



Perspective: The Lung, Particles, Fibers, Nanomaterials, and Autoimmunity

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Studies have shown that a wide range of factors including drugs, chemicals, microbes, and other environmental agents can induce pre-clinical autoimmunity. However, only a few have been confidently linked to autoimmune diseases. Among these are exposures to inhaled particulates that are known to be associated with autoimmune diseases such as lupus and rheumatoid arthritis. In this article, the potential of particle, fiber, and nanomaterial exposures to induce autoimmunity is discussed. It is hypothesized that inhalation of particulate material known to be associated with human autoimmune diseases, such as cigarette smoke and crystalline silica, results in a complex interplay of a number of pathological processes, including, toxicity, oxidative stress, cell and tissue damage, chronic inflammation, post-translational modification of self-antigens, and the formation of lymphoid follicles that provide a milieu for the accumulation of autoreactive B and T cells necessary for the development and persistence of autoimmune responses, leading to disease. Although experimental studies show nanomaterials are capable of inducing several of the above features, there is no evidence that this matures to autoimmune disease. The procession of events hypothesized here provides a foundation from which to pursue experimental studies to determine the potential of other environmental exposures to induce autoimmunity and autoimmune disease.

Keywords: lung, autoimmunity, particle, nanoparticle, fiber

INTRODUCTION

Numerous drugs, chemicals, microbes, and other environmental factors have been identified as possible causes of pre-clinical autoimmunity (1). However, very few have been confidently associated with human autoimmune diseases (2). Among these are respirable particulates such as cigarette smoke and silica dust (2) which have been linked to systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), and systemic sclerosis (SSc) (2, 3). Asbestos, a fibrous silicate mineral, has also been associated with autoimmunity, although linkage to specific autoimmune diseases is less well established (2, 4). In contrast, a role for nanomaterial [materials with at least one dimension between 1 and 100 nanometers (5, 6)] exposure as a causative agent in human autoimmunity remains to be established.

In reviewing the etiopathogenesis of environment-induced autoimmunity, we (7) proposed that the toxic insult of an environmental exposure initiates a multi-step process characterized by tissue

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damage, and the release of nucleic acids and other damage associated molecular patterns (DAMPs) including self- and modified self-antigens, which induces an innate inflammatory response, leading to adaptive immunity involving the presentation of self- and modified self-antigens to non-tolerant lymphocytes. Included within this process is the engagement of Toll-like receptors (TLRs) and other innate sensors, the production of inflammatory mediators, the expansion of autoreactive B and T effector cell populations, and the production of autoantibodies. These events often occur at the site of exposure, where they can lead to the development of tertiary lymphoid structures (TLSs), and/or germinal centers in secondary lymphoid organs draining the site of exposure. In either case the expansion of autoreactive lymphocytes and their migration to target tissues, such as the kidney in SLE or the joints in RA, results in autoimmune disease (7).

To include an additional perspective to the above hypothesis, this article compares the current state of knowledge regarding the response of the immune system to selected particle, fiber, and nanomaterial exposures. Specific particulate and fibrous materials are discussed because of their known role in preclinical autoimmunity and autoimmune diseases (2). This is contrasted with the immunological responses to nanomaterials, particularly engineered nanoparticles (ENPs), where evidence for induction of autoimmunity is less convincing.

PARTICULATE MATERIAL

Cigarette Smoke

Cigarette smoke is a complex mixture of particulate-phase and chemical compounds (8, 9), and is a significant risk factor for chronic obstructive pulmonary disease (COPD) (10). Smoking is also a significant risk factor for RA (11), and has been linked to other autoimmune diseases including SLE, multiple sclerosis, and Crohn's disease (2). The linkage of smoking and RA is strongest for seropositive disease particularly patients with anticitrullinated protein antibody (ACPA) (2) which is itself strongly linked to the HLA shared epitope (12). The association between smoking, RA, and the HLA-linked ACPA response identifies an important gene-environment interaction for autoimmune diseases (13).

The particulate material of cigarette smoke consists of partially combusted plant material between >0.25 to <1 um in size (8, 9, 14). The chronic inflammation resulting from cigarette smoke exposure is thought to be due to this particulate material and gas-phase chemicals but may also involve bacterial and fungal contaminants of tobacco (15). Cigarette smoke also contains reactive oxygen species (ROS), that may contribute to post-translational modification of proteins and other markers of oxidative stress (16). Tissue damage from inhaled smoke leads to activation of lung epithelial cells and alveolar macrophages (AM) *via* interaction between DAMPs and pattern recognition receptors (PRRs) such as TLRs (16). This initiates an innate immune response, characterized by inflammasome formation and the production of IL-1 β , IL-18, and other proinflammatory mediators including IL-6 and TNF- α (16). The resulting

infiltration of neutrophils and monocytes further exacerbates oxidative stress and inflammation. Additional processes, such as neutrophil death, mediated by NETosis, also leads to cell and tissue damage (17).

The innate inflammatory response progresses to adaptive immunity consisting of T helper cell responses, including Th1 and Th17 (18, 19), and B cells that can form lymphoid aggregates in the lung (10). These structures are similar to TLSs found in target organs of autoimmune diseases where they are argued to provide a microenvironment for the survival of autoreactive lymphocytes (20, 21). The presence of plasma cells in lung TLSs (21) suggests that the lungs are a source of autoantibodies, and this is supported by the presence of autoantibodies in bronchoalveolar lavage (BAL) fluid (21, 22). Moreover, autoantibodies to modified self-antigens have been described in COPD (21, 23, 24) including autoantibodies known to be important in seropositive RA [rheumatoid factor (RF) and ACPA] (11, 25). These events may occur years before clinical diagnosis of RA (26). It has been hypothesized that localized chronic inflammatory processes in the lungs following smoke exposure are important in the pre-clinical phase of not only diseases such as COPD but also autoimmune diseases such as RA (25, 27). Importantly, experimental studies have shown that TLSs and autoantibodies persist after ceasing smoking and are dependent upon IL-1 receptor 1 (28).

While COPD and RA are comorbidities, their pathological relationship remains under investigation (26, 29). Development of COPD in RA patients (29) is consistent with the known pulmonary involvement in RA (25, 30). Conversely, a recent study identified COPD as a risk factor for RA with the strongest association between COPD and seropositive RA in older smokers (31). Interestingly, individuals who were ACPA positive before RA diagnosis were at increased risk of developing COPD (32). This suggests a linkage between appearance of ACPA and susceptibility to respiratory disease. ACPA may also be an outcome of inhalation of other forms of particulate matter, and may identify inhalant exposures that increase the risk of RA or other autoimmune diseases (33).

Silica Dust

Exposure to crystalline silica as a result of the breakdown of quartz (e.g. mining, sandblasting, quarrying, ceramics), or during the fabrication of artificial stone (34, 35), can lead to silicosis (36), COPD (36), lung cancer (37), and autoimmune diseases including SLE, RA, SSc, and antineutrophil cytoplasmic antibody (ANCA)-related vasculitis (2, 35, 38). The occurrence of COPD in silica-exposed individuals may also reflect coincident cigarette smoke exposure (39). Significantly, when silica exposure and smoking occur together it results in a synergistic interaction which, for example, greatly increases the risk of ACPA-positive RA in Asian (40) and Caucasian (41) populations. Silica dust exposure is most often via occupational inhalation with deposition of respirable particles [$<10\mu m$ (42)] in the alveoli of the lungs leading to chronic inflammation and development of fibrotic nodules (36). Although the mechanistic aspects of the pathogenic process of silicosis have not been completely defined, a number of important checkpoints have been

identified (36, 43, 44). The cumulative dose and physicochemical properties of silica dust are important in inflammatory and fibrogenic responses (43, 45). Inhaled respirable silica particles can be deposited throughout the lung including the distal airways and alveoli leading to direct cytotoxicity (46). Silica particles also interact with endogenous molecules leading to the formation of a surface corona (43). Subsequent phagocytosis by AM leads to lysosomal damage resulting in activation of inflammatory and cell death pathways. A significant contributor to these processes is oxidative stress due to ROS originating from both silica particles and lung cells (47). The chronic persistence of silica particles in the lung results in fibrosis, silicosis (43, 48), and development of silica-induced autoimmunity (49).

An important observation in the initiation of pulmonary inflammation following crystalline silica inhalation is the release of IL-1a from AM following silica induced cellular damage (50). Release of IL-1 α from dying cells is known as an important mediator of sterile inflammation (51) following interaction between IL-1a released from crystal damaged cells and IL-1 receptor on surrounding macrophages (48, 50). Silicainduced cell death also results in the release of numerous types of DAMPs that are recognized by TLRs and other PRR on the cell surface as well as within cytoplasmic components such as endosomes. These interactions lead to intracellular signaling, activation of transcription factors such as NF-KB and AP-1, and the expression of inflammatory cytokines such as IL-1β, IL-6, TNF-a, and interferons (IFNs) (48). Phagocytosis of silica particles via scavenger receptors such as MARCO (SR-A6) and SR-A1 leads to lysosomal destabilization and release of proteases including cathepsins that contribute to the activation of the NLRP3 inflammasome and cleavage of IL-1ß and IL-18 into their bioactive forms (52-54). Generation of ROS, either from the surface of silica crystals or phagocytosis and lysosomal/ mitochondrial damage, also contributes to these events (45, 47). The end result is an inflammatory response consisting of neutrophils, monocytes and lymphocytes that exhibits acute and chronic phases which have differing molecular and cellular features (35). The failure to clear silica particles from the lungs exacerbates this chronic inflammatory process leading to fibroblast proliferation and collagen production resulting in fibrosis and silicosis (35, 36, 44).

Epidemiological studies have found associations between occupational silica exposure and SLE, SSc, and ANCA-related vasculitis (2, 55, 56). More recently, exposure to silica-containing dust from artificial stone during fabrication of countertops has been linked to both silicosis and autoimmune disease (34, 57). Pre-clinical features of autoimmunity, such as autoantibodies, as well as autoimmune diseases can occur in the absence of silicosis (35, 55) suggesting that events prior to fibrosis are important for silica-induced autoimmunity. The fibrosis following particulate exposure, including silica, may be the result of exaggerated and persistent immunosuppression due to TGF- β and IL-10, rather than chronic inflammation (58).

A significant outcome of the chronic pulmonary inflammation induced by crystalline silica, in animal models, is the formation of TLS (59). These structures comprise accumulations of B and T cells as well as follicular dendritic cells (FDC) and plasma cells (59-61). The presence of immunoglobulin including autoantibodies in the BALF of silica-exposed mice (60, 61) suggests that the lung is a site of autoantibody production. A variety of autoantibody specificities are found in both humans (62) and experimental models (59, 60, 63) after silica exposure. These include anti-DNA, -SS-A/Ro, -SS-B/La, -Scl70, -Sm, and -RNP. ACPA have also been described in RA patients exposed to silica (40). A possible mechanism for citrullination may involve silica-induced increases in peptidylargininedeiminase (PAD) activity (64). This is consistent with the formation of neutrophil extracellular traps (NETs) during NETosis induced by silica (65) which requires PAD mediated protein citrullination (66). Although autoantibody responses in silica exposed patients with autoimmune diseases are often consistent with those of idiopathic autoimmune diseases, it is unclear if they play a pathogenic role. As with smoking, it is also unclear how expression of silica-induced autoreactivity in the lung results in pathogenesis in organs such as the kidney in SLE or the joints in RA.

FIBROUS MATERIAL

Asbestos

Asbestos describes six naturally occurring fibrous silicate minerals categorized into two groups, serpentine (chrysotile) and amphibole (crocidolite, amosite, tremolite, anthophyllite, actinolite) (67). While there are significant differences in chemical composition and crystalline structure, serpentine fibers are more curvilinear and softer than the needle-like, brittle amphibole fibers (67). Occupational exposures, primarily from mining, construction, and automotive industries, can result in asbestosis and malignancy (68). The properties of asbestos fibers that cause toxicity and pathology, especially inflammation, are uncertain (69), consequently very little is known about the fiber properties necessary for autoimmunity. Short fibers (<5 um), although abundant in the lungs (70), are considered unlikely to cause malignancy (71), but it is unclear if this applies to autoimmunity.

Evidence from both human and experimental animal studies suggest that amphibole asbestos exposure can lead to autoimmunity (4, 72). Several different cohorts including Libby, Montana (73), Wittenoom, Western Australia (74), and Biancavilla, Sicily (75), have linked autoantibodies, including ANA, to asbestos exposure. Studies of the communities of Libby and Troy in Montana have found that almost 14% have been diagnosed with an autoimmune disease other than diabetes (76, 77) and mortality to autoimmune diseases is higher than expected (78). However, a causal role for asbestos exposure in autoimmune disease has not been established due to study limitations (4), and other confounders, including the possibility of concurrent silica and asbestos exposures (2), and smoking history (79).

Asbestos inhalation results in chronic pulmonary inflammation (48), mediated in part by NLRP3 inflammasome activation, and expression of IL-1 β and other inflammatory cytokines (80, 81). This is facilitated by asbestos fiber-induced

damage to lung epithelial cells, phagocytosis of asbestos particles by macrophages, lysosomal destabilization, subsequent cellular stress and damage leading to production of ROS and oxidative injury, culminating in a fibrogenic response including release of TGF- β , TNF- α , and IL-1 β , that promotes collagen deposition and fibrosis (82, 83). Pulmonary inflammation can include increases in AMs and neutrophils, as well as CD4⁺ T cells that spontaneously release IFN- γ , identified as diffuse lymphocyticmacrophage alveolitis (84).

Autoimmune diseases associated with Libby Asbestiform Amphibole (LAA) exposure include SLE, SSc, and RA (76, 78). ANA are common and include autoantibodies against dsDNA, SS-A/Ro52, Scl70, Sm, and RNP (76). Anti-mesothelial cell antibodies, and ANA, have been linked to pleural abnormalities (85). Autoantibodies against cyclic citrullinated peptide (CCP) and RF are not elevated (86) even though there is evidence of increased PAD2/4 and citrullination in the lung following asbestos exposure (87). ANA have also been found in subjects exposed to asbestos without systemic autoimmune diseases (75). This may be an indicator of pre-clinical autoimmunity, similar to the appearance of ACPA prior to COPD or RA (11, 25), as appearance of autoantibodies can precede diagnosis of autoimmune disease (88, 89). Whether autoantibodies can be linked to TLS in the lung in asbestosis, as has been observed for cigarette smoke and crystalline silica exposures, remains to be investigated.

NANOMATERIALS

Nanomaterials constitute two major groups, ambient ultrafine particles (UFPs) and ENPs (5, 6, 90). Numerous nanomaterial approaches are being developed to modulate immune responses (91, 92). This includes nanocarriers for drugs, vaccines, antigens and adjuvants (91–93) as therapies for chronic inflammation (91), infection (92), and autoimmunity (91, 92, 94) including specific tolerance approaches for treating autoimmunity (94, 95). The safety of these nanomedicines is dependent upon their ability to avoid toxicity and immune recognition

(96, 97) which may result in adverse toxicological (98) and immunological outcomes (99). Although UPFs and ENPs display a range of physicochemical properties (6), inhalation is a common route of exposure (90, 100) and can lead to tissue damage, protein corona, oxidative stress, inflammasome activation, proinflammatory mediators, and inflammation (5, 6, 99, 101), as well as pathological outcomes including fibrosis (58, 102). However, a role for NPs, particularly ENPs, in the causation of pre-clinical human autoimmunity or autoimmune diseases has not been established.

Nanodiamond NPs induce lysosomal damage and NETosis (103) but this leads to resolution of inflammation (103, 104) presumably by sequestration of the offending particles (105). There is evidence for this as NET-related proteins were found in BAL fluid in the acute phase response to TiO₂ NPs but disappeared over time, and were not associated with histopathological changes (106). Alternatively, solubility or clearance by phagocytes may lead to resolution of NP induced inflammation (103, 107). Nonetheless, nanoparticles appear capable of inducing post-translational protein modification and autoantibodies. Several nanoparticles including, SiO₂ NPs, cadmium NPs, ultrafine carbon black, and single-wall carbon nanotubes (SWCNT), elicited in vitro and/or in vivo increases in PAD activity and/or protein citrullination (64, 87, 108), and nickel nanowires stimulated anti-cyclic citrullinated (anti-CCP3) autoantibodies in female C57BL/6 mice (87). However, histological images do not show evidence of pulmonary TLSs following SWCNT exposure (109) or at sites of protein citrullination (64). Thus, although ENPs appear capable of inducing features of experimental pre-clinical autoimmunity (Table 1), studies have yet to show that this can mature to autoimmune disease.

DISCUSSION

Particulate exposures are among the initiating events most closely linked to human autoimmune diseases (2, 3), with the lungs as a major site of pathological and immunological events

TABLE 1 | Comparison of hypothesized steps leading to autoimmune disease following exposure to inhaled materials.

Features of Exposure	Silica dust	Cigarette smoke	Asbestos	Nanomaterial
Exposure site	Lung	Lung	Lung	Lung
Inflammatory response components	ROS	ROS	ROS	ROS
	Inflammasome	Inflammasome	Inflammasome	Inflammasome
	IL-1α, IL-1β, IL-18, IL-6, TNF-α	IL-1β, IL-18, IL-6,	IL-1β, TGF-β, TNF-α, IFN-γ	NETosis
	Interferons	TNF-α		
	NETosis	NETosis		
Self-protein citrullination	Yes	Yes	Yes	Yes
Tertiary lymphoid structure in lung	Yes	Yes	Alveolitis	ND
Autoantibodies	ACPA, DNA, SS-A/Ro, SS-B/La, Scl70,	RF, ACPA	DNA, SS-A/R052, Scl70, Sm, RNP	ACPA [×]
	Sm, RNP		Mesothelial cell	
Autoimmune diseases associated with exposure	SLE, SSc, RA, ANCA-related vasculitis	Seropositive RA SLE, MS	Rheumatological symptoms of SLE, SSc, RA	ND

Comparison of the hypothesized steps, as described in the text, leading to autoimmune disease following exposure to different inhaled materials. Silica dust and cigarette exposures are linked to the autoimmune diseases indicated. Exposure to asbestos is linked to features of autoimmunity, although evidence to support causation of human autoimmune disease is insufficient. There is no evidence that nanomaterial exposure leads to human autoimmune disease. X, evidence from experimental study only. ND, not determined.

leading to clinical disease (26, 27, 33, 110). Such exposures can be protracted and involve particulates resistant to degradation, both of which contributes to the chronic pulmonary inflammation associated with these exposures (48, 79). A complex interplay between pathological processes, including, cellular toxicity, oxidative stress, tissue damage, persistent inflammation, posttranslational modification of self-antigens, and the formation of TLSs, promote the generation of autoantibodies that contribute to development of autoimmunity. An unresolved issue is how expression of particulate-induced autoreactivity in the lung results in disease specific pathogenesis in distant organs (e.g. kidney in SLE or joints in RA). However, a recent hypothesis suggests that activated B cells act on preinflamamtory mesenchymal (PRIME) cells which then migrate to the joint in RA (111). The role of pulmonary mesenchymal cells in inflammation (112) supports the possibility of their interaction with B cells in lung TLSs.

Fibrous (asbestos) and non-fibrous (cigarette smoke, crystalline silica) fine (PM_{2.5}) particulates have been linked to pre-clinical autoimmunity and autoimmune diseases (2, 76). However, there is little information to support a role for ultrafine (PM_{0.1}) particulate matter in human autoimmune disease. This may reflect a difference in size which allows NPs to be cleared from the lung more readily. However, other aspects of pulmonary inflammation including the chronicity and/or resolution of the response, and the apparent absence of TLS, are likely to limit the severity of the adaptive autoimmune response and subsequent development of pathology in target organs other than the lung.

The pathological processes discussed above provide a foundation from which to determine the potential of other

REFERENCES

- Germolec D, Kono DH, Pfau JC, Pollard KM. Animal models used to examine the role of the environment in the development of autoimmune disease: Findings from an NIEHS Expert Panel Workshop. J Autoimmun (2012) 39(4):285–93. doi: 10.1016/j.jaut.2012.05.020
- Miller FW, Alfredsson L, Costenbader KH, Kamen DL, Nelson LM, Norris JM, et al. Epidemiology of environmental exposures and human autoimmune diseases: Findings from a National Institute of Environmental Health Sciences Expert Panel Workshop. J Autoimmun (2012) 39(4):259–71. doi: 10.1016/j.jaut.2012.05.002
- Parks CG, Miller FW, Pollard KM, Selmi C, Germolec D, Joyce K, et al. Expert panel workshop consensus statement on the role of the environment in the development of autoimmune disease. *Int J Mol Sci* (2014) 15 (8):14269–97. doi: 10.3390/ijms150814269
- Pfau JC. Immunotoxicity of asbestos. Curr Opin Toxicol (2018) 10:1–7. doi: 10.1016/j.cotox.2017.11.005
- Madl AK, Plummer LE, Carosino C, Pinkerton KE. Nanoparticles, lung injury, and the role of oxidant stress. *Annu Rev Physiol* (2014) 76:447–65. doi: 10.1146/annurev-physiol-030212-183735
- Yang W, Wang L, Mettenbrink EM, DeAngelis PL, Wilhelm S. Nanoparticle Toxicology. Annu Rev Pharmacol Toxicol (2020). doi: 10.1146/annurevpharmtox-032320-110338
- Pollard KM, Christy JM, Cauvi DM, Kono DH. Environmental Xenobiotic Exposure and Autoimmunity. *Curr Opin Toxicol* (2018) 10:15–22. doi: 10.1016/j.cotox.2017.11.009
- 8. John G, Kohse K, Orasche J, Reda A, Schnelle-Kreis J, Zimmermann R, et al. The composition of cigarette smoke determines inflammatory cell

particles and fibers to induce autoimmunity. Such studies will provide a better understanding of the physical and chemical properties of particulate matter that lead to the induction and propagation of autoimmune diseases.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

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recruitment to the lung in COPD mouse models. Clin Sci (Lond) (2014) 126(3):207-21. doi: 10.1042/CS20130117

- Braun M, Koger F, Klingelhofer D, Muller R, Groneberg DA. Particulate Matter Emissions of Four Different Cigarette Types of One Popular Brand: Influence of Tobacco Strength and Additives. *Int J Environ Res Public Health* (2019) 16(2):263. doi: 10.3390/ijerph16020263
- Brandsma CA, Van den Berge M, Hackett TL, Brusselle G, Timens W. Recent advances in chronic obstructive pulmonary disease pathogenesis: from disease mechanisms to precision medicine. *J Pathol* (2020) 250(5):624– 35. doi: 10.1002/path.5364
- Ishikawa Y, Terao C. The Impact of Cigarette Smoking on Risk of Rheumatoid Arthritis: A Narrative Review. *Cells* (2020) 9(2)475. doi: 10.3390/cells9020475
- Bowes J, Barton A. Recent advances in the genetics of RA susceptibility. *Rheumatol (Oxford)* (2008) 47(4):399–402. doi: 10.1093/rheumatology/ken005
- Lee YH, Bae SC, Song GG. Gene-environmental interaction between smoking and shared epitope on the development of anti-cyclic citrullinated peptide antibodies in rheumatoid arthritis: a meta-analysis. *Int J Rheum Dis* (2014) 17(5):528–35. doi: 10.1111/1756-185X.12307
- Kant N, Muller R, Braun M, Gerber A, Groneberg D. Particulate Matter in Second-Hand Smoke Emitted from Different Cigarette Sizes and Types of the Brand Vogue Mainly Smoked by Women. *Int J Environ Res Public Health* (2016) 13(8):479. doi: 10.3390/ijerph13080799
- Pauly JL, Smith LA, Rickert MH, Hutson A, Paszkiewicz GM. Review: Is lung inflammation associated with microbes and microbial toxins in cigarette tobacco smoke? *Immunol Res* (2010) 46(1-3):127–36. doi: 10.1007/s12026-009-8117-6
- Hikichi M, Mizumura K, Maruoka S, Gon Y. Pathogenesis of chronic obstructive pulmonary disease (COPD) induced by cigarette smoke. *J Thorac Dis* (2019) 11 (Suppl 17):S2129–S40. doi: 10.21037/jtd.2019.10.43

- 17. Grabcanovic-Musija F, Obermayer A, Stoiber W, Krautgartner WD, Steinbacher P, Winterberg N, et al. Neutrophil extracellular trap (NET) formation characterises stable and exacerbated COPD and correlates with airflow limitation. *Respir Res* (2015) 16:59. doi: 10.1186/s12931-015-0221-7
- Shan M, Cheng HF, Song LZ, Roberts L, Green L, Hacken-Bitar J, et al. Lung myeloid dendritic cells coordinately induce TH1 and TH17 responses in human emphysema. *Sci Trans Med* (2009) 1(4):4ra10. doi: 10.1126/ scitranlsmed.3000154
- Lee SH, Goswami S, Grudo A, Song LZ, Bandi V, Goodnight-White S, et al. Antielastin autoimmunity in tobacco smoking-induced emphysema. *Nat Med* (2007) 13(5):567–9. doi: 10.1038/nm1583
- Pipi E, Nayar S, Gardner DH, Colafrancesco S, Smith C, Barone F. Tertiary Lymphoid Structures: Autoimmunity Goes Local. *Front Immunol* (2018) 9:1952. doi: 10.3389/fimmu.2018.01952
- Polverino F, Seys LJ, Bracke KR, Owen CA. B cells in chronic obstructive pulmonary disease: moving to center stage. *Am J Physiol Lung Cell Mol Physiol* (2016) 311(4):L687–L95. doi: 10.1152/ajplung.00304.2016
- 22. Reynisdottir G, Karimi R, Joshua V, Olsen H, Hensvold AH, Harju A, et al. Structural changes and antibody enrichment in the lungs are early features of anti-citrullinated protein antibody-positive rheumatoid arthritis. *Arthritis Rheumatol* (2014) 66(1):31–9. doi: 10.1002/art.38201
- Wen L, Krauss-Etschmann S, Petersen F, Yu X. Autoantibodies in Chronic Obstructive Pulmonary Disease. *Front Immunol* (2018) 9:66. doi: 10.3389/ fimmu.2018.00066
- Byrne R, Todd I, Tighe PJ, Fairclough LC. Autoantibodies in chronic obstructive pulmonary disease: A systematic review. *Immunol Lett* (2019) 214:8–15. doi: 10.1016/j.imlet.2019.08.007
- Sparks JA, Karlson EW. The Roles of Cigarette Smoking and the Lung in the Transitions Between Phases of Preclinical Rheumatoid Arthritis. *Curr Rheumatol Rep* (2016) 18(3):15. doi: 10.1007/s11926-016-0563-2
- Friedlander HM, Ford JA, Zaccardelli A, Terrio AV, Cho MH, Sparks JA. Obstructive lung diseases and risk of rheumatoid arthritis. *Expert Rev Clin Immunol* (2020) 16(1):37–50. doi: 10.1080/1744666X.2019.1698293
- Pentony P, Duquenne L, Dutton K, Mankia K, Gul H, Vital E, et al. The initiation of autoimmunity at epithelial surfaces: a focus on rheumatoid arthritis and systemic lupus erythematosus. *Discov Med* (2017) 24(133):191–200.
- Morissette MC, Jobse BN, Thayaparan D, Nikota JK, Shen P, Labiris NR, et al. Persistence of pulmonary tertiary lymphoid tissues and anti-nuclear antibodies following cessation of cigarette smoke exposure. *Respir Res* (2014) 15:49. doi: 10.1186/1465-9921-15-49
- Gergianaki I, Tsiligianni I. Chronic obstructive pulmonary disease and rheumatic diseases: A systematic review on a neglected comorbidity. J Comorb (2019) 9:1–01 doi: 10.1177/2235042X18820209
- Spagnolo P, Lee JS, Sverzellati N, Rossi G, Cottin V. The Lung in Rheumatoid Arthritis: Focus on Interstitial Lung Disease. Arthritis Rheumatol (2018) 70(10):1544–54. doi: 10.1002/art.40574
- 31. Ford JA, Liu X, Chu SH, Lu B, Cho MH, Silverman EK, et al. Asthma, Chronic Obstructive Pulmonary Disease, and Subsequent Risk for Incident Rheumatoid Arthritis Among Women: A Prospective Cohort Study. *Arthritis Rheumatol* (2020) 72(5):704–13. doi: 10.1002/art.41194
- 32. Zaccardelli A, Liu X, Ford JA, Cui J, Lu B, Chu SH, et al. Elevated anticitrullinated protein antibodies prior to rheumatoid arthritis diagnosis and risks for chronic obstructive pulmonary disease or asthma. *Arthritis Care Res (Hoboken)* (2020). doi: 10.1002/acr.24140
- Prisco LC, Martin LW, Sparks JA. Inhalants other than personal cigarette smoking and risk for developing rheumatoid arthritis. *Curr Opin Rheumatol* (2020) 32(3):279–88. doi: 10.1097/BOR.000000000000705
- Barnes H, Goh NSL, Leong TL, Hoy R. Silica-associated lung disease: An old-world exposure in modern industries. *Respirology* (2019) 24(12):1165– 75. doi: 10.1111/resp.13695
- Pollard KM. Silica, Silicosis, and Autoimmunity. Front Immunol (2016) 7:97. doi: 10.3389/fimmu.2016.00097
- Leung CC, Yu IT, Chen W. Silicosis. Lancet (2012) 379(9830):2008–18. doi: 10.1016/S0140-6736(12)60235-9
- Steenland K, Ward E. Silica: a lung carcinogen. CA Cancer J Clin (2014) 64 (1):63–9. doi: 10.3322/caac.21214
- Turner MT, Samuel SR, Silverstone EJ, Yates DH. Silica Exposure and Connective Tissue Disease: An Under-Recognised Association in Three

Australian Artificial Stone Workers. Am J Respir Crit Care Med (2020) 201(3):378-80 doi: 10.1164/rccm.201905-1057LE

- 39. Fan Y, Xu W, Wang Y, Wang Y, Yu S, Ye Q. Association of occupational dust exposure with combined chronic obstructive pulmonary disease and pneumoconiosis: a cross-sectional study in China. *BMJ Open* (2020) 10(9): e038874. doi: 10.1136/bmjopen-2020-038874
- 40. Yahya A, Bengtsson C, Larsson P, Too CL, Mustafa AN, Abdullah NA, et al. Silica exposure is associated with an increased risk of developing ACPApositive rheumatoid arthritis in an Asian population: evidence from the Malaysian MyEIRA case-control study. *Mod Rheumatol* (2013). doi: 10.1007/s10165-013-0890-3
- Stolt P, Yahya A, Bengtsson C, Kallberg H, Ronnelid J, Lundberg I, et al. Silica exposure among male current smokers is associated with a high risk of developing ACPA-positive rheumatoid arthritis. *Ann Rheum Dis* (2010) 69 (6):1072–6. doi: 10.1136/ard.2009.114694
- Chubb LG, Cauda EG. Characterizing Particle Size Distributions of Crystalline Silica in Gold Mine Dust. Aerosol Air Qual Res (2017) 17 (1):24–33. doi: 10.4209/aaqr.2016.05.0179
- Pavan C, Fubini B. Unveiling the Variability of "Quartz Hazard" in Light of Recent Toxicological Findings. *Chem Res Toxicol* (2017) 30(1):469–85. doi: 10.1021/acs.chemrestox.6b00409
- 44. Kawasaki H. A mechanistic review of silica-induced inhalation toxicity. Inhal Toxicol (2015) 27(8):363–77. doi: 10.3109/08958378.2015.1066905
- 45. Pavan C, Delle Piane M, Gullo M, Filippi F, Fubini B, Hoet P, et al. The puzzling issue of silica toxicity: are silanols bridging the gaps between surface states and pathogenicity? *Part Fibre Toxicol* (2019) 16(1):32. doi: 10.1186/ s12989-019-0315-3
- Lopes-Pacheco M, Bandeira E, Morales MM. Cell-Based Therapy for Silicosis. Stem Cells Int (2016) 2016:5091838. doi: 10.1155/2016/5091838
- Fubini B, Hubbard A. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. *Free Radic Biol Med* (2003) 34(12):1507–16. doi: 10.1016/S0891-5849(03)00149-7
- Franklin BS, Mangan MS, Latz E. Crystal Formation in Inflammation. Annu Rev Immunol (2016) 34:173–202. doi: 10.1146/annurev-immunol-041015-055539
- Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, et al. Chronic inflammation in the etiology of disease across the life span. *Nat Med* (2019) 25(12):1822–32. doi: 10.1038/s41591-019-0675-0
- Rabolli V, Badissi AA, Devosse R, Uwambayinema F, Yakoub Y, Palmai-Pallag M, et al. The alarmin IL-1alpha is a master cytokine in acute lung inflammation induced by silica micro- and nanoparticles. *Part Fibre Toxicol* (2014) 11:69. doi: 10.1186/s12989-014-0069-x
- Rider P, Voronov E, Dinarello CA, Apte RN, Cohen I. Alarmins: Feel the Stress. J Immunol (2017) 198(4):1395–402. doi: 10.4049/jimmunol.1601342
- Borthwick LA. The IL-1 cytokine family and its role in inflammation and fibrosis in the lung. *Semin Immunopathol* (2016) 38(4):517–34. doi: 10.1007/ s00281-016-0559-z
- Sayan M, Mossman BT. The NLRP3 inflammasome in pathogenic particle and fibre-associated lung inflammation and diseases. *Part Fibre Toxicol* (2016) 13(1):51. doi: 10.1186/s12989-016-0162-4
- Hornung V, Bauernfeind F, Halle A, Samstad EO, Kono H, Rock KL, et al. Silica crystals and aluminum salts activate the NALP3 inflammasome through phagosomal destabilization. *Nat Immunol* (2008) 9(8):847–56. doi: 10.1038/ni.1631
- Hoy RF, Chambers DC. Silica-related diseases in the modern world. Allergy (2020) 75(11):2805–17 doi: 10.1111/all.14202
- Caplan A. Certain unusual radiological appearances in the chest of coalminers suffering from rheumatoid arthritis. *Thorax* (1953) 8(1):29–37. doi: 10.1136/thx.8.1.29
- Shtraichman O, Blanc PD, Ollech JE, Fridel L, Fuks L, Fireman E, et al. Outbreak of autoimmune disease in silicosis linked to artificial stone. Occup Med (Lond) (2015) 65(6):444–50. doi: 10.1093/occmed/kqv073
- Huaux F. Emerging Role of Immunosuppression in Diseases Induced by Micro- and Nano-Particles: Time to Revisit the Exclusive Inflammatory Scenario. Front Immunol (2018) 9:2364. doi: 10.3389/fimmu.2018.02364
- 59. Bates MA, Brandenberger C, Langohr I, Kumagai K, Harkema JR, Holian A, et al. Silica Triggers Inflammation and Ectopic Lymphoid Neogenesis in the Lungs in Parallel with Accelerated Onset of Systemic Autoimmunity and

Glomerulonephritis in the Lupus-Prone NZBWF1 Mouse. *PloS One* (2015) 10(5):e0125481. doi: 10.1371/journal.pone.0125481

- 60. Foster MH, Ord JR, Zhao EJ, Birukova A, Fee L, Korte FM, et al. Silica Exposure Differentially Modulates Autoimmunity in Lupus Strains and Autoantibody Transgenic Mice. *Front Immunol* (2019) 10:2336. doi: 10.3389/fimmu.2019.02336
- 61. Bates MA, Akbari P, Gilley KN, Wagner JG, Li N, Kopec AK, et al. Dietary Docosahexaenoic Acid Prevents Silica-Induced Development of Pulmonary Ectopic Germinal Centers and Glomerulonephritis in the Lupus-Prone NZBWF1 Mouse. Front Immunol (2018) 9:2002. doi: 10.3389/fimmu.2018.02002
- Conrad K, Mehlhorn J. Diagnostic and prognostic relevance of autoantibodies in uranium miners. *Int Arch Allergy Immunol* (2000) 123 (1):77-91. doi: 10.1159/000024426
- Mayeux JM, Escalante GM, Christy JM, Pawar RD, Kono DH, Pollard KM. Silicosis and Silica-Induced Autoimmunity in the Diversity Outbred Mouse. *Front Immunol* (2018) 9:874. doi: 10.3389/fimmu.2018.00874
- 64. Mohamed BM, Verma NK, Davies AM, McGowan A, Crosbie-Staunton K, Prina-Mello A, et al. Citrullination of proteins: a common post-translational modification pathway induced by different nanoparticles in vitro and in vivo. *Nanomedicine (Lond)* (2012) 7(8):1181–95. doi: 10.2217/nnm.11.177
- Li Y, Cao X, Liu Y, Zhao Y, Herrmann M. Neutrophil Extracellular Traps Formation and Aggregation Orchestrate Induction and Resolution of Sterile Crystal-Mediated Inflammation. *Front Immunol* (2018) 9:1559. doi: 10.3389/fimmu.2018.01559
- 66. Wong SL, Wagner DD. Peptidylarginine deiminase 4: a nuclear button triggering neutrophil extracellular traps in inflammatory diseases and aging. *FASEB J* (2018) 32:6358–70 doi: 10.1096/fj.201800691R
- Sporn TA. Mineralogy of asbestos. Recent Results Cancer Res (2011) 189:1– 11. doi: 10.1007/978-3-642-10862-4_1
- Solbes E, Harper RW. Biological responses to asbestos inhalation and pathogenesis of asbestos-related benign and malignant disease. J Invest Med (2018) 66(4):721–7. doi: 10.1136/jim-2017-000628
- Aust AE, Cook PM, Dodson RF. Morphological and chemical mechanisms of elongated mineral particle toxicities. J Toxicol Environ Health B Crit Rev (2011) 14(1-4):40–75. doi: 10.1080/10937404.2011.556046
- Dodson RF, Atkinson MA, Levin JL. Asbestos fiber length as related to potential pathogenicity: a critical review. *Am J Ind Med* (2003) 44(3):291–7. doi: 10.1002/ajim.10263
- Barlow CA, Grespin M, Best EA. Asbestos fiber length and its relation to disease risk. *Inhal Toxicol* (2017) 29(12-14):541-54. doi: 10.1080/ 08958378.2018.1435756
- Ferro A, Zebedeo CN, Davis C, Ng KW, Pfau JC. Amphibole, but not chrysotile, asbestos induces anti-nuclear autoantibodies and IL-17 in C57BL/6 mice. *J Immunotoxicol* (2014) 11(3):283–90. doi: 10.3109/1547691X.2013.847510
- Marchand LS, St-Hilaire S, Putnam EA, Serve KM, Pfau JC. Mesothelial cell and anti-nuclear autoantibodies associated with pleural abnormalities in an asbestos exposed population of Libby MT. *Toxicol Lett* (2012) 208(2):168– 73. doi: 10.1016/j.toxlet.2011.10.024
- Reid A, Franklin P, de Klerk N, Creaney J, Brims F, Musk B, et al. Autoimmune antibodies and asbestos exposure: Evidence from Wittenoom, Western Australia. Am J Ind Med (2018) 61(7):615–20. doi: 10.1002/ajim.22863
- Ledda C, Caltabiano R, Loreto C, Cina D, Senia P, Musumeci A, et al. Prevalence of anti-nuclear autoantibodies in subjects exposed to natural asbestiform fibers: a cross-sectional study. *J Immunotoxicol* (2018) 15(1):24– 8. doi: 10.1080/1547691X.2017.1415398
- Diegel R, Black B, Pfau JC, McNew T, Noonan C, Flores R. Case series: rheumatological manifestations attributed to exposure to Libby Asbestiform Amphiboles. J Toxicol Environ Health A (2018) 81(15):734–47. doi: 10.1080/ 15287394.2018.1485124
- Noonan CW, Pfau JC, Larson TC, Spence MR. Nested case-control study of autoimmune disease in an asbestos-exposed population. *Environ Health Perspect* (2006) 114(8):1243–7. doi: 10.1289/ehp.9203
- Larson TC, Williamson L, Antao VC. Follow-Up of the Libby, Montana Screening Cohort: A 17-Year Mortality Study. J Occup Environ Med (2020) 62(1):e1–6. doi: 10.1097/JOM.000000000001760
- Wong J, Magun BE, Wood LJ. Lung inflammation caused by inhaled toxicants: a review. *Int J Chron Obstruct Pulmon Dis* (2016) 11:1391–401. doi: 10.2147/COPD.S106009

- Dostert C, Petrilli V, Van Bruggen R, Steele C, Mossman BT, Tschopp J. Innate immune activation through Nalp3 inflammasome sensing of asbestos and silica. *Science* (2008) 320(5876):674–7. doi: 10.1126/science.1156995
- Cassel SL, Eisenbarth SC, Iyer SS, Sadler JJ, Colegio OR, Tephly LA, et al. The Nalp3 inflammasome is essential for the development of silicosis. *Proc Natl Acad Sci U S A* (2008) 105(26):9035–40. doi: 10.1073/pnas.0803933105
- Liu G, Cheresh P, Kamp DW. Molecular basis of asbestos-induced lung disease. Annu Rev Pathol (2013) 8:161–87. doi: 10.1146/annurev-pathol-020712-163942
- Pailes WH, Judy DJ, Resnick H, Castranova V. Relative effects of asbestos and wollastonite on alveolar macrophages. J Toxicol Environ Health (1984) 14(4):497–510. doi: 10.1080/15287398409530601
- Rom WN, Travis WD. Lymphocyte-macrophage alveolitis in nonsmoking individuals occupationally exposed to asbestos. *Chest* (1992) 101(3):779–86. doi: 10.1378/chest.101.3.779
- Pfau JC, McNew T, Hanley K, Swan L, Black B. Autoimmune markers for progression of Libby amphibole lamellar pleural thickening. *Inhal Toxicol* (2019) 31(11-12):409–19. doi: 10.1080/08958378.2019.1699616
- Pfau JC, Barbour C, Black B, Serve KM, Fritzler MJ. Analysis of autoantibody profiles in two asbestiform fiber exposure cohorts. *J Toxicol Environ Health* A (2018) 81(19):1015–27. doi: 10.1080/15287394.2018.1512432
- Mohamed BM, Boyle NT, Schinwald A, Murer B, Ward R, Mahfoud OK, et al. Induction of protein citrullination and auto-antibodies production in murine exposed to nickel nanomaterials. *Sci Rep* (2018) 8(1):679. doi: 10.1038/s41598-017-19068-1
- Aberle T, Bourn RL, Munroe ME, Chen H, Roberts VC, Guthridge JM, et al. Clinical and serological features distinguish patients with incomplete lupus classification from systemic lupus erythematosus patients and controls. *Arthritis Care Res (Hoboken)* (2017) 69(12):1780–8 doi: 10.1002/acr.23201
- Wither J, Johnson SR, Liu T, Noamani B, Bonilla D, Lisnevskaia L, et al. Presence of an interferon signature in individuals who are anti-nuclear antibody positive lacking a systemic autoimmune rheumatic disease diagnosis. *Arthritis Res Ther* (2017) 19(1):41. doi: 10.1186/s13075-017-1243-y
- Xia T, Li N, Nel AE. Potential health impact of nanoparticles. Annu Rev Public Health (2009) 30:137–50. doi: 10.1146/annurev.publhealth.031308.100155
- Prosperi D, Colombo M, Zanoni I, Granucci F. Drug nanocarriers to treat autoimmunity and chronic inflammatory diseases. *Semin Immunol* (2017) 34:61–7. doi: 10.1016/j.smim.2017.08.010
- Singha S, Shao K, Ellestad KK, Yang Y, Santamaria P. Nanoparticles for Immune Stimulation Against Infection, Cancer, and Autoimmunity. ACS Nano (2018) 12(11):10621–35. doi: 10.1021/acsnano.8b05950
- Kelly HG, Kent SJ, Wheatley AK. Immunological basis for enhanced immunity of nanoparticle vaccines. *Expert Rev Vaccines* (2019) 18(3):269– 80. doi: 10.1080/14760584.2019.1578216
- Serra P, Santamaria P. Nanoparticle-based approaches to immune tolerance for the treatment of autoimmune diseases. *Eur J Immunol* (2018) 48(5):751– 6. doi: 10.1002/eji.201747059
- Pearson RM, Podojil JR, Shea LD, King NJC, Miller SD, Getts DR. Overcoming challenges in treating autoimmuntity: Development of tolerogenic immune-modifying nanoparticles. *Nanomedicine* (2019) 18:282–91. doi: 10.1016/j.nano.2018.10.001
- Boraschi D, Castellano LRC, Italiani P. Editorial: Interaction of Nanomaterials with the Immune System: Role in Nanosafety and Nanomedicine. Front Immunol (2017) 8:1688. doi: 10.3389/fimmu.2017.01688
- Leso V, Fontana L, Iavicoli I. Nanomaterial exposure and sterile inflammatory reactions. *Toxicol Appl Pharmacol* (2018) 355:80–92. doi: 10.1016/j.taap.2018.06.021
- Wu T, Tang M. Review of the effects of manufactured nanoparticles on mammalian target organs. J Appl Toxicol (2018) 38(1):25–40. doi: 10.1002/ jat.3499
- Pallardy MJ, Turbica I, Biola-Vidamment A. Why the Immune System Should Be Concerned by Nanomaterials? *Front Immunol* (2017) 8:544. doi: 10.3389/fimmu.2017.00544
- 100. Leikauf GD, Kim SH, Jang AS. Mechanisms of ultrafine particle-induced respiratory health effects. *Exp Mol Med* (2020) 52(3):329–37. doi: 10.1038/ s12276-020-0394-0
- 101. You R, Lu W, Shan M, Berlin JM, Samuel EL, Marcano DC, et al. Nanoparticulate carbon black in cigarette smoke induces DNA cleavage

and Th17-mediated emphysema. *Elife* (2015) 4:e09623. doi: 10.7554/ eLife.09623

- Ma Q. Polarization of Immune Cells in the Pathologic Response to Inhaled Particulates. Front Immunol (2020) 11:1060. doi: 10.3389/fimmu.2020.01060
- 103. Munoz LE, Bilyy R, Biermann MH, Kienhofer D, Maueroder C, Hahn J, et al. Nanoparticles size-dependently initiate self-limiting NETosis-driven inflammation. *Proc Natl Acad Sci USA* (2016) 113(40):E5856–E65. doi: 10.1073/pnas.1602230113
- Biermann MH, Podolska MJ, Knopf J, Reinwald C, Weidner D, Maueroder C, et al. Oxidative Burst-Dependent NETosis Is Implicated in the Resolution of Necrosis-Associated Sterile Inflammation. *Front Immunol* (2016) 7:557. doi: 10.3389/fimmu.2016.00557
- 105. Yang H, Marion TN, Liu Y, Zhang L, Cao X, Hu H, et al. Nanomaterial Exposure Induced Neutrophil Extracellular Traps: A New Target in Inflammation and Innate Immunity. *J Immunol Res* (2019) 2019:3560180. doi: 10.1155/2019/3560180
- 106. Chezeau L, Kohlstaedt LA, Le Faou A, Cosnier F, Rihn B, Gate L. Proteomic analysis of bronchoalveolar lavage fluid in rat exposed to TiO2 nanostructured aerosol by inhalation. J Proteomics (2019) 207:103451. doi: 10.1016/j.jprot.2019.103451
- Napierska D, Thomassen LC, Lison D, Martens JA, Hoet PH. The nanosilica hazard: another variable entity. *Part Fibre Toxicol* (2010) 7(1):39. doi: 10.1186/1743-8977-7-39
- 108. Hutchinson D, Muller J, McCarthy JE, Gun'ko YK, Verma NK, Bi X, et al. Cadmium nanoparticles citrullinate cytokeratins within lung epithelial cells: cadmium as a potential cause of citrullination in chronic obstructive

pulmonary disease. Int J Chron Obstruct Pulmon Dis (2018) 13:441-9. doi: 10.2147/COPD.S152028

- 109. Lam CW, James JT, McCluskey R, Hunter RL. Pulmonary toxicity of singlewall carbon nanotubes in mice 7 and 90 days after intratracheal instillation. *Toxicol Sci* (2004) 77(1):126–34. doi: 10.1093/toxsci/kfg243
- 110. Holers VM, Demoruelle MK, Kuhn KA, Buckner JH, Robinson WH, Okamoto Y, et al. Rheumatoid arthritis and the mucosal origins hypothesis: protection turns to destruction. *Nat Rev Rheumatol* (2018) 14 (9):542–57. doi: 10.1038/s41584-018-0070-0
- 111. Orange DE, Yao V, Sawicka K, Fak J, Frank MO, Parveen S, et al. RNA Identification of PRIME Cells Predicting Rheumatoid Arthritis Flares. N Engl J Med (2020) 383(3):218–28. doi: 10.1056/NEJMoa2004114
- 112. Alkhouri H, Poppinga WJ, Tania NP, Ammit A, Schuliga M. Regulation of pulmonary inflammation by mesenchymal cells. *Pulm Pharmacol Ther* (2014) 29(2):156–65. doi: 10.1016/j.pupt.2014.03.001

Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

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