

Research Article

***Blastomyces dermatitidis* Yeast Lysate Antigen Combinations: Antibody Detection in Dogs with Blastomycosis**

Alex R. Boyd, Jamie L. VanDyke, and Gene M. Sclarone

Department of Biological Sciences, Idaho State University, P.O. Box 8007 921 S. 8th Avenue, Pocatello, ID 83209, USA

Correspondence should be addressed to Alex R. Boyd; boydale2@isu.edu

Received 31 July 2013; Revised 16 September 2013; Accepted 18 September 2013

Academic Editor: Remo Lobetti

Copyright © 2013 Alex R. Boyd et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The systemic fungal infection, blastomycosis, which infects both humans and animals has presented a diagnostic challenge for clinicians for many years. The aim of this study was to evaluate the diagnostic sensitivity of *Blastomyces dermatitidis* yeast lysate antigens with respect to antibody detection in dogs with blastomycosis. Lysate antigens were prepared from *B. dermatitidis* isolates T-58 and T-66 (dogs, Tennessee) and WI-R and WI-J (dogs, Wisconsin). Based on results obtained from a preliminary comparative study, five combinations of these isolates and one individual isolate were tested against 92 serum specimens from dogs with culture-proven or histologically-confirmed blastomycosis, using the indirect enzyme-linked immunosorbent assay (ELISA). Mean absorbance values obtained from the sera ranged from 0.905 with the individual T-58 antigen to 1.760 using an antigen combination (T-58 + T-66 + WI-R). All of the 6 antigenic preparations were able to detect antibody in the serum specimens, but the antigen combinations detected antibody to a higher degree than the individual antigen. This study provides evidence that combinations of the yeast lysate reagents seem to be more efficacious for antibody detection in dog sera, but our laboratory is continuing to evaluate antigen lysate combinations for detection of antibodies in blastomycosis.

1. Introduction

Blastomycosis, a systemic fungal infection of humans and animals, is produced by the dimorphic fungal organism *Blastomyces dermatitidis*. The infection is initiated by the inhalation of spores produced by the filamentous phase of the fungus. The organism exists in this stage in nature or in the laboratory at 25°C and has the ability to convert to the yeast phase at 37°C in the lungs of the infected host. The disease may be self-resolving or it may exist as an acute or chronic state in the pulmonary tissue. If the disease is untreated while in the lungs, it may become invasive and disseminate to other organs and possibly to the central nervous system where fatal meningitis may develop [1–5]. Blastomycosis, as well as other systemic mycoses, are termed “emerging fungal threats” since they not only infect persons with normal immune systems but are also a cause for concern in immunocompromised individuals [3, 4].

The geographic distribution of blastomycosis has been associated with southeastern and south-central states that

border the Ohio and Mississippi Rivers and upper midwestern states including areas in Wisconsin and Minnesota, which are highly endemic for the disease [6, 7].

Due to the increase in systemic fungal diseases, researchers have begun to devote more attention to developing ways of diagnosing, preventing, and treating these mycoses [8–17]. For the past several years the thrust of research in our laboratory has been associated with studies on various strains of *B. dermatitidis* from human, animal, or environmental specimens from many geographical locations in an effort to better understand the immunobiology of the organism [18–27]. Diagnosis of the disease has presented major problems. In some instances, culturing or histopathological examination may be beneficial, but in some patients these methods may not yield the desired results. This has led to more and more research being done to improve immunological assays which tend to provide a more rapid diagnosis, but we, as well as many other investigators, recognize that problems still exist with regard to the sensitivity and specificity of immunoassays.

Our current studies are aimed at evaluating combinations of the various *B. dermatitidis* yeast lysate antigens for the detection of antibodies in serum specimens from dogs with blastomycosis.

2. Materials and Methods

2.1. Antigens. Yeast phase lysate reagents (T-58, dog Tennessee; T-66, dog Tennessee; WI-R, dog Wisconsin; and WI-J, dog Wisconsin) were prepared by a method similar to one that was previously used for the production of antigen from *Histoplasma capsulatum* [28–30] and modified in our laboratory for *B. dermatitidis* lysate antigen production [23]. The yeast phase cells were grown for 7 days at 37°C in a chemically defined medium (glucose, 10.0 g; potassium phosphate monobasic, 1.5 g; calcium chloride dehydrate, 0.15 g; magnesium sulfate, 0.5 g; ammonium sulfate, 2.0 g; L-asparagine, 2.0 g; L-cysteine, 0.2 g; and pH adjusted to 6.2 with 5 N sodium hydroxide) in an incubator shaker, harvested by centrifugation (700 ×g; 5 min) followed by washing with distilled water, resuspended in distilled water, and then allowed to lyse for 7 days at 37°C in water with shaking. The preparations were centrifuged, filter sterilized, merthiolate added (1:10,000), and stored at 4°C. Protein determinations were performed on the lysates using the BCA protein assay kit (Pierce Chemical Company, Rockford, IL, USA), and dilutions of the antigenic reagents used in the ELISA assays were based on protein concentration. Combinations of the above four antigenic reagents as well as T-58 (not combined with others) were used for antibody detection. A previous preliminary comparative evaluation was performed [27] using a number of individual and combinations of the above lysate preparations to assess their ability to detect antibodies in 24 sera from dogs with blastomycosis. This study indicated that 6 of the preparations showed the greatest degree of sensitivity. Therefore, this present study, with a much greater number of serum specimens, was initiated to further evaluate the 6 optimal lysate reagents (T-58 + T-66 + WI-R; T-66 + WI-R; T-58 + WI-J; T-66 + WI-R + WI-J; T-58 + T-66, and the one individual antigen T-58) from the earlier study for antibody detection in 92 sera from dogs with diagnosed blastomycosis but with varying amount of antibody in the specimens.

2.2. Serum Specimens. Ninety-two serum specimens from dogs with diagnosed blastomycosis were provided by Dr. A. M. Legendre (University of Tennessee College of Veterinary Medicine, Knoxville, TN, USA). Negative (normal) sera were not included in this study since we were interested in comparing reactivity and not correcting for background with negative controls.

2.3. Enzyme-Linked Immunosorbent Assay (ELISA). The ability of each of the 6 (individual or combination preparations) yeast lysate reagents to detect antibody in the above serum specimens was determined using the indirect enzyme-linked immunosorbent assay (ELISA). Each lysate antigen was diluted (2000 ng of protein/mL) in a carbonate-bicarbonate

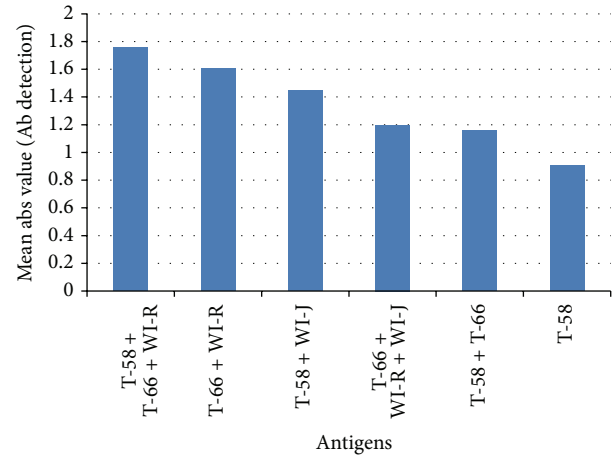


FIGURE 1: Comparison of the 6 *B. dermatitidis* yeast lysate antigens for the detection of antibodies in serum specimens from 92 dogs with diagnosed blastomycosis.

coating buffer (pH 9.6; equal amounts of each lysate were admixed in preparing the combinations and 2000 ng of protein/mL of the individual T-58 antigen resulting in 200 ng total protein/100 uL in each well) and then added to triplicate wells (100 uL) of a NUNC 96-well microplate (Fisher-Thermo). The plates were then incubated overnight at 4°C in a humid chamber followed by washing three times with phosphate buffered saline containing 0.15% Tween 20 (PBS-T). The serum specimens (1:2500 dilution; 100 uL) were added to the microplate wells and incubated for 30 min at 37°C in a humid chamber. Following this incubation the wells were washed as above and 100 uL of goat anti-dog IgG (H & L) peroxidase conjugate (Kirkegaard and Perry, Gaithersburg, MD, USA) was added to each well and incubated for 30 min at 37°C. The plates were again washed as above and 100 uL of TMB peroxidase substrate (Pierce/Fisher-Thermo) was added to each well and incubated for approximately 2 min at room temperature. The reaction was stopped by the addition of sulfuric acid and the absorbance read at 450 nm using a BIO-RAD 2550 EIA reader.

3. Results/Discussion

The mean absorbance values of the six *B. dermatitidis* lysate antigens, when used in the ELISA to detect antibodies in 92 dog sera, are shown in Figure 1. The five reagent combinations exhibited mean absorbance values greater than one, ranging from 1.158 to 1.760, while the single antigenic reagent (T-58) exhibited a mean absorbance value of 0.905. The most reactive reagent was T-58 + T-66 + WI-R, a mixture of two southern isolates and one northern isolate. All of the reagents were able to detect antibodies against blastomycosis with the optimal reagent detecting antibody at twice the rate of the single antigen.

The sensitivity of the 6 lysate preparations, when evaluated on their ability to detect antibody at mean absorbance values ranging from 0.400 to 2.800, is shown in Table 1. The

TABLE 1: The detection of antibody in the dog serum specimens at the different mean absorbance value ranges with each of the *B. dermatitidis* lysate preparations (number of positive specimens) and the sensitivity percentages of each lysate at the absorbance ranges.

(a)

Absorbance range	T-58 + T-66 + WI-R	T-66 + WI-R	T-58 + WI-J	T-66 + WI-J + WI-R	T-58 + T-66	T-58
0.400–0.800	4	2	12	14	22	47
0.801–1.200	12	20	25	35	29	21
1.201–1.600	20	22	15	28	23	15
1.601–2.000	17	25	20	12	17	9
2.010–2.400	32	22	16	3	1	0
2.401–2.800	7	1	4	0	0	0

(b)

Absorbance range	T-58 + T-66 + WI-R	T-66 + WI-R	T-58 + WI-J	T-66 + WI-J + WI-R	T-58 + T-66	T-58
>0.4	100%	100%	100%	100%	100%	100%
>0.8	96%	98%	87%	85%	76%	49%
>1.2	83%	76%	60%	47%	45%	26%
>1.6	61%	52%	44%	16%	20%	10%
>2.0	42%	25%	22%	37%	19%	0%

sensitivity percentages with the optimal lysate antigen (T-58 + T-66 + WI-R) range from 100% at the 0.400–0.800 absorbance range to 42% at the highest absorbance value (2.101–2.800). Sensitivity percentages ranged from 25% to 0% with the other 5 lysates at this highest absorbance value level.

Most previous studies have evaluated only single antigens for the detection of antibodies in blastomycosis. However, one previous study tested 14 antigens or antigen combinations against 24 dog sera to determine sensitivity using the ELISA [27]. In this study, we used the 6 most reactive reagents from that study to further examine sensitivity in this comparative assay.

4. Conclusion

This study indicates that certain combinations of antigens are preferable as immunodiagnostic reagents for antibody detection of *B. dermatitidis* in sera from dogs. The T-58 + T-66 + WI-R antigen combination is a promising candidate for future studies based on this comparative study. Therefore, our laboratory is continuing studies in an effort to develop a reagent that will aid in the reliable immunodiagnosis of blastomycosis in humans and animals.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

This research was supported by the Department of Biological Sciences, Idaho State University, Pocatello, no. 83209-8007.

References

- [1] R. W. Bradsher, S. W. Chapman, and P. G. Pappas, "Blastomycosis," *Infectious Disease Clinics of North America*, vol. 17, no. 1, pp. 21–40, 2003.
- [2] J. R. Bariola and K. S. Vyas, "Pulmonary blastomycosis," *Seminars in Respiratory and Critical Care Medicine*, vol. 32, no. 6, pp. 745–753, 2011.
- [3] J. A. McKinnell and P. G. Pappas, "Blastomycosis: new insights into diagnosis, prevention, and treatment," *Clinics in Chest Medicine*, vol. 30, no. 2, pp. 227–239, 2009.
- [4] M. Saccente and G. L. Woods, "Clinical and laboratory update on blastomycosis," *Clinical Microbiology Reviews*, vol. 23, no. 2, pp. 367–381, 2010.
- [5] J. A. Smith and C. A. Kauffman, "Blastomycosis," *Proceedings of the American Thoracic Society*, vol. 7, no. 3, pp. 173–180, 2010.
- [6] A. F. DiSalvo, "Blastomycosis," in *Topley and Wilson's Microbiology and Microbial Infections*, L. Collier, Ed., pp. 337–355, Arnold Publishers, London, UK, 9th edition, 1998.
- [7] B. S. Klein, J. M. Vergeront, R. J. Weeks et al., "Isolation of *Blastomyces dermatitidis* in soil associated with a larger outbreak of blastomycosis in Wisconsin," *Journal of Infectious Diseases*, vol. 155, pp. 262–268, 1986.
- [8] D. R. Allton, R. G. Rivard, P. A. Connolly et al., "Detection of Latin American strains of histoplasma in a murine model by use of a commercially available antigen test," *Clinical and Vaccine Immunology*, vol. 17, no. 5, pp. 802–806, 2010.
- [9] J. R. Bariola, C. A. Hage, M. Durkin et al., "Detection of *Blastomyces dermatitidis* antigen in patients with newly diagnosed blastomycosis," *Diagnostic Microbiology and Infectious Disease*, vol. 69, no. 2, pp. 187–191, 2011.
- [10] R. W. Bradsher and P. G. Pappas, "Detection of specific antibodies in human blastomycosis by enzyme immunoassay," *Southern Medical Journal*, vol. 88, no. 12, pp. 1256–1259, 1995.
- [11] P. Connolly, C. A. Hage, J. R. Bariola et al., "Blastomyces dermatitidis antigen detection by quantitative enzyme immunoassay," *Clinical and Vaccine Immunology*, vol. 19, no. 1, pp. 53–56, 2012.

- [12] M. Durkin, L. Estok, D. Hospenthal et al., "Detection of coccidioides antigenemia following dissociation of immune complexes," *Clinical and Vaccine Immunology*, vol. 16, no. 10, pp. 1453–1456, 2009.
- [13] C. A. Hage, J. A. Ribes, N. L. Wengenack et al., "A multicenter evaluation of tests for diagnosis of histoplasmosis," *Clinical Infectious Diseases*, vol. 53, no. 5, pp. 448–454, 2011.
- [14] B. S. Klein and J. M. Jones, "Isolation, purification, and radiolabeling of a novel 120-kD surface protein on *Blastomyces dermatitidis* yeasts to detect antibody in infected patients," *Journal of Clinical Investigation*, vol. 85, no. 1, pp. 152–161, 1990.
- [15] B. S. Klein, R. A. Squires, J. K. Lloyd, D. R. Ruge, and A. M. Legendre, "Canine antibody response to *Blastomyces dermatitidis* WI-1 antigen," *American Journal of Veterinary Research*, vol. 61, no. 5, pp. 554–558, 2000.
- [16] D. Spector, A. M. Legendre, J. Wheat et al., "Antigen and antibody testing for the diagnosis of blastomycosis in dogs," *Journal of Veterinary Internal Medicine*, vol. 22, no. 4, pp. 839–843, 2008.
- [17] K. S. Vyas, J. R. Bariola, and R. W. Bradsher, "Advances in the serodiagnosis of blastomycosis," *Current Fungal Infection Reports*, vol. 2, pp. 227–231, 2008.
- [18] T. R. Allison, J. C. Wright, and G. M. Scalarone, "*Blastomyces dermatitidis*: stability studies on different yeast lysate antigens," *Open Journal of Immunology*, vol. 3, pp. 98–102, 2013.
- [19] D. Andrae, K. Birch, T. Bybee, T. Ritcher, J. Werth, and G. M. Scalarone, "Antigen detection in canine blastomycosis: comparison of different antibody-antigen combinations in two competitive ELISAs," *Open Journal of Medical Microbiology*, vol. 2, pp. 110–114, 2012.
- [20] J. L. Bono, A. M. Legendre, and G. M. Scalarone, "Detection of antibodies and delayed hypersensitivity with Rotofor preparative IEF fractions of *Blastomyces dermatitidis* yeast phase lysate antigens," *Journal of Medical and Veterinary Mycology*, vol. 33, no. 4, pp. 209–214, 1995.
- [21] M. A. Fisher, J. L. Bono, R. O. Abuodeh, A. M. Legendre, and G. M. Scalarone, "Sensitivity and specificity of an isoelectric focusing fraction of *Blastomyces dermatitidis* yeast lysate antigen for the detection of canine blastomycosis," *Mycoses*, vol. 38, no. 5-6, pp. 177–182, 1995.
- [22] W. O. Hatch and G. M. Scalarone, "Comparison of colorimetric and chemiluminescent ELISAs for the detection of antibodies to *Blastomyces dermatitidis*," *Journal of Medical and Biological Sciences*, vol. 3, no. 1, pp. 1–6, 2009.
- [23] S. M. Johnson and G. M. Scalarone, "Preparation and ELISA evaluation of *Blastomyces dermatitidis* yeast phase lysate antigens," *Diagnostic Microbiology and Infectious Disease*, vol. 11, no. 2, pp. 81–86, 1988.
- [24] C. M. Sestero and G. M. Scalarone, "Detection of IgG and IgM in sera from canines with blastomycosis using eight *Blastomyces dermatitidis* yeast phase lysate antigens," *Mycopathologia*, vol. 162, no. 1, pp. 33–37, 2006.
- [25] J. F. Shurley and G. M. Scalarone, "Isoelectric focusing and ELISA evaluation of a *Blastomyces dermatitidis* human isolate," *Mycopathologia*, vol. 164, no. 2, pp. 73–76, 2007.
- [26] J. F. Shurley, A. M. Legendre, and G. M. Scalarone, "*Blastomyces dermatitidis* antigen detection in urine specimens from dogs with blastomycosis using a competitive binding inhibition ELISA," *Mycopathologia*, vol. 160, no. 2, pp. 137–142, 2005.
- [27] J. VanDyke, A. Boyd, J. Sorensen et al., "Detection of antibodies in serum specimens from dogs with blastomycosis with lysate antigens prepared from four *Blastomyces dermatitidis* dog isolates: individual antigens vs antigen combinations," *Open Journal of Veterinary Medicine*, vol. 3, no. 4, pp. 235–239, 2013.
- [28] H. B. Levine, G. M. Scalarone, and S. D. Chaparas, "Preparation of fungal antigens and vaccines: studies on *Coccidioides immitis* and *Histoplasma capsulatum*," *Contributions to Microbiology and Immunology*, vol. 3, pp. 106–125, 1977.
- [29] H. B. Levine, G. M. Scalarone, G. D. Campbell, R. C. Graybill, and S. D. Chaparas, "Histoplasmin-CYL, a yeast phase reagent in skin test studies with humans," *American Review of Respiratory Disease*, vol. 119, no. 4, pp. 629–636, 1979.
- [30] G. M. Scalarone, H. B. Levine, and S. D. Chaparas, "Delayed hypersensitivity responses of experimental animals to histoplasmin from the yeast and mycelial phases of *Histoplasma capsulatum*," *Infection and Immunity*, vol. 21, no. 3, pp. 705–713, 1978.