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Gut micobiota alteration by *Lactobacillus* rhamnosus reduces pro-inflammatory cytokines and glucose level in the adult model of Zebrafish

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Abstract

Objective: Type 2 diabetes mellitus (T2DM) is still a challenge for physicians to manage patient's circumstances. It is assumed that alterations in the normal flora may be involved in the pathogenesis of T2DM through inducing chronic inflammation. To investigate the effect of *Lactobacillus rhamnosus* as a common probiotic on T2DM, we induced an experimental model of T2DM in adult male Zebrafish by gradient hyper-glucose accumulation methodology.

Results: In this trial 3-month old male adult Zebrafish were divided in to four groups including two control groups and T2DM induced groups with or without probiotic treatment. After 5 days of acclimation, T2DM was induced by a gradient hyper-glucose accumulation methodology. Diabetic fishes had statistically abnormal blood glucose and proinflammatory cytokine levels compared to control group (p = 0.0001). These results suggest that probiotic intervention decreased the blood glucose level in the T2DM-P group by decreasing pro-inflammatory cytokines responsible for signaling in T2DM therapeutic modalities.

Keywords: Cytokine, Lactobacillus rhamnosus, Probiotic, Type 2 diabetes mellitus (T2DM), Zebrafish

Introduction

Metabolic syndrome has steadily increased globally and many etiological factors including altered inflammatory states, adipose abnormalities and insulin resistance, contribute to their pathogenesis and thus should be considered in related basic studies [1–3]. Diabetes mellitus (DM) which is known by glycemic disturbances is usually categorized to various categories. Type 1 DM patients are characterized by destruction of insulin-producing

pancreatic beta-cells and type 2 diabetes mellitus (T2DM) patients with lowered response to insulin. In T2DM, an individual's lifestyle and genetic coding are key risk factors [4, 5]. In obese T2DM patients, it was shown that losing weight reduces the glucose levels and can also improve glycemic control [6, 7]. However, there are still multiple challenges for physicians to manage patients' condition, including side effects and toxicity of current therapeutics and medication failures [8]. Recently, several studies have found that changes in gut microbiota composition play a role in the development of T2DM and obesity [1, 9]. Furthermore, nowadays it is useful to realize the effects of probiotics on diabetes management.

Probiotics are defined as live microorganisms with healthy beneficial to host if up taken in adequate amounts [10, 11]. Recent advances have highlighted the beneficial

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outcome in the pathology of inflammatory disorders after using gastrointestinal bacteria therapeutically [12]. Much evidence suggests that probiotic uptake has beneficial effects on blood glucose through different mechanisms [13–16]. Since intestinal microbiota and their metabolites directly contribute in insulin resistance; probiotic could have a strong effect on physiological function by altering gastrointestinal bacterial community [17, 18]. Consumption of a sufficient amount of probiotic is considered a therapeutic method for weight reduction and control of T2DM [19]. Furthermore, there is a direct correlation between the anti-diabetic drugs and gut microbiome community [9].

T2DM and obesity in the context of epidemiological data provide a solid platform for in vivo investigation of the disease. Nowadays, zebrafish (*Danio rerio*) has emerged as a powerful tool for scientific community among vertebrates. In the beginning of embryogenesis, zebrafish has transparent embryos with more than 85% of genetic similarity with human activity and thus as an attractive model organism for biomedical research [20]. Besides, it is also widely used for investigations on probiotics and host interaction with different gut microbiota and infectious diseases [21, 22].

Here in, we have developed a diet-induced zebrafish model for diabetes, to investigate the potential effect of oral administration of probiotic *Lactobacillus rhamnosus* as a diet supplement for inflammation caused by T2DM. If probiotic intervention could hinder the blood glucose elevation in the T2DM model via impact on cytokine levels, it might be considered a novel therapeutic approach in T2DM treatment.

Main text

Materials and methods

Animals

Three months old adult male fish as a gift from Zebrafish core facility, University of Tampere, Finland were divided into four groups including healthy control (HC), healthy control received probiotic (HC-P), T2DM group (T2DM) and T2DM group received probiotic (T2DM-P). Fish acclimation, induction diabetes and biometric analysis are described in Additional file 1 [23, 24].

Probiotic administration

Administration of *purchased L. rhamnosus* GG (ATCC: 53103) capsule from Culturelle Probiotics Co., Canada; is described in Additional file 1.

Blood glucose measurements

The blood collection used for glucometery using Match $^{\text{TM}}$ (OK Biotech Co, Taiwan) is described in Additional file 1.

Histological staining

After collecting biometric results, the small intestine of same fish used for histological staining procedures [25, 26] which is described in Additional file 1.

Quantitative real-time polymerase chain reaction analysis

Relative expression of IL-1 β and TNF- α (primer sequences in Additional file 2: Table S1) levels were calculated using $\Delta\Delta Ct$ method [27] using reverse transcription technique which is described in Additional file 1.

Statistical analysis

Data are presented as mean \pm standard error. Student's t-test was used for comparison between the two experimental groups using the statistical software package SPSS Statistics version 16 (IBM Corp., Armonk, NY, USA) with significance accepted at p < 0.001.

Results

Zebrafish biometric analysis

As summarized in Additional file 3: Table S2, a significant decrease in the length of the diabetic group (T2DM) and probiotic-treated diabetics (T2DM-P) was observed compared to controls (HC). Moreover, we observed a slight increase in the length of the probiotic-treated controls (HC-P vs. HC, P=0.013). Although both diabetic groups (T2DM and T2DM-P) showed lower weight compared to controls; the statistical analysis showed a significant difference between T2DM and HC (P=0.0001). As shown in Additional file 3: Table S2, a significant increase in the BMI of the HC-P group (1.255-fold) and a significant decrease in the T2DM group (0.343-fold) compared with HC (p<0.05) was observed.

Impact of probiotics on blood glucose

We observed significant blood glucose elevation in the T2DM group compared with HC and HC-P groups (P < 0.0001, Fig. 1). After 10 days treatment with probiotic, blood glucose has been decreased in the HC-P group in correlation with the supplemented probiotic (P = 0.011).

Influence of probiotics on zebrafish intestine

Histological analysis revealed some visible changes in the villus width and length during the probiotic treatment. Villus length has increased slightly in the T2DM-P group compared to T2DM; conversely, villus width has slightly

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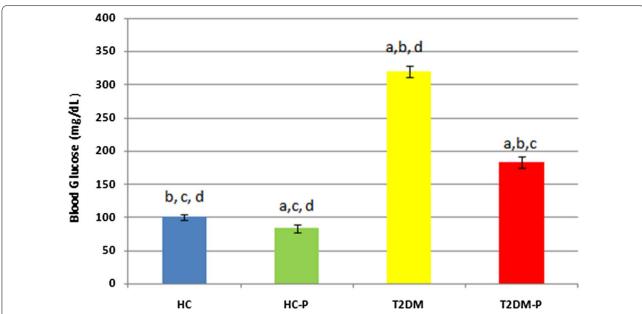


Fig. 1 This diagram represents the result of blood glucose from all trail groups. Data represents the mean + SE of the results for three independent assays. Statistical analysis compares difference of the mean of results between T2DM-P and HC, HC-P and T2DM. **a** Significant difference of other groups vs. HC-**b** Significant difference of other groups vs. HC-P. **c** Significant difference of other groups vs. T2DM-P Difference with p < 0.001 was considered significant. Each group contained 6 zebrafish

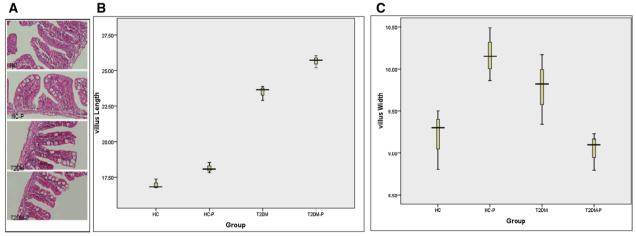


Fig. 2 A Histopathology evaluation of zebrafish small intestine. Intestinal tissues were stained with H&E and studied by microscopy (× 400 resolution). Healthy control group (HC), Healthy control group supplemented with probiotics (HC-P), Diabetic group (T2DM), Diabetic group supplemented with probiotic (T2DM-P). **B** Villus length increased slightly in the T2DM-P group following probiotic supplementation compared to T2DM group. **C**. Villus width was slightly higher in the T2DM group compared to T2DM-P group. Data shows mean ± SE for three independent assays

increased in the T2DM group compared with the T2DM-P (Fig. 2). Histological analysis of the intestinal tissues indicated a wide hyperplasia in the goblet cells located in microvilli's of the T2DM and T2DM-P groups.

Influence of probiotics on zebrafish pro-inflammatory cytokine expression

The relative expression of pro-inflammatory cytokines, IL-1 β and TNF- α was depicted in Fig. 3A, B. Compared to the control group (HC and HC-P, Fig. 3A), both cytokines were over-expressed in the T2DM group

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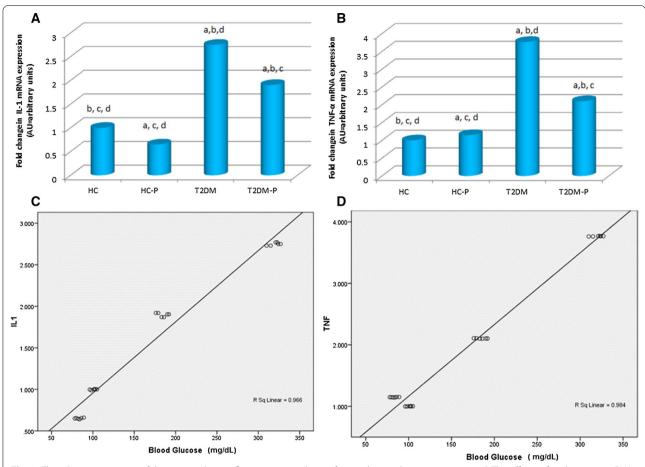


Fig. 3 The relative expression of the intestinal pro-inflammatory cytokines after 10 days probiotic intervention. **A** The effects of probiotic on mRNA expression of IL-1 β were determined by PCR. **B** The relative expression of TNF-α from all trail groups represent. The expression levels of β -actin were used as a loading control. **C** The Correlation of IL-1 β to blood glucose. **D** The Correlation of TNF-α vs. blood glucose. Control group (HC), Control + Probiotic (HC-P), Diabetic group (T2DM), Diabetic group with probiotic(T2DM-P) groups. a Significant difference of other groups vs. HC-P. c Significant difference of other groups vs. T2DM. d Significant difference of other groups vs. T2DM-P The mean difference is significant at p < 0.001

(P < 0.0001). The expression levels of IL-1β and TNF-α increased by 2.75 and 3.77-fold, respectively, in the T2DM group compared with HC. We found that probiotic supplementation resulted in a significant decrease in the expression levels of pro-inflammatory cytokines in T2DM-P group compare to T2DM (P < 0.001).

Correlation between blood glucose and pro-inflammatory cytokine

The correlation between blood glucose in various groups of zebrafish and pro-inflammatory cytokines (IL-1 β and TNF- α) expression was r=-0.966, p=0.001 and r=-0.984, p=0.001, respectively (Fig. 3C, D).

Discussion

A variety of genetic and feeding models of diabetes have been established, nevertheless, the majority of them have shown an inconsistency in pathological defects compared to human disease. Therefore, among creatures, zebrafish gained a growing platform for developmental research on diseases modeling, based on high similarity of digestive tract and comparable microbial and gut colonization with human [28–30]. Although, adult zebrafish has not been generally applied as an experimental model to study diabetes; here, we have well established it by overfeeding at adult stage, to study the effects of probiotic supplements on T2DM in vivo model.

To generate this model, we used a gradient hyper-glucose accumulation methodology. Physiologically, adult zebrafish absorb molecules from water to make a hyperosmotical internal environment, therefore, immersing them in a glucose containing solution (starting from 50 mM) and rising to final 200 mM could be beneficial to obtain a model of hyperglycemia in a much shorter time

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than other stepwise elevating glucose concentration of T2DM model [31]. According to our findings, this protocol increases blood glucose up to 300 mg/dL in T2DM Zebrafish which is in accordance to Gleeson et al. who showed a up to 400 mg/dL increase in blood glucose of adult zebrafish immersed in a 1% glucose solution [32]. Moreover, this method provides micro-environment stability and prevents fish fatality.

In the biometric result, probiotic-supplemented diet improved fish weight compared with the standard diet in both T2DM and HC groups similar to observations of Valcarce et al. [30]. Although, we did not expect to see significant changes on the length of the fish during the treatment periods, our data indicated a slight increase in the length and calculated body mass index (BMI) of HC-P and T2DM-P groups. Typically BMI as an excess of body fat is a marker of obesity and obesity related diseases. Although, the pathogenesis of obesity and its association to metabolic disorders like T2DM is complicated, adipose tissue polarized macrophages to increase secretion of pro-inflammatory cytokines such as TNF- α . Moreover, gut microbiota alterations through probiotic is considered to induce the host inflammatory response. As we shown in Fig. 3B, TNF- α was increased 1.149 in healthy group treated by probiotic. It is assumed that such a small increment of cytokine expression is not able to completely activate the entire inflammatory pathway like chronic inflammation process induced by obesity [33]. Falcinelli et al. exposed adult zebrafish to different amounts of lipid to understand the correlation between microbiota and obesity. They indicated that high lipid contents reduced community diversity which affected the genes transcription which involved in appetite control and supplementation of the diet with Lactobacillus rhamnosus decreases total body cholesterol in zebrafish [34]. In addition to TNF-α, IL-1 is another pro-inflammatory cytokines which released in obese animal via M1 macrophage stimulation [35]. According to our data, a significant decrement of IL-1 was also occurred in HC-P group compared to HC (1.533) which might be explained the probiotic modality on whole cytokine profile alteration in experimental model. This finding remarkably points out to the strong value of probiotic and contributes with the physiological condition which enhances effective fish development and growth [14, 19].

Our histological analysis clearly showed some visible changes in the villus length and width during the probiotic treatment. Since the T2DM fish groups were under harsh condition due to high concentration of glucose, their homeostasis tries to adapt the condition to survive by extension of the villus diameter to increase the absorption. Additionally, result of intestinal staining specified hyperplasia in goblet cells located in microvillis'

in the T2DM group. As, one of the primary characteristic sign of obesity is hyperplasia and hypertrophy; administration of probiotic had such an ability to prevent goblet cells disruption due to high glucose in digestion system of our zebrafish diabetic model [36, 37].

Alterations in the intestinal homoeostasis may play a major role in the development of systemic inflammatory diseases including diabetes in which blood glucose management is challenging [38, 39]. In parallel with previous research, we showed that the consumption of probiotic based foods had significantly decreased blood glucose [40, 41]. Moreover, we identified the blood glucose level slightly lower in the HC-P group than the others. Since probiotic can strongly affect the growth, development and immune system improvement; thus, current results specified the probiotic *Lactobacillus rhamnosus* capability to improve tolerance in high glucose concentration [40].

Recently, the contribution of the mucosal immune system and the gut microbiome in metabolic disease including T2DM has been highly concerned [38, 42]. Our findings showed that relative mRNA expression levels of IL1- β and TNF- α were down-regulated in fish with probiotic supplementation despite the induction of innate immune-related cytokine genes by probiotic *Chromobacterium aquaticum* reported by Yi et al. [43]. This apparent inconsistency in cytokine profile was seen among the genus of *Lactobacillus* whereas *L. sakei* induced proinflammatory cytokines including IL-1 β and TNF- α ; and *L. johnsonii* promoted the production of TGF- β in cellular models [44].

Moreover, there was a robust positive correlation between both IL1- β and TNF- α and blood glucose levels (Fig. 3C, D). Delgadillo-Silva et al. showed that altered composition of the gut microbiome stimulate the intestinal residing innate and adaptive immune cells and induce a cytokine-mediated inflammation which is accompanied by hyperglycemia [45]. TNF as an inhibitor of insulin signaling is a major contributor towards obesityrelated diseases. Indeed, it has been demonstrated that mentioned cytokines may play role in the inflammatory destruction of insulin-producing β-cells in human T2DM [39, 46, 47]. Due to their synergistic effect, pharmacological blockage of mentioned cytokines has been clinically modulating inflammatory diseases; however, a therapeutic gap for managing islet inflammation and cytokine production in T2DM is present [48].

Finally, we assumed that probiotic bacteria, like *L. rhamnosus*, through their pathogen associated molecular pattern signaling pathway and bioactive components might reduce the immune-cell infiltration, decrease pro-inflammatory cytokines and ameliorate the hyperglycemic phenotype n fish models. Hence, probiotic

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intervention might be useful in blood glucose management in T2DM patients.

Conclusion

In summary, this study elucidates that probiotics such as *Lactobacillus rhamnosus* may hinder the blood glucose elevation in the T2DM group by their immunemodulatory effects. Thereby development of a probiotic based therapeutic formulation could promote healthiness in patients with T2DM.

Limitations

The limitation of this study includes lack of data on time dependent changes in blood glucose levels, cytokines levels and comparing different type of probiotics. Therefore, the effectiveness of various probiotic and their long-term impact on glucose levels should be investigated in this useful experimental model.

Abbreviations

ATCC: American Type Culture Collection; BMI: Body mass index; BP: Base pair; CFU: Colony forming unit; EMRI: Endocrinology and Metabolism Research Institute; HC: Healthy control group received no probiotic treatment; HC-P: Healthy control group treated with probiotic; H&E: Hematoxylin and eosin; IL-1β: Interleukin 1 beta; Real-time PCR: Real-time polymerase chain reaction; SE: Standard error; T2DM: Type 2 diabetes mellitus; T2DM: T2DM group received no probiotic supplement; T2DM-P: T2DM group received probiotic supplements; TGF-β: Transforming growth factor beta; TNF-α: Tumor necrosis factor alpha.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13104-021-05706-5.

Additional file 1. Complementary information of animals, probiotic administration, histological staining and quantitative real-time polymerase chain reaction analysis.

Additional file 2: Table S1. Primer sequences used for amplification of specific genes through real-time quantitative PCR.

Additional file 3: Table S2. Summary of the effects of probiotic supplementation on length, weight gain and BMI of experimental groups.

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Authors' contributions

FB contributed during the experimental process, performed zebrafish diabetic modeling, animals tissue sampling and wrote the first manuscript. FS1 conducted probiotic dosing and control of treatments, performed statistical analysis and depictions and revised the manuscript. RF edited all drafts and rewrote the final version of the paper. HM1 provided special tools and reagents for animal husbandry and performed control of incubation conditions. RC performed statistical analysis and conducted probiotic dosing and control of treatments. HM2 provided reagents for animal husbandry and conducted probiotic dosing and control of treatments. FS2 consulted in special measures for animal husbandry and assisted in interpretation of histology data. MRK conceived, directed, and designed the study, obtained funding, edited

all drafts and rewrote the final version of the paper. All authors read and approved the final manuscript.

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Availability of data and materials

The data supporting our findings be presented within the manuscript and additional supporting files. Moreover, other datasets used during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

All animal experiments were performed according to the ethical guidelines and standards of fish treating in Zebrafish Fish Core Facility (ethical code number: ZDB-LAB-190117-2).

Consent for publication

Not applicable. As all images are entirely unidentifiable and there are no details on individuals reported within the manuscript, consent for publication of images may not be required.

Competing interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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