

) Taylor & Francis

OPEN ACCESS Check for updates

An IncP-2 plasmid sublineage associated with dissemination of bla_{IMP-45} among carbapenem-resistant Pseudomonas aeruginosa

Xuefei Zhang^a, Leilei Wang^a, Dan Li^a, Pei Li^a, Lili Yuan^a, Fan Yang^{a,b}, Qinglan Guo^a* and Minggui Wang ^{Da}*

^aInstitute of Antibiotics, Huashan Hospital, Fudan University, Shanghai, People's Republic of China; ^bInfection Control Unit, Huashan Hospital, Fudan University, Shanghai, People's Republic of China

ABSTRACT

IMP-45, a variant of IMP-9, is one of the dominant metallo-β-lactamases (MBLs) in clinical carbapenem-resistant Pseudomonas aeruginosa (CRPA) isolates in China. The aim of this study was to investigate the distribution and mechanism of dissemination of bla_{IMP-45}. MBL genes were detected by PCR in 173 non-duplicate CRPA isolates collected from Hospital HS in Shanghai and 605 P. aeruginosa isolates from a multicenter surveillance of bla_{IMP-45} in China. In total, 17 IMP-45-producers (14 from Hospital HS and 3 from other hospitals) were identified. Molecular typing identified an outbreak of 11 IMP-45-producing ST508 CRPA in the ICU of Hospital HS. Conjugation assays and whole genome sequencing were conducted among IMP-45-producers. Genomic comparison revealed that 16 bla_{IMP-} 45-carrying plasmids (9 from this study and 7 from GenBank) shared a similar backbone with IncP-2 bla_{IMP-9}-carrying plasmid pOZ176 but lacked repA-oriV-parAB region. repA2 gene was presented in pOZ176, bla_{IMP-45}-carrying plasmids (17 from this study and 7 from GenBank) and 15 megaplasmids from GenBank. Phylogenetic analysis of repA2 showed that most bla_{IMP-45}-carrying plasmids were clustered into a sublineage separate from the one containing pOZ176. This IncP-2 plasmid sublineage contributed to the dissemination of bla_{IMP-45} among genetically diverse P. aeruginosa and recruited multiple resistance genes during its evolution.

ARTICLE HISTORY Received 4 December 2020; Revised 18 February 2021; Accepted 21 February 2021

KEYWORDS Carbapenem-resistant Pseudomonas aeruginosa; outbreak; blaIMP-45; InCP-2 plasmid; sublineage; dissemination

Introduction

Pseudomonas aeruginosa is prone to be resistant to β lactams, aminoglycosides and quinolones. Production of metallo- β -lactamases (MBLs) is one of the primary carbapenem resistance mechanisms in this species, among which IMP and VIM are the most prevalent [1]. IMP-1, IMP-4, IMP-6, IMP-8, IMP-9, IMP-10 and IMP-45 have been reported in China [2]. IMP-9 was initially identified in P. aeruginosa isolates from Guangzhou, China [3]. Afterwards, outbreaks of IMP-9-producing P. aeruginosa were observed in this area in 2000 and from 2005 to 2007 [3,4]. IMP-45, a single amino acid substitution variant (G214S) of IMP-9, was first reported in a canine-origin P. aeruginosa from Beijing, China in 2014, showing higher level resistance to meropenem than to imipenem [5]. Afterwards, more clinical isolates of IMP-45-producing Pseudomonas were discovered [6-8], including one isolated from a French patient who had been repatriated from an ICU in Shanghai, China [8], implying the risk of worldwide spread of *bla*_{IMP-45} by such dissemination.

IMP-9-encoding plasmid pOZ176 (500 kb), the only full sequenced IncP-2 plasmid before 2013, contained bla_{IMP-9} , bla_{OXA-10} and aacA4 genes conferring β -lactam and aminoglycoside resistance. Two replication genes, repA and repA2 were identified in pOZ176 [3,9]. IncP-2 plasmids, generally >300 kb and in single copy, exhibit tellurite resistance and are narrow-host-range for Pseudomonas spp [10]. Previous studies have characterized two *bla*_{IMP-45}-carrying megaplasmids from clinical isolates (P. putida and P. aeruginosa) in China [6,7], but information about the dissemination of *bla*_{IMP-45} gene remains largely limited and the role of plasmids in the dissemination of *bla*_{IMP-45} is poorly understood.

In this study, we report an outbreak of carbapenem-resistant P. aeruginosa (CRPA) co-carrying *bla*_{IMP-45}, *qnrVC1* and *armA* in a tertiary hospital of Shanghai and the disappearance of outbreak clones after strengthened infection control measures. Subsequently, we carried out a multicenter surveillance of *bla*_{IMP-45} in *P. aeruginosa* clinical isolates in China and explored the role of IncP-2 plasmids in the dissemination of *bla*_{IMP-45} among *P. aeruginosa*.

CONTACT 🛛 Qinglan Guo 🖾 qinglanguo@fudan.edu.cn Minggui Wang 🖾 mgwang@fudan.edu.cn 🖻 Institute of Antibiotics, Huashan Hospital, Fudan University, 12 M. Wulumuqi Road, Shanghai 200040, People's Republic of China *These authors contributed equally to this study.

Supplemental data for this article can be accessed at https://doi.org/10.1080/22221751.2021.1894903

^{© 2021} The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group, on behalf of Shanghai Shangyixun Cultural Communication Co., Ltd

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Clinical isolates and antimicrobial susceptibility testing

One hundred and seventy-three non-duplicate CRPA isolates were collected from a tertiary hospital (Hospital HS) in Shanghai between January 2015 and April 2018. CRPA was defined as *P. aeruginosa isolate* resistant to either imipenem or meropenem. Additionally, a multicenter surveillance was performed with 605 non-duplicate *P. aeruginosa* isolates collected consecutively from 11 hospitals in 8 provinces/municipalities across China, including 3 hospitals in Shanghai, 2 in Beijing and 1 in each of the other 6 provinces (July, 2018 to February, 2019, Supplementary Table S1).

Minimal inhibitory concentrations (MICs) were determined for 13 antimicrobial agents by broth microdilution method and interpretation was according to recommendations of the CLSI [11].

MBLs screening and identification

PCR amplification was performed to screen for MBL genes (bla_{IMP} , bla_{VIM} , bla_{NDM} , bla_{SPM} , bla_{SIM} , bla_{GIM} , bla_{DIM} , bla_{AIM} and bla_{FIM}) [12,13]. bla_{IMP} -positive isolates were further amplified with primers specific for various subtypes (Supplementary Table S2).

Molecular typing of P. aeruginosa isolates

Pulsed-field gel electrophoresis (PFGE) was performed using SpeI (TaKaRa Bio, Dalian, China) as the restriction enzyme and with a switch time of 2 s-40 s [14,15]. The PFGE patterns were analysed by Bio-Numerics (version 4.0; Applied Maths, Inc.) and a dendrogram was generated by the UPGMA method based on the Dice coefficient.

Multilocus sequence typing (MLST) was performed according to the instructions in the *P. aeruginosa* MLST website (http://pubmlst.org/paeruginosa/). STs were compared with those in the MLST *P. aeruginosa* database by goeBURST [16]. A neighbor-joining tree from concatenated seven housekeeping genes (*acsA*, *aroE*, *guaA*, *mutL*, *nuoD*, *ppsA*, and *trpE*) was generated using MEGA 7.0 [17].

Conjugation assay

Transfer of plasmid carrying bla_{IMP-45} was performed by filter mating with *P. aeruginosa* PAO1^{Rif} (rifampin resistant) as recipient. Transconjugants were selected on LB plates supplemented with meropenem (2 mg/ L) and rifampin (500 mg/L), and further confirmed by PCR amplification and sequencing of bla_{IMP-45} and *guaA*, one of the seven housekeeping genes for MLST of *P. aeruginosa*. The transconjugants harbouring bla_{IMP-45} were tested for antimicrobial susceptibility. Conjugation was also carried out using *Escherichia coli* J53 (azide resistant) as recipient.

Whole genome sequencing (WGS) and sequence assembly

Sequencing of 9 IMP-45-producers, including 6 *P. aeruginosa* clinical isolates (HS15-106, HS17-127, HS18-41, GZ18-2, KM18-18 and RJ19-28) and 3 transconjugants (TcHS15-101, TcHS15-158 and TcHS15-172) were performed on Hiseq X-ten or Novaseq platforms (Illumina Inc., San Diego, CA, USA). De novo assembly was performed using Velvet version 1.2.03 [18] or SOAPdenovo2 [19]. To obtain the complete genomic sequence, clinical isolate of *P. aeruginosa* HS17-127 was further subjected to Pacbio sequencing and assembled with HGAP [20].

For the 8 transconjugants of outbreak isolates without WGS data, the *repA2* gene and genetic context surrounding bla_{IMP-45} were amplified by PCR and sequenced with a series of primers designed according to the genome sequence of the transconjugant of *P. aeruginosa* outbreak isolate HS15-101 (Table S2).

Bioinformatics analysis

The contigs were annotated using RAST (http://rast. nmpdr.org/), screened for insertion sequences with ISfinder [21], and analysed for STs, antibiotic resistance genes and plasmid typing at the Centre for Genomic Epidemiology web site (http://www.genomicepi demiology.org/). BLASTN searches were conducted to find the complete sequenced *bla*_{IMP-45}-harbouring plasmids in GenBank database using *bla*_{IMP-45} as the query sequence. BLAST Ring Image Generator (BRIG) [22] were used in the comparative analysis of the plasmids.

Nucleotide sequence accession numbers

The sequences of transconjugants TcHS15-101, TcHS15-158 and TcHS15-172, and *P. aeruginosa* clinical isolates HS15-106, HS17-127, HS18-41, GZ18-2, KM18-18 and RJ19-28 were submitted to GenBank with Bioproject ID PRJNA631492 (Supplementary Table S3). The accession numbers for the chromosome of *P. aeruginosa* clinical isolate HS17-127 and plasmid pHS17-127 are CP061376 and CP061377, respectively.

Results

Outbreak of bla_{IMP-45}-bearing P. aeruginosa in Hospital HS

Fourteen out of 173 CRPA from Hospital HS were positive for bla_{IMP-45} : 12 CRPA isolated from 2015, 1 from 2017 and 1 from 2018. All of them exhibited

resistance to antipseudomonal β -lactams excluding aztreonam, β -lactamase inhibitor combinations and aminoglycosides. The MICs of aztreonam ranged from 4 to 16 mg/L in 7 strains whereas the remaining strains were highly resistant to aztreonam (from 64 and >128 mg/L). All the *bla*_{IMP-45}-carriers were resistant to quinolones except isolate HS17-127 (Table 1).

Eleven out of 12 bla_{IMP-45} -bearing CRPA from 2015 were isolated from ICU patients and shared the same sequence type, ST508 (Figure 1(A)). They presented indistinguishable or closely related PFGE patterns, different from that of HS15-106 (ST3014) from an outpatient. All the 12 bla_{IMP-45} -bearing CRPA from 2015 were discovered in the first nine months this year as indicated by the timeline of the isolation date of the first bla_{IMP-45} -carrying *P. aeruginosa* from each patient (Figure 1(B)). Taken together, an outbreak of bla_{IMP-45} -bearing ST508 CRPA occurred in the ICU of Hospital HS in 2015. The other two bla_{IMP-45} -bearing CRPA, HS17-127 (ST369) and HS18-41 (ST357), were clonally distinct from the previous outbreak bla_{IMP-45} -carriers in Hospital HS.

Multicenter surveillance of bla_{IMP-45}

To investigate the prevalence of bla_{IMP-45} , a multicenter surveillance was carried out among 605 *P. aeruginosa* clinical isolates collected throughout China, including 226 CRPA. Genotypic characterization found 8 isolates positive for bla_{IMP} (4 isolates positive for bla_{IMP-45} and 1 for bla_{IMP-3}) and 6 isolates carrying bla_{VIM-2} . No additional MBL-producing isolates were found. These 3 bla_{IMP-45} -carriers belonged to ST1420, ST274 and ST708, respectively (Table 1).

Transferability of bla_{IMP-45}

To examine the transferability of bla_{IMP-45} , all the 17 bla_{IMP-45} -carrying CRPA were performed conjugation with *P. aeruginosa* PAO1^{Rif} as recipients. Fifteen transconjugants harbouring bla_{IMP-45} were obtained from the 11 ST508 outbreak strains and HS17-127, HS18-41 from Hospital HS as well as KM18-18 and RJ19-28 from another two hospitals. All the transconjugants contained the same *guaA* allele with the recipient *P. aeruginosa* PAO1^{Rif}. Transconjugants displayed similar antimicrobial susceptibility profiles with their donors except that 3 transconjugants (TcHS15-158, TcHS15-172 and TcHS15-209) were susceptible to quinolones (Supplementary Table S4). However, transfer of bla_{IMP-45} to *E. coli* failed.

General features of bla_{IMP-45}-harbouring plasmids

P. aeruginosa clinical isolate HS17-127 was fully sequenced resulting in one chromosome and one

														gnrVC6					fampin.
	Resistance gene	bla _{iMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , bla _{PER-1} , bla _{AEM-1} , armA, gnrVC6	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	bla _{IMP-45} , bla _{OXA-1} , bla _{AFM-1} , qnrVC6	P: cefepime; ATM: aztreonam; IPM: imipenem; MEM: meropenem; CIP: ciprofloxacin; LEV: levofloxacin; AMK: amikacin; CT: colistin; RIF: rifampin
	RIF	256	256	384	512	512	384	512	512	512	256	512	512	32	384	512	384	16	n; LEV: le
	Ե	-	0.5	0.5	0.5	-	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	-	0.5	-	-	ofloxaci
	AMK	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	16	>128	>128	>128	CIP: cipro
	LEV	16	∞	16	64	64	16	64	64	64	16	16	64	2	∞	32	128	16	penem;
	CIP	8	4	∞	16	16	∞	16	16	16	∞	∞	16	-	4	16	64	16	A: merol
MICs (mg/L) ^b	MEM	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	128	>128	>128	8	>128	iem; MEN
	ЫМ	32	32	32	32	64	32	64	64	64	32	64	32	128	32	64	2	>128	A: imiper
	ATM	8	8	8	64	32	8	>128	>128	>128	8	16	64	64	4	4	16	>128	onam; IPN
	FEP	>128	128	>128	>128	>128	128	>128	>128	>128	>128	>128	>128	>128	>128	128	128	>128	TM: aztre
	CZA	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	fepime; A
	CAZ	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	>128	n; FEP: ce
	TZP	128	64	128	128	128	128	>128	>128	>128	128	128	>128	128	128	128	64	128	'avibactar
	PIP	128	64	128	128	128	128	>128	>128	>128	128	128	>128	128	128	128	64	>128	ftazidime,
	Sequence type	ST508	ST3014	ST508	ST369	ST357	ST274	ST1420	ST708	center surveillance. ceftazidime; CZA: ce									
	Transconjugant	TcHS15-101	Failed	TcHS15-109	TcHS15-110	TcHS15-111	TcHS15-141	TcHS15-146	TcHS15-158	TcHS15-172	TcHS15-176	TcHS15-209	TcHS15-215	TcHS17-127	TcHS18-41	TcKM18-18	Failed	TcRJ19-28	bla _{lmP 45} -carrying <i>P. aeruginosa</i> isolates from multicenter surveillance. PIP: piperacillin; TZP: piperacillin/tazobactam; CAZ: ceftazidime; CZA: ceftazidime/avibactam; FE
	Source	Urine	Sputum	urine	sputum	sputum	sputum	sputum	wound fluid	ing <i>P. aeruginosa</i> n; TZP: piperacilli									
	Clinical isolate	HS15-101	HS15-106	HS15-109	HS15-110	HS15-111	HS15-141	HS15-146	HS15-158	HS15-172	HS15-176	HS15-209	HS15-215	HS17-127	HS18-41	KM18-18 ^a	GZ18-2 ^a	RJ 19-28 ^a	^a bla _{IMP-45} -carryi ^b PIP: piperacilli

Table 1. Characteristics of 17 *Pseudomonas aeruginosa* clinical isolates carrying *bla*_{IMP-45}

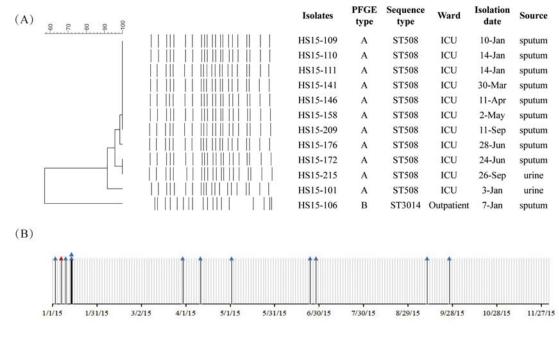


Figure 1. Twelve carbapenem-resistant *Pseudomonas aeruginosa* (CRPA) isolates carrying bla_{IMP-45} at Hospital HS in 2015. (A) PFGE of 12 bla_{IMP-45} -bearing CRPA isolates. ST508 isolates belonged to pattern A and HS15-106 typed as pattern B. (B) Timeline of the first positive cultures of the bla_{IMP-45} -bearing CRPA isolates for the 12 patients. The blue triangle indicates the isolation date of the bla_{IMP-45} -carrying *P. aeruginosa* from ICU patients and the red indicates that of the outpatient.

plasmid pHS17-127. The sequencing analysis revealed that repA2 and bla_{IMP-45} genes were on the plasmid pHS17-127 (486,963 bp), which shared a highly similar backbone with IncP-2 plasmid pOZ176 (Figure 2).

Whole genome sequences of 3 transconjugants and 5 clinical isolates in this study (Supplementary Table S5) were compared to the fully sequenced plasmid pHS17-127 and the 7 completely sequenced bla_{IMP}. 45-harbouring plasmids in GenBank (up to April, 2020). The 7 plasmids, varying in size from 374 kb to nearly 514 kb, were isolated in China and from P. aeruginosa except for 1 from P. putida [6] (Table 2). Comparative genome analysis revealed that these 16 bla_{IMP-45}-carrying plasmids (Table 2) shared an overall similar backbone, including genes essential for replication (repA2, 1188 bp), partition (par) and conjugal transfer (tra) (Figure 2). Moreover, they contained an operon terABCDEZ conferring tellurite resistance, which is a uniform property of IncP-2 plasmids. Virulence factors, such as pilus biogenesis gene pilD and chemotaxis gene cluster, cheABRWXZ, were also identified in the backbones.

An IncP-2 plasmid sublineage associated with dissemination of bla_{IMP-45}

In order to explore the mechanism underlying dissemination of bla_{IMP-45} , 16 bla_{IMP-45} -carrying plasmids were compared with IncP-2 bla_{IMP-9} -harbouring plasmid pOZ176. Comparative analysis revealed that they shared similar plasmid backbones (Figure 2). However, the IncP-2 *repA-oriV-parAB* region of pOZ176 was absent in all *bla*_{IMP-45}-harbouring plasmids except that pHS18-41 and pGZ18-2 contained a region with only about 84% identity. On the contrary, a second replication/partitioning system *repA2/parAB-parB2* was shared by both *bla*_{IMP-9}- and *bla*_{IMP-45}-harbouring plasmids with an identity of >98%.

When BLASTN with the repA2 gene of pOZ176 was performed for its homologs (100% query coverage), a total of 23 fully sequenced megaplasmids were identified in NCBI database, including pOZ176 and the above 7 *bla*_{IMP-45}-harbouring plasmids (Figure 3). They shared a similar backbone with pOZ176 as well as the *bla*_{IMP-45}-carrying plasmids in this study, even though these megaplasmids were absent of a large fragment containing repA-oriV-parAB of pOZ176 (Figure 2). Additionally, repA2 genes were confirmed by PCR and sequencing in the remaining 8 transconjugants of outbreak strains without WGS data in this study (Supplementary Table S2). Phylogenetic analysis of the 40 repA2 genes from 17 bla_{IMP-45}carrying CRPA in this study and 23 megaplasmids in GenBank revealed 4 distinct subgroups (Figure 3). All the *bla*_{IMP-45}-carrying plasmids with the exception of RJ19-28 were in a sublineage separate from the one containing *bla*_{IMP-9}-harbouring pOZ176. IncP-2 plasmids and other plasmid lineages, such as IncP-7 and IncP-9, were phylogenetically analysed on the basis of replication genes, revealing that IncP-2 plasmid lineage was clearly seperated from other plasmid lineages (Figure S2).

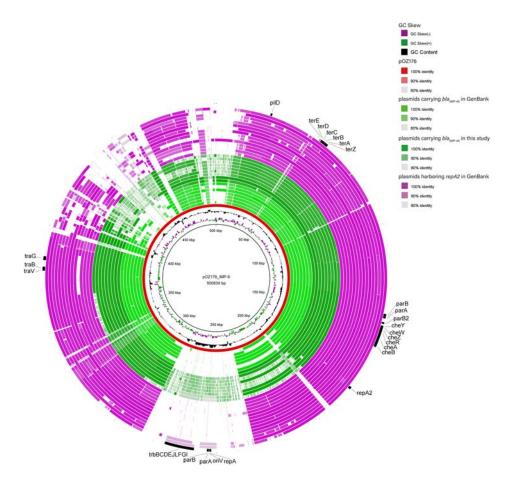


Figure 2. Genome comparison of plasmids containing a *repA2* gene with IncP-2 plasmid pOZ176. This map was constructed with BRIG. The various colour levels indicate a BLAST result with sequence identity ranging from 80% to 100%. The inner red circle represents the pOZ176 reference sequence. The light green, dark green and purple circles (from innermost to outermost) indicate genome sequence of 7 *bla*_{IMP-45}-carrying megaplasmids in GenBank (p727-IMP, pA681-IMP, pBM413, pBM908, pPAG5, pR31014-IMP and pSY153-MDR), 9 *bla*_{IMP-45}-carrying plasmids in this study (pHS17-127, pRJ19-28, pHS15-101, pHS15-106, pHS15-158, pHS15-172, pKM18-18, pHS18-41 and pGZ18-2) and 16 completely sequenced megaplasmids harbouring *repA2* in GenBank (pBT2101, pPABL048, AR441_unnamed3, AR_0356_unnamed2, RW109 plasmid 1, p12939-PER, pCF39S, pNK546-KPC, AR439_unnamed2, pRBL16, p1, pBT2436, p12969-DIM, pJB37 and pTTS12), respectively Backbone genes of pOZ176 are indicated in the figure.

Coexistence of bla_{IMP-45}, arma and qnrVC1 or qnrVC6

Apart from 1 plasmid carrying only bla_{IMP-45} and 1 with both bla_{IMP-45} and qnrVC6, 14 out of 16 plasmids co-carried bla_{IMP-45} and armA, including 9 together with qnrVC1 and 3 with qnrVC6 (Table 2). The genetic structures harbouring these resistant determinants were confirmed by WGS analysis or PCR and sequencing (Supplementary Table S2). As in the previously reported IMP-45 producers [5,8], bla_{IMP-45} was located in the variable region of In786, adjacent to a Tn1548derivative containing armA with or without qnrVC6(Figure S1). qnrVC1 was in class 1 integron In1237 that coexisted with the Tn1548-derivative on the megaplasmids. However, In1237 was not transferred to the recipient strain together with bla_{IMP-45} -harbouring plasmids from HS15-158, HS15-172 and HS15-209.

Discussion

In this study, 17 IMP-45-producers were discovered and belonged to 7 dissimilar STs. All these STs were

different from previously reported IMP-45-producers, such as ST308, ST235 and ST389 [5,7,8], demonstrating the diverse population structure of IMP-45-producing *P. aeruginosa*.

Outbreaks of MBL-producing P. aeruginosa have been reported in hospitals worldwide [23-25]. Recently, an outbreak of IMP-19-producing ST235 and IMP-29-producing ST111 of clinical P. aeruginosa was reported in France [26]. Here we report an outbreak caused by IMP-45-producing ST508 CRPA isolates in Hospital HS from January to September, 2015. Since around September, 2015, strengthened infection control measures were implemented in the ICU, including improved hospital-wide sanitation, hand and environmental hygiene, contact precautions, changing disinfection to sterilization for reusable ventilator accessories (exhalation valve and respiratory humidifier) and using disposable ventilator circuits instead of recycled ones. The outbreak clone subsequently disappeared and distinct clonal complex lineages carrying *bla*_{IMP-45} emerged sporadically in 2017 and 2018, suggesting a shift of

Table 2. Characteristics of 16 *bla*_{IMP-45}-carrying plasmids (9 in this study and 7 in GenBank).

Plasmid	Host strain	ST	Location	Resistance gene ^c	Size (bp) ^d	Resource	Accession number ^e
pHS15-101	P. aeruginosa HS15- 101	ST508	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	ND	This study	Ν
pHS15-106	P. aeruginosa HS15- 106	ST3014	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	ND	This study	Ν
oHS15-158	P. aeruginosa HS15- 158	ST508	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , armA	ND	This study	Ν
oHS15-172	P. aeruginosa HS15- 172	ST508	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , armA	ND	This study	Ν
oHS17-127	P. aeruginosa HS17- 127	ST369	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , bla _{PER-1} , bla _{AFM-1} , armA, gnrVC6	486,963	This study	CP061377
oHS18-41	P. aeruginosa HS18- 41	ST357	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	ND	This study	Ν
oGZ18-2	P. aeruginosa GZ18-2	ST1420	Guangzhou, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	ND	This study	Ν
oKM18-18	P. aeruginosa KM18- 18	ST274	Kunming, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	ND	This study	Ν
oRJ19-28	P. aeruginosa RJ19- 28	ST708	Shanghai, China	bla _{IMP-45} , bla _{OXA-1} , bla _{AFM-1} , qnrVC6	ND	This study	Ν
oR31014- IMP	P. aeruginosa	unknown	China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	374,000	GenBank	MF344571
5727-IMP	P. aeruginosa	unknown	China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	430,173	GenBank	MF344568
oSY153- MDR	P. putida SY153	unknown	Hainan, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	468,170	GenBank	KY883660
DA681-IMP	P. aeruginosa	ST274	China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC6	397,519	GenBank	MF344570
BM908	P. aeruginosa PA298	ST277	China	bla _{IMP-45} , bla _{OXA-1}	395,774	GenBank	CP040126
PAG5	P. aeruginosa PAG5	unknown	China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	513,322	GenBank	CP045003
BM413	P. aeruginosa PA121617	ST389	Guangzhou, China	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC6	423,017	GenBank	CP016215
3	P. aeruginosa M140A	ST308	Beijing, China	bla _{IMP-45} , bla _{OXA-1}	NA	GenBank	KJ510410
b	P. aeruginosa 14.1819	ST235	France	bla _{IMP-45} , bla _{OXA-1} , armA, qnrVC1	NA	GenBank	KU984333

^abla_{IMP-45} was located on the chromosome.

^bDetailed information of plasmid carrying *bla*_{IMP-45} in *P. aeruginosa* 14.1819 was not available.

^cResistance genes referring to β -lactamase genes, *armA* and *qnrVC* here.

^dN: Putative plasmids in this study without complete sequence. NA: Not available.

^eN: Plasmids in this study without complete sequence.

the bla_{IMP-45} -carrying CRPA clones during the survey period.

Although at least 14 incompatible groups (IncP-1~IncP-14) of plasmids have been identified in Pseudomonas species, there is no well-established scheme for the plasmid typing of this species as of Enterobacteriaceae [27,28]. Similar to IncP-2 plasmids reported previously [9,10], the *bla*_{IMP-45}-carrying plasmids are tellurite resistant, conjugative, and transfer between P. aeruginosa and P. putida but not to E. coli. Plasmids usually undergo continuous rearrangement and mutations, sometimes occurring in regions for plasmid typing [29], resulting in novel untypeable plasmids or new plasmid lineages evolving from currently well-studied plasmid types. The incompatibility group of bla_{IMP-45}-carrying plasmids has not been fully clarified since they lack the region containing IncP-2 repA-oriV-parAB [9]. A previous study reported that IncP-2 repA-oriV-parAB, an auxiliary replicon, was located in an integrative and conjugative element Tn6398a [30]. Moreover, homologs of IncP-2 repA gene were seldom present in Pseudomonas spp. but more frequently found in putative genomic islands on the chromosome of a variety of other species, such as Azotobacter, Burkholderia, Stenotrophomonas and Xanthomonas [9].

In contrast, IncP-2 *repA2* gene and its close relatives (>96%) have been exclusively discovered on plasmids from *Pseudomonas spp.*, indicating that the *repA2* gene is probably the actual one encoding the IncP-2 replication initiator protein and contributing to the narrow-host-range of IncP-2 plasmids for *Pseudomonas spp*.

Based on the phylogenetic analysis of repA2 and the comparative genome analysis in this study, all the plasmids involved are closely related genetically, belonging to the same plasmid family as IncP-2 pOZ176. The phylogenetic tree of repA2 grouped almost all bla_{IMP-45} -carrying plasmids into a different subgroup from the one containing pOZ176.

In summary, clonal diversity was observed in the 17 IMP-45-producing CRPA isolates identified in this study except for outbreak clone ST508 from Hospital HS. All the bla_{IMP-45} -carrying plasmids were related to IncP-2 plasmid pOZ176, and contributing to the dissemination of bla_{IMP-45} . Moreover, this IncP-2 plasmid sublineage has undergone multiple evolutionary events, recruiting bla_{IMP-45} , *armA* and *qnrVC1/qnrVC6*, thus acting as a vehicle for the dissemination of carbapenem, aminoglycoside and quinolone resistance among *Pseudomonas spp.*, with consequent compromise of therapeutic options.

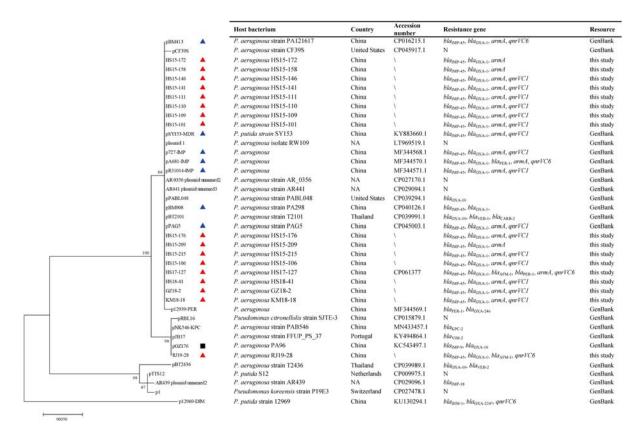


Figure 3. A maximum likelihood phylogenetic tree of 40 *repA2* genes and features of their hosts. Red solid triangle, blue solid triangle and black solid square indicate bla_{IMP-45} -carrying plasmids in this study, 7 bla_{IMP-45} -carrying megaplasmids in GenBank and IncP-2 plasmid pOZ176, respectively. Resistance genes refer to β -lactamase genes, *armA* and *qnrVC* here. "N" denotes that the plasmid harbours no β -lactamase genes, *armA* or *qnrVC*.

Acknowledgements

The authors are grateful to Dr. George A. Jacoby for his critical review of the manuscript.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by grants from the National Natural Science Foundation of China [grant numbers 81773785, 81991531, 81673479, 81872909], Shanghai Municipal Science and Technology Commission [grant number 18411950600].

ORCID

Minggui Wang D http://orcid.org/0000-0001-7682-5859

References

- Cornaglia G, Giamarellou H, Rossolini GM. Metalloβ-lactamases: a last frontier for β-lactams? Lancet Infect Dis. 2011 May;11(5):381–393.
- [2] Hong DJ, Bae IK, Jang IH, et al. Epidemiology and Characteristics of metallo-β-lactamase-producing *Pseudomonas aeruginosa*. Infect Chemother. 2015 Jun;47(2):81–97.

- [3] Xiong J, Hynes MF, Ye H, et al. bla(IMP-9) and its association with large plasmids carried by *Pseudomonas aeruginosa* isolates from the People's Republic of China. Antimicrob Agents Chemother. 2006 Jan;50(1):355–358.
- [4] Chao Z, Xiao-Feng W, Dan-Hong S, et al. Outbreak of *Pseudomonas aeruginosa* producing IMP-9-type metallo-beta-lactamase in guangzhou, China. Int J Antimicrob Agents. 2008 Oct;32(4):363–365.
- [5] Wang Y, Wang X, Schwarz S, et al. IMP-45-producing multidrug-resistant *Pseudomonas aeruginosa* of canine origin. J Antimicrob Chemother. 2014 Sep;69 (9):2579–2581.
- [6] Yuan M, Chen H, Zhu X, et al. pSY153-MDR, a p12969-DIM-related mega plasmid carrying bla (IMP-45) and armA, from clinical *Pseudomonas putida*. Oncotarget. 2017 Sep;8(40):68439–68447.
- [7] Liu J, Yang L, Chen D, et al. Complete sequence of pBM413, a novel multidrug resistance megaplasmid carrying qnrVC6 and bla(IMP-45) from *Pseudomonas aeruginosa*. Int J Antimicrob Agents. 2018 Jan;51(1):145–150.
- [8] Janvier F, Otto MP, Jove T, et al. A case of multiple contamination with methylase ArmA-producing pathogens. J Antimicrob Chemother. 2017 Feb;72(2):618–620.
- [9] Xiong J, Alexander DC, Ma JH, et al. Complete sequence of pOZ176, a 500-kilobase IncP-2 plasmid encoding IMP-9-mediated carbapenem resistance, from outbreak isolate *Pseudomonas aeruginosa* 96. Antimicrob Agents Chemother. 2013 Aug;57(8):3775–3782.
- [10] Jacoby GA, Sutton L, Knobel L, et al. Properties of IncP-2 plasmids of Pseudomonas spp. Antimicrob Agents Chemother. 1983 Aug;24(2):168–175.

- [11] Institute CaLS. Performance standards for antimicrobial susceptibility testing—twenty-seventh edition: M100. Wayne (PA): CLSI; 2017; 2020.
- [12] Pollini S, Maradei S, Pecile P, et al. FIM-1, a new acquired metallo- β -lactamase from a *Pseudomonas aeruginosa* clinical isolate from Italy. Antimicrob Agents Chemother. 2013 Jan;57(1):410–416.
- [13] Poirel L, Walsh TR, Cuvillier V, et al. Multiplex PCR for detection of acquired carbapenemase genes. Diagn Microbiol Infect Dis. 2011 May;70(1):119–123.
- [14] Ribot EM, Fair MA, Gautom R, et al. Standardization of pulsed-field gel electrophoresis protocols for the subtyping of Escherichia coli O157:H7, salmonella, and shigella for PulseNet. Foodborne Pathog Dis. 2006 Spring;3(1):59–67.
- [15] Selim S, El Kholy I, Hagagy N, et al. Rapid identification of *Pseudomonas aeruginosa* by pulsed-field gel electrophoresis. Biotechnol Biotechnol Equip. 2015 Jan;29(1):152–156.
- [16] Francisco AP, Bugalho M, Ramirez M, et al. Global optimal eBURST analysis of multilocus typing data using a graphic matroid approach. BMC Bioinformatics. 2009 May;10(152). DOI:10.1186/ 1471-2105-10-152.
- [17] Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol. 2016 Jul;33(7):1870–1874.
- [18] Zerbino DR, Birney E. Velvet: algorithms for de novo short read assembly using de bruijn graphs. Genome Res. 2008 May;18(5):821–829.
- [19] Luo R, Liu B, Xie Y, et al. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. Gigascience. 2012 Dec;1(1):18.
- [20] Chin CS, Alexander DH, Marks P, et al. Nonhybrid, finished microbial genome assemblies from longread SMRT sequencing data. Nat Methods. 2013 Jun;10(6):563–569.
- [21] Siguier P, Perochon J, Lestrade L, et al. ISfinder: the reference centre for bacterial insertion sequences.

Nucleic Acids Res. 2006 Jan;34(Database issue):D32–D36.

- [22] Alikhan NF, Petty NK, Ben Zakour NL, et al. BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. BMC Genomics. 2011 Aug;12 (402). DOI:10.1186/1471-2164-12-402.
- [23] Lee K, Lim JB, Yum JH, et al. . bla(VIM-2) cassettecontaining novel integrons in metallo-beta-lactamase-producing *Pseudomonas aeruginosa* and *Pseudomonas putida* isolates disseminated in a Korean hospital. Antimicrob Agents Chemother. 2002 Apr;46(4):1053–1058.
- [24] Hirakata Y, Yamaguchi T, Nakano M, et al. Clinical and bacteriological characteristics of IMP-type metallo-beta-lactamase-producing *Pseudomonas aeruginosa*. Clin Infect Dis. 2003 Jul;37(1):26–32.
- [25] Mavroidi A, Tsakris A, Tzelepi E, et al. Carbapenemhydrolysing VIM-2 metallo- beta-lactamase in *Pseudomonas aeruginosa* from Greece. J Antimicrob Chemother. 2000 Dec;46(6):1041–1042.
- [26] Gbaguidi-Haore H, Varin A, Cholley P, et al. A bundle of measures to control an Outbreak of *Pseudomonas aeruginosa* associated With P-trap contamination. Infect Control Hosp Epidemiol. 2018 Feb;39(2):164–169.
- [27] Boronin AM. Diversity of Pseudomonas plasmids: to what extent? FEMS Microbiol Lett. 1992 Dec;100(1-3):461-467.
- [28] Jacoby GA. Resistance plasmids of pseudomonas. In: Sokatch JR, editor. The bacteria, Vol. X. The biology of pseudomonas. Orlando, FL: Academic Press; 1986. p. 265–293.
- [29] Rozwandowicz M, Brouwer MSM, Fischer J, et al. Plasmids carrying antimicrobial resistance genes in enterobacteriaceae. J Antimicrob Chemother. 2018 May;73(5):1121–1137.
- [30] Jiang X, Yin Z, Yuan M, et al. Plasmids of novel incompatibility group IncpRBL16 from Pseudomonas species. J Antimicrob Chemother. 2020 Aug 1;75(8):2093–2100.