

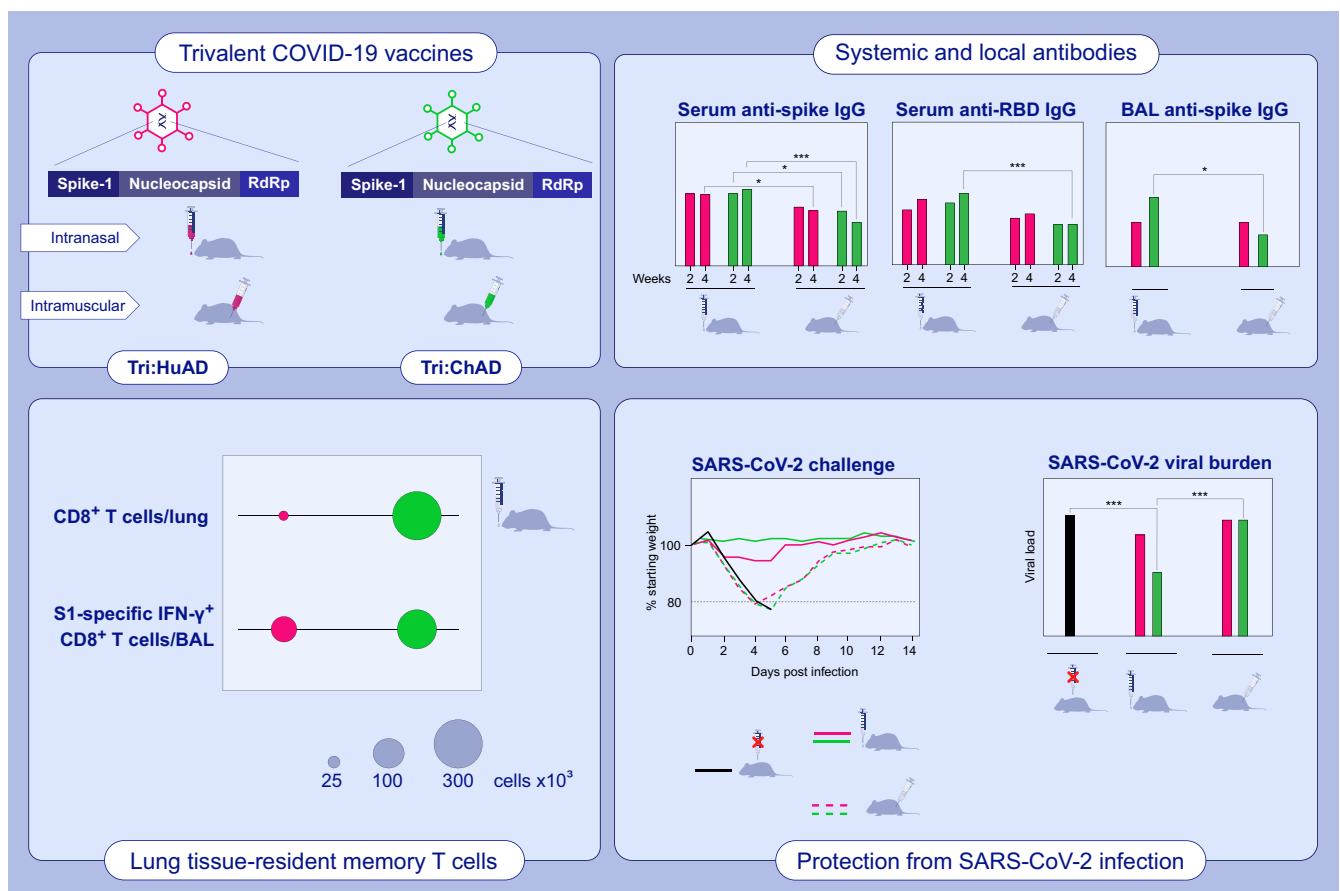
## NEWS AND VIEWS

# A potential immunological silver bullet for COVID-19: The trivalent chimpanzee adenoviral serotype-68 vector (Tri:ChAd)

Humanity has been facing a major challenge recently from a microbe called SARS-CoV-2. Although aged people are more susceptible to COVID-19, the vaccine has been administered even in 5-year-old children so far to control and prevent disease.<sup>1,2</sup> Several strains of SARS-CoV-2 have been reported within two years. Some of these have attracted public attention for their contagiousness, or level of mutations, and are termed variants of concern (VOCs).<sup>3,4</sup> The number of vaccines registered to the World Health Organization in clinical and preclinical development is 144 and 195, respectively.<sup>5</sup> While the generation of different types of vaccines is important for

protection effectiveness, the route of vaccine administration is also a key factor. For example, the respiratory mucosal immunity induced by the inhaled aerosol modified vaccinia Ankara Tuberculosis vaccine in humans was not achieved by intradermal injection of the same vaccine.<sup>6</sup>

Providing long-term protection against current and future variants and sustained vaccine-induced herd immunity is a great challenge. Due to the short duration of humoral protection and immunity of the first generation of commercial COVID-19 vaccines, it is imperative to develop a vaccine that provides long-term



**FIGURE 1** Intranasal Tri:ChAd single-dose vaccine is the best vaccine candidate against COVID-19 compared to intramuscular immunization. Intramuscular (i.m.), intranasal (i.n.), trivalent human adenovirus serotype 5 ( $5 \times 10^7$  PFU/Tri:HuAd), trivalent chimpanzee adenovirus serotype 68 ( $1 \times 10^7$  PFU/Tri:ChAd), bronchoalveolar lavage fluid (BAL), interferon- $\gamma$  (IFN- $\gamma$ ), RNA-dependent RNA polymerase (RdRp)

protection against existing and emerging variants. Janssen Ad26. COV2.S ( $5 \times 10^{10}$  viral particles) and Oxford/AstraZeneca ChAdOx1 ( $5 \times 10^{10}$  viral particles) are administered as single and two doses, respectively, via intramuscular (i.m.) injection. However, Ad26. COV2.S and ChAdOx1 have generated concern because of some side effects.<sup>7,8</sup> To overcome these problems, Afkhami et al.<sup>9</sup> recently showed that respiratory mucosal delivery of next-generation COVID-19 vaccine provides robust protection against both ancestral and variant strains of SARS-CoV-2.

Afkhami et al.<sup>9</sup> developed adenoviral vectors of trivalent human serotype 5 ( $5 \times 10^7$  PFU/Tri:HuAd) or trivalent chimpanzee serotype 68 ( $1 \times 10^7$  PFU/Tri:ChAd) which are suitable for single-dose intranasal (i.n.) administration in the mouse model (BALB/c, C57BL/6J, B6. Cg-Tg (K18-ACE2) 2PrImn/J, B cell-deficient mice C. Cg-Igh-J<sup>tm1Dhu</sup>, BALB/c mice depleted of T cells) and evaluated responses at 2, 4, and 8 weeks post-immunization (Figure 1). Vectors included the full-length S1 domain of spike (S) which contains the N-terminal domain, receptor-binding domain (RBD), and numerous T-cell epitopes. S1 was fused to the vesicular stomatitis virus G protein transmembrane/trimerization domain. This domain was used to provide membrane anchor and facilitation of trimerization and exosomal targeting for enhanced antibody responses. Furthermore, to broaden T-cell immunity against additional viral antigens, nucleocapsid and truncated nsp12 RNA-dependent RNA polymerase proteins were included in vaccine design as a single polyprotein downstream of a porcine teschovirus 2A sequence.

In comparison with i.m. vaccination, i.n. Tri:ChAd vaccine provides significantly greater induced S- and RBD-specific IgG responses in serum and significant amounts of anti-S IgA, superior airway T-cell responses, multifunctional CD8+ T cells with cytotoxic potential within the respiratory tract, multifunctional respiratory mucosal tissue-resident memory T-cell responses, and trained airway macrophages. Concerning memory B cells in systemic lymphoid and local lung tissues, high levels of RBD-specific B cells were formed by i.n. Tri:ChAd compared to i.n. Tri:HuAd. In the lung tissue, only i.n. Tri:ChAd induces these cells. The existence of S1-, N-, and RdRp-specific CD4+ T cells were observed in the airways and spleen following a Tri:ChAd vaccination by i.n. administration but not i.m. I.n. Tri:ChAd vaccine also empowers the Th1-skewed S-specific IgG2a antibody response without Th2 skewing of antibody responses. In addition, i.n. Tri:ChAd vaccine protects against lethal infection by SARS-CoV-2 VOCs which are B.1.1.7 and B.1.351. Surprisingly, the authors found that clinical outcomes/illnesses do not always corroborate with the viral burden. Indeed, while i.n. Tri:ChAd vaccine protected wild-type and B cell- and T cell-deficient mice in clinical outcomes, lack of B or T cells led to partially impaired viral clearance in the lung.

A superior protection against a mouse-adapted SARS-CoV-2 is provided by i.n. immunization rather than i.m. vaccination. Desirable protection of i.n. immunization develops only when humoral and T-cell immunity exist. Vaccine-induced trained innate immunity

improves clinical outcomes but it partially controls the viral burden. It is promising to design a vaccine without the vulnerabilities of first-generation vaccines and to develop a cheaper and more attractive vaccine for protection-control in the fight against COVID-19. Thus, this vaccine exhibits extraordinary results for COVID-19 and will be awaited with great interest in primates and clinical phase studies.

## KEYWORDS

adenoviral vectors, mucosal immunity, next-generation vaccine, SARS-CoV-2, trained innate immunity


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## CONFLICT OF INTEREST

The authors declare that they have no potential conflict of interest.

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