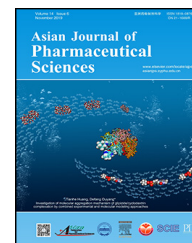


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Review

Cancer nanotechnology: Enhancing tumor cell response to chemotherapy for hepatocellular carcinoma therapy



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ABSTRACT

Hepatocellular carcinoma (HCC) is one of the deadliest cancers due to its complexities, reoccurrence after surgical resection, metastasis and heterogeneity. In addition to sorafenib and lenvatinib for the treatment of HCC approved by FDA, various strategies including transarterial chemoembolization, radiotherapy, locoregional therapy and chemotherapy have been investigated in clinics. Recently, cancer nanotechnology has got great attention for the treatment of various cancers including HCC. Both passive and active targetings are progressing at a steady rate. Herein, we describe the lessons learned from pathogenesis of HCC and the understanding of targeted and non-targeted nanoparticles used for the delivery of small molecules, monoclonal antibodies, miRNAs and peptides. Exploring current efficacy is to enhance tumor cell response of chemotherapy. It highlights the opportunities and challenges faced by nanotechnologies in contemporary hepatocellular carcinoma therapy, where personalized medicine is increasingly becoming the mainstay. Overall objective of this review is to enhance our understanding in the design and development of nanotechnology for treatment of HCC.

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1. Introduction

Hepatocellular carcinoma (HCC) is the sixth most malignant human cancer and rank third in cancer-related deaths globally among cancer patients. The incidence also is geographically related, as is the mortality, with Eastern and South-Eastern Asia and Western Africa having a high incidence [1–3]. HCC usually occurs in the established background of liver cirrhosis—due to the endopathic cause and exopathic cause often varies considerably across areas and populations. Normally, HCC can be treated curatively with surgical resection or liver transplantation if diagnosed early [4]. However, since the majority of HCC patients are diagnosed at an advanced stage, their median survival times are generally less than 1 year, leading to a poor prognosis [5]. One reason is surveillance programs are not widely implemented, despite being recommended in national guidelines [6]. On the other hand, prevention of alcohol intake, vaccination against hepatitis B virus (HBV) infection, intake of vitamin D and calcium can prevent HCC in many cases [7]. However, they could not be used to obliterate the disease. Furthermore, many investigations have shown that the occurrence of this multi-stage carcinogenesis evolves through dysregulation of multiple signaling pathways, genetic alterations with the initiation of inflammatory responses leading to the formation of a heterogeneous solid tumorous mass [8]. Implementation of such programs has only resulted in an early diagnosis in some rich places [9].

To date, there are six treatments can extend life expectancy of patients with HCC: surgical resection, liver transplantation, radiofrequency ablation, transcatheter arterial chemoembolization (TACE), sorafenib and lenvatinib [10,11]. One third of patients are eligible for potentially curative therapies, which include resection, transplantation or local ablation, and such curative treatment can extend median survival times beyond 60 months [12]. Unfortunately, most patients are still diagnosed at advanced disease stages. In this setting, only treatment with sorafenib (the first FDA approved molecularly targeted drug for HCC treatment) is able to improve overall survival in the last few decades [13]. In 2018, lenvatinib-capsules (Lenvima, Eisai Inc.) was approved by FDA for the first-line treatment of un-resectable hepatocellular carcinoma and showed better subordinate clinical endpoints when compared with sorafenib [14]. Although the anti-cancer potency of sorafenib, lenvatinib and some other drugs have been proven, poor aqueous solubility, pharmacokinetics and undesirable side effects make the approach difficult to apply widely in HCC patients [15]. The major side effects of these drugs treatment are elevated blood pressure, diarrhea, fatigue, hand-foot syndrome, skin rash/desquamation, and nausea [16]. The poor aqueous solubility and side effects might be overcome by the use of nanotechnologies that can control and target the release of effective drugs to the specific tumor site [17].

With increasing numbers of nanotherapeutics and nanodiagnosics being commercialized or having reached clinical stage in recent years [18]. Plenty of drug delivery vehicles have been developed from different biomaterials, such as synthetic polymers [19–24], natural polymers [25–28], microsphere liposomes [29–31], peptide [32–36], small molecules [37–41], and protein [42–46] et al. Specific cancer nanotechnologies

have been proven to be efficacious with respect to better targeting and minimizing the non-specific cytotoxic effect to the surrounding organs [47–51]. Targeting the receptors by drug delivery nanoparticles could significantly alleviate the adversities of chemotherapy [52,53]. Significant achievements also have been witnessed in the field of HCC drug delivery systems [52,54,55]. Understanding the biological processes of the distribution or/and retention of nanoparticles inside the HCC microenvironment is therefore essential to the development of personalized nanomedicine approaches. Herein, we examine the fundamentals behind targeting of nanoparticles to HCC and cancer cells. We will discuss the causes and recent treatment modalities of HCC, with an emphasis on cancer nanotechnology for enhancing HCC cell response to chemotherapy. With the perspective of developing new therapeutic nanotechnologies, we will also examine how the physicochemical parameters of the nanoparticles affect their localization, retention, cell binding, internalization, efficacy and toxicity.

2. Current treatment strategies for hepatocellular carcinoma

HCC has been well investigated as a complex multi-step process arising from combination of genetic and epigenetic alterations, somatic mutations, genomic instability and environmental factors [56]. There are multiple factors responsible for the initiation and progression of HCC including virus-induced [56,57], alcohol-induced, fungi-induced obesity and type II diabetes [58,59]. Common treatment strategies are composed of surgical (resection and liver transplantation) and nonsurgical approaches (transarterial chemoembolization, transarterial radiation, percutaneous local ablation, microwave ablation and systemic therapy). In nonsurgical drug therapy, not only small molecule drugs such as sorafenib and lenvatinib, but also gene therapy, immunotherapy and combination therapies have been used for HCC therapy [60–62]. Among them, systemic therapies which using small molecule drugs to target various signaling pathways following surgical resection and liver transplantation have been applied particularly where locoregional therapy has been confirmed invalid [59,63]. However, small molecular chemotherapy in HCC also suffers from the drawbacks of dose-limiting toxicities, development of multidrug resistance (MDR) and unfavorable side-effects like other cancers [64–67]. Although various multikinase inhibitors have been used for systemic treatment of HCC, only few drugs such as sorafenib (orally active multikinase inhibitor) and lenvatinib (oral multi-targeted tyrosine kinase inhibitor) have been approved as deserves special mention drug for treatment of advanced HCC. Many investigations have indicated that it could prolong overall survival in patients and delay the time to progression of advanced HCC by using sorafenib or lenvatinib. However, fatigue, diarrhea, hypertension, skin toxicity, weight loss, hypophosphatemia are almost the frequent symptoms amongst patients with these two drugs' treatment. Further, various first line and second line therapies are currently under evaluation for the HCC treatment, however, potential randomized trial with another drugs such as sunitinib, tivantinib, brivanib and linifanib have

been disappointing due to the adverse effects and not superior to sorafenib. Only lenvatinib capsules (Lenvima, Eisai Inc.) have been demonstrated that were noninferior than sorafenib for overall survival. In view of the failures of clinical trials for most of the chemotherapeutic drugs, elaborate and extensive information of the underlying molecular mechanisms undergoing in each patient's tumor progression is critically necessary. Other small molecule targeted therapies such as regorafenib, doxorubicin, gefitinib, vatalanib, cediranib, bortezomib (proteasome inhibitor), rapamycin, sirolimus, and gemcitabine as single agent or in combination with mAb have been evaluated in clinical trials targeting pathways which are activated in HCC. Recently, combination therapies also have been widely studied and shown many positive results [65,68,69].

In addition, two other types of molecular-based therapies are currently under development for the treatment of HCC: epigenetic modifying therapies [70] and microRNAs (miRNAs) [71]. Epigenetic modifications in certain genes have been proposed to drive progression of HCC. Therapies targeting epigenetic modifiers, such as DNA methyltransferases and histone deacetylases, are attractive approaches [72,73]. A multicenter phase I/II trial of the histone deacetylase inhibitor belinostat for treatment of patients with HCC is currently being developed [74]. Some other researches also showed that those with high immunoreactivity to the UV excision repair protein RAD23 homologue B (HR23B) to HDAC inhibitors had a higher rate of disease stabilization [75]. Bitzer et al. have studied the resminostat plus sorafenib as the second line therapy and showed potential activity in patients with HCC [76]. MiRNAs-based strategies that block the interaction of HDAC with its substrates have also been studied [77–79]. The results showed that many kinds of miRNAs are important regulators of gene expression and associated with the development of HCC [80,81].

Otherwise, their value in clinical management has been investigated. Functional screens of miRNAs have identified key regulators of glypican-3, a surface proteoglycan that acts as a co-receptor, controls cell apoptosis, proliferation and is a potential target for treatment of HCC [78]. In a word, evidence suggests that miRNA-based therapies are worthwhile approaches to cancer treatment. Shuai and his co-workers have investigated the folate-targeted theranosticsiRNA (Fa-PEG-gPEI-SPION/psiRNA-TBLR1) nanoparticles for enhanced HCC chemotherapy and obtained the positive results (Fig. 4).

Recently, cancer immunotherapy rises rapidly and also plays a critical role in HCC treatment due to the liver as the main immune organ of the lymphatic system [79,82,83]. Three kinds of hepatic cells named liver sinusoidal endothelial cells, Kupffer cells and dendritic cells (DCs) are main roles for the immune response in the HCC microenvironment [84]. However, only few immunotherapy trials for HCC have been studied so far with contrasting results, suggesting that significant improvements are needed. It is beyond a doubt that such an inherent immune tumor environment needs to be taken into account and counterbalanced when immunotherapeutic approaches are designed and implemented in HCC. Therefore, combination therapy of monoclonal antibodies or chemotherapies along with transarterial chemoembolization is a promising approach in increasing the overall survival of HCC patients whose chances of relapse after surgery are evident [85,86]. Pre-

vious studies have shown promising safety, efficacy and tumor targeting of the 131I-labeled metuximab and transcatheter arterial chemoembolization combination for unresectable HCC [87,88]. Combination of low dose cyclophosphamide treatment with a vaccine (e.g. telomerase vaccine) in HCC has been evaluated in a single clinical trial [89,90]. Unfortunately, the combination did not show antitumor efficacy in respect to tumor response and time to progression. Nearly 5 years, PD-1 and its ligand PD-L1 have been widely used as an immune regulating checkpoint with a well-established role in the development of HCC progression. A single clinical trial in HCC patients has been reported combining anti-PD-1 Ab and a GPC3 peptide vaccine. Results showed improved antitumor effects of a peptide vaccine correlating with the increased levels of vaccine-specific CTLs and reduced tumor-infiltrating T cells [88]. Nivolumab is an immunotherapy that inhibits PD-1. It has been used as a second line systemic treatment in HCC patients who have been treated with or intolerant to sorafenib and has been granted approval by FDA in 2017. To increase responses to immunotherapy, combination of PD-1 or PDL1 and tyrosine kinase inhibitors are currently investigated significant improved clinical outcomes. Xu et al. generated and used SHR-1210 (anti-PD-1 antibody) and apatinib (VEGFR2 inhibitor) to study combination therapy treatment with advanced HCC. It was found that combination therapy demonstrated manageable toxicity and encouraging clinical activity in patients [91]. Markus Joerger et al. also provided data from this clinical case to support the potential of combination treatment of the oral multi-kinase inhibitor regorafenib with PD-1 or PDL1 targeted monoclonal antibodies to advanced HCC therapy [92]. These discoveries can be used to promote the development of HCC immunotherapy. After the completion of genome-wide association studies (GWAS) and pharmacogenomics, personalized medicine has gradually become possible in HCC. Personalized medicine has been the mainstay for the treatment of HCC. Personal genetic analysis can hold the promise of identifying patients and family members, who would benefit from personalized medicine, modifying risk factors and so on [93,94].

3. Application of nanomedicine for hepatocellular carcinoma therapy

Although the efficiency of these few chemotherapeutic molecules in the management of HCC, the drugs are not usually delivered at high concentrations into the malignant tissues. Thereby, hydrophobicity, toxicity and bioavailability of these small molecular drugs caused dose-limiting side effects are still the deliver challenge. Nanotechnology is a powerful tool for the delivery and targeting of therapeutics in HCC [95–97]. Since HCC cells undergo genetic and phenotypic changes compared to other hepatic cells, targeting of HCC cells is an obvious avenue for treatment of HCC (Fig. 1). Various targeting and delivery strategies have been explored for nanoparticles (NPs) [98–100], micelles [101,102], liposomes [103] both passively and receptor-mediated active targeting in HCC. Normally, nanotechnologies could overcome the unfavorable side-effects of systemic administration of chemotherapeutics by improving the pharmacokinetics, biodistribution, accumulating cytotoxic agents in tumor site and elevating

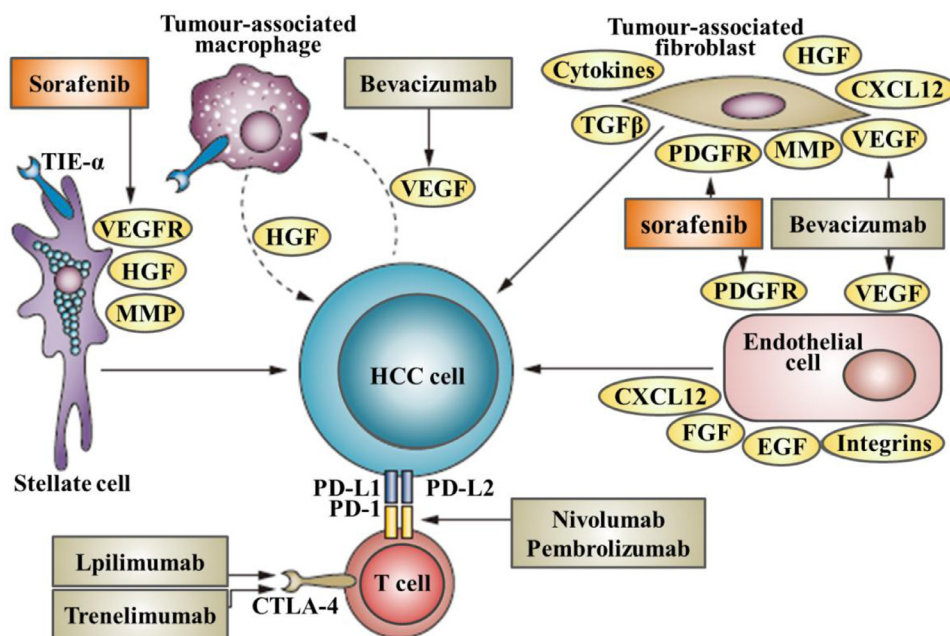


Fig. 1 – This diagram summarizes known therapies for treatment of HCC receptors, and the cellular locations of drug-target interactions.

the effectiveness of treatment through drug delivery nanosystems [104–107]. Nanometer size range of nanosystems could help drugs reach the tumor cells through the leaky vasculature and enhance site-specific enhanced delivery [108]. However, degradability, stability, circulation, metabolism as well as the balance between side effects and curative effect still have to be carefully considered for the design of efficient deliver nanosystems. Otherwise, HCC patients almost developed based on prolonged inflammatory processes, which emerged as a result of genetic and epigenetic alterations. Tumor suppressor genes, in particular p53 and retinoblastoma, are the frequently altered in HCC. Further, as a kind of highly vascular tissues, HCC progression is almost accompanied by abnormal angiogenesis at different stage and etiology. Thereby, angiogenesis related molecules such as VEGFR, RAF and EGFR are extremely useful targets in the development of selective therapeutics. Hence, the specific surface molecules of liver cells including ASGPR and endocytic cell surface receptors are highly expressed by HCC cells that are distinguished from other tissues. These specific related genotypic and phenotypic alterations have been utilized for HCC targeting diagnosis and therapy. Herein, some of the recent designs and findings in the field of nano-based medicines for passive and receptor-specific active targeting for the treatment of hepatocellular carcinogenesis would be discussed.

3.1. Passive targeting nanotechnology for hepatocellular carcinoma therapy

In case of passive targeting, nanomedicines can reach tumor via the leaky vasculature of the tumors by the enhanced permeability and retention (EPR) effect [109–111]. Nanoparticles can be delivered to specific sites by passive targeting. It also has been reported that nanoparticles injected intravenously

are taken up by the liver after only a few minutes due to the opsonization process. To date, various stimuli including pH, enzymatic, redox, light and heat have been used for passive targeting of nanoparticles carrying drugs in HCC [112,113]. Recently, nanoassemblies based on supramolecular complexes of nonionic amphiphilic cyclodextrin and sorafenib as effective weapons to kill HCC cells have been developed (Fig. 2) [114]. Sorafenib robustly interacts with nonionic amphiphilic cyclodextrin, forming supramolecular complexes that behave as building passive targeting nanoassemblies. These nanoassemblies are highly stable in aqueous medium, retaining sorafenib up to 2 weeks. Interestingly, these passive targeting systems show very low hemolytic activity and a high efficiency to inhibit the growth of three different HCC cell lines, similarly to free sorafenib, which indicated the preferential *in vivo* accumulation of nanoassemblies could result in a superior therapeutic efficacy than the free drug. Antisense-miRNA-21 and gemcitabine (GEM) co-encapsulated PEGylated-PLGA NPs have been developed to overcome drug resistance and improve their therapeutic efficacy *in vitro*. For lenvatinib, Yuan et al. have developed aliposome-encapsulated contrast agent comprising lenvatinib-bound nanoparticles for screening of early-phase non-small cell lung cancer. However, nanotechnology has not been used for enhancing the HCC therapy of lenvatinib.

3.2. Active targeting nanotechnology for hepatocellular carcinoma therapy

Although nanosystems have gained tremendous success for delivering chemotherapeutic agents, their rapid clearance by the reticuloendothelial system (RES) after systemic administration restricts them from their delivery to hepatocytes which are the main site for HCC origination [115,116]. As a result,

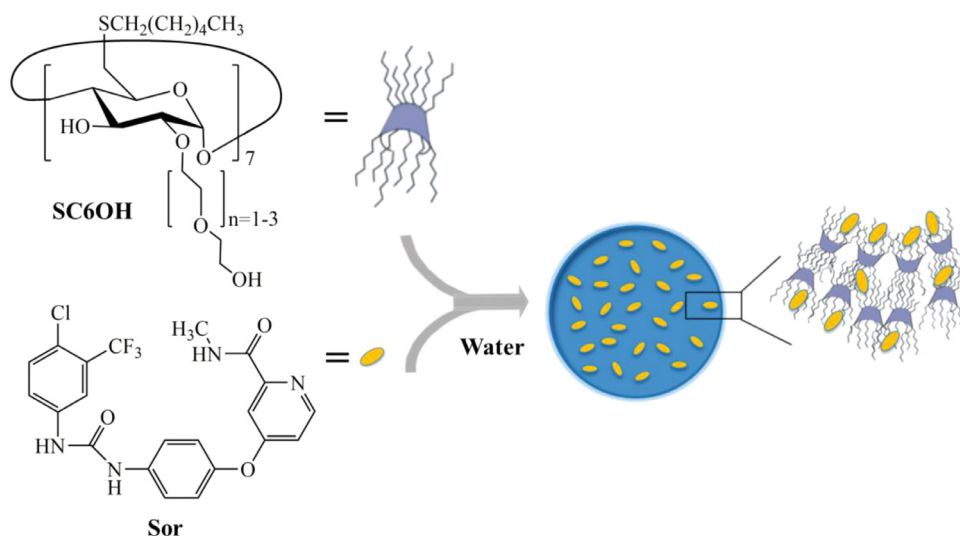


Fig. 2 – Schemed view of nanoassemblies formation in aqueous solution (Reproduced with permission from [67]. Copyright 2017 Shenyang Pharmaceutical University).

the nanotechnological approaches of site specific and/or targeted drug delivery amenable to distinct hepatic cells and their receptors are being studied extensively by researchers [117–119]. Further, active targeting was almost accompanied with passive targeting when nanosystems were designed [120–122].

Asialoglycoprotein receptor (ASGPR) has been identified as a commonly found lectin receptor which is profoundly expressed in mammalian liver cells [123,124]. Using natural ligands such as asialofetuin as well as synthetic ligands could achieve specific liver ASGPR targeting [58]. For example, nanoparticles and liposomes incorporating various lactosylated strategies have been evaluated for targeting efficacy to hepatic tumor cells. Polyethylene sebacate (PES)-Gantrez® AN 119 Dox NPs have been developed with ASGPR ligands by Sandhya et al. and showed the high efficacy coupled with greater safety as a promising nanocarrier for improved therapy of HCC [125]. Meng and colleagues successfully demonstrated the galactose-installed photo-crosslinked pH-sensitive degradable micelles (Gal-CLMs) for active targeting chemotherapy have a great potential for hepatocellular carcinoma chemotherapy (Fig. 3) [126]. In another study, cholesterol arabinogalactan anchored liposomes (CHOL-AL-AG) carrying DOX targeting ASGPR receptors on mammalian hepatocytes in HCC were studied where improved delivery of DOX was reported in terms of stability, loading efficiency, tumor regression both *in vitro* and *in vivo* compared to unmodified liposomes [127].

Integrins are transmembrane receptors which are responsible for cell–cell and cell–extracellular matrix interactions [128]. Hepatic stellate cells almost express integrins when it suffers fibrotic liver [129]. For instance, the peptide sequence RGD acts as a targeting ligand for collagen VI and RGD-coupled liposomes has been extensively applied for targeting integrin receptors in HCC [130]. To deliver the poor water soluble drug paclitaxel (PTX) effectively in HCC, Chen et al. have developed a kind of integrin- $\alpha v \beta 3$ receptor targeted liposomal PTX [131]. RGD-motif was successfully conjugated

to DSPE-PEG to prepare RGD-modified liposomes (RGD-LP). Both *in vitro* and *in vivo* studies demonstrated RGD-LP-PTX had enhanced anti-proliferative and tumor growth inhibitory effects activity against HepG2 cells and HepG2-bearing mice tumor respectively than RGD-LP or free PTX. Moreover, combination of DOX and sorafenib in iRGD decorated lipid-polymer hybrid core-shell structured NPs also indicated enhanced antitumor efficacy in HCC *in vivo* models with improved bioavailability and longer circulation time [132]. Recently, RGD modified lipid-coated NPs for the co-delivery of the hydrophobic drugs have been used by Wang et al. to against hepatocellular carcinoma. The combination of sorafenib and quercetin formulations was more effective than the single drug formulations in both NPs and solution groups. This RGD modified NPs achieved the most significant tumor growth inhibition effect *in vitro* and *in vivo* [133].

Vandetanib is an oral inhibitor of VEGFR, EGFR, RET-tyrosine kinase and has been approved for medullary thyroid cancer [134]. It has been explored for HCC as encapsulated NPs with targeting moiety with enhanced antitumor efficacy [135].

HCC cells overexpressed transferrin (Tf) receptors which have become useful targeting avenues for potential treatment [136]. The ideal of sorafenib in albumin nano-shell was used and DOX was loaded in poly (vinyl alcohol) as nano-core, the interactions were simulated and then the nanomedicines were formed by a sequential freeze-thaw/coacervation method (Fig. 5) [137]. It indicated that rationally designed core-shell nanoparticles can effectively combine clinically relevant single-agent drugs for exerting synergistic activity against liver cancer.

CL4 RNA aptamer specific to EGFR and CD133 targeted aptamers was conjugated to PLGA nanoparticles was studied for salinomycin delivery to cancer stem cells in HCC treatment [138].

Like other cancers, folate receptors are significantly overexpressed in HCC and to target these receptors, its natural ligand, folic acid (FA) has been explored for nanoparticle drug delivery to the cancer cells [3,139]. Xu et al. studied the

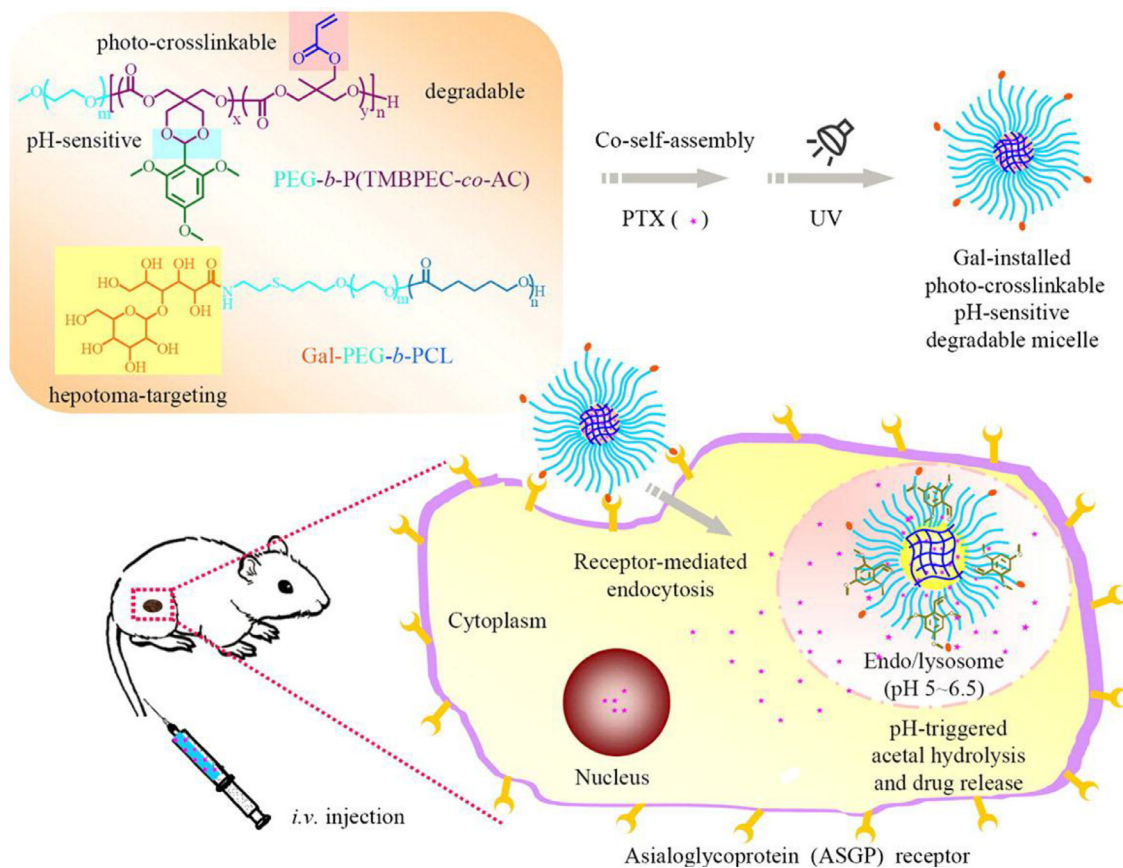


Fig. 3 – Illustration of pH-sensitive degradable micelles as an integrative nanovehicle for active targeting hepatocellular carcinoma chemotherapy (Reproduced with permission from [114]. Copyright 2015 American Chemical Society).

antitumor potency of the folate receptor-targeted liposomes (FA-PEG-DSPE) co-delivery of antitumor agent docetaxel and iSur-pDNA *in vitro* and *in vivo* [140].

VEGF receptors are found to be overexpressed in HCC due to the receptor activation via multiple signaling pathways [140,141]. Recently, iRGD-decorated polymeric NPs for the efficient delivery of vandetanib to HCC have been investigated. Wang et al. studied the biodegradable PEG-PLA to nanoprecipitate this potent agent to form water-soluble NPs that are suitable for intravenous administration [142]. Active targeting by aptamer-mediated delivery showed significant anti-tumor efficacy when tested both *in vitro* and *in vivo*. The results also demonstrate that reformulating targeted therapeutic agents in NPs permits their systemic administration and thus significantly improves the potency of currently available, orally delivered agents.

Glycyrrhetic acid (GA) receptor is overexpressed on the hepatocyte surface and has been extensively studied for drug targeting [143]. Qi et al. reviewed the roles of this receptor and GA-mediated drug delivery in HCC [144]. GA-modified nanoparticles formulations of DOX exhibited a much higher level of tumor accumulation than nontargeted NPs at 1 h after injection in hepatoma-bearing Balb/c mice. Zhang and colleagues studied poly (L-Histidine) (PHIS) mediated polymeric system (GA-PEG-PHIS-PLGA) with GA for targeted delivery of the anti-cancer agent and rographolide [145].

Passive and active targeting nanotechnologies have shown some potential for hepatocellular carcinoma therapies. However, tumor recurrence needs deeper or more thorough therapies, such as combination targeting, long circulation and so on. There are still many challenges for nanotechnology to fabricate more ideal delivery systems.

4. Influence of the architecture of nanomedicine for hepatocellular carcinoma therapy

The conjugation of ligands on the liver cells surface of nanosystems changes not only the properties of the targeting molecules but also the nanocarriers. From a chemical point of view, while ligands bind to the surface of NPs, it would lose the rotational and translational freedom bestowed to free molecules, otherwise, the new targeted entity achieves improved avidity due to the increased valency. Similarly, the size, shape, surface properties (charge, density and hydrophobicity), and composition of NPs can also be the influence factors [146,147]. Fig. 6 shows the various properties of NPs which could change the delivery behavior of drugs from NPs to HCC cells. As we know, targeted NPs have shown benefits that go beyond the simple delivery of anti HCC drugs in many cases. To fully understand the properties of targeted NPs, it is impor-

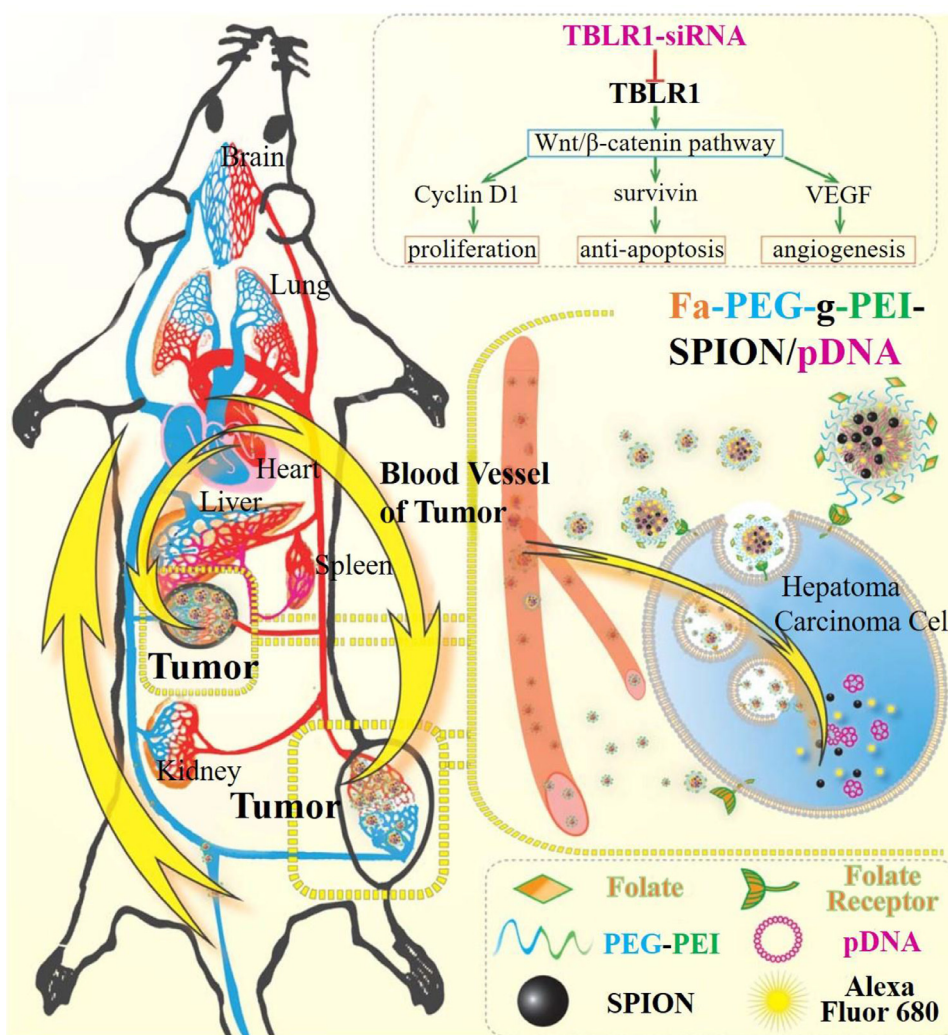


Fig. 4 – Illustration of the folate-targeted theranostic small interfering RNA (siRNA) nanomedicine Fa-PEG-g-PEI-SPION/psRNA-TBLR1 hepatocellular carcinoma chemotherapy.

tant to determine how the physicochemical properties of the NPs affect the interactions with their targets. Herein, we will discuss the main factors such as the ligand density, size and shape, surface and ligand charge and surface hydrophobicity of NPs to influence the behaviors of them.

4.1. The ligand density

For all delivery, the density of the targeting molecules on the surface of NPs impacts their affinity for the substrate due to increased cooperative effects. According to the thermodynamics, the more binding of a ligand to its substrate would promote the subsequent binding of its neighbors [148]. Biologically, increasing the ligand density would aggrandize the interactions of the NPs with the cell membrane so that force the local concentration of receptors and lead to internalization of cell membranes. These synergistic effects impede the detachment of the NPs from the cell surface and result in increased avidity [149].

The increasing ligand density usually results in improved cellular uptake *in vitro* [150]. However, targeting ligand density

is critical to maximizing the efficiency of targeting NPs to cancer cells, and further increasing in ligand density will have deleterious effects on cell binding beyond the cooperative effect of the ligand get saturate. The reason of this effect might due to the improper orientation of the ligand, steric hindrance of neighboring molecules or competitive behaviors for the binding of the receptor [151]. Similar negatively cooperative effect has been observed in ligand-clustered NPs where the ligands are arranged in patchy clusters by Hammond group [152]. Valencia et al. have investigated and showed that the use of high densities of hydrophobic ligands can increase the macrophage uptake of the NPs without providing significant advantages in terms of receptor-mediated internalization [153].

Similar effects have been observed *in vivo* where higher densities do not always result in improved efficacy. Many studies have shown that the alteration of the NPs surface to incorporate the ligands can modify the blood circulation and biodistribution profiles of the NPs. Gu et al. have developed the aptamer-targeted polymeric NPs and shown that increasing the ligand density above 5% would cause the increasing

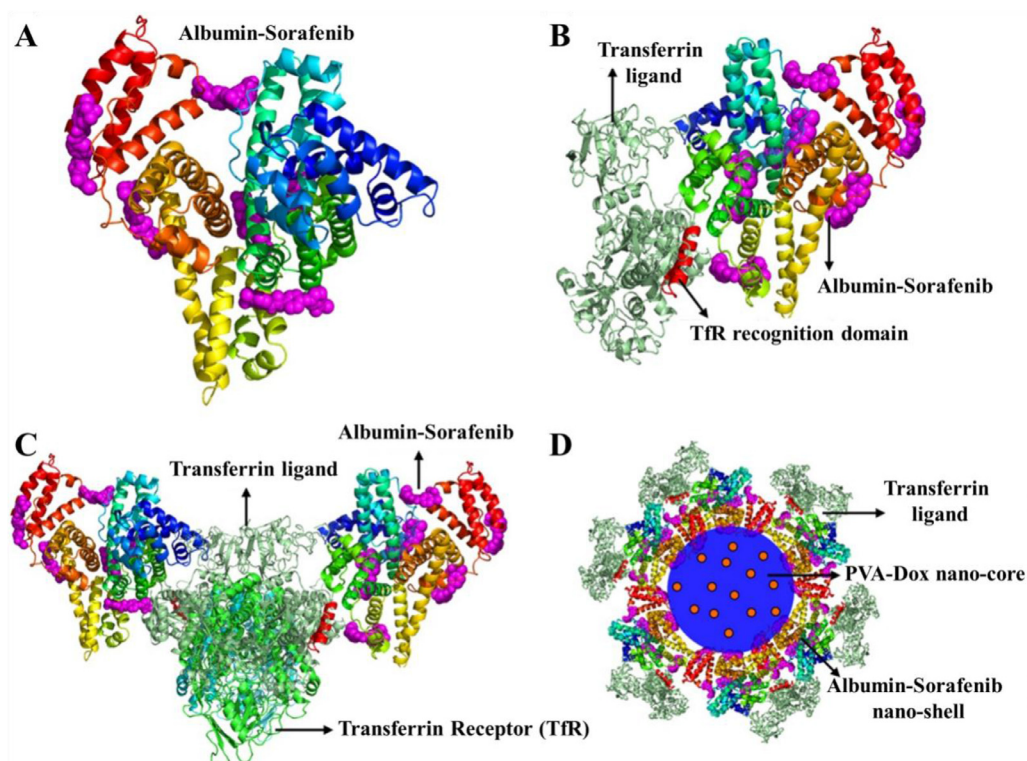


Fig. 5 – In silico docking simulation of core-shell nanomedicine (Reproduced with permission from [85]. Copyright 2016 Lin et al.).

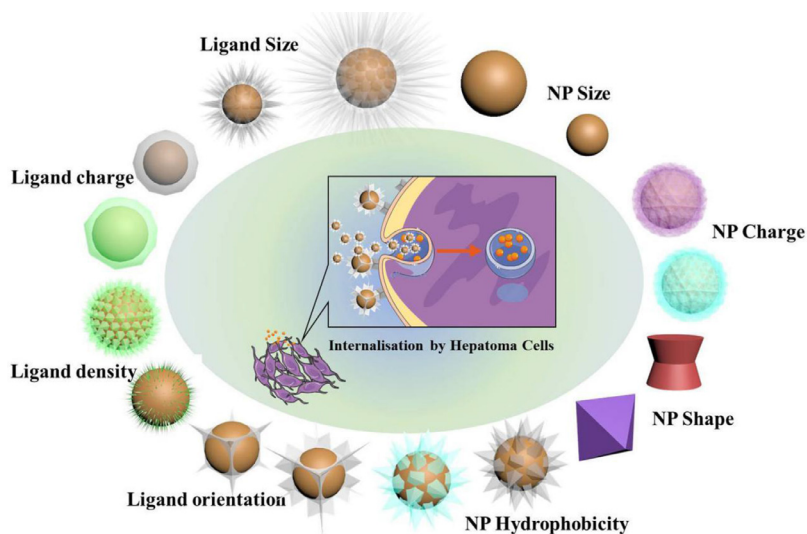


Fig. 6 – The physicochemical properties of the ligand and the NP affect their blood circulation profiles, their biodistribution and their ability to be internalized by cancer cells.

clearance of the NPs by the liver and the spleen and impeded the distribution to the tumor [154].

Finally, many studies indicated that tumor selective antibodies could suffer from a binding-site barrier preventing their in-depth diffusion in the tumor [155,156]. This phenomenon is observed when high affinity molecules rapidly bind perivascular cells surrounding the cancer cells upon their

extravasation to the tumor. The binding-site barrier also limits the in-depth diffusion of small molecular weight drugs with high affinity for HCC cells. This effect has been recently targeted to the receptors NPs which showed not enough HCC cells penetration compared to their non-targeted counterpart. Interestingly in that case, the binding-site barrier effect was not observed with larger NPs, presumably because of differ-

ences in the rates of HCC cells penetration and/or cell internalization. Therefore, it seems probable that adequately tuning the ligand density on the NP surface could mitigate the binding-site barrier effect and translate into adequate retention time and maximal cellular uptake throughout the HCC [157].

4.2. Size and shape

The size and shape of the nanomaterial must be taken into consideration early in the design of targeted NPs. For HCC, smaller sizes and spherical shape represent higher curvatures might promote hepatic targeting but can be problematic for post synthesis ligand functionalization. In addition, the tethering of high molecular weight ligands on the surface of NPs would increase their hydrodynamic radius [158]. This increase in size must always be considered in light of the possible size restrictions involved in tumor accumulation. Besides the above mentioned effects, size can also affect cellular uptake. Although, the avidity of the particle increased with size range from 2 to 70 nm, the maximum uptake is a compromise between high avidity and optimal cell membrane wrapping around the NPs. Additionally, it is not clear how these conclusions can be expanded to other systems, as the interactions between NPs and cell highly depend on the physicochemical properties of each material. *In vivo*, Lee et al. have reported that the size of actively targeted NPs could affect the intracellular deposition, not only in liver. In this case, although the intratumoral accumulation of smaller NPs was decreased due to shorter blood circulation times, the cytoplasmic and nuclear distribution of the 25-nm NPs was superior to that of the 60-nm colloids [158,159]. Moreover, the shape of NPs might influence the cell uptake kinetics and internalization pathways through modulating the nanomaterial interact with the cell surface [160].

4.3. Surface and ligand charge

Chemically, the charge of unfunctionalized NPs and that of the ligand can affect the conjugation yield and the spatial display of the ligand on the surface. Repulsive or attractive forces between the surface of the NPs and the ligand can interfere with the conjugation or affect the final ligand structure and conformation. A chemical spacer with reasonable length, such as PEG, can be helpful to reduce the effect, but might simultaneously complicate synthesis and increase the final particle size [161,162]. Previous study also showed that the final surface charge will affect the efficacy of the targeted NPs [163]. For HCC, due to the interaction between cationic NPs and negatively charged cell membranes, positively-charged NPs show increased cellular binding and uptake, in a first pass effect and non-specific manner [164]. Because most ligands are charged molecules, the NPs surface charge is determined by the combinations of ligand densities, materials, NP formulation strategies and so on. Although recent work was carried out to address how charge density affects interactions of actively targeted NPs with HCC cells and NPs charge can affect cellular uptake, it remains unclear what parameters offer the best hepatic targeting *in vivo*.

4.4. Surface hydrophobicity

Except surface charge and above factors, hydrophobicity can also affect the architecture of the ligand display. Further, this property can have serious effects since most polymeric NPs have hydrophobic cores. Previous studies have shown that during the self-assembly of polymeric hydrophobic particles, hydrophobic ligand such as folic acid could remain trapped in the particle core without being properly displayed on the surface. In some cases, the final surface hydrophobicity of NPs can also affect nonspecific interactions with HCC cells. Actively targeted NPs without steric stabilization seem to lose their substrate-binding capacity when proteins adsorb on their surface. Surface functionalization can delay adsorption of plasma proteins and has been demonstrated *in vitro* and *in vivo* where the efficacy is dependent on both circulation times and ligand–substrate interactions.

Overall, although the aforementioned factors hold true for the majority of therapeutic NPs, no general strategies have been found suitable to address the limitation of the nano-systems. For HCC therapy, assessing the efficacy of a treatment still requires time and resources, and providing matching targeted delivery nano-systems remain therefore unlikely. However, most studies have demonstrated the targeting ligand might be key factor to decide the *in vivo* profiles of nanoparticles. More and more targeted liposomes and polymeric NPs have been made to clinical development stages indicating that the decisive role of ligands. With the aim to transport in depth of tumor, multi-ligands or multi-functional ligands have been used to alternate the negative effects of mono-high ligand density.

5. Conclusion and future prospect

Since HCC is a complex multi-step process influenced by many factors for its initiation and progression, therapeutic interventions are challenging for this disease. With the rising incidence of HCC and high morbidity and mortality rates associated with this cancer, there is an urgent need to develop effective treatment and prevention strategies. We have reviewed the various targeted receptors and recent treatment modalities of HCC with emphasis on nanotechnology strategies in the field of HCC. While surgical resection and liver transplantation are the only strategies available for treating advanced stage patients, nanomedicines and immunotherapy might be promising approach for successful treatment of HCC in the future. Not only safety, toxicity, preclinical efficacy studies are required thoroughly before any clinical application of these delivery strategies are further explored. As GWAS and more comprehensive data sets become available, the clinical advantages of using nanomedicine for HCC treatment might be better understood. Based on the personalized medicine, combined nanomedicines with immunotherapy would be a fine choice for HCC therapy. Immunotherapy strategies have exhibited encouraging results for various cancer therapies. However, the dynamic nature of immune responses at the HCC sites and some other challenging such as complication of multi-immune and individual difference would lead to combination nanotechnology and immunotherapy an opportunity

and challenge coexist in clinic. With the development of immunotherapy, all together these will forecast a bright future of nanomedicine for hepatocellular carcinoma therapy when it could control and regulate the immune response from exogenous immune stimulation.

Conflicts of interest

The authors confirm that this article content has no conflict of interest. The authors alone are responsible for the content and writing of this article.

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Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ajps.2019.04.005.

REFERENCES

- Abeylath SC, Turoso E. Glycosylated polyacrylate nanoparticles by emulsion polymerization. *Carbohydr Polym* 2007;70(1):32–7.
- Arzumanyan A, Reis HMGPV, Feitelson MA. Pathogenic mechanisms in HBV- and HCV-associated hepatocellular carcinoma. *Nat Rev Cancer* 2013;13(2):123–35.
- Xia W, Low PS. Folate-targeted therapies for cancer. *J Med Chem* 2010;53(19):6811–24.
- Serper M, Taddei TH, Mehta R, et al. Association of provider specialty and multidisciplinary care with hepatocellular carcinoma treatment and mortality. *Gastroenterology* 2017;152(8):1954–64.
- Bruix J, Reig M, Sherman M. Evidence-based diagnosis, staging, and treatment of patients with hepatocellular carcinoma. *Gastroenterology* 2016;150(4):835–53.
- Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. *Hepatology* 2011;53(3):1020–2.
- Chang MH, Chen TH, Hsu HM, et al. Prevention of hepatocellular carcinoma by universal vaccination against hepatitis B virus: the effect and problems. *Clin Cancer Res* 2005;11(21):7953–7.
- Dutta R, Mahato RI. Recent advances in hepatocellular carcinoma therapy. *Pharmacol Therapeut* 2017;173:106–17.
- Bruix J, Gores GJ, Mazzaferro V. Hepatocellular carcinoma: clinical frontiers and perspectives. *Gut* 2014;63(5):844–55.
- Bruix J, Takayama T, Mazzaferro V, et al. Adjuvant sorafenib for hepatocellular carcinoma after resection or ablation (STORM): a phase 3, randomised, double-blind, placebo-controlled trial. *Lancet Oncol* 2015;16(13):1344–54.
- Bruix J, Qin S, Merle P, et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2017;389(10064):56–66.
- Llovet JM, Ricci S, Mazzaferro V, et al. Sorafenib in advanced hepatocellular carcinoma. *New Engl J Med* 2008;359(23):218–19.
- Escudier B, Eisen T, Stadler WM, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. *New Engl J Med* 2007;356(2):125–34.
- Kudo M, Finn RS, Qin S, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet* 2018;391(10126):1163–73.
- Thapa RK, Choi JY, Poudel BK, et al. Multilayer-coated liquid crystalline nanoparticles for effective sorafenib delivery to hepatocellular carcinoma. *ACS Appl Mater Inter* 2015;7(36):20360–8.
- Wu SH, Chen JJ, Kudelka A, Lu J, Zhu XL. Incidence and risk of hypertension with sorafenib in patients with cancer: a systematic review and meta-analysis. *Lancet Oncol* 2008;9(2):117–23.
- Wan J, Qiao Y, Chen X, et al. Structure-guided engineering of cytotoxic cabazitaxel for an adaptive nanoparticle formulation: enhancing the drug safety and therapeutic efficacy. *Adv Funct Mater* 2018:1804229.
- Mizrahy S, Peer D. Polysaccharides as building blocks for nanotherapeutics. *Chem Soc Rev* 2012;41(7):2623–40.
- Ferrari M. Cancer nanotechnology: opportunities and challenges. *Nat Rev Cancer* 2005;5(3):161–71.
- Gao ST, Tang GS, Hua DW, et al. Stimuli-responsive bio-based polymeric systems and their applications. *J Mater Chem B* 2019;7(5):709–29.
- Chen QL, Yang YY, Lin X, et al. Platinum (IV) prodrugs with long lipid chains for drug delivery and overcoming cisplatin resistance. *Chem Comm* 2018;54(42):5369–72.
- Feng XR, Ding JX, Gref R, Chen XS. Poly(β -cyclodextrin)-mediated polylactide-cholesterol stereocomplex micelles for controlled drug delivery. *Chinese J Polym Sci* 2017;35(6):693–9.
- Ding JX, Shi FH, Xiao CS, et al. One-step preparation of reduction-responsive poly(ethylene glycol)-poly (amino acid)s nanogels as efficient intracellular drug delivery platforms. *Polym Chem UK* 2011;2(12):2857–64.
- Hu CY, Chen Z, Wu SJ, et al. Micelle or polymersome formation by PCL-PEG-PCL copolymers as drug delivery systems. *Chinese Chem Lett* 2017;28(9):1905–9.
- Hua DW, Liu ZC, Wang F, et al. pH responsive polyurethane (core) and cellulose acetate phthalate (shell) electrospun fibers for intravaginal drug delivery. *Carbohydr Polym* 2016;151:1240–4.
- Farazi PA, Depinho RA. Hepatocellular carcinoma pathogenesis: from genes to environment. *Natur Rev Cancer* 2006;6(9):674–87.
- Zhang SH, Xin PK, Ou QM, Hollett G, Gu ZP, Wu J. Poly (ester amide)-based hybrid hydrogels for efficient transdermal insulin delivery. *J Mater Chem B* 2018;6(42):6723–30.
- Liao WZ, Yu ZQ, Lin ZH, et al. Biofunctionalization of selenium nanoparticle with dictyophora indusiata polysaccharide and its antiproliferative activity through death-receptor and mitochondria-mediated apoptotic pathways. *Sci Rep UK* 2015;5:18629–42.
- Su CW, Chiang MY, Lin YL, et al. Sodium dodecyl sulfate-modified doxorubicin-loaded chitosan-lipid nanocarrier with multi polysaccharide-lecithin nanoarchitecture for augmented bioavailability and stability of oral administration *in vitro* and *in vivo*. *J Biomed Nanotechnol* 2016;12(5):962–72.
- Li BW, Liu PL, Wu H, et al. A bioorthogonal nanosystem for imaging and *in vivo* tumor inhibition. *Biomaterials* 2017;138:57–68.

- [31] Liu YR, Kim YJ, Siriwon N, Rohrs JA, Yu ZQ, Wanga P. Combination drug delivery via multilamellar vesicles enables targeting of tumor cells and tumor vasculature. *Biotechnol Bioeng* 2018;115(6):1403–15.
- [32] Sakon M, Nagano H, Dono K, et al. Combined intraarterial 5-fluorouracil and subcutaneous interferon- α therapy for advanced hepatocellular carcinoma with tumor thrombi in the major portal branches. *Cancer* 2002;94(2):435–42.
- [33] Yu ZQ, Cai Z, Chen QL, et al. Engineering β -sheet peptide assemblies for biomedical applications. *Biomater Sci* 2016;4(3):365–74.
- [34] Liao WZ, Lai T, Chen LY, et al. Synthesis and characterization of a walnut peptides-zinc complex and its antiproliferative activity against human breast carcinoma cells through the induction of apoptosis. *J Agr Food Chem* 2016;64(7):1509–19.
- [35] Yang YY, Wang XF, Liao GC, et al. iRGD-decorated red shift emissive carbon nanodots for tumor targeting fluorescence imaging. *J Colloid Interf Sci* 2017;509:515–21.
- [36] Yu ZQ, Xu Q, Dong CB, et al. Self-assembling peptide nanofibrous hydrogel as a versatile drug delivery platform. *Curr Pharm Des* 2015;21(29):4342–54.
- [37] Yu ZQ, Schmaltz RM, Bozeman TC, et al. Selective tumor cell targeting by the disaccharide moiety of bleomycin. *J Am Chem Soc* 2013;135(8):2883–6.
- [38] Yang CB, Wang ZY, Ou CW, Chen MS, Wang L, Yang ZM. A supramolecular hydrogelator of curcumin. *Chem Commun* 2014;50(66):9413–15.
- [39] Yao C, Tian J, Wang H, et al. Loading-free supramolecular organic framework drug delivery systems(sof-DDSs) for doxorubicin:normal plasm and multidrug resistant cancer cell-adaptive delivery and release. *Chinese Chem Lett* 2017;28(4):893–9.
- [40] Xiao YF, An FF, Chen JX, Xiong SY, Zhang XH. The impact of light irradiation timing on the efficacy of nanoformula-based photo/chemo combination therapy. *J Mater Chem B* 2018;6(22):3692–702.
- [41] Zhang JF, An FF, Li YA, et al. Simultaneous enhanced diagnosis and photodynamic therapy of photosensitizer-doped perylene nanoparticles via doping, fluorescence resonance energy transfer, and antenna effect. *Chem Commun* 2013;49(73):8072–4.
- [42] Pan Y, Long MJC, Lin HC, Hedstrom L, Xu B. Magnetic nanoparticles for direct protein sorting inside live cells. *Chem Sci* 2012;3(12):3495–9.
- [43] Pan Y, Long MJC, Li XM, Shi JF, Hedstrom L, Xu B. Glutathione (GSH)-decorated magnetic nanoparticles for binding glutathione-S-transferase (GST) fusion protein and manipulating live cells. *Chem Sci* 2011;2(5):945–8.
- [44] Wang HX, Wu JP, Xu L, Xie K, Chen C, Dong YH. Albumin nanoparticle encapsulation of potent cytotoxic therapeutics shows sustained drug release and alleviates cancer drug toxicity. *Chem Commun* 2017;53(17):2618–21.
- [45] An FF, Deng ZJ, Ye J, et al. Aggregation-induced near-infrared absorption of squaraine dye in an albumin nanocomplex for photoacoustic tomography *in vivo*. *ACS Appl Mater Interfaces* 2014;6(20):17985–92.
- [46] An FF, Zhang XH. Strategies for preparing albumin-based nanoparticles for multifunctional bioimaging and drug delivery. *Theranostics* 2017;7(15):3667–89.
- [47] Bertrand N, Wu J, Xu XY, Kamaly N, Farokhzad OC. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Delivery Rev* 2014;66:2–25.
- [48] Kuang TR, Liu YR, Gong TT, Peng XF, Hu XL, Yu ZQ. Enzyme-responsive nanoparticles for anticancer drug delivery. *Curr Nanosci* 2016;12(1):38–46.
- [49] Yu YJ, Xu Q, He SS, et al. Recent advances in delivery of photosensitive metal-based drugs. *Coord Chem Rev* 2019;387:154–79.
- [50] You XR, Gu ZP, Huang J, Kang Y, Chu CC, Wu J. Arginine-based poly (ester amide) nanoparticle platform: from structure-property relationship to nucleic acid delivery. *Acta Biomater* 2018;74:180–91.
- [51] Guo H, Xu WG, Chen JJ, et al. Positively charged polypeptide nanogel enhances mucoadhesion and penetrability of 10-hydroxycamptothecin in orthotopic bladder carcinoma. *J Control Release* 2017;259:136–48.
- [52] Chen Z, Wu C, Zhang ZF, Wu WP, Wang XF, Yu ZQ. Synthesis, functionalization, and nanomedical applications of functional magnetic nanoparticles. *Chinese Chem Lett* 2018;29(11):1601–8.
- [53] Chen JJ, Ding JX, Xu WG, et al. Receptor and microenvironment dual-recognizable nanogel for targeted chemotherapy of highly metastatic malignancy. *Nano Lett* 2017;17(7):4526–33.
- [54] Turato C, Balasso A, Carloni V, et al. New molecular targets for functionalized nanosized drug delivery systems in personalized therapy for hepatocellular carcinoma. *J Control Release* 2017;268:184–97.
- [55] Lo A, Lin CT, Wu HC. Hepatocellular carcinoma cell-specific peptide ligand for targeted drug delivery. *Mol Cancer Ther* 2008;7(3):579–89.
- [56] Pei YF, Zhang T, Renault V, Zhang XG. An overview of hepatocellular carcinoma study by omics-based methods. *Acta Bioch Bioph Sin* 2009;41(1):1–15.
- [57] Kew MC. Epidemiology of chronic hepatitis B virus infection, hepatocellular carcinoma, and hepatitis B virus-induced hepatocellular carcinoma. *Pathol Biol* 2010;58(4):273–7.
- [58] Sanhueza CA, Baksh MM, Thuma B, et al. Efficient liver targeting by polyvalent display of a compact ligand for the asialoglycoprotein receptor. *J Am Chem Soc* 2017;139(9):3528–36.
- [59] Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. *Hepatology* 2011;53(3):1020–35.
- [60] Llovet JM, Villanueva A, Lachenmayer A, Finn RS. Advances in targeted therapies for hepatocellular carcinoma in the genomic era. *Nat Rev Clin Oncol* 2015;12(7):408–24.
- [61] Prieto J, Melero I, Sangro B. Immunological landscape and immunotherapy of hepatocellular carcinoma. *Nat Rev Gastro and Hepat* 2015;12(12):681–700.
- [62] Huang Y, Yang X, Xu TR, et al. Overcoming resistance to TRAIL-induced apoptosis in solid tumor cells by simultaneously targeting death receptors, c-FLIP and IAPs. *Int J Oncol* 2016;49(1):153–63.
- [63] Xu WG, Ding JX, Xiao CS, Li LY, Zhuang XL, Chen XS. Versatile preparation of intracellular-acidity-sensitive oxime-linked polysaccharide-doxorubicin conjugate for malignancy therapeutic. *Biomaterials* 2015;54:72–86.
- [64] Szakács G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov* 2006;5(3):219–34.
- [65] Wang HX, Lu ZJ, Wang LJ, et al. New generation nanomedicines constructed from self-assembling small-molecule prodrugs alleviate cancer drug toxicity. *Cancer Res* 2017;77(24):6963–74.
- [66] Wang HX, Chen JM, Xu C, et al. Cancer nanomedicines stabilized by π - π stacking between heterodimeric prodrugs enable exceptionally high drug loading capacity and safer delivery of drug combinations. *Theranostics* 2017;7(15):3638–52.
- [67] Wang LQ. Preparation and *in vitro* evaluation of an acidic environment-responsive liposome for paclitaxel tumor targeting. *Asian J Pharm Sci* 2017;12(5):470–7.

- [68] Wu F, Wang ZB, Chen WZ, et al. Advanced hepatocellular carcinoma: treatment with high-intensity focused ultrasound ablation combined with transcatheter arterial embolization. *Radiology* 2005;235(2):659–67.
- [69] Hu XL, Zhai SD, Liu GH, Xing D, Liang HJ, Liu SY. Concurrent drug unplugging and permeabilization of polyprodrug-gated crosslinked vesicles for cancer combination chemotherapy. *Adv Mater* 2018;30(21):1706307–14.
- [70] Anestopoulos I, Voulgaridou GP, Georgakilas AG, Franco R, Pappa A, Panayiotidis MI. Epigenetic therapy as a novel approach in hepatocellular carcinoma. *Pharmacol Therapeut* 2015;145:103–19.
- [71] Gonzálezvallinas M, Breuhahn K. MicroRNAs are key regulators of hepatocellular carcinoma (HCC) cell dissemination—what we learned from microRNA-494. *Hepatobil Surg Nutr* 2016;5(4):372–6.
- [72] Lyko F, Brown R. DNA methyltransferase inhibitors and the development of epigenetic cancer therapies. *J Natl Cancer I* 2005;97(20):1498–506.
- [73] Feinberg AP, Koldobskiy MA, Göndör A. Epigenetic modulators, modifiers and mediators in cancer aetiology and progression. *Nat Rev Genet* 2016;17(5):284–99.
- [74] Yeo W, Chung HC, Chan SL, et al. Epigenetic therapy using belinostat for patients with unresectable hepatocellular carcinoma: a multicenter phase i/ii study with biomarker and pharmacokinetic analysis of tumors from patients in the mayo phase ii consortium and the cancer therapeutics research group. *J Clin Oncol* 2012;30(27):3361–7.
- [75] Yokoi M, Hanaoka F. Two mammalian homologs of yeast Rad23, HR23A and HR23B, as multifunctional proteins. *Gene* 2017;597:1–9.
- [76] Bitzer M, Horger M, Giannini EG, et al. Resminostat plus sorafenib as second-line therapy of advanced hepatocellular carcinoma – the SHELTER study. *J Hepatol* 2016;65(2):280–8.
- [77] Karakatsanis A, Papaconstantinou I, Gazouli M, Lyberopoulou A, Polymeneas G, Voros D. Expression of microRNAs, miR-21, miR-31, miR-122, miR-145, miR-146a, miR-200c, miR-221, miR-222, and miR-223 in patients with hepatocellular carcinoma or intrahepatic cholangiocarcinoma and its prognostic significance. *Mol Carcino* 2013;52(4):297–303.
- [78] Miao HL, Lei CJ, Qiu ZD, et al. MicroRNA-520c-3p inhibits hepatocellular carcinoma cell proliferation and invasion through induction of cell apoptosis by targeting glypican-3. *Hepatol Res* 2014;44(3):338–48.
- [79] Chen YC, Ramjiawan RR, Reiberger T, et al. CXCR4 inhibition in tumor microenvironment facilitates anti-programmed death receptor-1 immunotherapy in sorafenib-treated hepatocellular carcinoma in mice. *Hepatology* 2015;61(5):1591–602.
- [80] Kim HS, Lee KS, Bae HJ, et al. MicroRNA-31 functions as a tumor suppressor by regulating cell cycle and epithelial-mesenchymal transition regulatory proteins in liver cancer. *Oncotarget* 2015;6(10):8089–102.
- [81] Yang NN, Ekanem NR, Sakyi CA, Ray SD. Hepatocellular carcinoma and microRNA: new perspectives on therapeutics and diagnostics. *Adv drug deliver rev* 2015;81:62–74.
- [82] Xu WG, Ding JX, Li LY, Xiao CS, Zhuang XL, Chen XS. Acid-labile boronate-bridged dextran-bortezomib conjugate with up-regulated hypoxic tumor suppression. *Chem Commun* 2015;51(31):6812–15.
- [83] Wang C, Ye YQ, Hochu GM, Sadeghifar H, Gu Z. Enhanced cancer immunotherapy by microneedle patch-assisted delivery of anti-PD1 antibody. *Nano Lett* 2016;16(4):2334–40.
- [84] Freitas-Lopes MA, Mafra K, David BA, Carvalho-Gontijo R, Menezes GB. Differential location and distribution of hepatic immune cells. *Cells* 2017;6(4):48–70.
- [85] Lin JZ, Wu LC, Bai X, et al. Combination treatment including targeted therapy for advanced hepatocellular carcinoma. *Oncotarget* 2016;7(43):71036–51.
- [86] Wang C, Ye YQ, Hu QY, Bellotti A, Gu Z. Tailoring biomaterials for cancer immunotherapy: emerging trends and future outlook. *Adv Mater* 2017;29(29):1606036–60.
- [87] He Q, Lu WS, Liu Y, Guan YS, Kuang AR. 131I-labeled metuximab combined with chemoembolization for unresectable hepatocellular carcinoma. *World J Gastroentero* 2013;19(47):9104–10.
- [88] Sawada YU, Yoshikawa T, Shimomura M, Iwama T, Endo I, Nakatsura T. Programmed death-1 blockade enhances the antitumor effects of peptide vaccine-induced peptide-specific cytotoxic T lymphocytes. *Int J Oncol* 2015;46(1):28–36.
- [89] Buonaguro L, Consortium H. Developments in cancer vaccines for hepatocellular carcinoma. *Cancer Immunol Immun* 2016;65(1):93–9.
- [90] Tagliamonte M., Tornesello M.L., Buonaguro F.M., Luigi B. Vaccine approaches in hepatocellular carcinoma. *Springer* 2017:1–17.
- [91] Xu JM, Zhang Y, Jia R, et al. Anti-PD-1 antibody SHR-1210 combined with apatinib for advanced hepatocellular carcinoma, gastric, or esophagogastric junction cancer: an open-label, dose escalation and expansion study. *Clin Cancer Res* 2019;25(2):515–23.
- [92] Joerger M, Güller U, Bastian S, Driessen C, von Moos R. Prolonged tumor response associated with sequential immune checkpoint inhibitor combination treatment and regorafenib in a patient with advanced pretreated hepatocellular carcinoma. *J Gastrointest Oncol* 2019;10(2):373–8.
- [93] Galun D, Srdic-Rajic T, Bogdanovic A, Loncar Z, Zuvella M. Targeted therapy and personalized medicine in hepatocellular carcinoma: drug resistance, mechanisms, and treatment strategies. *J Hepatocell Carcinoma* 2017;4:93–103.
- [94] Agarwal PD, Lucey MR. Genetic variations linked to hepatocellular carcinoma: personalized medicine takes a step forward. *Am J Gastroenterol* 2018;113(10):1435–6.
- [95] Depalo N, Iacobazzi RM, Valente G, et al. Sorafenib delivery nanopatform based on superparamagnetic iron oxide nanoparticles magnetically targets hepatocellular carcinoma. *Nano Res* 2017;10(7):2431–48.
- [96] Lamprecht A. Nanomedicines in gastroenterology and hepatology. *Nat Rev Gastro Hepat* 2015;12(4):195–204.
- [97] Hu XL, Hu JM, Tian J, et al. Polyprodrug amphiphiles: hierarchical assemblies for shape-regulated cellular internalization, trafficking and drug delivery. *J Am Chem Soc* 2013;135(46):17617–29.
- [98] Xu ZH, Chen LL, Gu WW, et al. The performance of docetaxel-loaded solid lipid nanoparticles targeted to hepatocellular carcinoma. *Biomaterials* 2009;30(2):226–32.
- [99] Chen JJ, Ding JX, Wang YC, et al. Sequentially responsive shell-stacked nanoparticles for deep penetration into solid tumors. *Adv Mater* 2017;29(32):1701170–1.
- [100] Ding JX, Xu WG, Zhang Y, et al. Self-reinforced endocytoses of smart polypeptide nanogels for "on-demand" drug delivery. *J Control Release* 2013;172(2):444–55.
- [101] Jin C, Qian NS, Zhao W, et al. Improved therapeutic effect of DOX-PLGA-PEG micelles decorated with bivalent fragment HAb18 F(ab')₂ for hepatocellular carcinoma. *Biomacromolecules* 2010;11(9):2422–31.

- [102] Ding JX, Chen LH, Xiao CS, Chen L, Zhuang XL, Chen XS. Noncovalent interaction-assisted polymeric micelles for controlled drug delivery. *Chem Commun* 2014;50(77):11274–90.
- [103] Terada T, Iwai M, Kawakami S, Yamashita F, Hashida M. Novel PEG-matrix metalloproteinase-2 cleavable peptide-lipid containing galactosylated liposomes for hepatocellular carcinoma-selective targeting. *J Control Release* 2006;111(3):333–42.
- [104] Perry JL, Reuter KG, Luft JC, Pecot CV, Zamboni W, DeSimone JM. Mediating passive tumor accumulation through particle size, tumor type and location. *Nano Lett* 2017;17(5):2879–86.
- [105] Huang Y, Yang X, Xu TR, et al. Overcoming resistance to TRAIL-induced apoptosis in solid tumor cells by simultaneously targeting death receptors, c-FLIP and IAPs. *Int J Oncol* 2016;49(1):153–63.
- [106] Hu XL, Liu GH, Li Y, Wang XR, Liu SY. Cell-penetrating hyperbranched polyprodrug amphiphiles for synergistic reductive milieu-triggered drug release and enhanced magnetic resonance signals. *J Am Chem Soc* 2015;137(1):362–8.
- [107] Wang HX, Xie HY, Wang JG, et al. Self-assembling prodrugs by precise programming of molecular structures that contribute distinct stability, pharmacokinetics and antitumor efficacy. *Adv Funct Mater* 2015;25(31):4956–65.
- [108] Kanapathipillai M, Brock A, Ingber DE. Nanoparticle targeting of anti-cancer drugs that alter intracellular signaling or influence the tumor microenvironment. *Adv Drug Deliver Rev* 2014;79–80:107–18.
- [109] Bazak R, Hourri M, Achy SE, Hussein W, Refaat T. Passive targeting of nanoparticles to cancer: a comprehensive review of the literature. *Mol and Clin Oncol* 2014;2(6):904–8.
- [110] Hu XL, Liu SY. Recent advances towards fabrication and biomedical applications of responsive polymeric assemblies and nanoparticle hybrid superstructures. *Dalton T* 2015;44(9):3904–22.
- [111] Zhao N, Wu BY, Hu XL, Xing D. NIR-triggered high-efficient photodynamic and chemo-cascade therapy using caspase-3 responsive functionalized upconversion nanoparticles. *Biomaterials* 2017;141:40–9.
- [112] Wang LN, Su WJ, Liu Z, et al. CD44 antibody-targeted liposomal nanoparticles for molecular imaging and therapy of hepatocellular carcinoma. *Biomaterials* 2012;33(20):5107–14.
- [113] Devulapally R, Foygel K, Sekar TV, Willmann JK, Paulmurugan R. Gemcitabine and antisense-microRNA co-encapsulated PLGA-PEG polymer nanoparticles for hepatocellular carcinoma therapy. *ACS Appl Mater Inter* 2016;8(49):33412–22.
- [114] Bondi ML, Scala A, Sortino G, et al. Nanoassemblies based on supramolecular complexes of non ionic amphiphilic cyclodextrin and sorafenib as effective weapons to kill human HCC cells. *Biomacromolecules* 2015;16(12):3784–91.
- [115] Liu XS, Li H, Chen YJ, Jin Q, Ren KF, Ji J. Mixed-charge nanoparticles for long circulation, low reticuloendothelial system clearance and high tumor accumulation. *Adv Healthc Mater* 2014;3(9):1439–47.
- [116] Brannonpeppas L, Blanchette JO. Nanoparticle and targeted systems for cancer therapy. *Adv Drug Deliver Rev* 2012;64:206–12.
- [117] Byrne JD, Betancourt T, Brannon-Peppas L. Active targeting schemes for nanoparticle systems in cancer therapeutics. *Adv Drug Deliver Rev* 2008;60(15):1615–26.
- [118] Marcucci F, Lefoulon F. Active targeting with particulate drug carriers in tumor therapy: fundamentals and recent progress. *Drug Discov Today* 2004;9(5):219–28.
- [119] Cai LL, Gu ZP, Zhong J, et al. Advances in glycosylation-mediated cancer-targeted drug delivery. *Drug Discov Today* 2018;23(5):1126–38.
- [120] Zhao YY, Chen HL, Chen X, et al. Targeted nanoparticles for head and neck cancers: overview and perspectives. *Wires Nanomed Nanobi* 2017;9(6):1469–82.
- [121] You XR, Kang Y, Hollett G, et al. Polymeric nanoparticles for colon cancer therapy: overview and perspectives. *J Mater Chem B* 2016;4(48):7779–92.
- [122] Song ZH, Chen X, You XR, et al. Self-assembly of peptide amphiphiles for drug delivery: the role of peptide primary and secondary structures. *Biomater Sci UK* 2017;5(12):2369–80.
- [123] D'Souza AA, Devarajan PV. Asialoglycoprotein receptor mediated hepatocyte targeting - Strategies and applications. *J Control Release* 2015;203:126–39.
- [124] Witzigmann D, Quagliata L, Schenk SH, Quintavalle C, Terracciano LM, Huwyler J. Variable asialoglycoprotein receptor 1 expression in liver disease: implications for therapeutic intervention. *Hepatol Res* 2016;46(7):686–96.
- [125] Pranatharthi S, Patel MD, Malshe VC, et al. Asialoglycoprotein receptor targeted delivery of doxorubicin nanoparticles for hepatocellular carcinoma. *Drug Deliv* 2017;24(1):20–9.
- [126] Zou Y, Song Y, Yang WJ, Meng FH, Liu HY, Zhong ZY. Galactose-installed photo-crosslinked pH-sensitive degradable micelles for active targeting chemotherapy of hepatocellular carcinoma in mice. *J Control Release* 2014;193:154–61.
- [127] Pathak PO, Nagarsenker MS, Barhate CR, et al. Cholesterol anchored arabinogalactan for asialoglycoprotein receptor targeting: synthesis, characterization, and proof of concept of hepatospecific delivery. *Carbohydr Res* 2015;408:33–43.
- [128] Thomas D, O'Brien T, Pandit A. Toward customized extracellular niche engineering: progress in cell-entrapment technologies. *Adv Mater* 2018;30(1):1703948–67.
- [129] Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. *Nat Rev Gastroenterol Hepatol* 2017;14(7):397–411.
- [130] Bartneck M, Warzecha KT, Tacke F. Therapeutic targeting of liver inflammation and fibrosis by nanomedicine. *Hepatobiliary Surg Nutr* 2014;3(6):364–76.
- [131] Chen LY, Liu YB, Wang WY, Liu K. Effect of integrin receptor-targeted liposomal paclitaxel for hepatocellular carcinoma targeting and therapy. *Oncol Lett* 2015;10(1):77–84.
- [132] Zhang J, Hu J, Chan HF, et al. iRGD decorated lipid-polymer hybrid nanoparticles for targeted co-delivery of doxorubicin and sorafenib to enhance anti-hepatocellular carcinoma efficacy. *Nanomed Nanotechnol* 2016;12:1303–11.
- [133] Wang C, Su L, Wu CS, Wu JL, Zhu CB, Yuan GY. RGD peptide targeted lipid-coated nanoparticles for combinatorial delivery of sorafenib and quercetin against hepatocellular carcinoma. *Drug Dev Ind Pharm* 2016;42(12):1938–44.
- [134] Bronte G, Bronte E, Novo G, et al. Conquests and perspectives of cardio-oncology in the field of tumor angiogenesis-targeting tyrosine kinase inhibitor-based therapy. *Expert Opin Drug Saf* 2015;14(2):253–67.
- [135] Qiu SY, Jiao LR. Improving detection combined with targeted therapy for small hepatocellular carcinoma. *Ann Trans Med* 2019. doi:10.21037/atm.2019.01.19.
- [136] Sciort R, Paterson AC, Eyken PV, Callea F, Kew MC, Desmet VJ. Transferrin receptor expression in human hepatocellular carcinoma: an immunohistochemical study of 34 cases. *Histopathology* 1988;12(1):53–63.

- [137] Malarvizhi GL, Retnakumari AP, Nair S, Koyakutty M. Transferrin targeted core-shell nanomedicine for combinatorial delivery of doxorubicin and sorafenib against hepatocellular carcinoma. *Nanomedicine* 2014;10(8):1649–59.
- [138] Jiang JX, Chen HW, Yu C, et al. The promotion of salinomycin delivery to hepatocellular carcinoma cells through EGFR and CD133 aptamers conjugation by PLGA nanoparticles. *Nanomedicine* 2015;10(12):1863–79.
- [139] Minami K, Hiwatashi K, Ueno S, et al. Prognostic significance of CD68, CD163 and folate receptor- β positive macrophages in hepatocellular carcinoma. *Exper Ther Med* 2018;15(5):4465–76.
- [140] Xu ZH, Zhang ZW, Chen Y, Chen LL, Lin LP, Li YP. The characteristics and performance of a multifunctional nanoassembly system for the co-delivery of docetaxel and iSur-pDNA in a mouse hepatocellular carcinoma model. *Biomaterials* 2010;31(5):916–22.
- [141] Zhu AX, Dan GD, Sahani DV, Jain RK. HCC and angiogenesis: possible targets and future directions. *Nat Rev Clin Oncol* 2011;8(5):292–301.
- [142] Wang J, Wang H, Li J, et al. iRGD-decorated polymeric nanoparticles for the efficient delivery of vandetanib to hepatocellular carcinoma: preparation and *in vitro* and *in vivo* evaluation. *ACS Appl Mater Inter* 2016;8:19228–37.
- [143] Sun YQ, Dai CM, Zheng Y, Shi SD, Hu HY, Chen DW. Binding effect of fluorescence labeled glycyrrhetic acid with GA receptors in hepatocellular carcinoma cells. *Life Sci* 2017;188:186–91.
- [144] Qi WW, Yu HY, Guo H, et al. Doxorubicin-loaded glycyrrhetic acid modified recombinant human serum albumin nanoparticles for targeting liver tumor chemotherapy. *Mol Pharm* 2015;12(3):675–83.
- [145] Zhang JM, Zhang M, Ji J, et al. Glycyrrhetic acid-mediated polymeric drug delivery targeting the acidic microenvironment of hepatocellular carcinoma. *Pharm Res Dordr* 2015;32(10):3376–90.
- [146] Blanco E, Shen H, Ferrari M. Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nat Biotechnol* 2015;33(9):941–51.
- [147] Jo DH, Kim JH, Lee TG, Kim JH. Size, surface charge, and shape determine therapeutic effects of nanoparticles on brain and retinal diseases. *Nanomed Nanotechnol* 2015;11(7):1603–11.
- [148] Rechlin C, Scheer F, Terwesten F, et al. Price for opening the transient specificity pocket in human aldose reductase upon ligand binding: structural, thermodynamic, kinetic, and computational analysis. *ACS Chem Biol* 2017;12(5):1397–415.
- [149] Weissleder R, Kelly K, Sun EY, Shtatland T, Josephson L. Cell-specific targeting of nanoparticles by multivalent attachment of small molecules. *Nat Biotechnol* 2005;23(11):1418–23.
- [150] Reuter KG, Perry JL, Kim D, Luft JC, Liu RH, DeSimone JM. Targeted PRINT hydrogels: the role of nanoparticle size and ligand density on cell association, biodistribution, and tumor accumulation. *Nano Lett* 2015;15(10):6371–8.
- [151] Chen H, Paholak H, Ito M, et al. ‘Living’ PEGylation on gold nanoparticles to optimize cancer cell uptake by controlling targeting ligand and charge densities. *Nanotechnology* 2013;24(35):355101–8.
- [152] Poon Z, Chen S, Engler AC, et al. Ligand-clustered “patchy” nanoparticles for modulated cellular uptake and *in vivo* tumor targeting. *Angew Chem Int Ed* 2010;49(40):7266–70.
- [153] Valencia PM, Hanewichhollatz MH, Gao W, et al. Effects of ligands with different water solubilities on self-assembly and properties of targeted nanoparticles. *Biomaterials* 2011;32(26):6226–33.
- [154] Gu F, Zhang L, Teply BA, et al. Precise engineering of targeted nanoparticles by using self-assembled biointegrated block copolymers. *P Natl Acad Sci USA* 2008;105(7):2586–91.
- [155] Miao L, Newby JM, Lin CM, et al. The binding site barrier elicited by tumor-associated fibroblasts interferes disposition of nanoparticles in stroma-vessel type tumors. *ACS Nano* 2016;10(10):9243–58.
- [156] Liu D, Auguste DT. Cancer targeted therapeutics: from molecules to drug delivery vehicles. *J Control Release* 2015;219:632–43.
- [157] Park JO, Stephen Z, Sun C, et al. Glypican-3 targeting of liver cancer cells using multifunctional nanoparticles. *Mol Imaging* 2011;10(1):69–77.
- [158] Jiang W, Kim BYS, Rutka JT, Chan WCW. Nanoparticle-mediated cellular response is size-dependent. *Nat Nanotechnol* 2008;3(3):145–50.
- [159] Lee H, Fonge H, Hoang B, Reilly RM, Allen C. The effects of particle size and molecular targeting on the intratumoral and subcellular distribution of polymeric nanoparticles. *Mol Pharm* 2010;7(4):1195–208.
- [160] Gratton SEA, Ropp PA, Pohlhaus PD, et al. The effect of particle design on cellular internalization pathways. *Proc Natl Acad Sci USA* 2008;105(33):11613–18.
- [161] Stefanick JF, Ashley JD, Kiziltepe T, Bilgicer B. A systematic analysis of peptide linker length and liposomal polyethylene glycol coating on cellular uptake of peptide-targeted liposomes. *ACS Nano* 2013;7(4):2935–47.
- [162] Krishnan G, Subramaniyan J, Subramani PC, Muralidharan B, Thiruvengadam D. Hesperetin conjugated PEGylated gold nanoparticles exploring the potential role in anti-inflammation and anti-proliferation during diethylnitrosamine-induced hepatocarcinogenesis in rats. *Asian J Pharm Sci* 2017;12(5):442–55.
- [163] Zhao F, Zhao Y, Liu Y, Chang XL, Chen CY, Zhao YL. Cellular uptake, intracellular trafficking, and cytotoxicity of nanomaterials. *Small* 2011;7(10):1322–37.
- [164] Davis ME, Chen ZG, Shin DM. Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nat Rev Drug Discov* 2008;7(9):771–82.