Check for updates

EDITORIALS

8 Preempting Bronchopulmonary Dysplasia: Time to Focus on the Placenta?

Bronchopulmonary dysplasia (BPD), a chronic lung disease described more than 50 years ago, remains one of the main complications in preterm infants born before 28 weeks' gestational age (1) with longterm consequences beyond childhood (2-4). One of the challenges in finding effective therapies for BPD is the multifactorial aspect of the disease. Accumulating evidence now clearly suggests that adverse antenatal factors are potent determinants of postnatal respiratory outcomes (5, 6). Not surprisingly, disorders of the placenta-which could be designated as the fetal lung in utero-have profound effects on lung development and can influence postnatal lung development. Pioneering work by Mestan and colleagues clearly demonstrated that histological and cord blood biomarkers indicative of placental maternal vascular underperfusion—as can occur in preeclampsia—are predictive of BPD (7, 8), supporting the vascular hypothesis of this lung disease (9). Likewise, placental inflammation and/or infection in the course of chorioamnionitis impedes normal lung growth (10) and can worsen outcome (11). These observations raise interesting questions about BPD endotypes. Inferring those endotypes based on dysfunctional placentation may have important therapeutic implications (12). Early recognition of risk factors and interventions specifically targeting the pathogenic mechanisms at play may prove effective in preventing BPD.

There is much excitement in the neonatal field about the lung protective potential of mesenchymal stromal cells (MSCs) because of their putative pleiotropic effects and their capability of targeting the multiple pathogenic mechanisms contributing to BPD (13–15). MSCs seem to orchestrate wound repair in part through the release of exosomes or small extracellular vesicles (16, 17). These MSC-derived small extracellular vesicles (MEx) improve lung structure and function in oxygen-induced neonatal lung injury in mice by polarizing lung macrophages toward an antiinflammatory "M2-like" state (18).

In this issue of the *Journal*, Taglauer and colleagues (pp. 86–95) take advantage of an interesting preeclampsia model in the heme oxygenase-1-null mouse $(Hmox1^{-/-})$ to demonstrate the deleterious effects of the preeclamptic intrauterine environment on fetal and postnatal lung development (19). Their data further confirm the adverse impact of prenatal conditions and position this mouse model as a novel and useful tool to explore the impact of preeclampsia on postnatal complications. Next, the authors tested the therapeutic potential of weekly antenatal intravenous injections of MEx. Antenatal MEx were able to attenuate the proinflammatory preeclamptic amniotic fluid proteomic profile and restore angiogenic and lung developmental pathways, as well as branching morphogenesis of E17 lungs and postnatal alveolar structure. Together with biodistribution studies revealing that MEx traffic to the uterus, these data suggest that the mechanism of action of MEx is likely through the utero-placental interface. Indeed, direct application of MEx had no effect on fetal lung explants, although these *ex vivo* experiments may not mimic the clinical scenario. It is probable that the effect of MEx is through immune modulation. In previous work, Taglauer and colleagues assessed utero–placental leukocytes using mass cytometry and showed that a single MEx injection in $Hmox1^{-/-}$ mice affected the abundance, surface marker repertoire, and cytokine profiles of multiple immune cell populations (20).

However, the pleiotropic effects of MSCs generate the most excitement for this potential breakthrough therapy capable of curbing complications of extreme preterm birth. Identifying the contents of MEx may reveal a rich therapeutic cargo that could eventually be tailored to BPD endotypes. For example, MEx can be highly enriched in the proangiogenic miR-486-5p and thereby improve myocardial infarction (21). MEx-derived miR-486-5p also attenuates oxidative stress (22), another mechanism contributing to BPD.

Finally, it will be important to follow up on these exciting new data in other models of preeclampsia and chorioamnionitis. Large animal studies (23) will also be critical to confirm the antenatal therapeutic potential of MEx, to more precisely pinpoint their distribution, and to explore clinically relevant administration protocols as well as safety and efficacy endpoints.

Although the clinical translation of postnatal MSC therapy for BPD has already begun (24), Taglauer and colleagues here propose a provocative avenue of intervention before birth. The respiratory community has good experience with antenatal therapies since the breakthrough discovery of Liggins and Howie demonstrating the potent effect of antenatal corticosteroid treatment in reducing the incidence of respiratory distress syndrome and mortality among preterm neonates (25). More than 50 years after the description of BPD, we may finally be focusing on one of the important culprits of the disease: *the placenta*.

Author disclosures are available with the text of this article at www.atsjournals.org.

Bernard Thébaud, M.D., Ph.D. Regenerative Medicine Program Ottawa Hospital Research Institute Ottawa, Ontario, Canada

Department of Cellular and Molecular Medicine University of Ottawa Ottawa, Ontario, Canada

and Department of Pediatrics Children's Hospital of Eastern Ontario (CHEO) and CHEO Research Institute Ottawa, Ontario, Canada

³This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0. For commercial usage and reprints, please e-mail Diane Gern (dgern@thoracic.org).

Originally Published in Press as DOI: 10.1165/rcmb.2021-0472ED on November 8, 2021

EDITORIALS

ORCID ID: 0000-0003-1844-7145 (B.T.).

References

- 1. Thébaud B, Goss KN, Laughon M, Whitsett JA, Abman SH, Steinhorn RH, et al. Bronchopulmonary dysplasia. Nat Rev Dis Primers 2019;5:78.
- Goss KN, Beshish AG, Barton GP, Haraldsdottir K, Levin TS, Tetri LH, et al. Early pulmonary vascular disease in young adults born preterm. Am J Respir Crit Care Med 2018;198:1549–1558.
- Lewandowski AJ, Augustine D, Lamata P, Davis EF, Lazdam M, Francis J, et al. Preterm heart in adult life: cardiovascular magnetic resonance reveals distinct differences in left ventricular mass, geometry, and function. Circulation 2013;127:197–206.
- Wong PM, Lees AN, Louw J, Lee FY, French N, Gain K, et al. Emphysema in young adult survivors of moderate-to-severe bronchopulmonary dysplasia. Eur Respir J 2008;32:321–328.
- Manuck TA, Levy PT, Gyamfi-Bannerman C, Jobe AH, Blaisdell CJ. Prenatal and perinatal determinants of lung health and disease in early life: a National Heart, Lung, and Blood Institute workshop report. JAMA Pediatr 2016;170:e154577.
- Morrow LA, Wagner BD, Ingram DA, Poindexter BB, Schibler K, Cotten CM, et al. Antenatal determinants of bronchopulmonary dysplasia and late respiratory disease in preterm infants. Am J Respir Crit Care Med 2017;196:364–374.
- Mestan KK, Check J, Minturn L, Yallapragada S, Farrow KN, Liu X, et al. Placental pathologic changes of maternal vascular underperfusion in bronchopulmonary dysplasia and pulmonary hypertension. *Placenta* 2014;35:570–574.
- Mestan KK, Gotteiner N, Porta N, Grobman W, Su EJ, Ernst LM. Cord blood biomarkers of placental maternal vascular underperfusion predict bronchopulmonary dysplasia-associated pulmonary hypertension. J Pediatr 2017;185:33–41.
- Abman SH. Bronchopulmonary dysplasia: "a vascular hypothesis". Am J Respir Crit Care Med 2001;164:1755–1756.
- Hirsch K, Taglauer E, Seedorf G, Callahan C, Mandell E, White CW, et al. Perinatal hypoxia-inducible factor stabilization preserves lung alveolar and vascular growth in experimental bronchopulmonary dysplasia. Am J Respir Crit Care Med 2020;202:1146–1158.
- Hartling L, Liang Y, Lacaze-Masmonteil T. Chorioamnionitis as a risk factor for bronchopulmonary dysplasia: a systematic review and metaanalysis. Arch Dis Child Fetal Neonatal Ed 2012;97:F8–F17.
- Pierro M, Villamor-Martinez E, van Westering-Kroon E, Alvarez-Fuente M, Abman SH, Villamor E. Association of the dysfunctional placentation endotype of prematurity with bronchopulmonary dysplasia: a systematic review, meta-analysis and meta-regression. *Thorax* [online ahead of print] 23 Jul 2021; DOI: 10.1136/thoraxjnl-2020-216485.

- Aslam M, Baveja R, Liang OD, Fernandez-Gonzalez A, Lee C, Mitsialis SA, et al. Bone marrow stromal cells attenuate lung injury in a murine model of neonatal chronic lung disease. Am J Respir Crit Care Med 2009;180:1122–1130.
- Augustine S, Cheng W, Avey MT, Chan ML, Lingappa SMC, Hutton B, et al. Are all stem cells equal? Systematic review, evidence map, and meta-analyses of preclinical stem cell-based therapies for bronchopulmonary dysplasia. Stem Cells Transl Med 2020;9: 158–168.
- 15. van Haaften T, Byrne R, Bonnet S, Rochefort GY, Akabutu J, Bouchentouf M, *et al.* Airway delivery of mesenchymal stem cells prevents arrested alveolar growth in neonatal lung injury in rats. *Am J Respir Crit Care Med* 2009;180:1131–1142.
- Lesage F, Thébaud B. Nanotherapies for micropreemies: Stem cells and the secretome in bronchopulmonary dysplasia. *Semin Perinatol* 2018;42:453–458.
- Mahida RY, Matsumoto S, Matthay MA. Extracellular vesicles: a new frontier for research in acute respiratory distress syndrome. *Am J Respir Cell Mol Biol* 2020;63:15–24.
- Willis GR, Fernandez-Gonzalez A, Anastas J, Vitali SH, Liu X, Ericsson M, et al. Mesenchymal stromal cell exosomes ameliorate experimental bronchopulmonary dysplasia and restore lung function through macrophage immunomodulation. Am J Respir Crit Care Med 2018;197:104–116.
- Taglauer ES, Fernandez-Gonzalez A, Willis GR, Reis M, Yeung V, Liu X, et al. Antenatal mesenchymal stromal cell extracellular vesicle therapy prevents preeclamptic lung injury in mice. Am J Respir Cell Mol Biol 2022;66:86–95.
- Taglauer ES, Fernandez-Gonzalez A, Willis GR, Reis M, Yeung V, Liu X, et al. Mesenchymal stromal cell-derived extracellular vesicle therapy prevents preeclamptic physiology through intrauterine immunomodulation. *Biol Reprod* 2021;104:457–467.
- Li Q, Xu Y, Lv K, Wang Y, Zhong Z, Xiao C, et al. Small extracellular vesicles containing miR-486-5p promote angiogenesis after myocardial infarction in mice and nonhuman primates. *Sci Transl Med* 2021;13:eabb0202.
- 22. Ma N, Li S, Lin C, Cheng X, Meng Z. Mesenchymal stem cell conditioned medium attenuates oxidative stress injury in hepatocytes partly by regulating the miR-486-5p/PIM1 axis and the TGF-β/Smad pathway. *Bioengineered* 2021;12:6434–6447.
- Rozance PJ, Anderson M, Martinez M, Fahy A, Macko AR, Kailey J, et al. Placental insufficiency decreases pancreatic vascularity and disrupts hepatocyte growth factor signaling in the pancreatic islet endothelial cell in fetal sheep. *Diabetes* 2015;64: 555–564.
- 24. Ahn SY, Chang YS, Lee MH, Sung SI, Lee BS, Kim KS, et al. Stem cells for bronchopulmonary dysplasia in preterm infants: a randomized controlled phase II trial. *Stem Cells Transl Med* 2021;10: 1129–1137.
- Liggins GC, Howie RN. A controlled trial of antepartum glucocorticoid treatment for prevention of the respiratory distress syndrome in premature infants. *Pediatrics* 1972;50:515–525.