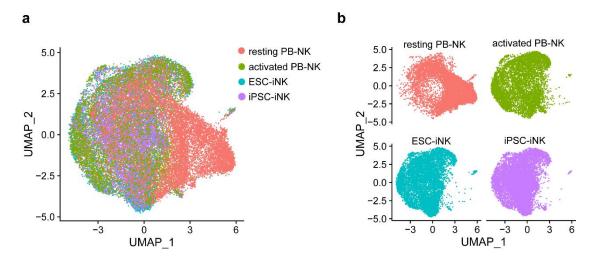
a The proportion of CD144⁺ cells in CD73⁻DLL4⁺CD31⁺CD34⁺ iHECs on Day 9. Data were collected from four independent experiments and analyzed by two-tailed independent t-test. n = 4. **b** The proportion of CD43⁺ cells in CD235a⁻CD45⁺CD34⁺ iHPCs on Day 16. Data were collected from four independent experiments and analyzed by Mann-Whitney U test. n = 4. **c** The proportion of CD56⁺ cells in CD45⁺CD3⁻ cells on Day 27. Data were collected from four independent experiments and analyzed by two-tailed independent t-test. n = 4. **d** 5 × 10⁵ OP9, OP9-hDLL1, and OP9-hDLL4 feeder cells were combined with 2 × 10⁴ mesoderm cells separately to form organoid aggregates on Day 2. Data were collected and analyzed on Day 27 (n = 4 each group). Statistics: two-tailed independent t-test.



Supplementary Fig. S3 Comparison of the induction efficiency, maturation, and cytotoxicities of iNK cells from ESCs and iPSCs using OP9 and OP9-hDLL4 feeder cells. **a-b** Immuno-phenotypes (CD45⁺CD3⁻CD56⁺) of iNK cells from ESC (**a**) and iPSCs (**b**) on Day 27. **c-d** Statistics of the maturation of iNK cells from ESC (**c**) and iPSCs (**d**) on Day 27 (n = 6 each group). **e-f** Statistics of the number of CD45⁺CD3⁻CD56⁺ iNK cells on Day 27 (n = 6 each group). **g-h** Cytotoxicity analysis of ESC-iNK cells (**g**) and iPSC-iNK cells (**h**) against A1847 cell line (n = 4 each group). **i-j** Cytotoxicity analysis of ESC-iNK cells (**i**) and iPSC-iNK cells (**j**) against k562 cell line (n = 4 each group). Statistics: two-tailed independent t-test (**c-j**) and Mann-Whitney U test (**g**).

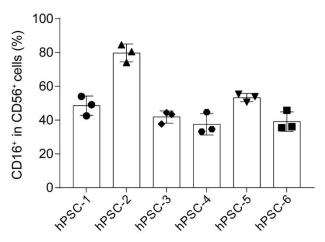
Supplementary Fig.S4

cells respectively.



Supplementary Fig. S4 Integration of scRNA-seq data generated from ESC-iNK, iPSC-iNK, resting PB-NK, and activated PB-NK cells.
a Projection of ESC-iNK, iPSC-iNK, resting PB-NK, and activated PB-NK cells.
b UMAP visualization of ESC-iNK, iPSC-iNK, resting PB-NK, and activated PB-NK

Supplementary Fig.S5



Supplementary Fig. S5 CD16 molecule expression pattern of iNK cells.

CD16 expression levels of iNK cells and NK cells from human embryonic cell line hPSC-1, hPSC-2, hPSC-3, hPSC-4, hPSC-5, and human iPSC line hPSC-6. CD16⁺ cells were gated on CD45⁺CD3⁻CD56⁺ iNK cells. Data were collected from three independent experiments. All samples were collected on Day 27.

Supplementary Table S1

Primer sequences for real-time quantitative PCR of selected genes	
Primer name	Primer sequence $(5'\rightarrow 3')$
β-ACTIN-F	CATGTACGTTGCTATCCAGGC
β -ACTIN-R	CTCCTTAATGTCACGCACGAT
<i>TBXT-</i> F	TATGAGCCTCGAATCCACATAGT
TBXT-R	CCTCGTTCTGATAAGCAGTCAC
APLNR-F	CTCTGGACCGTGTTTCGGAG
APLNR-R	GGTACGTGTAGGTAGCCCACA
<i>HAND1-</i> F	GAGAGCATTAACAGCGCATTCG
<i>HAND1-</i> R	CGCAGAGTCTTGATCTTGGAGAG
<i>MSGN1-</i> F	AACCTGCGCGAGACTTTCC
<i>MSGN1</i> -R	GTCTGTGAGTTCCCCGATGTA