Synthesis and Structural Modification of Marine Natural Products

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Abstract: In the last decades, marine natural products (MNPs), have attracted extensive interest from both chemists and pharmacologists due to their chemical and bioactive diversities. This special issue, collecting total synthesis and structural modification of six different type of bioactive MNPs, is expected to inspire and attract more research effects invested into MNP research.

1. Introduction

It is not surprising that marine organisms can produce secondary metabolites that are very different from those produced by terrestrial plants, given their special marine living environment conditions including high concentration of salts, high pressure, low concentration of oxygen, and dark condition [1]. In recent years, the capability of marine organisms to produce highly potent bioactive metabolites with unique structures has attracted wide interest of chemists and pharmacologists aiming to find marine natural products (MNPs) with potential clinical value [2,3].

In the projects aiming at discovering drug leads/candidates derived from MNPs, a number of novel bioactive MNPs and their analogs have been identified. In this special issue, the total synthesis and structural modification of six different types of bioactive MNPs associated with Guo's, and other, groups will be discussed.

## 2. The Synthesis and Modification of Bioactive NMPs

2.1. Phidianidines A (**1a**) and B (**1b**)

1,2,4-Oxadiazole is a common, but very important, pharmacophore widely found in synthetic molecules. However, natural products possessing a 1,2,4-oxadiazole ring system are extremely rare. To the best of our knowledge, phidianidines A (**1a**) and B (**1b**) (Figure 1) are the first natural representatives of 1,2,4-oxadiazole derivatives, isolated from Chinese aeolid opisthobranch *Phidiana militaris* [4]. Both novel tryptophan-derived alkaloids showed robust cytotoxicity against C6 and HeLa cells, within nanomolar range. In addition, phidianidine A was found to be a new antagonist of CXCR4 [5], an important therapeutic target for the treatment of diseases like HIV infection, rheumatoid arthritis and cancer.
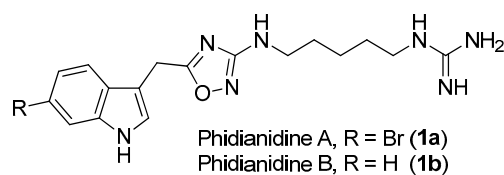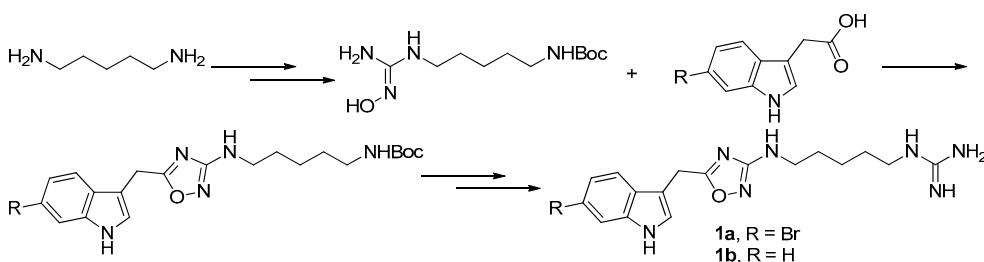
Figure 1. The structures of phidianidines A (**1a**) and B (**1b**).

    To provide novel phidianidines derivatives for further pharmacological study, several research groups have launched projects aiming at their total syntheses. The first synthesis of phidianidines A (**1a**) and B (**1b**) was reported by Snider et al. (Figure 2) [6]. Generally, the synthesis started with 1,5-diazidopentane and the key 1,2,4-oxadiazole intermediates were generated by the reaction of indole-3-acetyl chloride with *N*-5-azidopentyl-*N*′-hydroxyguanidine. Then, reduction of the terminal azide afforded the corresponding amine, which was smoothly transformed to guanidine. Finally, the syntheses of phidianidines were completed in 19% overall yield. At almost the same time, Lindsley and coworkers described the synthesis of **1a** and **1b** (Figure 3), with a similar approach to that of Snider [7]. In their work, both alkaloids were found to be selective inhibitors of the dopamine transporter (**1a**: $IC_{50} = 390$ nM; **1b**: $IC_{50} = 680$ nM) and selective agonists of the μ opioid receptor (**1a**: $EC_{50} = 17\%$ nM; **1b**: $EC_{50} = 12\%$). However, there are some critical aspects, such as the use of toxic reagents (i.e., BrCN) and the formation of unstable intermediates in the above protocols. In order to avoid these inconveniences, Manzo and coworkers reported a simpler and easier synthetic route, which is based on the coupling of 3-indolacetic acid methyl ester and the amino-alkyl hydroxy guanidine intermediate (Figure 4) [8]. The latest concise synthetic approach reported by Chamberland et al. was inspired by the proposed biosynthetic hypothesis of phidianidines. In their protocol, the azide was not used as an amine protecting group based on safety concern and the guanidine was installed first to eliminate the unnecessary steps of deprotection (Figure 5) [9].
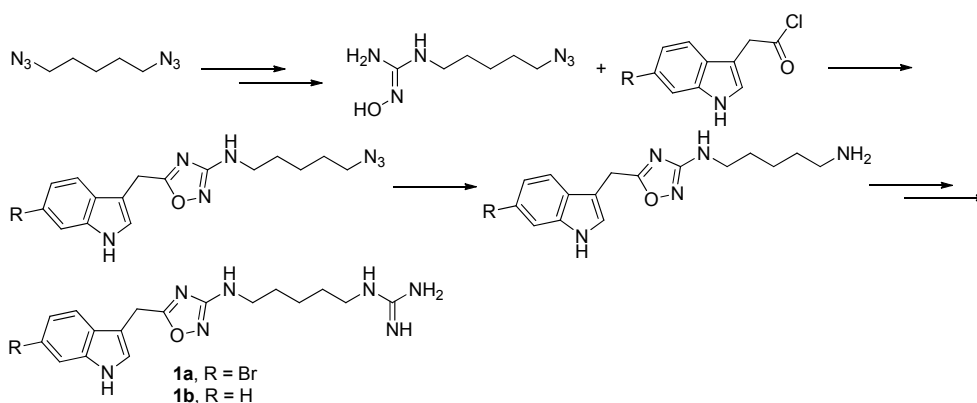
Figure 2. Sinder's protocol [6] for the synthesis of phidianidines.

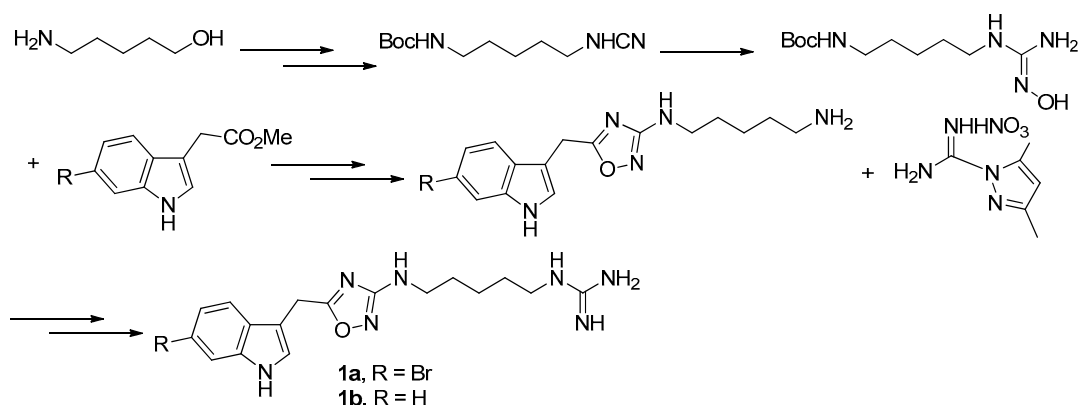**Figure 3.** Lindsley's protocol [7] for the synthesis of phidianidines.

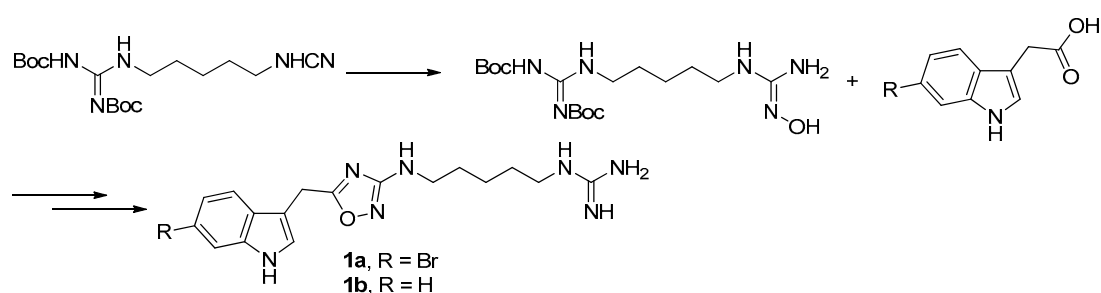**Figure 4.** Manzo's protocol [8] for the synthesis of phidianidines.

Figure 5. Chamberland's protocol [9] for the synthesis of phidianidines.

Since there was no structure-activity relationship (SAR) study on phidianidines, the first mimic synthesis and SAR study of phidianidine B (**1b**) was carried out by Guo's group (Figure 6). Based on a scaffold hopping strategy, the benzene and other aromatic ring systems were introduced to modify the alkyl-guanidinium side chain, resulting in a series of phidianidine B-based analogs as shown in Figure 6. First, the alkylguanidinium side chain was modified by the introduction of the furan ring moiety in part A and substituted phenyl or aminopyrimidine in part B; the furan ring moiety was then replaced by thiophene, benzene, or pyridine. All of these analogs were evaluated for bioactivities in different screen models. Among these analogs, derivative **1c** showed good in vitro neuroprotective effects against amyloid-β_{25-35}-, hydrogenperoxide-, and oxygen-glucose deprivation-induced neurotoxicity in SH-SY5Y cells, indicating that **1c** is a promising drug candidate for treating Alzheimer's disease [10]. In addition, derivative **1d** was found to be a novel potent selective inhibitor of protein tyrosine phosphatase 1B (PTP1B, IC_{50} = 8.96 μM), a potential target for treating type II diabetes and obesity [11].

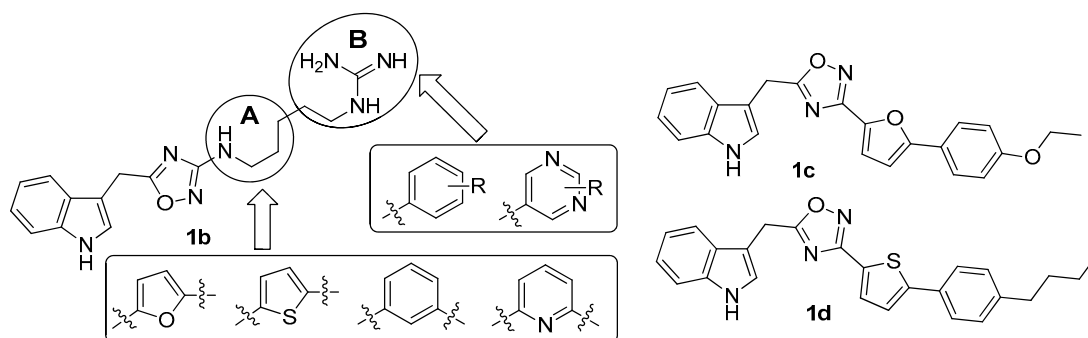**Figure 6.** The synthesis of phidianidine B mimic in Guo's group [10,11].

2.2. *Gymnorrhizol* (**2a**) *and Bruguiesulfurol* (**2b**)

Marine disulfide- and multisulfide-containing metabolites are a special and important class of natural products, which exhibited promising bioactivities, including antitumor, antibiotic, and enzyme-inhibitory activities. In particular, the disulfide bond was found to play an important role for their bioactivities [12]. The chemical constituent investigation on the Chinese marine mangrove *Bruguiera gymnorrhiza* resulted in the isolation of several cyclic disulfides [13–16]. Among these disulfides, an unusual 15-membered macyrocylic polydisulfide, gymnorrhizol (**2a**), and its known monomer, bruguiesulfurol (**2b**) (Figure 7) [17], were first found to be promising PTP1B inhibitors, with IC$_{50}$ values of 14.9 and 17.5 μM, respectively.
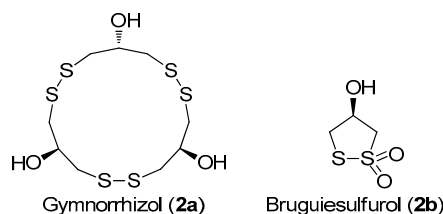
Figure 7. The structures of gymnorrhizol (**2a**) and bruguiesulfurol (**2b**).

The first total synthesis of gymnorrhizol (**2a**) was prepared by Guo's group in only three steps, starting from (*R*)-1-bromo-3-chloroisopropanol and 1,3-dichloropropan-2-ol as shown in Figure 8. The 1,3-dichloropropan-2-ol was converted into its bispotassium salt, which then reacted with Bunte salt produced from (*R*)-1-bromo-3-chloroisopropanol to give the target compound **2a** in 34% overall yield [16]. Later we started a project to synthesize bruguiesulfurol (**2b**) with epibromohydrin as starting material. As hydroxyl-protecting group, 4-bromobenzoyl was initially selected since it makes small-molecule compound easy to be purified and crystallized. However, the target compound **2b** was not obtained from **2d** in a variety of deprotection conditions, because **2d** cannot be hydrolyzed in acidic conditions and rapidly decomposed in basic conditions. Then, the 2-tetrahydropyranyl group was used as protecting group, which was easily removed under acidic conditions. Finally, the total synthesis of **2b** was successfully achieved for the first time [18], with a key step of mCPBA oxidation and deprotection in one pot with an overall yield of 30% (Figure 8).
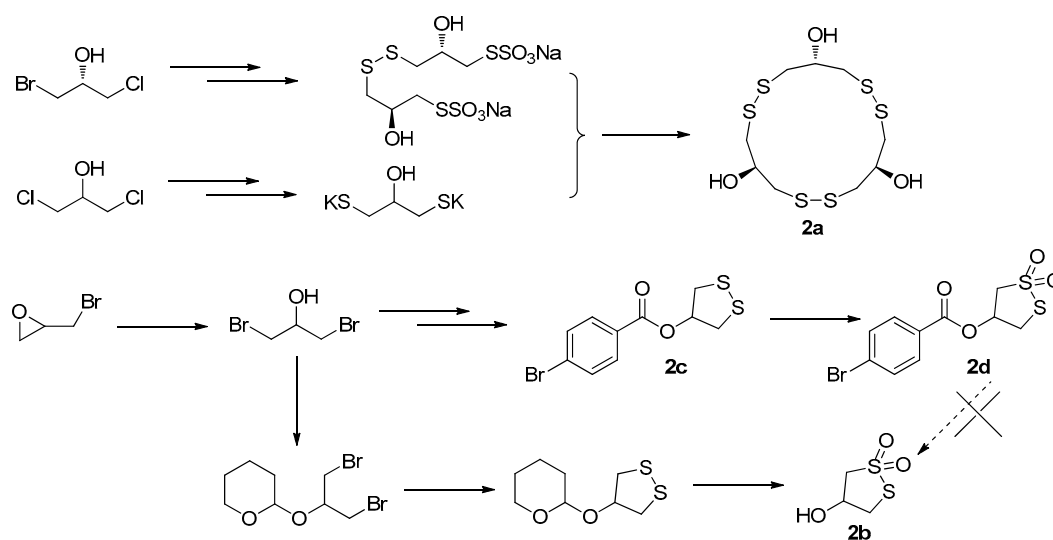
Figure 8. The total synthesis of **2a** and **2b** in Guo's group [16,18].

During the synthesis of **2b**, a key intermediate **2c** with unoxidized disulfide ring was found to exhibit stronger PTP1B inhibitory activity (IC$_{50}$ = 11.01 μM) than that of **2b** and significant selectivity toward PTP1B versus other PTPs. Based on this observation, a series of 1,2-dithiolane-4-ol derivatives **2e** with different benzoyl group and its oxidative products **2f** and **2g** were prepared [18]. The bioassay results indicated that the compound **2c** still showed the best PTP1B inhibitory and the cyclic dithiolane moiety was essential for PTP1B inhibition. Since the current PTP1B inhibitors usually have undesirable cell permeability and oral bioavailability due to the presence of highly negatively-charged polar pharmacophores in their structures [19], a further structural optimization on **2c** was carried out by the introduction of less polar substituted benzoate in 4-position of benzene with a different aliphatic chain as a linkage (**2h**) aiming at increasing its activity. When the bromine group in **2c** was replaced by the hex-5-yn-1-yl 2,5-dibromobenzoate fragment, the PTP1B inhibitory activity was increased by almost 20-fold (IC$_{50}$ = 0.59 μM) (Figure 9) [20].
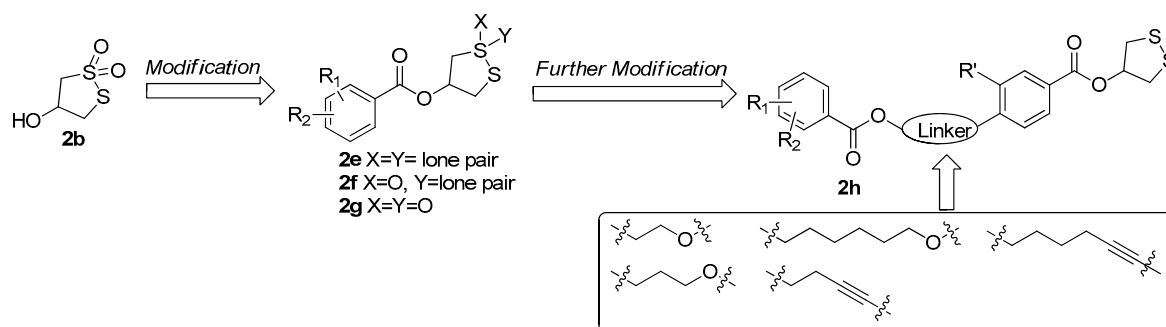

Figure 9. The synthesis of bruguiesulfurol derivatives in Guo's group [18,20].

2.3. Paracaseolide A (3)

Paracaseolide A (**3**), a novel α-alkylbutenolide dimer characterized by an unusual tetraquinane oxa-cage bislactone skeleton bearing two linear alkyl chains, was isolated from the Chinese mangrove plant *Sonneratia paracaseolaris* (Figure 10) [21]. This compound exhibited significant inhibitory activity against dual specificity phosphatase CDC25, a key enzyme for cell cycle progression and associated with tumor aggressiveness, with an IC$_{50}$ value of 6.44 μM.
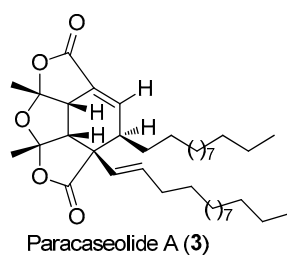

Paracaseolide A (**3**)

Figure 10. The structure of paracaseolide A (**3**).

The fascinating, skeletally-unique structure quickly captured the attention of synthetic chemists after a short period of its reveal. The first total synthesis of paracaseolide A (**3**) was reported by Noutsias and Vassilikogiannakis [22]. The synthetic strategy was based on a bioinspired [4 + 2]-dimerization/ketalization/epimerization of a 4-hydroxybutenolide precursor **3a**, generated by singlet oxygen-mediated oxidation of a furan intermediate formed from tridecan-1-ol and furan-2-carbaldehyde (Figure 11). In order to obtain suitable quantities of **3** for SAR study, Kraus and Guney developed a concise synthetic route allowing strategically-distinct generation of **3** and its analogs [23]. Their syntheses began with ene-2,5-dione, which was converted into the key tetraquinane oxa-cage bis-lactone **3b** characterized by a great potential to be introduced functionality into its

molecular skeleton. Finally, the target **3** was synthesized after a tandem vicinal difunctinalization of a α,β-unsaturated lactone **3c** to form **3d**, and followed by elimination to achieve the requisite unsaturation (Figure 12). In addition, researchers from other five different groups have also reported the total synthesis of **3** involving the same key step as that of Vassilikogiannakis's protocol, namely the [4 + 2]-dimerization of 4-hydroxybutenolide (**3a**) [24–28].
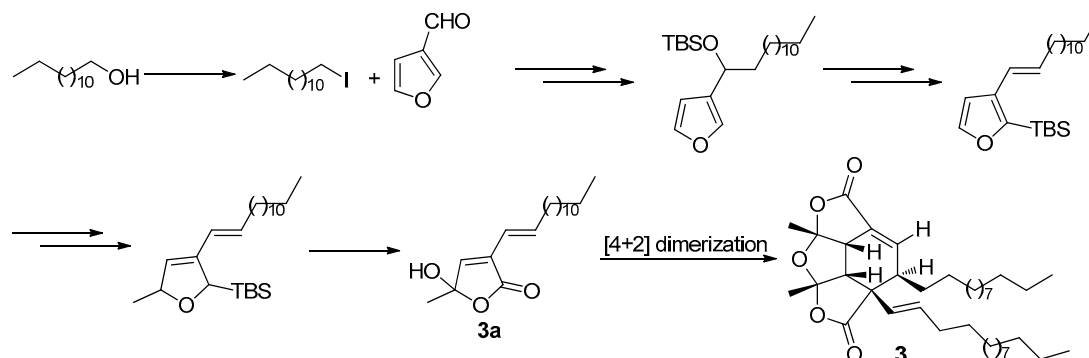

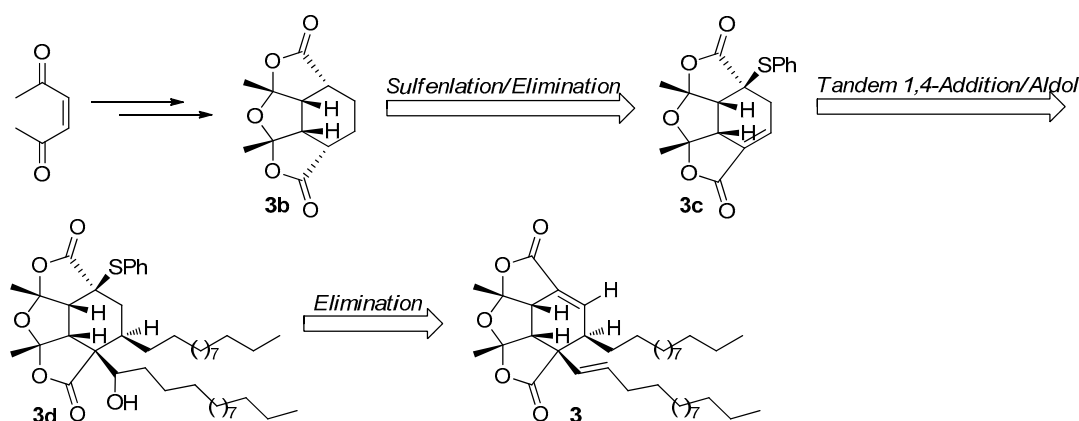**Figure 11.** Vassilikogiannakis's protocol [22] for the synthesis of **3**.

Figure 12. Kraus's protocol [23] for the synthesis of **3**.

2.4. (R)-de-O-Methyllasiodiplodin (**4**)

The twelve-membered macrolide (*R*)-de-*O*-methyllasiodiplodin (**4**, Figure 13), previously isolated from plants and fungus, was found to show various bioactivities, such as inhibition of prostaglandin biosynthesis, antimicrobial, and significant antitumor activities [29]. This macrolide was found to be a potent nonsteroidal antagonist of mineralocorticoid receptor (MR), a validated target for treating hypertension and other cardiovascular diseases, with an IC_{50} value of 8.93 μM [30].

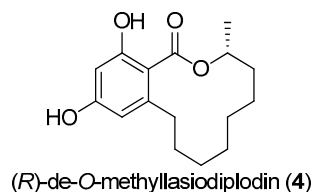(*R*)-de-*O*-methyllasiodiplodin (**4**)

Figure 13. The structure of (*R*)-de-*O*-methyllasiodiplodin (**4**).

The first asymmetric total synthesis of **4** was published in 1990 by Ruano et al. [31]. The orcinol monohydrate was used as a starting material and the key chiral center was smoothly created in

the last synthetic steps by asymmetric induction of a chiral sulfoxide group as shown in Figure 14. However, this synthetic route was somewhat complicated and low-yielding with 18 steps in 0.8% overall yield. In 1996, a short and flexible route toward **4**, starting with 3,5-dimethoxyphenol which was subsequently carboxylated via a kolbe-Schmitt reactions, was developed by Alois Fürstner and coworkers [32]. Finally, the macrolide ring was formed in the key ring closing metathesis (RCM) reaction catalyzed by Gurbbs I generation catalyst (Figure 15). In 2009, a more facile and efficient preparation for **4** suitable for the general synthetic laboratories was completed [33]. The synthetic route to **4** shown in Figure 16 was similar with Fürstner's protocol, but started from orcinol monohydrate and used Gurbbs II catalyst in the RCM reaction. Noticeably, the yield of the last demethylation step was increased to 57% by performing the quenching under reduced pressure, compared with Ruano's procedure (17% yield).

To optimize the activity of **4**, a series of hydroxyl-derivatives **4a** were prepared and evaluated for their antagonistic activity against MR. The bioassay result indicated that the diacetylated compound exhibited a more potent antagonistic effect against MR that **4**, with an IC_{50} value of 2.78 μM. Further, another series of derivatives were prepared by preserving diacetylate moiety and introducing different ring size of the lactone without chiral methyl. Finally, the IC_{50} value for MR antagonistic activity was increased to 0.58 μM, when the n was 5. The current SAR study indicated that the acetylation at the phenolic hydroxyl group increases the MR antagonistic effect and the ring size of the lactone was very crucial for its activity [30].
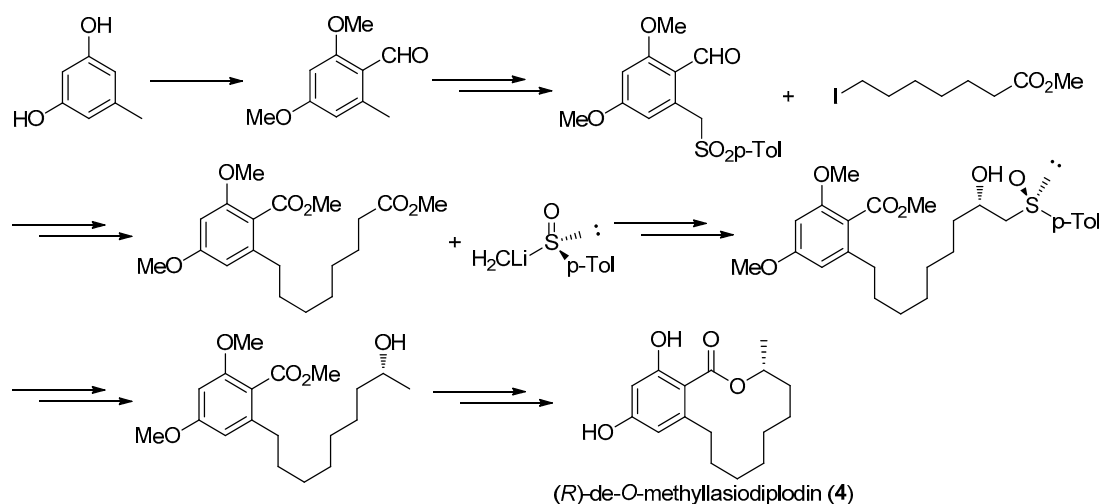

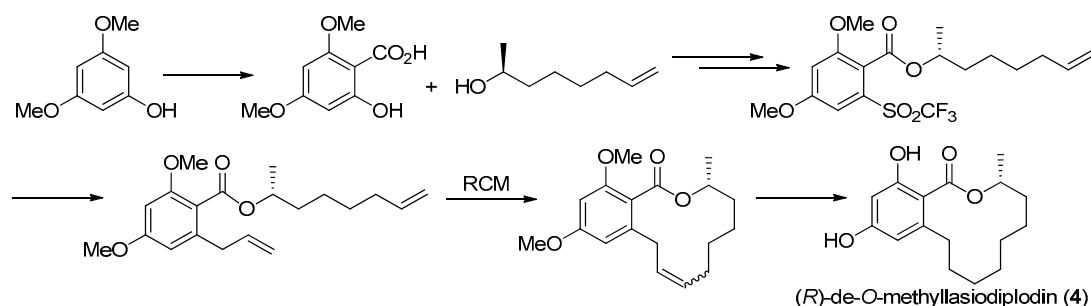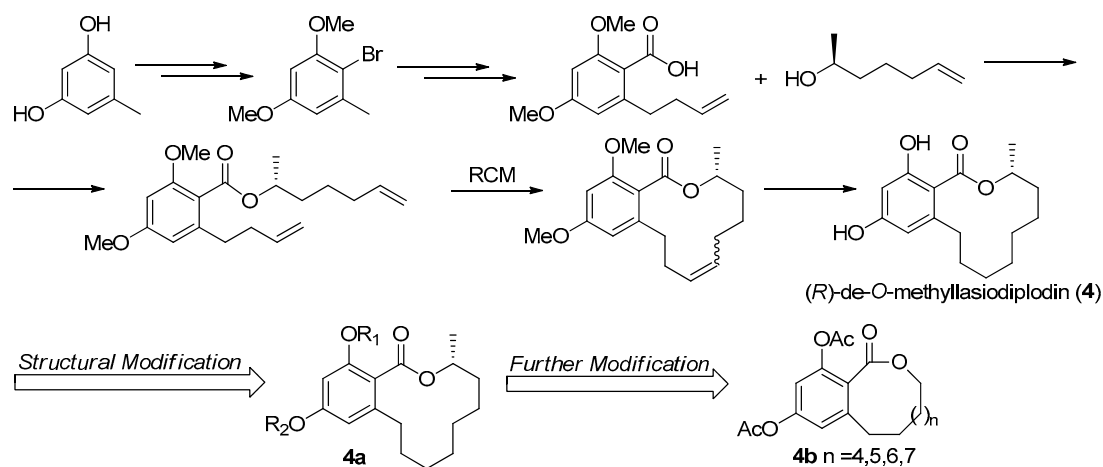**Figure 14.** Ruano's protocol [31] for the synthesis of **4**.

Figure 15. Alois Fürstner's protocol [32] for the synthesis of **4**.

Figure 16. Synthesis and structural modification of **4** in Guo's group [30,33].

2.5. Methyl Spongoate (**5**)

Methyl spongoate (**5**) (Figure 17) is a novel steroid with an uncommon C-20 methoxycarbonyl group with *R* absolute configuration, which was isolated from the Sanya soft coral *Spongodes* sp. This compound showed significant cytotoxicity against BEL-7402 (IC$_{50}$ = 0.33 μM), P388 (IC$_{50}$ = 8.9 μM), A-549 (IC$_{50}$ = 11.7 μM), and HT-29 cell lines (IC$_{50}$ = 11.7 μM) [34]. However, its low natural yield with 2.5 mg obtained from 456 g (dry weight) of the *Spongodes* sp. limited further potential pharmacological studies. In order to supply sufficient amount of **5**, its first stereoselective synthesis was carried out by Gong et al. [35]. As shown in Figure 17, the total synthesis of **5** started from the pregnenolone acetate and was successfully achieved in 14 steps with an 11% overall yield, which is amenable to scale-up in an average chemical laboratory.
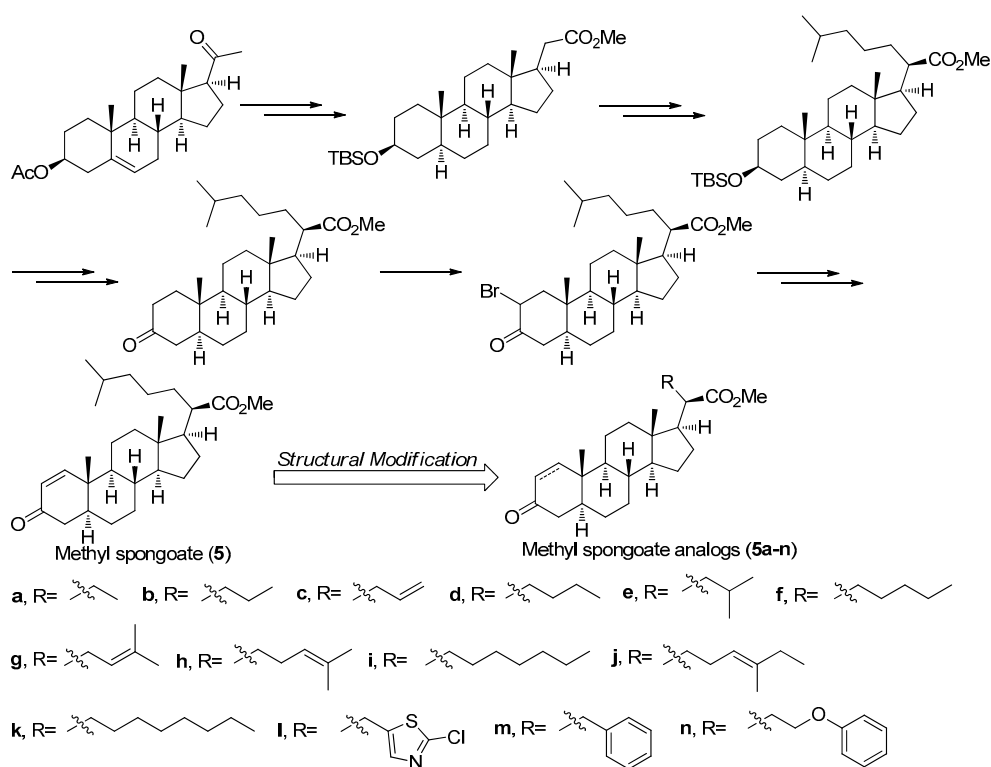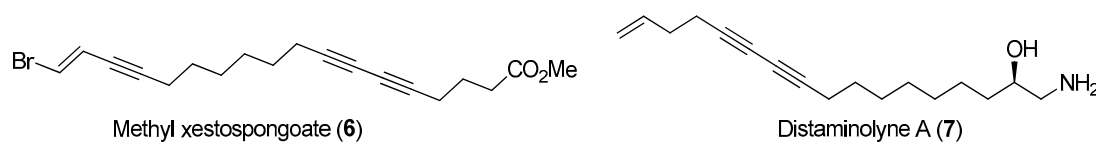

Figure 17. The structure, synthesis and SAR study of **5** in Guo's group [35,36].

Based on a similar synthetic strategy, a series of related derivatives **5a-n** with different C-20 side-chains were prepared for further anti-tumor SAR studies [36]. All of the analogs and **5** were evaluated against a panel of cancer cell lines including A549, HCT-116, HepG2, SW-1990, MCF-7, and NCI-H460. The results showed that compound **5i** having a heptyl side chain had the best inhibition against HepG2, SW-1990, MCF-7, and NCI-H460 cell lines with IC_{50} values ranging from 6 to 10 μM, much better that that of **5** (IC_{50} values ranging from 17 to 30 μM). The preliminary SAR study not only have demonstrated that the side-chain has an important role in determining antitumor activity but also confirmed the significance of the α,β-unsaturated carbonyl moiety of A ring in the steroid as an electrophilic Michael acceptor in the antitumor activity.

### 2.6. Methyl Xestospongoate (**6**) and Distaminolyne A (**7**)

Polyacetylenic natural products isolated from marine algae and invertebrates often displayed a broad array of biological properties, including cytotoxicity, antimicrobial activity, and various enzyme inhibitory activities [37]. During the investigation of components of the extract of marine sponge *Xestospongia testudinaria*, methyl xestospongoate (**6**, Figure 18), a brominated polyacetylenic fatty acid, was found to exhibit strong pancreatic lipase (PL) inhibitory activity with an IC_{50} value of 3.1 μM [38,39].

Figure 18. The structures of methyl xestospongoate (**6**) and distaminolyne A (**7**).

The first total synthesis of **6** was achieved in five steps with 30% overall yield, based on the retrosynthesis analysis. The key synthetic steps include the Sonogashira coupling reaction of 1,2-dibromoethylene with intermediated **6a**, and the Cu(I) catalyzed cross-coupling reaction of 1,9-decadiyne with methyl 6-bromo-5-hexynate (Figure 19).

To better understand its SAR information, a following structural modification on **6** was carried out to produce a series of halogenated analogs **6b** with different chain length. The bioassay results indicated that the PL inhibitory activity of the terminal brominated ones are better than that of the chlorinated ones, and the chain length with 16-20 C-atom might be the most optimal for their PL inhibitory activity. However, the structural modification made activity of all these analogs decrease or lose, with inhibition percentage at 50 μM less than 44.53%, compared with that of **6** with 68.84% inhibition at 50 μM [40].
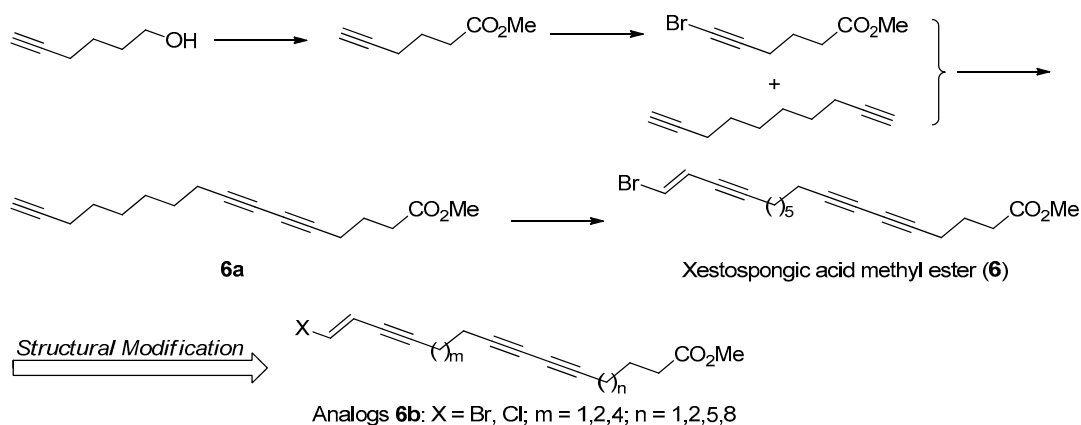
Figure 19. The synthesis and structural modification of **6** in Guo's group [40].

　　　Distaminolyne A (**7**, Figure 18) was an antibacterial acetylenic amino alcohol isolated from New Zealand ascidian *Pseudodistoma opacum* [41]. The structure of this compound is similar to that of methyl xestospongoate (**6**), making it a possible PL inhibitor. In addition, the chiral 1,2-amino alcohol group is an important structural function group in bioactive compounds. All these observations led us to embark on the total synthesis of distaminolyne A (**7**) and its enantiomer (**7a**). The first total synthesis of both compounds were achieved from the commercially-available undec-10-en-1-ol (Figure 20) [42]. A key proline-catalyzed asymmetric α-amino-oxylation of an aldehyde intermediate was used to introduce the chiral center en route to the enantiomerically-pure 1,2-amino alcohols. Bioassay result revealed that both enantiomers displayed weak PL inhibitory activity, with 36.7% and 24.7% at 50 μM, respectively. Interestingly, the [α]$_D$ values of both synthesized enantiomers indicated that the absolute configuration of the natural distaminolyne A should be revised as 2*R*.
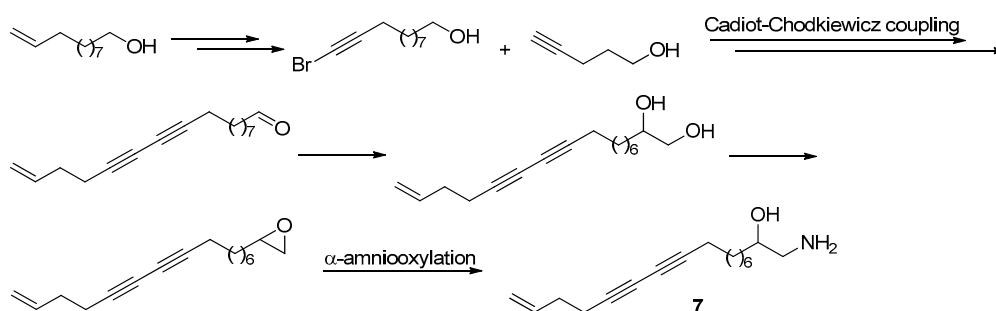

Figure 20. The synthesis of **7** in Guo's group [42].

3. Conclusions

　　　Marine natural products (MNPs) have become a valuable resource for the development of novel drugs. However, their low natural yield extremely limits the further druggability research and few of them can be directly used in clinical applications. Therefore, the total syntheses of MNPs are essentially necessary for studying their biological activities, while the structural modifications are useful for yielding safer, more potent and selective molecules, with improved physicochemical and pharmacokinetic properties. The examples above briefly illustrate the synthesis and structural modification of some selected MNPs, which is only a small basic part during the development of marine product-derived drugs in recent years. To promote the development of efficient synthetic and modified strategies for MNPs, closer cooperation between organic chemists, medicinal chemists, and pharmacologists is desired.

Author Contributions: C.-S.J. and Y.-W.G. designed the manuscript and significantly participated in its writing; J.Z. and H.Z. contributed to writing of the article; and L.A.M. was significantly engaged in correcting the text.

References

1.　　Prinsep, M.R. Sulfur-containing natural products from marine invertebrates. In *Studies in Natural Products Chemistry*; Rahman, A.U., Ed.; Elsevier Science B.V.: Amsterdam, The Netherlands, 2003; Volume 28, pp. 617–751.

2.　　Mayer, A.M.S.; Rodriguez, A.D.; Taglialatela-Scafati, O.; Fusetani, N. Marine pharmacology in 2009–2011: Marine Compounds with Antibacterial, Antidiabetic, Antifungal, Anti-inflammatory, Antiprotozoal, Antituberculosis and Antiviral Activities; affecting the Immune and Nervous System, and other Miscellaneous Mechanisms of Action. *Mar. Drugs* **2013**, *11*, 2510–2573. [PubMed]

3. Sawadogo, W.R.; Boly, R.; Cerella, C.; Teiten, M.H.; Dicato, M.; Diederich, M. A Survey of Marine Natural Compounds and Their Derivatives with Anti-cancer Activity Reported in 2012. *Molecules* **2015**, *20*, 7097–7142. [CrossRef] [PubMed]

4. Carbone, M.; Li, Y.; Irace, C.; Mollo, E.; Castelluccio, F.; Di Pascale, A.; Cimino, G.; Santamaria, R.; Guo, Y.W.; Gavagnin, M. Structure and cytotoxicity of phidianidines A and B: First finding of 1,2,4-oxadiazole system in a marine natural product. *Org. Lett.* **2011**, *13*, 2516–2519. [CrossRef] [PubMed]

5. Vitale, R.M.; Gatti, M.; Carbone, M.; Barbieri, F.; Felicità, V.; Gavagnin, M.; Florio, T.; Amodeo, P. Minimalist hybrid ligand/receptor-based pharmacophore model for CXCR4 applied to a small-library of marine natural products led to the identification of phidianidine a as a new CXCR4 ligand exhibiting antagonist activity. *ACS Chem. Biol.* **2013**, *8*, 2762–2770. [CrossRef] [PubMed]

6. Lin, H.Y.; Snider, B.B. Synthesis of phidianidines A and B. *J. Org. Chem.* **2012**, *77*, 4832–4836. [CrossRef] [PubMed]

7. Brogan, J.T.; Stoops, S.L.; Lindsley, C.W. Total synthesis and biological evaluation of phidianidines A and B uncovers unique pharmacological profiles at CNS targets. *ACS Chem. Neurosci.* **2012**, *3*, 658–664. [CrossRef] [PubMed]

8. Manzo, E.; Pagano, D.; Carbone, M.; Ciavatta, M.L.; Gavagnin, M. Synthesis of phidianidine B, a highly cytotoxic 1,2,4-oxadiazole marine metabolite. *ARKIVOC* **2012**, 220–228.

9. Buchanan, J.C.; Petersen, B.P.; Chamberland, S. Concise total synthesis of phidianidine A and B. *Tetrahedron Lett.* **2013**, *54*, 6002–6004. [CrossRef]

10. Jiang, C.S.; Fu, Y.; Zhang, L.; Gong, J.X.; Wang, Z.Z.; Xiao, W.; Zhang, H.Y.; Guo, Y.W. Synthesis and biological evaluation of novel marine-derived indole-based 1,2,4-oxadiazoles derivatives as multifunctional neuroprotective agents. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 216–220. [CrossRef] [PubMed]

11. Zhang, L.; Jiang, C.S.; Gao, L.X.; Gong, J.X.; Wang, Z.H.; Li, J.Y.; Li, J.; Li, X.W.; Guo, Y.W. Design, synthesis and in vitro activity of phidianidine B derivatives as novel PTP1B inhibitors with specific selectivity. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 778–781. [CrossRef] [PubMed]

12. Jiang, C.S.; Müller, W.E.; Schröder, H.C.; Guo, Y.W. Disulfide- and multisulfide-containing metabolites from marine organisms. *Chem. Rev.* **2012**, *112*, 2179–2207. [CrossRef] [PubMed]

13. Sun, Y.Q.; Guo, Y.W. Gymnorrhizol, an unusual macrocyclic polydisulfide from the Chinese mangrove *Bruguiera gymnorrhiza*. *Tetrahedron Lett.* **2004**, *45*, 5533–5535. [CrossRef]

14. Lui, H.L.; Shen, X.; Jiang, H.L.; Guo, Y.W. Structural Studies on an Unusual Novel Macrocyclic Polydisulfide from the Chinese Mangrove *Bruguiera gymnorrhiza*. *Chin. J. Org. Chem.* **2008**, *28*, 246–251.

15. Huang, X.Y.; Wang, Q.; Liu, H.L.; Zhang, Y.; Xin, G.R.; Shen, X.; Dong, M.L.; Guo, Y.W. Diastereoisomeric macrocyclic polydisulfides from the mangrove *Bruguiera gymnorrhiza*. *Phytochemistry* **2009**, *70*, 2096–2100. [CrossRef] [PubMed]

16. Gong, J.X.; Shen, X.; Yao, L.G.; Jiang, H.; Krohn, K.; Guo, Y.W. Total synthesis of gymnorrhizol, an unprecedented 15-membered macrocyclic polydisulfide from the Chinese mangrove *Bruguiera gymnorrhiza*. *Org. Lett.* **2007**, *9*, 1715–1716. [CrossRef] [PubMed]

17. Homhual, S.; Zhang, H.J.; Bunyapraphatsara, N.; Kondratyuk, T.P.; Santarsiero, B.D.; Mesecar, A.D.; Herunsalee, A.; Chaukul, W.; Pezzuto, J.M.; Fong, H.H. Bruguiesulfurol, a new sulfur compound from *Bruguiera gymnorrhiza*. *Planta Med.* **2006**, *72*, 255–260. [CrossRef] [PubMed]

18. Chen, J.; Jiang, C.S.; Ma, W.Q.; Gao, L.X.; Gong, J.X.; Li, J.Y.; Li, J.; Guo, Y.W. The first synthesis of natural disulfide bruguiesulfurol and biological evaluation of its derivatives as a novel scaffold for PTP1B inhibitors. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 5061–5065. [CrossRef] [PubMed]

19. Iversen, L.F.; Moller, K.B.; Pedersen, A.K.; Peters, G.H.; Petersen, A.S.; Andersen, H.S.; Branner, S.; Mortensen, S.B.; Moller, N.P. Structure determination of T cell protein-tyrosine phosphatase. *J. Biol. Chem.* **2002**, *277*, 19982–19990. [CrossRef] [PubMed]

20. Chen, J.; Gao, L.X.; Gong, J.X.; Jiang, C.S.; Yao, L.G.; Li, J.Y.; Li, J.; Xiao, W.; Guo, Y.W. Design and synthesis of novel 1,2-dithiolan-4-yl benzoate derivatives as PTP1B inhibitors. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 2211–2216. [CrossRef] [PubMed]

21. Chen, X.L.; Liu, H.L.; Li, J.; Xin, G.R.; Guo, Y.W. Paracaseolide A, first α-alkylbutenolide dimer with an unusual tetraquinane oxa-cage bislactone skeleton from Chinese mangrove *Sonneratia paracaseolaris*. *Org. Lett.* **2011**, *13*, 5032–5035. [CrossRef] [PubMed]

22. Noutsias, D.; Vassilikogiannakis, G. First total synthesis of paracaseolide A. *Org. Lett.* **2012**, *14*, 3565–3567. [CrossRef] [PubMed]

23. Guney, T.; Kraus, G.A. Total synthesis of paracaseolide A. *Org. Lett.* **2013**, *15*, 613–615. [CrossRef] [PubMed]

24. Vasamsetty, L.; Khan, F.A.; Mehta, G. Total synthesis of a novel oxa-bowl natural product paracaseolide A via a "putative" biomimetic pathway. *Tetrahedron Lett.* **2013**, *54*, 3522–3525. [CrossRef]

25. Giera, D.S.; Stark, C.B.W. Total synthesis of (±)-paracaseolide A and initial attempts at a Lewis acid mediated dimerization of its putative biosynthetic precursor. *RCS Adv.* **2013**, *3*, 21280–21284. [CrossRef]

26. Boukouvalas, J.; Jean, M.A. Streamlined biomimetic synthesis of paracaseolide A via aerobic oxidation of a 2-silyloxyfuran. *Tetrahedron Lett.* **2014**, *55*, 4248–4250. [CrossRef]

27. Wang, T.; Hoye, T.R. Diels-Alderase-free, bis-pericyclic, [4 + 2] dimerization in the biosynthesis of (±)-paracaseolide A. *Nat. Chem.* **2015**, *7*, 641–645. [CrossRef] [PubMed]

28. Yin, J.P.; Tang, C.L.; Gao, L.X.; Ma, W.P.; Li, J.Y.; Li, Y.; Li, J.; Nan, F.J. Design and synthesis of paracaseolide A analogues as selective protein tyrosine phosphatase 1B inhibitors. *Org. Biomol. Chem.* **2014**, *12*, 3441–3445. [CrossRef] [PubMed]

29. Xu, J.; Jiang, C.S.; Zhang, Z.L.; Ma, W.Q.; Guo, Y.W. Recent progress regarding the bioactivities, biosynthesis and synthesis of naturally occurring resorcinolic macrolides. *Acta Pharmacol. Sin.* **2014**, *35*, 316–330. [CrossRef] [PubMed]

30. Jiang, C.S.; Zhou, R.; Gong, J.X.; Chen, L.L.; Kurtán, T.; Shen, X.; Guo, Y.W. Synthesis, modification, and evaluation of (*R*)-de-*O*-methyllasiodiplodin and analogs as nonsteroidal antagonists of mineralocorticoid receptor. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 1171–1175. [CrossRef] [PubMed]

31. Solladié, G.; Rubio, A.; Carreño, M.C.; Ruano, J.L.G. Asymmetric synthesis of orsellinic acid type macrolides: The example of lasiodiplodin. *Tetrahedron Asymmetry* **1990**, *1*, 187–198. [CrossRef]

32. Fürstner, A.; Kindler, N. Macrocycle formation by ring-closing-metathesis. 2. An efficient synthesis of enantiomerically pure (*R*)-(+)-lasiodiplodin. *Tetrahedron Lett.* **1996**, *37*, 7005–7008.

33. Guo, Y.W.; Wang, H.Y.; Gong, J.X.; Zhang, X.D.; Jiang, C.S. A Macrolide Fused with Benzene Derivative with Pancreatic Inhibitory Activity. Patent CN102101854A, 18 December 2009.

34. Yan, X.H.; Lin, L.P.; Ding, J.; Guo, Y.W. Methyl spongoate, a cytotoxic steroid from the Sanya soft coral *Spongodes* sp. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2661–2663. [CrossRef] [PubMed]

35. Gong, J.X.; Miao, Z.H.; Yao, L.G.; Ding, J.; Kurtán, T.; Guo, Y.W. Stereoselective synthesis of methyl spongoate, a new steroid with potent antitumor activities. *Synlett* **2010**, *3*, 480–482.

36. Jiang, C.S.; Huang, C.G.; Feng, B.; Li, J.; Gong, J.X.; Kurtán, T.; Guo, Y.W. Synthesis and antitumor evaluation of methyl spongoate analogs. *Steroids* **2010**, *75*, 1153–1163. [CrossRef] [PubMed]

37. Zhou, Z.F.; Menna, M.; Cai, Y.S.; Guo, Y.W. Polyacetylenes of marine origin: Chemistry and bioactivity. *Chem. Rev.* **2015**, *115*, 1543–1596. [CrossRef] [PubMed]

38. Liang, L.F.; Wang, T.; Cai, Y.S.; He, W.F.; Sun, P.; Li, Y.F.; Huang, Q.; Taglialatela-Scafati, O.; Wang, H.Y.; Guo, Y.W. Brominated polyunsaturated lipids from the Chinese sponge *Xestospongia testudinaria* as a new class of pancreatic lipase inhibitors. *Eur. J. Med. Chem.* **2014**, *79*, 290–297. [CrossRef] [PubMed]

39. Gong, J.X.; Wang, H.Y.; He, W.F.; Wang, Z.Z.; Xiao, W.; Guo, Y.W. A concise synthesis of xestospongic acid methyl ester with pancreatic lipase inhibitory activity. *J. Asian Nat. Prod. Res.* **2013**, *15*, 916–919. [CrossRef] [PubMed]

40. Gong, J.X.; He, W.F.; Liu, H.L.; Jiang, C.S.; Wang, T.; Wang, H.Y.; Guo, Y.W. synthesis and evaluation of pancreatic lipase inhibitory halogenated polyunsaturated lipids from marine natural products: Methyl xestospongoate and analogs. *Helv. Chim. Acta* **2016**, *99*, 78–82. [CrossRef]

41. Wang, J.Y.; Pearce, N.; Chan, S.T.S.; Taylor, R.B.; Page, M.J.; Valentin, A.; Bouguet-Kondracki, M.-L.; Dalton, J.P.; Wiles, S.; Copp, B.R. Biologically Active Acetylenic Amino Alcohol and *N*-Hydroxylated 1,2,3,4-Tetrahydro-β-carboline Constituents of the New Zealand Ascidian *Pseudodistoma opacum*. *J. Nat. Prod.* **2016**, *79*, 607–610. [CrossRef] [PubMed]

42. Sun, D.Y.; Han, G.Y.; Gong, J.X.; Nay, B.; Li, X.W.; Guo, Y.W. Asymmetric total synthesis of distaminolyne A and revision of its absolute configuration. *Org. Lett.* **2017**, *19*, 714–717. [CrossRef] [PubMed]