


# The Role of Circulating A $\beta$ Seeds in the Progression of Cerebral Amyloidosis

Nazaret Gamez<sup>1,2</sup> and Rodrigo Morales<sup>1,3</sup> 

<sup>1</sup>Department of Neurology, The University of Texas Health Science Center at Houston, Houston, TX, USA. <sup>2</sup>Dpto. Biología Celular, Genética y Fisiología, Instituto de Investigación Biomédica de Málaga-IBIMA, Facultad de Ciencias, Universidad de Málaga, Málaga, Spain. <sup>3</sup>Centro Integrativo de Biología y Química Aplicada (CIBQA), Universidad Bernardo O'Higgins, Santiago, Chile.

Neuroscience Insights  
Volume 17: 1–4  
© The Author(s) 2022  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/26331055221123072



**ABSTRACT:** While understudied, it is suspected that peripheral A $\beta$  peptides affect Alzheimer's disease (AD)-associated pathological changes in the brain. The peripheral sink hypothesis postulates that the central and peripheral pools of A $\beta$  co-exist in equilibrium. As such, cerebral amyloid levels may be modulated by intervening circulating A $\beta$ . In this commentary, we discuss relevant literature supporting the potential role of peripheral A $\beta$  in exacerbating brain amyloidosis in both humans and mouse models of AD. Moreover, we highlight the need to further understand the mechanisms by which circulating A $\beta$  peptides may reach the brain and contribute to neuropathology. Finally, we discuss the implications of targeting peripheral A $\beta$  as a therapeutic approach in treating AD.

**KEYWORDS:** Alzheimer's disease, neurodegeneration, prion, amyloid-beta, peripheral tissues

**RECEIVED:** July 9, 2022. **ACCEPTED:** August 15, 2022.

**TYPE:** Commentary

**FUNDING:** The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: Research reported in this publication was supported by grants from the NIH (RF1AG059321, RF1AG072491) and the Alzheimer's Association (NIRGD-15-363554) to RM.

**DECLARATION OF CONFLICTS OF INTEREST:** The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

**CORRESPONDING AUTHOR:** Rodrigo Morales, Department of Neurology, The University of Texas Health Science Center at Houston, 6431 Fannin St., Houston, TX 77030, USA. Email: Rodrigo.MoralesLoyola@uth.tmc.edu

**COMMENT ON:** Morales R, Bravo-Alegria J, Moreno-Gonzalez I, Duran-Aniotz C, Gamez N, Edwards III G, Soto C. Transmission of cerebral amyloid pathology by peripheral administration of misfolded A $\beta$  aggregates. *Mol Psychiatry*. 2021 Oct;26(10):5690-5701. doi: 10.1038/s41380-021-01150-w. Epub 2021 May 17. PMID: 34002023; PMCID: PMC8595465.

## Background

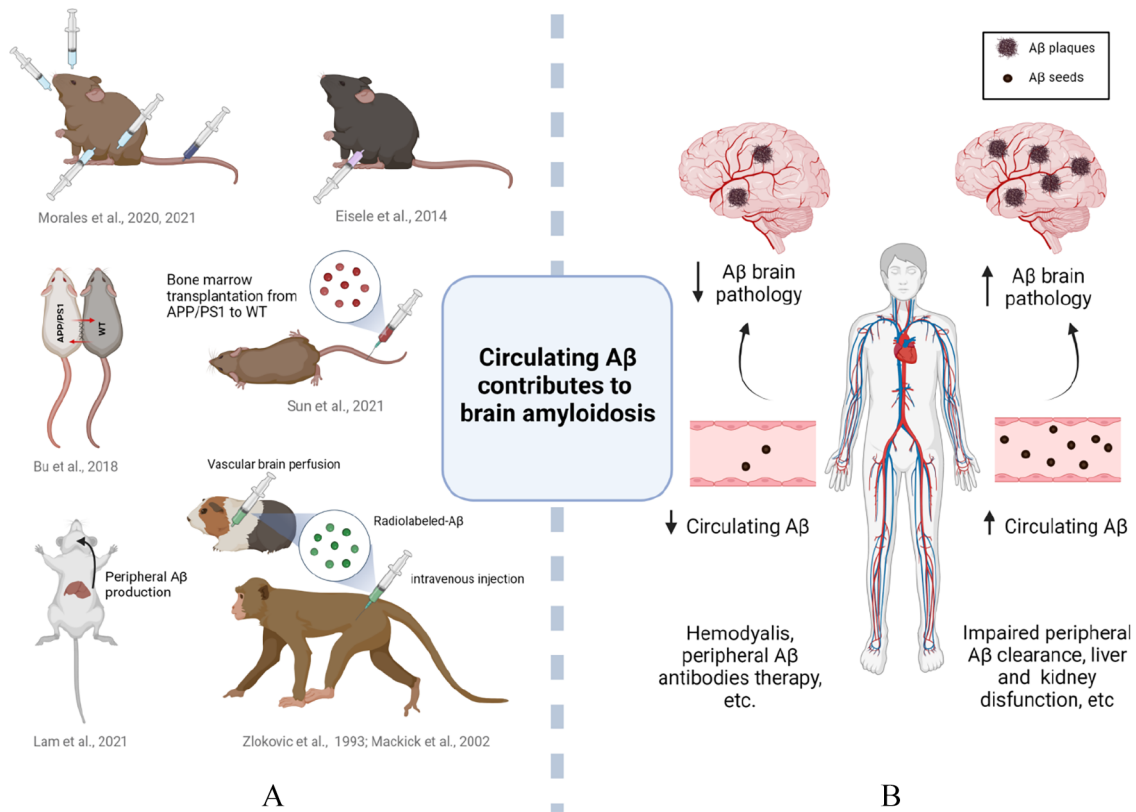
Alzheimer's disease (AD) is a proteinopathy characterized by the abnormal accumulation of misfolded proteins, such as amyloid-beta (A $\beta$ ) and tau, in the brain. While the etiology of AD is still unknown, A $\beta$  aggregates are considered the triggers of a cascade of events leading to tau pathology, neuroinflammation, synaptic loss, neurodegeneration, and cognitive decline. The progression of A $\beta$  and tau pathology throughout the brain may be explained by their prion-like properties. As such, it is thought that the spreading of brain pathology in AD is mediated by the self-propagation of misfolded protein aggregates, or "seeds," that template the further misfolding and aggregation of natively folded proteins across brain regions. The protein "seeding" concept has gained special interest due to findings showing that cerebral amyloidosis can be exogenously exacerbated after the intracerebral administration of A $\beta$  seeds into transgenic (Tg) mouse models of AD<sup>1,2</sup> and humans.<sup>3,4</sup> Moreover, additional studies have experimentally demonstrated that brain amyloidosis can be induced through seeding in mice that normally do not do it.<sup>5</sup> This "seeding" mechanism resembles that observed with infectious prions.<sup>5</sup> Cumulatively, these findings suggest that disease mechanisms associated with protein misfolding can be initiated and/or accelerated by the exogenous administration of similar proteinaceous particles.

### *Role of peripheral A $\beta$ seeds in the progression of brain amyloidosis*

Intriguingly, A $\beta$  aggregates have been found not only in the brain of AD patients but also throughout the periphery,

including vascular tissue, skeletal muscle, pancreas, and liver.<sup>6</sup> Whether peripheral A $\beta$  seeds play a role in exacerbating cerebral amyloidosis via seeding remains contentious. In this context, our recent study<sup>7</sup> demonstrated that intraperitoneal, intramuscular, and ocular (eye drops) administration of A $\beta$ -seed laden brain extracts accelerated brain amyloidosis in a mouse model of AD. Not only do our results confirm the efficacy of intraperitoneal seeding, as observed by Eisele et al.,<sup>8</sup> but also bring to light two other relevant peripheral routes previously shown to be involved in prion infections.<sup>9,10</sup> These results strongly suggest that A $\beta$  aggregates present in peripheral tissues may have an effect on the progression of disease-associated pathological changes. Moreover, Eisele et al. were able to detect intraperitoneally inoculated A $\beta$  in peripheral tissues, including the liver and spleen, and, more interestingly, in blood monocytes up to 1-week post-administration, suggesting a potential mechanism of A $\beta$  seed transportation from the periphery to the brain via circulating macrophages.<sup>8</sup> In support of these findings, in our study,<sup>7</sup> we observed a significant increase in seeded A $\beta$  deposits associated with cerebral blood and meningeal vessels in all the experimental animals that underwent peripheral induction of brain amyloidosis. Interestingly, blood cells have been implicated as a major avenue of peripheral A $\beta$  production.<sup>11</sup> In a recent study, bone marrow cell transplantation from APP<sup>swe</sup>/PS1<sup>dE9</sup> mice into wild-type (WT) mice resulted in continuous production of human A $\beta$  in the blood and caused AD-associated neuropathology in WT recipients.<sup>12</sup> Likewise, Lam et al., who developed a Tg mouse model in which the expression of human





**Figure 1.** Peripheral A $\beta$  contributes to AD pathology in the brain. (A) Summary of experimental animal bioassays supporting the role of peripheral A $\beta$  in brain amyloidosis (further discussed in the main text). (B) Schematic representation depicting how alterations at peripheral A $\beta$  concentrations may alter brain pathology.

APP is absent in the brain and largely limited to the liver, found that aged mice displayed human A $\beta$  deposition in the brain, suggesting that liver-derived A $\beta$  peptides can enter the CNS and facilitate pathological changes.<sup>13</sup> These data highlight the possible role of the vascular system in not only generating but also transporting peripheral A $\beta$  aggregates to the central nervous system (CNS), where protein seeding mechanisms likely result in parenchymal amyloidosis (Figure 1A).

The peripheral sink hypothesis postulates that central and peripheral levels of A $\beta$  are in equilibrium. As such, it is believed that an increase/reduction of peripheral A $\beta$  may result in higher/lower levels of A $\beta$  in the brain.<sup>14</sup> In this context, numerous studies have evaluated the role of circulating A $\beta$  on brain amyloidosis. One report by Bu et al. supports this notion by using a parabiosis model linking APP<sup>swe</sup>/PS1<sup>dE9</sup> Tg and WT littermates.<sup>15</sup> They observed that parabiotic WT animals developed parenchymal and vascular A $\beta$  deposition as well as other AD-associated pathologies. Interestingly, circulating fluor-labeled A $\beta$  was detected in the brain of parabiotic mice and thought to be the trigger of AD pathogenesis.<sup>15</sup> Blood A $\beta$  levels have been extensively examined in the Tg2576 animal model.<sup>16</sup> To further elucidate whether circulating A $\beta$  seeds present in the blood of old Tg2576 are able to promote amyloid pathology progression in non-disease-affected animals, we performed blood infusion experiments in young Tg mice.<sup>17</sup> Our results demonstrated that the recipient animals displayed

a significant acceleration of cerebral amyloidosis after receiving 2 transfusions of old-Tg blood. Unexpectedly, intravenously infused mice did not display vascular amyloid deposits in the brain.<sup>17</sup> This might be explained by the fact that circulating A $\beta$  were suspected to be composed mostly by oligomeric species, which may penetrate the CNS of recipient animals, diffuse throughout the brain parenchyma, and then seed amyloid pathology. Nevertheless, the isolation of misfolded A $\beta$  in the blood of animal models and humans remains a challenge and several groups are actively researching to develop methods able to identify these disease-relevant particles in this bodily fluid.

Although the aforementioned studies support the idea that circulating A $\beta$  modifies AD pathology in the brain, there are some reports that have not been able to recapitulate these findings. For instance, Fukuchi et al. found that APP-C99 Tg mice do not develop cerebral amyloidosis despite presenting high levels of circulating A $\beta$ .<sup>18</sup> In a more recent publication, Brackhan et al. generated <sup>13</sup>C-lysine-labeled human A $\beta$  from Tg mouse brain extracts that were intraperitoneally injected into young Tg mice.<sup>19</sup> They detected the inoculated A $\beta$  in the liver and lymphoid tissues but not in the brain of treated animals. They concluded that intraperitoneally administered A $\beta$  does not reach the brain from the periphery nor does it play any significant role in exacerbating brain amyloidosis.<sup>19</sup> Conversely, studies in guinea pigs and nonhuman primate models of cerebral amyloidosis have found that peripherally administered

labeled-A $\beta$  reaches the brain, contributes to parenchymal amyloidosis, and co-localizes with A $\beta$  plaques<sup>20,21</sup> (Figure 1A). The discrepancies noted by the study from Brackhan et al. may be explained by the source of the inoculum and/or the utilization of specific Tg mouse models that might generate distinct A $\beta$  species that therefore result in differences in the rate of amyloid pathology progression.

Furthermore, our study utilized A $\beta$ -containing brain homogenate (BH) as inoculum. Considering this, it is plausible that other components of the BH (including lipopolysaccharides, inflammatory cytokines, cell-toxic products, etc.<sup>22</sup>) may influence cerebral amyloidosis in a manner distinct from A $\beta$  seeding. While the effect of these other factors remains under active investigation, studies with purified recombinant A $\beta$  have demonstrated a similar exacerbation in brain amyloidosis when intravenously inoculated into Tg mice (Morales et al.<sup>17</sup> and unpublished data).

Interestingly, the experimental animals described in our peripheral administration study<sup>7</sup> displayed a variety of A $\beta$ -neuropathological profiles and brain tropisms depending on the peripheral route of inoculation. While in prion diseases different tropisms have often been found to correlate to different prion strains, at present we lack the evidence to claim that specific peripheral routes favor different strains of A $\beta$  that may exist in the original inoculum. Alternatively, the distinct tropisms observed may be due to varying efficacies in reaching the CNS depending on the route of peripheral inoculation.

Importantly, our study<sup>7</sup> considered oral administration as an additional peripheral pathway. We performed oral gavages of highly concentrated A $\beta$ -containing BH from mouse and human origin into experimental mice. Although different dosages were tested, orally inoculated animals did not present significant neuropathological changes to that observed in non-injected control mice.<sup>7</sup> We hypothesize that orally administered A $\beta$  seeds may be susceptible to gastrointestinal degradation and/or unable to cross the epithelial layers of the gastrointestinal tract. Based on prion studies, prion inoculation into the tongue of hamsters is 100,000-fold more efficient than oral ingestion.<sup>23</sup> Current experiments in our laboratory are ongoing to study whether intra-tongue inoculation of Tg BH results in an effective route to transmitting AD phenotypes into Tg2576 animals. This may provide additional information on the capability of misfolded A $\beta$  particles to spread across different tissues.

Together, these findings shed light on the complexity of AD pathogenesis and the role that circulating A $\beta$  may play in neuropathology and disease progression. Further investigation is needed to decipher how systemic A $\beta$  reaches the CNS and the potential mechanisms underlying peripherally seeded brain amyloidosis.

### *Potential avenues for AD therapeutic interventions*

Reservoirs of A $\beta$  can be found in several non-neuronal peripheral tissues that likely contribute to an active interchange of

these peptides between the brain and the periphery.<sup>6</sup> It has been hypothesized that A $\beta$  levels in the CNS and the periphery maintain an equilibrium state.<sup>14</sup> This equilibrium is dependent, among other factors, on the capacity of A $\beta$  to cross the blood-brain barrier (BBB) from the brain to the circulatory system and vice versa. Moreover, the efflux of brain-derived A $\beta$  may be mediated by the blood-CSF barrier, the arachnoid villi, and the glymphatic pathway.<sup>14</sup> It is thought that an imbalance of A $\beta$  production and clearance may drive abnormal accumulation of A $\beta$  in the brain and AD pathology progression.

Considering what was discussed above, targeting peripheral A $\beta$  has emerged as an unconventional yet promising approach in the treatment of AD. The therapeutic implication of the peripheral sink hypothesis suggests that peripheral antibodies can be utilized to sequester circulating A $\beta$  thus lowering its concentration in the blood.<sup>24,25</sup> This shift in the dynamic equilibrium between the A $\beta$  concentrations in the brain and periphery may cause a net efflux of A $\beta$  from the brain which may reduce plaque load and ameliorate pathology. As such, the reduction of A $\beta$  peptides in the periphery may be a viable mechanism for reducing A $\beta$  related pathology in the brain.<sup>24,25</sup> Further support for this notion comes from the observation that patients who have undergone hemodialysis, effectively clearing peripheral A $\beta$ , exhibit decreased A $\beta$  deposition in the brain.<sup>26</sup> Congruently, multiple studies have demonstrated that impairment of peripheral A $\beta$  clearance exacerbates AD pathology in the brain<sup>27-29</sup> (Figure 1B). These studies suggest that the modulation of peripheral or circulating A $\beta$  merits further investigation as a promising therapeutic avenue to reduce amyloid burden in the brain.

### Concluding Remarks

Our original publication entitled “Transmission of cerebral amyloid pathology by peripheral administration of misfolded A $\beta$  aggregates,”<sup>7</sup> as well as many others,<sup>8,11-13,15,17</sup> provide evidence that peripheral A $\beta$  seeds play an important role in the progression of cerebral A $\beta$  amyloidosis. This is of special interest to further study AD pathogenesis, as not only brain-derived but also circulating A $\beta$  peptides may drive both central and systemic AD-associated features. Moreover, targeting A $\beta$  from the periphery might open novel avenues for promising diagnostic and therapeutic interventions.

### Author Contributions

NG wrote the manuscript and prepared Figure 1. RM reviewed and edited the manuscript, and approved its final version.

### ORCID iD

Rodrigo Morales  <https://orcid.org/0000-0001-7766-5770>

### REFERENCES

1. Kane MD, Lipinski WJ, Callahan MJ, et al. Evidence for seeding of  $\beta$ -amyloid by intracerebral infusion of Alzheimer brain extracts in  $\beta$ -amyloid precursor protein-transgenic mice. *J Neurosci*. 2000;20:3606-3611.

2. Morales R, Bravo-Alegria J, Duran-Aniotz C, Soto C. Titration of biologically active amyloid- $\beta$  seeds in a transgenic mouse model of Alzheimer's disease. *Sci Rep*. 2015;5:9349.
3. Jaunmuktane Z, Mead S, Ellis M, et al. Evidence for human transmission of amyloid- $\beta$  pathology and cerebral amyloid angiopathy. *Nature*. 2015;525:247-250.
4. Purro SA, Farrow MA, Linehan J, et al. Transmission of amyloid- $\beta$  protein pathology from cadaveric pituitary growth hormone. *Nature*. 2018;564:415-419.
5. Morales R, Duran-Aniotz C, Castilla J, Estrada LD, Soto C. De novo induction of amyloid- $\beta$  deposition in vivo. *Mol Psychiatry*. 2012;17:1347-1353.
6. Roher AE, Esh CL, Kokjohn TA, et al. Amyloid beta peptides in human plasma and tissues and their significance for Alzheimer's disease. *Alzheimer's Dement*. 2009;5:18-29.
7. Morales R, Bravo-Alegria J, Moreno-Gonzalez I, et al. Transmission of cerebral amyloid pathology by peripheral administration of misfolded  $a\beta$  aggregates. *Mol Psychiatry*. 2021;26:5690-5701.
8. Eisele YS, Fritschi SK, Hamaguchi T, et al. Multiple factors contribute to the peripheral induction of cerebral amyloidosis. *J Neurosci*. 2014;34:10264-10273.
9. Bosque PJ, Ryou C, Telling G, et al. Prions in skeletal muscle. *Proc Natl Acad Sci U S A*. 2002;99:3812-3817.
10. Orrù CD, Soldau K, Cordano C, et al. Prion seeds distribute throughout the eyes of sporadic Creutzfeldt-Jakob disease patients. *mBio*. 2018;9:e02095-18.
11. Chen M, Inestrosa NC, Ross GS, Fernandez HL. Platelets are the primary source of amyloid beta-peptide in human blood. *Biochem Biophys Res Commun*. 1995;213:96-103.
12. Sun HL, Chen SH, Yu ZY, et al. Blood cell-produced amyloid- $\beta$  induces cerebral Alzheimer-type pathologies and behavioral deficits. *Mol Psychiatry*. 2021;26:5568-5577.
13. Lam V, Takechi R, Hackett MJ, et al. Synthesis of human amyloid restricted to liver results in an Alzheimer disease-like neurodegenerative phenotype. *PLoS Biol*. 2021;19:e3001358.
14. Tarasoff-Conway JM, Carare RO, Osorio RS, et al. Clearance systems in the brain-implications for Alzheimer disease. *Nat Rev Neurol*. 2015;11:457-470.
15. Bu XL, Xiang Y, Jin WS, et al. Blood-derived amyloid- $\beta$  protein induces Alzheimer's disease pathologies. *Mol Psychiatry*. 2018;23:1948-1956.
16. Kawarabayashi T, Younkin LH, Saido TC, Shoji M, Ashe KH, Younkin SG. Age-dependent changes in brain, CSF, and plasma amyloid (beta) protein in the tg2576 transgenic mouse model of Alzheimer's disease. *J Neurosci*. 2001;21:372-381.
17. Morales R, Duran-Aniotz C, Bravo-Alegria J, et al. Infusion of blood from mice displaying cerebral amyloidosis accelerates amyloid pathology in animal models of Alzheimer's disease. *Acta Neuropathol Commun*. 2020;8:213-216.
18. Fukuchi KI, Ho L, Younkin SG, et al. High levels of circulating beta-amyloid peptide do not cause cerebral beta-amyloidosis in transgenic mice. *Am J Pathol*. 1996;149:219-227.
19. Brackhan M, Calza G, Lundgren K, et al. Isotope-labeled amyloid- $\beta$  does not transmit to the brain in a prion-like manner after peripheral administration. *EMBO Rep*. 2022;23:e54405.
20. Zlokovic BV, Ghiso J, Mackic JB, McComb JG, Weiss MH, Frangione B. Blood-brain barrier transport of circulating Alzheimer's amyloid beta. *Biochem Biophys Res Commun*. 1993;197:1034-1040.
21. Mackic JB, Bading J, Ghiso J, et al. Circulating amyloid-beta peptide crosses the blood-brain barrier in aged monkeys and contributes to Alzheimer's disease lesions. *Vasc Pharmacol*. 2002;38:303-313.
22. Wendeln AC, Degenhardt K, Kaurani L, et al. Innate immune memory in the brain shapes neurological disease hallmarks. *Nature*. 2018;556:332-338.
23. Bartz JC, Kincaid AE, Bessen RA. Rapid prion neuroinvasion following tongue infection. *J Virol*. 2003;77:583-591.
24. DeMattos RB, Bales KR, Cummins DJ, Dodart JC, Paul SM, Holtzman DM. Peripheral anti-A $\beta$  antibody alters CNS and plasma aaA $\beta$  clearance and decreases brain aaA $\beta$  burden in mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2001;98:8850-8855.
25. Piazza F, Greenberg SM, Savoirdo M, et al. Anti-amyloid  $\beta$  autoantibodies in cerebral amyloid angiopathy-related inflammation: implications for amyloid-modifying therapies. *Ann Neurol*. 2013;73:449-458.
26. Sakai K, Senda T, Hata R, et al. Patients that have undergone hemodialysis exhibit lower amyloid deposition in the brain: Evidence supporting a therapeutic strategy for Alzheimer's disease by removal of blood amyloid. *J Alzheimers Dis*. 2016;51:997-1002.
27. Tian DY, Cheng Y, Zhuang ZQ, et al. Physiological clearance of amyloid-beta by the kidney and its therapeutic potential for Alzheimer's disease. *Mol Psychiatry*. 2021;26:6074-6082.
28. Mawuenyega KG, Sigurdson W, Ovod V, et al. Decreased clearance of CNS  $\beta$ -amyloid in Alzheimer's disease. *Science*. 2010;330:1774-1774.
29. Wang YR, Wang QH, Zhang T, et al. Associations between hepatic functions and plasma amyloid-beta levels-implications for the capacity of liver in peripheral amyloid-beta clearance. *Mol Neurobiol*. 2017;54:2338-2344.