SHORT REPORT



Comparison of whole genomes of tick-borne encephalitis virus from mountainous alpine regions and regions with a lower altitude

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Received: 15 June 2020 / Accepted: 16 December 2020 / Published online: 24 January 2021 © The Author(s) 2021

Abstract

Tick-borne encephalitis (TBE) has been a notifiable disease in Germany since 2001. Its causative agent, the TBE virus (TBEV), is the most important arbovirus in Europe and Northern Asia. The illness, caused by the European Subtype usually displays flu-like symptoms, but can result in sequelae and, in 2 % of all cases, in death. Over the last few decades, the virus has spread into new habitats, such as higher altitudes in the Alpine region. For this study, it was hypothesized that the environmental challenges that the virus might be exposed to at such altitudes could lead to the selection of viral strains with a higher resilience to such environmental factors. To determine whether strains identified at higher altitudes (> 500 m above sea level) (n=5) and lower altitudes (< 500 m above sea level) (n=4) was performed. No common phylogenetic ancestry or shared amino acid substitutions could be identified that differentiated the alpine from the lowland viral strains. These findings support the idea of many individual introductions of TBEV into the alpine region and the establishment of foci due to non-viral specific factors such as favorable conditions for vector species and host animals due to climate change.

Keywords Tick-borne encephalitis virus \cdot Mountains \cdot Genetic analysis \cdot Tick-borne encephalitis \cdot Tick-borne encephalitis virus strains

Introduction

TBEV is the agent of tick-borne encephalitis (TBE), a severe infection of the central nervous system that may result in sequelae and can possibly end in disability or death. Currently, the virus is divided into three subtypes, the European, the Siberian and the Far Eastern subtype [1-5]. TBEV is mostly transmitted via tick bites. However, especially in Eastern Europe, outbreaks caused by alimentary transmission through unpasteurized milk and soft cheese have been reported [6-12].

Edited by William Dundon.

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In recent years, the European subtype of TBEV has come more into focus since its distribution pattern changed significantly. The virus appeared in mountainous regions of the Alps previously considered free of natural foci of TBEV [13, 14]. It is unclear, whether certain/specific genetic traits of the virus are responsible for this sudden claim of new endemic areas or if the climatic conditions changed and became more suitable for the natural transmission cycle. It is possible that the mountainous strains were naturally selected due to some small nucleotide polymorphisms (SNPs) in their genome that made them more resistant to the alpine environment. To shed light on the question, 5 different virus strains, isolated from these new endemic areas, were thoroughly analysed regarding their genomic sequences and compared to strains isolated from long established foci in lower altitudes.

Results and discussion

For all TBEV-EU strains included in the study the whole genome sequences were generated. The phylogenetic tree based on the nucleotide sequences showed no evidence of a common origin of the mountainous strains (Fig. 1). The virus strains D15_33, K2 and HB171_11 had the closest phylogenetic relation to each other, despite coming from different elevation levels and collecting sites being 200 km apart.

In Table 1 the different strains have been compared in regard to nucleotide changes/amino acid differences. This analysis reveals that the strain K2 differs the most from strain

NE_1/7 (Neudoerfl) with 219 nucleotide changes (shown in *Italics*), whereas the strains D15_569 and D17_1044 show the fewest differences (shown in **Bold**). Regarding amino acid substitutions the biggest difference of 40 amino acid changes is between strains K2 and D17_1989 (highlighted in *Italics*) and the least amino acid difference can be observed between the strains HB171_11 and BaWa16_303 with only 19 amino acid changes (indicated in **Bold**).

Fig. 1 a Phylogenetic tree of whole genomes of several TBEV-Eu strains. The whole genomes were amplified in three DNA amplicons covering the whole genome [15]. For sequencing the Illumina MiSeq platform and the MiSeq reagent kit V3 (Illumina, Inc., San Diego, USA) was used, following the manufacturer's instructions. Assembly was performed using the software Spades v.3.12. For the phylogenetic comparison, available TBEV-Eu whole genome sequences published in the NCBI GenBank database were chosen. The tree was generated using the maximum likelihood approach and 1000 bootstraps were implemented for statistical support [16–19]. The strains used in this study are highlighted in red (alpine regions) and blue (lower altitudes). b Table containing the meta data for the TBEV strains used in the analysis



(B)	Strain	Location	Country	Year of Isolation	GPS - X	GPS- Y	Elevation	Sequencing material
	D14_97	Aschau	А	2014	47,2583	11,8871	710 m	tick homogenate
	D15_33	Egelsee	А	2015	47,6049	12,1707	563 m	tick homogenate
	D15_569	Wald	А	2015	47,0778	10,8329	1364 m	tick homogenate
	D17_1044	Mühlau	D	2017	47,7254	12,3938	611 m	tick homogenate
	D18_1133	Tres	I	2018	46,3173	11,1069	858 m	tick homogenate
	D17_1989	Petting	D	2017	47,9259	12,819	459 m	tick homogenate
	BaWa16_303	Haselmühl	D	2016	49,409	11,8831	430 m	tick homogenate
	NE_1/7	Neudoerfl	А	1975	47,7887	16,3051	266 m	babymouse brain
	K2	Karlsruhe	D	1980	49,032	8,4136	116 m	babymouse brain
	HB171_11	Heselbach	D	2011	49,2973	12,2006	432 m	VeroB4 P0/P1

	D14_97	$D15_{-33}$	D15_569	D17_1044	D18_1133	D17_1989	BaWa16_303	NE_1/7	K2	$HB171_{-11}$
D14_97		97,335 (273)	97,648 (241)	97,326 (274)	97,55 (251)	97,374 (269)	97,696 (236)	97,804 (225)	97,306 (276)	97,365 (270)
D15_33	99,268 (25)		97,726 (233)	97,413 (265)	97,599 (246)	97,443 (262)	97,384 (268)	97,394 (267)	97,433 (263)	97,628 (243)
D15_569	99,18 (28)	99,268 (25)		97,921 (213)	97,833 (222)	97,657 (240)	97,706 (235)	97,667 (239)	97,677 (238)	97,862 (219)
D17_1044	99,18 (28)	99,209 (27)	99,121 (30)		97,521 (254)	97,433 (263)	97,365 (270)	97,365 (270)	97,345 (272)	97,531 (253)
D18_1133	99,356 (22)	99,356 (22)	99,297 (24)	99,238 (26)		97,531 (253)	97,56 (250)	97,501 (256)	97,54 (252)	97,589 (247)
D17_1989	99,092 (31)	99,151 (29)	99,063 (32)	99,151 (29)	99,092 (31)		97,579 (248)	97,462 (260)	97,267 (280)	97,482 (258)
BaWa16_303	99,385 (21)	99,356 (22)	99,268 (25)	99,268 (25)	99,385 (21)	99,151 (29)		97,823 (223)	97,355 (271)	97,511 (255)
NE_1/7	99,238 (26)	99,209 (27)	99,121 (30)	99,238 (26)	99,238 (26)	99,033 (33)	99,385 (21)		97,257 (281)	97,384 (268)
K2	99,092 (31)	99,121 (30)	99,033 (33)	99,033 (33)	99,121 (30)	98,828 (40)	99,238 (26)	99,033 (33)		97,609 (245)
HB171_11	99,297 (24)	99,326 (23)	99,18 (28)	99,18 (28)	99,297 (24)	99,063 (32)	99,443 (19)	99,238 (26)	99,151 (29)	

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All ten amino acid sequences were aligned and analysed for substitutions. No common amino acid substitution differentiating the alpine and the lowland strains was found. All virus proteins were analysed separately. The capsid protein showed up to two individual changes in the amino acid sequence. The prM/M protein showed up to two individual changes in the sequence with only strain HB171_11 having a non-synonymous change of T141I. The strain D14_97 showed four amino acid changes in the E-gene sequence, two of which were heterologous and may affect the superficial charge of the virus membrane. Other non-synonymous changes were found in strain D15_569 with L459S, strain D17_1989 that has a Y130H and K2 with A83T. Y130H has already been correlated with increased neuroinvasiveness in immunodeficient mice [20].

The NS1 protein showed up to two individual amino acid changes. Strain D17_1044 showed a non-synonymous substitution of the non-polar I127T, K2 one of P103S and HB171_11 A41T. Compared to its length the most variable protein was NS2A. The three strains D15_33, K2 and HB171_11 showed the same synonymous substitution of V41I and the strains D17_1044 and D17_1989 shared a substitution of I53M. The latter is remarkable, since both belong to two different genetic clusters and share another substitution in their NS3 protein.

Within the NS2B amino acid alignment two non-synonymous changes could be identified for the strains D15_569 and HB171_11. For NS3 up to three individual substitutions were found and three of the strains (D15_33, D18_1133 and K2) showed non-synonymous changes. The NS4A showed one substitution in three viruses, with only the substitution of Gly100 for serine in strain D17_1044 being non-synonymous. Three strains (D17_1989, K2 and HB171_11) had up to two individual substitutions in the NS4B protein.

The NS5 showed up to fourteen amino acid substitutions ranging from one to seven individual changes. The virus with most individual changes was strain D17_1044, followed by strain D17_1989.

The virus strain with the fewest individual amino acid changes was strain D18_1133 with 23% changes in their amino acid sequence. It was followed by strain D14_97 with 33.3% individual changes. The virus strain with most individual changes was HB171_11 with 48%.

Therefore, a phylogenetic analysis often results in poor statistical support and has to be interpreted very cautiously. The comparison of our TBEV strains confirms this observation, since the genetic difference of the ten TBEV strains in our study was 0.028 substitutions per site in total. Over the last two decades a clear shift in the distribution of TBE virus from lower to higher altitudes has been observed. So far, the possible effects on the genomic characteristics of TBEV are still unclear. One working hypothesis was that the virus replication had to adapt to harsh environmental conditions. Therefore, these strains might exhibit certain specific changes in their genomes as a response to the conditions they were exposed in regions with higher altitudes. Another explanation is that every strain from a mountainous area is derived from a common ancestor adapted to the conditions found in mountains. After further distribution, they could form new natural foci. Furthermore, a change in climatic and therefore ecoepidemiological conditions in the higher altitudes might lead to better conditions for TBEV replication in ticks.

We were not able to find a common specific genetic trait shared by all strains from areas of an altitude above 500 m above sea level. According to the phylogenetic analysis, each virus strain was probably introduced individually into its location and found favourable environmental conditions (possibly due to climate changes). We observed some of the amino acid changes described in the paper by Formanová et al. [21] in all eight virus strains (namely Ile167V, E127D, V201I and G206R). In addition, T33S was found in HB171_11 and D17_1989 and I53M in D17_1044 and D17_1989. Furthermore, we could not determine a close phylogenetic relationship between the mountainous strains. In fact, strain D15 33 was genetically most closely related to the lower altitudinal region strains HB171 11 and K2 in our analysis. This is another indicator for the newly emerged strains to be distributed into new areas by chance.

NS2A is said to be involved in the shift between RNA packaging and RNA replication [22, 23], its high variability could be the reason for the different replication rates and infectivity of different TBEV strains. The same could apply to the high variability in NS2B, a protein suspected to be involved in modulating the membrane permeability during infection [24]. To confirm or disprove this hypothesis further experiments need to be undertaken.

Since we could not find any hints pointing towards specific genetic characteristics of TBEV strains from higher altitudes, we assume that other reasons for the new distributional pattern are responsible for the emerging alpine distribution of TBEV.

As shown by Rubel et al. in [25] the Alps, have undergone a severe climatic shift since 1876. It can be assumed that this shift contributes to transformation of the Alpine environment and therefore, the distribution of vector species, and host animals might change. This has yet to be proven by further research.

Acknowledgements We thank Prof. Dr. F.X. Heinz and Prof. Dr. Karin Stiasny (Institute of Virology, Medical University of Vienna, Austria) for kindly providing us with strain K2.

Funding Open Access funding enabled and organized by Projekt DEAL. G.L. and the study were funded by the grant TTU 01 801 FB2016 of the German Center for Infection Research (DZIF).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Chumakov MP, Zyaitlenok NA, Vorob'eva MS (1944) The studies of virus encephalitis. Report II. The geographical distribution and epidemiological characteristics of tick-borne encephalitis in European part of Soviet Union, Siberia and Kazachstan. Neuropathol Psychiatry 13:20–23
- Pogodina VV, Bochkova NG, Koreshkova GV (1981) Strain properties of the Aina/1448 serotype of the tick-borne encephalitis virus. VoprVirusol 6:741–746
- Heinz FX, Collett MS, Purcell RH, Gould EA, Howard CR, Houghton M, Moormann RJM, Rice CM, Thiel HJ (2000) Family Flaviviridae. In: van Regenmortel MHV, Fauquet CM, Bishop DHL, Carstens E, Estes JK, Lemon S, Maniloff J, Mayo MA, McGeogch D, Pringle CR, Wickner RB (eds) Virus taxonomy: classification and nomenclature of viruses. Seventh report of the International Committee on Taxonomy of Viruses. Academic Press, San Diego, pp 859–878
- Kovalev SY, Mukhacheva TA (2017) Reconsidering the classification of tick-borne encephalitis virus within the Siberian subtype gives new insights into its evolutionary history. Infect Genet Evol 55:159–165
- Dai X, Shang G, Lu S, Yang J, Xu J (2018) A new subtype of eastern tick-borne encephalitis virus discovered in Qinghai-Tibet Plateau. China Emerg Microbes Infect 7:74
- Gritsun TS, Frolova TV, Zhankov I, Armesto M, Turner SL, Frolova MP, Pogodina VV, Lashkevich VA, Gould EA (2003) Characterization of a Siberian virus isolated from a patient with progressive chronic tick-borne encephalitis. J Virol 77(1):25–36
- Moritsch H, Kovac W (1962) Investigations on pathogenesis of alimentary infection with tick-borne encephalitis virus in mice. In: Libíková H (ed) Biology, of viruses of the tick-borne encephalitis complex. Czech. Acad. Sci., Prague, pp 283–285
- Pogodina VV (1958) Resistance of tick-borne encephalitis virus to gastric juice. VoprVirusol 5:271–275
- Pogodina VV (1960) An experimental study on the pathogenesis of tick-borne encephalitis following alimentary infection. Part 1. The dynamics of distribution of the virus in white mice infected by the enteral route. VoprVirusol 5:272–279
- Shapoval, AN (1976) In: L. A. Ulitski (ed), Chronic forms of tickborne encephalitis. Medicine. Leningrad, Russia
- Pogodina, VV, Frolova MP, and Erman BA (1986) In: E. F. Bocharov (ed.), Chronic tick-borne encephalitis. Nauka. Moscow, Russia

- 12. Leonova GN (2009) Tick-borne encephalitis: current aspects. Balabanov Press, Moskow, p 168
- Robert Koch-Institut (RKI) (2020) FSME: Risikogebiete in Deutschland (Stand: January 2020). Epid Bull 8:3–19. https:// doi.org/10.25646/6510
- Heinz FX, Stiasny K, Holzmann H, Kundi M, Sixl W et al (2015) Emergence of tick-borne encephalitis in new endemic areas in Austria: 42 years of surveillance. Eurosurveillance 20:13
- Andersen NS, Bestehorn M, Chitimia-Dobler L, Kolmos HJ, Jensen PM, Dobler G, Skarphédinsson S (2019) Phylogenetic characterization of tick-borne encephalitis virus from Bornholm, Denmark. Ticks Tick-Borne Dis 10(3):533–539. https://doi. org/10.1016/j.ttbdis.2018.12.008
- Edgar RC (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res 32:1792– 1797. https://doi.org/10.1093/nar/gkh340
- Tamura K, Nei M (1993) Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. Mol Biol Evol 10:512–526. https://doi. org/10.1093/oxfordjournals.molbev.a040023
- Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. Evol Int J Org Evol 39:783–791. https://doi.org/10.1111/j.1558-5646.1985.tb00420.x
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol Evol 30:2725–2729
- 20. Pletnev AG (2001) Infectious cDNA clone of attenuated langat tick-borne flavivirus (strain E5) and a 3' deletion mutant

constructed from it exhibit decreased neuroinvasiveness in immunodeficient mice. Virology 282(2):288–300. https://doi.org/10.1006/viro.2001.0846

- Formanová P, Cerný J, Cerná BB, Valdés JJ, Kozlova I, Dzhioev Y, Ruzek D (2015) Full genome sequences and molecular characterization of tick-borne encephalitis virus strains isolated from human patients. Ticks Tick-Borne Dis 6:38–46
- 22. Khromykh AA, Varnavski AN, Sedlak PL, Westaway EG (2001) Coupling between replication and packaging of flavivirus RNA: evidence derived from the use of DNA-based full-length cDNA clones of Kunjin virus. J Virol 75:4633–4640
- Leung JY, Pijlman GP, Kondratieva N, Hyde J, Mackenzie JM, Khromykh AA (2008) Role of nonstructural protein NS2A in flavivirus assembly. J Virol 82(10):4731–4741. https://doi. org/10.1128/JVI.00002-08
- Chang YS, Liao CL, Tsao CH, Chen MC, Liu CI, Chen LK, Lin YL (1999) Membrane permeabilization by small hydrophobic nonstructural proteins of Japanese encephalitis virus. J Virol 73:6257–6264
- 25. Rubel F, Brugger K, Haslinger K, Auer I (2017) The climate of the European Alps: shift of very high resolution Köppen-Geiger climate zones 1800–2100. Meteorol Z 26(2):115–125

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