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Synthesis and Biological Evaluation of New 3-Phenyl-1-[(4-arylpiperazin-1-yl)alkyl]piperidine-2,6-diones

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Abstract

A set of 13 alkyl derivatives of 3-phenylpiperidine-2,6-dione were synthesized. Newly obtained compounds were investigated *in vitro* against HIV-1 and other selected viruses. The benzyl **3f** and fluorophenyl **3g** derivatives showed moderate protection against CVB-2 and the compound **3g** also against HSV-1. Derivatives were tested also for their antibacterial and antifungal activity. The molecular structures of **3a** and **3d** were determined by an X-ray analysis.

Keywords

Antiviral activity • Cytotoxicity • 3-Phenylpiperidine-2,6-dione • X-ray crystallography

Introduction

Literature survey shows that phenylpiperazinyl group is a nuclei of antiviral, antibacterial and antifungal [1–3] agents. It could be integrated with imidazole [4, 5], 1,8-naphthyridone [6], furan [7], 1,4-dithine [8] or quinolone [9] rings. N-alkyl and N-aryl piperazines are also present as phosphonate [10] and cyano [11] derivatives. Short-chain *p*-substituted aryl piperazines are found as active against *Staphylococcus aureus* and *Pseudomonas aeruginosa* [12, 13]. Recently, derivatives of diarylpiperidin-4-one were found as the new class of antimicrobial agents with activity against pathogenic bacterial species and fungal

strains [14]. There is also an increasing concern on substituted 4-arylpiperazines, 4-azatricyclodec-8-ene-3,5-dione derivatives, as some of them have been reported as antiviral agents, e.g. against Yellow Fever Virus (YFV) and Border Disease Virus (CVB-2), as well as antibacterial and antifungal compounds [1, 15]. Some anti-HIV-1 agents, as NNRTIs (non-nucleoside reverse transcriptase inhibitors) possess an arylpiperazine part (Atevirdine, Indinavir, Delavirdine and Vicriviroc). Their activity is also connected with the presence of pyridinylpiperazine or 2-hydroxypropyl fragments. Literature survey revealed that modifications of their structures lead to new potential antiviral drugs [3, 16–19].

This work describes synthesis and a wide spectrum of antiviral activity screening of some novel 3-phenylpiperidine-2,6-diones. Almost all piperazine derivatives presented in this study were tested on anti-HIV-1 activity. Antibacterial and antifungal activity of most of newly synthesized compounds was also evaluated. Due to its high 5-HT_{1A} receptor affinity and selectivity [20], the derivative **3a** was tested for its pharmacological properties in three behavioral tests in mice (data not shown). The molecular structures of **3a** and **3d** were determined by an X-ray analysis.

Results and discussion

The preparation of new thirteen 3-phenylpiperidine-2,6-dione derivatives is described. The general synthetic pathway and structure of the investigated compounds is given in Fig. 2. The starting imide **2** was obtained from 2-phenylglutaric anhydride in reaction with ammonium carbonate. Next the compound was subjected to the reaction with 1,4-dibromobutane and 1,3-dibromopropane in order to be transformed into the corresponding alkyl derivatives **3** and **4**. Next these compounds were condensed with appropriate amines to yield compounds **3a–3g** and **4a–4c**. Obtained N-substituted derivatives were purified by column chromatography. Elemental analysis and ¹H NMR spectra confirmed the identity of the product. The molecular structure of **3a** and **3d** was established by the crystal structure analysis (Fig. 1). Moreover, the two crystals are hemihydrates with a high degree of isostructurality, *viz*. they have the same symmetry, similar mode of molecular packing and noncovalent interactions. Very characteristic feature is: (a) the O–H…N hydrogen bond formed between the water molecule and the piperazine N1-atom, and (b) the lack of hydrophobic interactions of both the methoxy group (**3a**) and pyridine N-atom (**3d**).



Fig. 1. Molecular conformation of **3d** and the O-H...N hydrogen bond (dashed line) fromed between base and water molecule.



Fig. 2. Synthesis of 3-phenylpiperidine-2,6-dione derivatives.

	^a MT-4	^b HIV-1	^c MDBK	dBVDV	^e BHK-21	^f YFV	^f Reo-1
Ħ	CC ₅₀	EC ₅₀	CC ₅₀	EC ₅₀	CC ₅₀	EC_{50}	EC_{50}
2	>100	>100	>100	>100	>100	>100	>100
3a	>100	>100	>100	>100	>100	>100	>100
3c	>100	>100	>100	>100	>100	>100	>100
3d	>100	>100	>100	>100	>100	>100	>100
3e	74	>74	>100	>100	>100	>100	>100
3f	>100	>100	>100	>100	>100	>100	>100
3g	54	>54	>100	>100	>100	>100	>100
4a	>100	>100	>100	>100	>100	>100	>100
4b	>100	>100	>100	>100	>100	>100	>100
4c	>100	>100	>100	>100	>100	>100	>100
4d	>100	>100	>100	>100	>100	>100	>100
AZT [§]	50	0.01					
NM 108 [#]				1.8		2.5	
#	^g Vero-76	^ʰ HSV-1	^h VV	^h VSV	^h CVB-2	^h Sb-1	
#	CC_{50}	EC_{50}	EC_{50}	EC ₅₀	EC_{50}	EC_{50}	_
-	00	>98	>98	>98	>98	>98	
2	98						
2 3a	98 >100	>100	>100	>100	>100	>100	
2 3a 3c	98 >100 >100	>100 >100	>100 >100	>100 >100	>100 >100	>100 >100	
2 3a 3c 3d	98 >100 >100 98	>100 >100 >98	>100 >100 >98	>100 >100 >98	>100 >100 >98	>100 >100 >98	
2 3a 3c 3d 3e	98 >100 >100 98 >100	>100 >100 >98 >100	>100 >100 >98 >100	>100 >100 >98 >100	>100 >100 >98 >100	>100 >100 >98 >100	
2 3a 3c 3d 3e 3f	98 >100 >100 98 >100 96	>100 >100 >98 >100 >96	>100 >100 >98 >100 >96	>100 >100 >98 >100 >96	>100 >100 >98 >100 44	>100 >100 >98 >100 >96	
2 3a 3c 3d 3e 3f 3g	98 >100 >100 98 >100 96 92	>100 >100 >98 >100 >96 31	>100 >100 >98 >100 >96 >92	>100 >100 >98 >100 >96 >92	>100 >100 >98 >100 44 44	>100 >100 >98 >100 >96 >92	
2 3a 3c 3d 3e 3f 3g 4a	98 >100 >100 98 >100 96 92 >100	>100 >100 >98 >100 >96 31 >100	>100 >100 >98 >100 >96 >92 >100	>100 >100 >98 >100 >96 >92 >100	>100 >100 >98 >100 44 44 >100	>100 >100 >98 >100 >96 >92 >100	
2 3a 3c 3d 3e 3f 3g 4a 4b	98 >100 >100 98 >100 96 92 >100 98	>100 >100 >98 >100 >96 31 >100 >98	>100 >100 >98 >100 >96 >92 >100 >98	>100 >100 >98 >100 >96 >92 >100 >98	>100 >100 >98 >100 44 44 >100 >98	>100 >100 >98 >100 >96 >92 >100 >98	
2 3a 3c 3d 3e 3f 3g 4a 4b 4c	98 >100 98 >100 96 92 >100 98 >100	>100 >100 >98 >100 >96 31 >100 >98 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100	>100 >100 >98 >100 44 44 >100 >98 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100	
2 3a 3c 3d 3e 3f 3g 4a 4b 4c 4d	98 >100 98 >100 96 92 >100 98 >100 >100	>100 >100 >98 >100 >96 31 >100 >98 >100 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100	>100 >100 >98 >100 44 44 >100 >98 >100 >100	>100 >98 >100 >96 >92 >100 >98 >100 >100	
2 3a 3c 3d 3e 3f 3g 4a 4b 4c 4d NM 176*	98 >100 98 >100 96 92 >100 98 >100 >100	>100 >100 >98 >100 >96 31 >100 >98 >100 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100	>100 >100 >98 >100 44 44 >100 >98 >100 >100 23	>100 >98 >100 >96 >92 >100 >98 >100 >100 18	
2 3a 3c 3d 3e 3f 3g 4a 4b 4c 4d NM 176* M 5255**	98 >100 >100 98 >100 96 92 >100 98 >100 >100	>100 >100 >98 >100 >96 31 >100 >98 >100 >100	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100 1.8	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100	>100 >100 >98 >100 44 44 >100 >98 >100 >100 23	>100 >100 >98 >100 >96 >92 >100 >98 >100 >100 18	

Tab. 1. Cytotoxicity and antiviral activity of compounds 2, 3a–3g and 4a–4d.

Antiviral activity is given as EC_{50} (Median Effective Concentration – the concentration of a drug (μ M) required to induce a 50% effect), and cytotoxicity is given as CC_{50} (Cytotoxic Concentration – the amount of a drug (μ M) at which 50% of cells become dead). ^{a,c,e,g} Compd. concn. (μ M) required to reduce the viability of mock-infected MT-4 (CD4⁺

Human T-cells containing an integrated HTLV-1 genome, ^a) cells or MDBK (Bovine normal kidney, ^c) cells or BHK (Hamster normal kidney fibroblast, ^e) monolayers or VERO-76 (Monkey normal kidney) monolayers by 50%, as determined by the colorimetric MTT method.

^{b,d,f,h} Compd. concentration (μM) required to achieve 50% protection of MT-4 cells from the HIV-1-induced cytopathogenicity (^b) or MDBK cells from the BVDV (Bovine Viral Diarrhea Virus)-induced cytopathogenicity (^d) or BHK (Kidney fibroblast) cells from the YFV (Yellow Fever Virus) and Reo (Reovirus 1)-induced cytopathogenicity (^f) or to reduce the plaque number of HSV-1 (Herpesvirus 1), VV (Vaccinia Virus), VSV (Vesicular Stomatitis Virus), CVB-2 (Coxsackievirus B2), Sb-1 (Poliovirus 1) and RSV (Respiratory Syncytial Virus) by 50% in VERO-76 monolayers (^h), as determined by the MTT method.

[§] 3'-azidothymidine; [#] 2'-ß-methylguanosine; * 2'-ethynyl-D-cytidine; ** mycophenolic acid; *** acycloGuanosine.

For biochemical studies N-substituted alkyl derivatives were converted into their hydrochlorides. Compounds 3a and 3d were presented as monohydrochlorides and **3b**, **3c**, **3e**, **3f**, **3g**, **4a**, **4b**, **4d** as dihydrochlorides.

Ten compounds, N-substituted derivatives of 3-phenylpiperidine-2,6-dione, were evaluated *in vitro* against viruses, bacteria and fungi.

Title compounds were evaluated for antiviral activity against viruses representative of two of the three genera of Flaviviridae family, that is, Flaviviruses (Yellow Fever Virus, YFV) and Pestiviruses (Bovine Viral Diarrhoea Virus, BVDV), as Hepaciviruses can hardly be used in routine cell-based assays. Compounds were also tested against representatives of other virus families. Among ssRNA+ were a retrovirus (human immunodeficiency virus type 1, HIV-1) and two Picornaviruses (Coxsackie Virus type B2, CVB-2 and Poliovirus type-1, Sabin strain, Sb-1); among ssRNA- were a Rhabdoviridae (Vesicular Stomatitis Virus, VSV) representative. Among double-stranded RNA (dsRNA) viruses was a Reoviridae representative (Reo-1). Two representatives of DNA virus families were also included: Herpes Simplex type-1, HSV-1 (Herpesviridae) and Vaccinia Virus, VV (Poxviridae).

In addition to the antiviral activity, compounds were evaluated *in vitro* against representative strains of Gram-positive and Gram-negative bacteria (*Staphylococcus aureus, Pseudomonas aeruginosa*), yeasts and moulds (*Candida albicans and Aspergillus niger*).

AZT (3'-azidothymidine), NM 108 (2'-ß-methylguanosine), NM 176 (2'-ethynylcitidine), M 5255 (Mycophenolic Acid) and ACG (acycloGuanosine) were used as reference inhibitors of ssRNA+, ssRNA- and DNA viruses, respectively.

Two of tested derivatives presented moderate antiviral activity (Table 1). The CVB-2 cells were susceptible to benzyl (**3f**) and 1-(4-fluorophenyl)-substituted (**3g**) long-chain arylpiperazine derivatives. The fluorophenyl derivative was also active against HSV-1 virus. The 50% cytotoxic concentration (CC_{50}) of this compound in Vero-76 cells was 92 μ M. The range of cytotoxicity of compounds in MT-4 cells was from 100 to 54 μ M.

MIC ^a [µM]				
	S <i>. aureus</i> DSM 2569	<i>P. aeruginosa</i> DSM 1117	<i>C. albicans</i> DSM 1386	<i>A. niger</i> DSM 1988
2, 3a, 3c–3g, 4a–4d	>100	>100	>100	>100
Ciprofloxacin ^b	4	0.8	_	_
Miconazole ^b	_	_	0.8	20

 Tab. 2.
 Antibacterial and antifungal activities of 3-phenylpiperidine-2,6-dione and its derivatives.

^a The antimicrobial activity is given as Minimum inhibitory concentration (MIC) corresponding to the lowest concentration of an antimicrobial compound that showed complete growth inhibition.
 ^b Ciprofloxacin was solubilized in water (0.1 M solution) and Miconazole in DMSO (0.1 M solution), according to the British Society for Antimicrobial Chemotherapy (BSAC) protocol and stored at 4°C overnight. Reference Compounds were diluted from 100 to 0.0013 μM.

Gram-negative rods as well as gram-positive strains and fungal organisms were resistant to all tested agents. Their minimal inhibitory concentration (MIC) values for all compounds were above 100 μ M (Table 2). However, none of title compounds turned out to be active against HIV-1, BVDV or representatives of ssRNA- and dsRNA viruses.

Theoretical calculated lipophilicity (log*P*) of synthesized compounds ranged from 1.89 to 3.92. According to Clark and Lobell rules [21], all derivatives could cross the blood-brain barrier to act as ligands of receptors of central nervous system.

Experimental

Chemistry

All chemicals and solvents were purchased from Aldrich (Vienna, Austria). Melting points were determined on an Electrothermal Digital Melting Point Apparatus (Essex, UK) and are uncorrected. The ¹H-NMR spectra were recorded on a Bruker (Rheinstetten, Germany) spectrometer, operating at 400 or 300 MHz. The chemical shift values are expressed in ppm relative to TMS as an internal standard. Elemental analyses were recorded on a CHN model 2400 Perkin-Elmer (Hitachi, Tokyo, Japan). TLC was carried out using silica gel 60 F_{254} , layer thickness 0.25 mm (E. Merck, Darmstadt, Germany) and the results were visualized using UV lamp at 254 nm. Column chromatography was carried out using silica gel 60 (200–400 mesh, Merck).

The elemental analyses and ¹H-NMR spectra, as well as melting points are given for dihydrochlorides (except for compounds **3**, **4**, **3a** and **3d**). Yields are presented for crude products.

Log*P* values for arylpiperazine derivatives were calculated using ChemBioDrawUltra 12.0. (http://www.cambridgesoft.com).

Molecular structure of **3a** and **3d** was confirmed by an X-ray crystallography (Table 3). The intensity data were collected at room temperature with a KM4 diffractometer using graphite monochromated CuK α radiation ($\lambda = 1.54178$ Å) and $\omega - 2\theta$ scan mode. Both compounds crystallized as hemihydrates and their crystals were of poor quality. Structure was solved by the SHELXS-97 program and refined by full-matrix least-squares on F^2 using the SHELXL-97 program [18]. Non-hydrogen atoms were refined with anisotropic displacement parameters, except those of phenyl rings of **3a**. The H-atoms of molecules **3a** and **3d** were positioned geometrically and 'riding' model was used in the refinement, while H-atoms of water molecules were located on difference maps. Crystallographic data had been deposited with the Cambridge Crystallographic Data Center.

3-Phenylpiperidine-2,6-dione (2)

A mixture of 2-phenylglutaric anhydride (0.0165 mol) and $(NH_4)_2CO_3$ (0.125 mol) was reduced to powder and heated up to 185 °C using a metal bath of Wood's alloy. When CO_2 and NH_3 were completely liberated, 10 cm³ of paraffin oil was added and whole mixture was heated until the evolution of gas ceased. The residue was crystallized from hexane and ethyl acetate; mp 143 °C (mp 142–143 from methanol [23]).

Tab. 3.	Crystal data and parameters of the data collection and refinement for crystals of
	3a . $\frac{1}{2}$ H ₂ O and 3d . $\frac{1}{2}$ H ₂ O.

Identification code	3a. ¹ ⁄₂ H₂O	3d. ¹ / ₂ H ₂ O			
No CCDC*	770 127	770 128			
Formula weight	444.56	415.53			
Crystal system	monoclinic	monoclinic			
Space group	C2	C2			
Unit cell dimensions					
a (Å)	26.891(5)	18.908(4)			
b (Å)	6.344(1)	6.244(1)			
<i>c</i> (Å)	18.631(4)	19.087(4)			
β (°)	129.05(3)	93.56(3)			
Volume (Å ³); Z	2468.3(8); 4	2249.1(8); 4			
Density (calc) (g cm ⁻³)	1.196	1.227			
Absorption coeff. (mm ⁻¹)	0.639	0.646			
F(000)	956	892			
Crystal size (mm)	0.28 x 0.12 x 0.05	0.22 x 0.20 x 0.04			
Theta range for data collection (°)	3.05 to 70.16	4.69 to 72.13			
Index ranges	$-32 \le h \le 32, -7 \le k \le 7,$	$-23 \le h \le 23, -7 \le k \le 7,$			
	-16 ≤ / ≤ 22	-23 ≤ <i>I</i> ≤ 0			
Reflections collected	4852	4462			
Independent reflections	4436 [<i>R</i> (int) = 0.1722]	4328 [<i>R</i> (int) = 0.0489]			
Data / restraints / parameters	4436 / 1 / 223	4328 / 1 / 276			
Goodness-of-fit on <i>F</i> ²	1.000	0.942			
Final <i>R</i> indices [<i>I</i> >2s (<i>I</i>)]	<i>R</i> 1 = 0.0787,	<i>R</i> 1 = 0.0380,			
	<i>wR</i> 2 = 0.2201	<i>wR</i> 2 = 0.0961			
Δρ max.; min. (e Å ⁻³)	0.25; -0.29	0.18; -0.18			
These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via					

www.ccdc.cam.ac.uk.

General method for preparation of 1-(alkyl)-3-phenylpiperidine-2,6-diones 3 and 4

A mixture of imide **2** (1.5 g, 0.008 mol), 1,4-dibromobutane (0.02 mol) and 1,3-dibromopropane (0.02 mol), respectively, anhydrous K_2CO_3 (1.5 g) and catalytic amount of 98% 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) were refluxed in acetone for 60–70 h. Then, the solvent was removed on a rotary evaporator and the oily residue was purified by column chromatography (chloroform : methanol 9.5:0.5 vol).

1-(4-Bromobutyl)-3-phenylpiperidine-2,6-dione (3)

Yield: 78%, mp 138.5–140 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.32 (m, 5H, ArH), 3.85 (m, 3H), 3.43 (t, *J* = 6.4 Hz, 2H), 2.72 (m, 2H), 2.22 (m, 2H), 1.88 (m, 2H), 1.73 (m, 2H). Anal. Calcd. for C₁₅H₁₈BrNO₂: C, 55.57; H, 5.60; N, 4.32. Found: C, 55.47; H, 5.61; N, 4.33.

1-(3-Bromopropyl)-3-phenylpiperidine-2,6-dione (4)

Yield: 75%, oil. ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.34 (m, 4H), 7.17 (d, *J* = 7.6 Hz, 1H), 3.98 (t, *J* = 6.8 Hz, 2H), 3.84 (dd, *J*₁ = 5.6 Hz, *J*₂ = 8.8 Hz), 3.40 (t, *J* = 6.8 Hz, 2H), 2.73 (m, 2H), 2.26 (m, 2H), 2.17 (m, 2H). Anal. Calcd. for C₁₄H₁₆BrNO₂: C, 54.21; H, 5.20; N, 4.52. Found: C, 54.47; H, 5.24; N, 4.55.

General method for the preparation of 1-[4-aryl/heteroarylpiperazin-1-ylbutyl) (3a–3g) and 1-[3-aryl/heteroarylpiperazin-1-ylpropyl) (4a–4c) derivatives of 3-phenylpiperidine-2,6-dione

A mixture of derivative **3** (0.3 g, 0.001 mol) and **4** (0.3 g, 0.001 mol), respectively, the corresponding amine (0.002 mol), anhydrous K_2CO_3 (0.3 g) and catalytic amount of KI was refluxed in acetone for 30 h. Then the mixture was filtered off and the solvent was evaporated. The residue was purified by column chromatography (chloroform : methanol 9.5:0.5 vol) and/or crystallized from hexane.

Obtained compounds were converted into their hydrochlorides. The solid product was dissolved in methanol saturated with gaseous HCI. The hydrochloride was precipitated by addition of diethyl ether. The crude product was crystallized from methanol/ethyl ether.

1-{4-[4-(2-Methoxyphenyl)piperazin-1-yl]butyl}-3-phenylpiperidine-2,6-dione (3a)

Mp 210 °C from acetone (mp (salt) from acetone 209–210 °C) [20]. LogP = 3.64.

3-Phenyl-1-[4-(4-pyrimidin-2-ylpiperazin-1-yl)butyl]piperidine-2,6-dione (3b)

Yield: 70%, mp 210–212 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.30 (d, *J* = 4.8 Hz, 2H), 7.33 (m, 3H), 7.17 (d, *J* = 7.2 Hz, 2H), 6.48 (t, *J* = 4.8 Hz, 1H), 3.85 (m, 7H), 2.72 (m, 2H), 2.52 (m, 4H), 2.43 (m, 2H), 2.22 (m, 2H), 1.60 (m, 4H). Anal. Calcd. for C₂₃H₂₉N₅O₂ · 2HCl · $\frac{1}{2}$ H₂O: C, 55.34; H, 6.76; N, 14.67. Found: C, 55.07; H, 6.72; N, 14.87. Log*P* = 2.34.

1-{4-[4-(2-Hydroxyphenyl)piperazin-1-yl]butyl}-3-phenylpiperidine-2,6-dione (3c)

Yield: 67%, mp 234–236 °C. ¹H NMR (300 MHz, DMSO, TMS): δ 7.30 (m, 5H), 6.91 (m, 3H), 6.77 (t, *J* = 7.2 Hz, 1H), 5.20 (m, 6H), 4.00 (dd, *J*₁ = 4.8 Hz, *J*₂ = 11.2 Hz, 1H), 3.72 (t, *J* = 6.8 Hz, 2H), 3.14 (m, 4H), 2.82 (m, 1H), 2.65 (m, 1H), 2.23 (m, 1H), 2.06 (m, 1H), 1.73 (m, 2H), 1.73 (t, *J* = 6.8 Hz, 2H). Anal. Calcd. for C₂₅H₃₁N₃O₃ · 2HCl · ½ H₂O: C, 59.64; H, 6.81; N, 8.35. Found: C, 59.51; H, 6.71; N, 8.31. Log*P* = 3.37.

3-Phenyl-1-[4-(4-pyridin-2-ylpiperazin-1-yl)butyl]piperidine-2,6-dione (3d)

Yield: 62%, mp 102–104 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.19 (m, 1H), 7.47 (t, *J* = 7.2 Hz, 1H), 7.33 (m, 4H), 7.17 (d, *J* = 7.2 Hz, 1H), 6.63 (m, 2H, pyridine), 3.84 (m, 3H), 3.59 (m, 4H), 2.75 (m, 2H), 2.60 (m, 4H), 2.46 (m, 2H), 2.23 (m, 2H), 1.61 (m, 4H). Anal. Calcd. for C₂₄H₃₀N₄O₂ · 2HCl · 2H₂O: C, 55.92; H, 7.04; N, 10.87. Found: C, 56.09; H, 6.74; N, 10.55. Log*P* = 3.14. *Crystal data*: C₂₄H₃₀N₄O₂ · ¹/₂ H₂O, crystal system monoclinic, space group *C*2 with unit cell dimensions: *a* = 18.908(4) Å, *b* = 6.244(1) Å, *c* = 19.087(4)= Å, β = 93.56(3)°, *V* = 2249.1(8)Å³, *Z* = 4, D(calcd) = 1.227 g/cm³. Independent reflections 4328, final *R* indices [for 1293 reflections with *I* > 2 σ (*I*)] *R*1 = 0.038, *wR*2 = 0.0961.

3-Phenyl-1-[4-(4-phenylpiperazin-1-yl)butyl]piperidine-2,6-dione (**3e**)

Yield: 60%, mp 177–179 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.17 (m, 10H, ArH), 3.85 (m, 3H, CH₂-1' and H-3), 3.22 (m, 4H, piperidine, H-2,6), 2.74 (m, 2H, CH₂-4'), 2.63 (m, 4H, piperidine, H-3,5), 2.45 (m, 2H, H-4), 2.20 (m, 2H, H-5), 1.60 (m, 4H, CH₂-2' and CH₂-3'). Anal. Calcd. for C₂₅H₃₁N₃O₂ · 2HCl · H₂O: C, 60.48; H, 7.11; N, 8.47. Found: C, 60.31; H, 6.79; N, 8.37. Log*P* = 3.76.

1-[4-(4-Benzylpiperazin-1-yl)butyl]-3-phenylpiperidine-2,6-dione (**3f**)

Yield: 55%, mp 248–250 °C. ¹H NMR (300 MHz, DMSO, TMS): δ 7.64 (m, 2H, Ar, H-3,5), 7.46 (m, 3H, Ar, H-2,4,6), 7.30 (m, 5H, ArH), 4.36 (br.s, 2H, CH₂-1'), 4.00 (dd, J_1 = 4.8 Hz, J_2 = 11.2 Hz, 1H, H-3), 3.67 (m, 10H, piperidine, H-2,3,5,6 and H-4), 3.09 (m, 2H, CH₂-4'), 2.80 (m, 1H, CH₂-Ar), 2.63 (m, 1H, H-4), 2.19 (m, 1H, H-5), 2.03 (m, 1H, H-5), 1.67 (m, 2H, CH₂-3'), 1.50 (m, 2H, CH₂-2'). Anal. Calcd. for C₂₆H₃₃N₃O₂ · 2HCl · $\frac{1}{2}$ H₂O: C, 62.27; H, 7.24; N, 8.38. Found: C, 62.58; H, 6.92; N, 8.47. Log*P* = 3.42.

1-{4-[4-(4-Fluorophenyl)piperazin-1-yl]butyl}-3-phenylpiperidine-2,6-dione (3g)

Yield: 73%, mp 178–180 °C. ¹H NMR (300 MHz, DMSO, TMS): δ 7.12 (m, 9H), 4.00 (dd, J_1 = 4.8 Hz, J_2 = 11.2 Hz, 1H), 3.70 (m, 4H), 3.50 (m, 2H), 3.14 (m, 7H), 2.81 (m, 1H), 2.64 (m, 1H), 2.22 (m, 1H), 1.74 (m, 2H), 1.50 (t, J = 7.2 Hz, 2H). Anal. Calcd. for C₂₅H₃₀FN₃O₂ · 2HCl · ³/₄ H₂O: C, 58.88; H, 6.62; N, 8.24. Found: C, 58.86; H, 6.35; N, 8.16. Log*P* = 3.92.

1-{3-[4-(2-Methoxyphenyl)piperazin-1-yl]propyl}-3-phenylpiperidine-2,6-dione (4a)

Yield: 63%, mp 178–180 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.33 (m, 3H), 7.18 (d, *J* = 7.6 Hz, 2H), 7.01 (m, 1H), 6.93 (t, *J* = 6.8 Hz, 2H), 6.86 (m, 1H), 3.93 (t, *J* = 6.8 Hz, 2H), 3.85 (m, 4H), 3.20 (m, 4H), 2.74 (m, 8H), 2.24 (m, 2H), 1.93 (m, 2H). Anal. Calcd. for $C_{25}H_{31}N_3O_3 \cdot 2HCI \cdot 2H_2O$: C, 65.60; H, 7.03; N, 7.92. Found: C, 56.87; H, 6.69; N, 8.08. Log*P* = 3.18.

3-Phenyl-1-[3-(4-pyrimidin-2-ylpiperazin-1-yl)propyl]piperidine-2,6-dione (4b)

Yield: 60%, mp 214–216 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.32 (d, J = 4.8 Hz, 2H), 7.33 (m, 3H), 7.18 (d, J = 7.2 Hz, 2H), 6.52 (t, J = 4.0 Hz), 3.88 (m, 7H), 2.76 (m, 8H), 2.23 (m, 2H), 1.96 (m, 2H). Anal. Calcd. for C₂₂H₂₇N₅O₂ · 2HCl · ¹/₂ H₂O: C, 55.58; H, 6.36; N, 14.73. Found: C, 55.43; H, 6.40; N, 14.59. Log*P* = 1.89.

1-{3-[4-(2-Hydroxyphenyl)piperazin-1-yl]propyl}-3-phenylpiperidine-2,6-dione (**4c**)

Yield: 65%, mp 158–160 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.33 (m, 3H), 7.17 (m, 3H), 7.07 (t, *J* = 7.6 Hz, 1H), 6.94 (d, *J* = 7.6 Hz), 6.85 (t, *J* = 7.6 Hz, 1H), 3.93 (t, *J* = 7.2 Hz, 2H), 3.88 (dd, *J*₁ = 5.2 Hz, *J*₂ = 8.8 Hz, 1H), 3.01 (m, 4H), 2.76 (m, 8H), 2.24 (m, 2H), 1.91 (m, 2H). Anal. Calcd. for C₂₄H₂₉N₃O₃ · HCl · 2 ½ H₂O: C, 58.95; H, 7.21; N, 8.59. Found: C, 59.45; H, 6.84; N, 8.33. Log*P* = 2.92.

3-Phenyl-1-[3-(4-pyridin-2-ylpiperazin-1-yl)propyl]piperidine-2,6-dione (4d)

Yield: 70%, mp 116–118 °C. ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.19 (d, *J* = 3.2 Hz, 1H), 7.47 (m, 1H), 7.32 (m, 3H), 7.17 (d, *J* = 7.2 Hz, 2H), 6.63 (m, 2H), 3.92 (t, *J* = 7.6 Hz, 2H), 3.84 (dd, *J*₁ = 5.2 Hz, *J*₂ = 9.2 Hz, 1H), 3.56 (m, 4H), 2.72 (m, 2H), 2.58 (m, 4H), 2.47 (t, *J*

= 6.8 Hz, 2H), 2.21 (m, 2H), 1.83 (m, 2H). Anal. Calcd. for C₂₃H₂₈N₄O₂ · 2HCl · 2H₂O: C, 55.09; H, 6.83; N, 11.17. Found: C, 55.48; H, 6.54; N, 10.97. Log*P* = 2.69.

Microbiological assays

Compounds

Compounds were dissolved in DMSO at 100 mM and then diluted in culture medium.

Cells and Viruses

Cell lines were purchased from American Type Culture Collection (ATCC). The absence of mycoplasma contamination was checked periodically by the Hoechst staining method. Cell lines supporting the multiplication of RNA viruses were the following: CD4⁺ human T-cells containing an integrated HTLV-1 genome (MT-4); Madin Darby Bovine Kidney (MDBK); Baby Hamster Kidney (BHK-21) and Monkey kidney (Vero 76) cells.

Bacterial strains

The antibacterial activity of compounds was tested against collection strains representative of Gram-positive bacteria (*Staphylococcus aureus* DSM 2569) and Gram-negative bacteria (*Pseudomonas aeruginosa* DSM 1117). Antifungal activity was tested against collection strains representative of yeasts (*Candida albicans* DSM 1386) and moulds (*Aspergillus niger* DSM 1988).

Cytotoxicity Assays

For cytotoxicity tests, run in parallel with antiviral assays, MDBK, BHK and Vero 76 cells were resuspended in 96 multiwell plates at an initial density of 6×10^5 , 1×10^6 and 5×10^5 cells/mL, respectively, in maintenance medium, without or with serial dilutions of tested compounds. Cell viability was determined after 48–120 hrs at 37 °C in a humidified CO₂ (5%) atmosphere by the MTT method. The cell number of Vero 76 monolayers was determined by staining with the crystal violet dye.

For cytotoxicity evaluations, exponentially growing cells derived from human haematological tumors [CD4⁺ human T-cells containing an integrated HTLV-1 genome (MT-4)] were seeded at an initial density of 1 x 10⁵ cells/mL in 96 well plates in RPMI-1640 medium, supplemented with 10% fetal calf serum (FCS), 100 units/mL penicillin G and 100 µg/mL streptomycin. Cell cultures were then incubated at 37 °C in a humidified, 5% CO₂ atmosphere in the absence or presence of serial dilutions of test compounds. Cell viability was determined after 96 hrs at 37 °C by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) method [24].

Antiviral assay

Activity of compounds against Human Immunodeficiency virus type-1 (HIV-1) was based on inhibition of virus-induced cytopathogenicity in MT-4 cells acutely infected with a multiplicity of infection (m.o.i.) of 0.01. Briefly, 50 μ L of RPMI containing 1x10⁴ MT-4 were added to each well of flat-bottom microtitre trays containing 50 μ L of RPMI, without or with serial dilutions of test compounds. Then, 20 μ L of an HIV-1 suspension containing 100 $CCID_{50}$ were added. After a 4-day incubation, cell viability was determined by the MTT method.

Activity of compounds against Yellow Fever Virus (YFV) and Reo virus type-1 (Reo-1) was based on inhibition of virus-induced cytopathogenicity in acutely infected BHK-21 cells. Activities against Bovine Viral Diarrhoea Virus (BVDV), in infected MDBK cells, were also based on inhibition of virus-induced cytopathogenicity.

BHK and MDBK cells were seeded in 96-well plates at a density of 5 x 10^4 and 3 x 10^4 cells/well, respectively, and were allowed to form confluent monolayers by incubating overnight in growth medium at 37 °C in a humidified CO₂ (5%) atmosphere. Cell monolayers were then infected with 50 µL of a proper virus dilution (in serum-free medium) to give an m.o.i = 0.01. 1 hr later, 50 µL of MEM Earle's medium, supplemented with inactivated foetal calf serum (FCS), 1% final concentration, without or with serial dilutions of test compounds, were added. After 3-4 days of incubation at 37 °C, cell viability was determined by the MTT method.

Activity of compounds against Coxsackie virus, B-2 strain (CVB-2), Polio virus type-1 (Polio-1), Sabin strain, Vesicular Stomatitis Virus (VSV), Vaccinia Virus (VV) and Herpes Simplex Virus type-1 (HSV-1), in infected Vero 76 cells, was determined by plaque reduction assays in Vero 76 cell monolayers. To this end, Vero 76 cells were seeded in 24-well plates at a density of 2 x 10^5 cells/well and were allowed to form confluent monolayers by incubating overnight in growth medium at 37 °C in a humidified CO₂ (5%) atmosphere. Then, monolayers were infected with 250 µL of proper virus dilutions to give 50-100 PFU/well. Following removal of unadsorbed virus, 500 µL of Dulbecco's modified Eagle's medium, supplemented with 1% inactivated FCS and 0.75% methyl cellulose, without or with serial dilutions of test compounds, were added. Cultures were incubated at 37 °C for 2 (Sb-1 and VSV) or 3 (CVB-2, VV and HSV-1) and then fixed with PBS containing 50% ethanol and 0.8% crystal violet, washed and air-dried. Plaques were then counted. 50% effective concentrations (EC₅₀) were calculated by linear regression technique.

Antibacterial and antifungal assays

The antibacterial and antifungal activities were evaluated by determining the Minimum inhibitory concentration (MIC) by the broth microdilution procedure.

Bacterial strains were grown on Tryptic soy agar at 37 °C for 1 day. Cell suspensions of these recent cultures were prepared in sterile 0.85% saline solution by 4–5 colonies. The turbidity of the suspensions was adjusted to the McFarland 0.5 standard. Suspensions were diluted in cation-supplemented Mueller-Hinton broth. For each microorganism, 100 μ L of the fivefold serial dilutions of the compounds in cation-supplemented Mueller-Hinton broth and 100 μ L of *inoculum* were added to each well of a microdilution plate (final titre 5 x 10⁵ CFU/mL). The inoculated plates were incubated at 37 °C in non-CO₂ incubator and humid atmosphere. The MICs were determined after 16–20 h [25].

Fungal strains were grown on Sabouraud's dextrose agar at 35 °C for 1–5 days. Suspensions of these recent cultures were prepared in sterile saline solution (NaCl 0.85%). Suspensions were then diluted in Sabouraud's dextrose broth. 100 μ L of the

fivefold serial dilutions of the compounds in Sabouraud's dextrose broth and 100 μ L of *inoculum* were added to each well of a microdilution plate (*C. albicans* 1 x 10⁴ cell/mL; *A. niger* OD₆₀₀ 0.05). The inoculated plates were incubated at 35 °C in non-CO₂ incubator and humid atmosphere. The MICs were determined after 24 and 48 h.

The concentration of each *inoculum* was confirmed by viable counts on agar plates by plating the appropriate dilution of the growth control well, immediately after inoculation, and incubating until visible growth. MIC corresponded to the lowest concentration of an antimicrobial compound that showed complete growth inhibition.

Linear regression analysis

Viral and cell growth at each drug concentration was expressed as percentage of untreated controls and the concentrations resulting in 50% (EC_{50} , CC_{50}) growth inhibition were determined by linear regression analysis.

Authors' Statement

Competing Interests

The authors declare no conflict of interest.

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