

RESEARCH ARTICLE

Pneumocystis Pneumonia in Patients with Autoimmune Diseases: A Retrospective Study Focused on Clinical Characteristics and Prognostic Factors Related to Death

Minjiang Chen¹, Xinlun Tian^{1*}, Fang Qin², Jiong Zhou³, Jinjing Liu⁴, Mengzhao Wang¹, Kai-Feng Xu¹

1 Department of Respiratory Medicine, Peking Union Medical College Hospital, Beijing, China, **2** Department of Respiratory Medicine, China Meitan General Hospital, Beijing, China, **3** Offices for Infection Control, Peking Union Medical College Hospital, Beijing, China, **4** Department of Rheumatology, Peking Union Medical College Hospital, Beijing, China

* xinlun_t@sina.com



Abstract

Background

With the increasing use of immunosuppressive agents, the number of opportunistic infections has risen in patients with autoimmune diseases. Pneumocystis pneumonia (PCP) is one of these opportunistic infections that have a high mortality rate. However, only a few studies have described PCP in these patients, and these studies are limited in scope. We conducted this retrospective study to describe the clinical characteristics and factors associated with outcomes of PCP in patients with autoimmune diseases.

Methods

A retrospective study was performed in laboratory diagnosed PCP patients with autoimmune diseases in an academic hospital over a 10-year period. Patients with human immunodeficiency virus (HIV) infection were not included. Clinical characteristics were collected and the factors related to death were analysed.

Results

A total of 69 patients with PCP during the study period were included. Common clinical features included fever (81%), cough (56%), and dyspnea (35%). Ground glass opacity (81%) and reticulation (52%) were the most common radiological findings. Concurrent pulmonary infections including bacterium, aspergillus and cytomegalovirus were found in 34% of the patients. The overall in-hospital mortality rate was 32%. High mortality was associated with lower PaO₂/FiO₂ ratios and albumin levels. The lymphocyte count, CD4+ T cell count, previous usage of immunosuppressive agents, the duration and dose of glucocorticoids did not affect the outcome.

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Conclusions

The mortality rate in PCP patients with autoimmune diseases is high. Low PaO₂/FiO₂ ratios and albumin levels are independent prognostic factors of mortality.

Introduction

Pneumocystis jirovecii (*P. jirovecii*) is an opportunistic fungal pathogen that causes pneumonia in patients with immunosuppressive diseases. At present, pneumocystis pneumonia (PCP) remains a leading cause of death in patients with human immunodeficiency virus (HIV) infection. However, due to the increased use of chemotherapy and immunosuppressive agents, the incidence of PCP in HIV-negative patients has also increased. Unlike PCP in HIV patients, non-HIV PCP has a different clinical course and shows a higher mortality rate [1,2,3,4]. Recently, there are many reports about PCP in patients with autoimmune diseases such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA), especially in patients receiving prolonged usage of prednisone, immunosuppressive agents or biologic agents [5,6,7,8,9].

Several factors, such as female gender, co-infections, high D(A-a)O₂, high lactate dehydrogenase levels, increased BUN, pre-existing lung disease, more severe pneumonia and delay of initial treatment, have been reported to be associated with poor prognosis in non-HIV patients with PCP [10,11,12]. However, the factors related to death in non-HIV PCP patients with autoimmune diseases still need to be defined. Thus, we conducted this study to describe the clinical manifestations and outcomes and to evaluate the prognostic factors related with death in Chinese patients from an academic hospital. Our study is the largest single-centre study focusing on non-HIV PCP patients with autoimmune diseases in Mainland China. The results will provide additional information on these characteristics in Chinese patients.

Material and Methods

Patients

Medical records of Peking Union Medical College Hospital (PUMCH, an academic hospital in China) between January 2004 and December 2013 were retrospectively reviewed. Eligible patients had laboratory confirmed PCP and were diagnosed with autoimmune diseases. Other eligible criteria included an age of at least 12 years and a negative serum HIV test. Written informed consent was obtained from all patients (or from a caregiver in the case of children) for their clinical records to be used in this study. This study was approved by the PUMCH ethics review board.

Diagnosis

All cases of autoimmune diseases met the diagnostic criteria for their respective disease. The diagnosis of PCP was made on the basis of clinical symptoms and was confirmed by microbiological tests. Symptoms included one or more of the following: non-productive cough, dyspnea during exertion, fever, or chest pain. Computed tomography (CT) showed diffuse ground glass opacity (GGO), as well as inefficiency of antibiotics for non-PCP infections as previously described [13]. Respiratory specimens were obtained from bronchoalveolar lavage (BAL) fluid or hypertonic saline induced sputum. Tissue specimens were obtained from transbronchial lung biopsy or CT guided percutaneous lung puncture. A positive microbiological test was defined as the microscopic demonstration of *Pneumocystis jirovecii* through Gomorimethenamine silver (GMS) staining for the visualization of trophozoites and cysts in respiratory samples or lung tissue. Pneumocystis-specific nested polymerase chain reaction (PCR) was also performed in BAL.

specimens. PCP was confirmed if *Pneumocystis jirovecii* was found upon microscopic examination while positive results of PCR was regarded as infection only in patients with coinciding clinical symptoms of PCP, no evidence of other co-infections, and a response to anti-PCP therapy.

Data collection

Demographical details, underlying autoimmune diseases, associated medical conditions, clinical characteristics, radiology manifestations and laboratory results were retrospectively reviewed. Use of immunosuppressive agents, glucocorticoids and biological agents before the diagnosis of PCP were summarized. The corticosteroid dose was expressed as the prednisolone equivalent dose. Laboratory values included the cytomegalovirus (CMV) PCR results, and findings from bacterial, fungal and tuberculosis cultures from respiratory samples were also analysed.

Statistics

Descriptive statistics were used to calculate the means, standard deviations and frequencies. A univariate binary logistic regression analysis was performed. The continuous variables were analysed using the independent *t* test, and the categorical variables were compared by the *Chi*-square or Fisher's test. *P*-values ≤ 0.05 were considered statistically significant. All covariates with a *P* value ≤ 0.05 were included in multivariate analysis.

Results

Over the ten-year study period, a total of 264 patients were diagnosed with PCP in PUMCH. Among them, 69 patients with underlying autoimmune disease were included in this analysis.

Demographic data and underlying conditions

The patients consisted of 25 (36%) males and 44 (64%) females with a median age of 39 years (range 13 to 78 years) at initial diagnosis. The most common underlying diseases were SLE (39/57%), followed by dermatomyositis/myositis (11/16%), vasculitis (9/13%), RA (6/9%), and other autoimmune diseases (4/6%). Comorbidities included kidney disease, interstitial lung disease, heart failure and diabetes mellitus. Among those with chronic kidney disease, two patients had undergone renal replacement therapy ([Table 1](#)).

Immunosuppressive agents

Glucocorticoid was used in all patients. The mean dose was equivalent to 49 ± 25 mg (range 5 to 133mg) prednisone. The median duration of usage was 120 days (range 10 to 2670 days). In the majority of patients (63/91%), immunosuppressive agents were used 3 months before PCP was diagnosed. The most commonly used immunosuppressive agent was cyclophosphamide (40/63%). Biological agents were used in 5 (4%) patients (including rituximab in 2 patient and tumor necrosis factor alpha inhibitors in 3 patients).

Clinical, Radiologic and laboratory findings

The common clinical features included fever (56/81%), cough (39/56%), and dyspnea (24/35%). CT scans were performed in all 69 patients. The most common CT findings at initial diagnosis were ground glass opacity (56/81%) and reticulation (36/52%). The mean white blood cell count was 7335 ± 4528 cell/ μ l, while the mean total lymphocyte count was 529 ± 661 cell/ μ l. T cell subsets detection was performed in 38 patients. The mean CD4 positive T cell counts were 169 ± 196 cell/ μ l. Serum albumin (ALB), immunoglobulin, and lactate dehydrogenase levels were also tested ([Table 2](#)). Microorganisms found in respiratory specimens other

Table 1. Demographical details, underlying diseases, and diagnostic procedures of the patients.

Variables	No. of patients (%) or median (range)
Age, median years (range)	39(13–78)
Gender	
Male	25 (36%)
Female	44(64%)
Underlying disease	
Systemic Lupus Erythematosus (SLE)	39(57%)
Dermatomyositis/Myositis	11(16%)
Vasculitis*	9(13%)
RA	6(9%)
Other CTDs**	4(6%)
Use of glucocorticoid	69(100%)
Prednisone	39(57%)
Methylprednisolone	28(41%)
Other glucocorticoids	2(3%)
Dose of glucocorticoid	49(5–133)
Duration of glucocorticoid	120(10–2670)
Use of immunosuppressive agents [#]	63(91%)
Cyclophosphamide	40(63%)
Cyclosporin A	8(13%)
Mycophenlatemofetil	7(11%)
Methotrexate	6(10%)
Tripterygium Glycosides	7(11%)
Acetazolamide	2(3%)
Use of biological agents ^{&}	5 (4%)
≥ 2 immunosuppressive agents	11(17%)
Diagnostic procedure	
BAL	31 (45%)
Sputum	40 (58%)
Tissue	2 (3%)
Positive GMS stain	38 (55%)
Positive PCR	45 (65%)
Co-infections	24(34%)

* Vasculitis: Behcet's disease, microscopic polyangiitis, granulomatosis with polyangiitis

** Other CTDs: Sjogren syndrome (SS), undifferentiated connective tissue disease (UCTD), mixed connective tissue disease, scleroderma

Immunosuppressive agents: cyclophosphamide, cyclosporin A, mycophenlatemofetil, and tripterygium glycosides

& Biological agents: ritaximab, and antitumor necrosis factor α (infliximab, entanercept)

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than *P. jirovecii* included bacterium (5 patients) and *aspergillus* (7 patients). CMV had been detected in 13 patients, including 9 patients with positive qualitative detection of CMV DNA by PCR and 10 patients with positive CMV IgM.

Treatment and outcomes

Sulfamethoxazole (TMPco) was used as the first-line treatment in 59 (86%) patients. Among the 59 patients, 7 (10%) patients did not respond to TMPco. Clindamycin, primaquine and/or

Table 2. Clinical manifestations, Radiologic characters and laboratory findings of the patients.

	No. of patients (%)
Clinical manifestations	
Fever	56 (81%)
Cough	39 (56%)
Dyspnea	24 (35%)
Radiologic characters (CT scanning)	
Ground glass opacity (GGO)	56 (81%)
Reticulation	36 (52%)
Lymph node enlargement	25 (36%)
Patchy scattered infiltrates and parenchymal consolidations	22 (32%)
Pleural infusion	17 (25%)
Interlobular septa thicken	16 (23%)
Nodular consolidations	10 (14%)
Pneumothorax	5 (7%)
Cystic lesions	2 (3%)
Laboratory findings	
WBC count, cell/ μ l, mean \pm SD	7335 \pm 4528
Lymphocytes, cell/ μ l, mean \pm SD	529 \pm 661
CD4+ T cell, cell/ μ l	169 \pm 196
Albumin, g/L, mean \pm SD	30 \pm 4
Serum IgG, g/L, mean \pm SD	9.44 \pm 5.24
Serum IgA, g/L, mean \pm SD	1.90 \pm 0.75
Serum IgM, g/L, mean \pm SD	1.02 \pm 0.43
LDH, U/L	477 \pm 145
PaO ₂ /FiO ₂ ratios, mean \pm SD	217 \pm 98
Co-infections	
Cytomegalovirus DNA PCR positive	13(19%)
Bacterium in BAL*	5(7%)
<i>Aspergillus</i> in BAL	7(10%)

* Included *pseudomonas aeruginosa* in three specimens, *Acinetobacter baumannii* in one specimen, *Klebsiella pneumonia* in one specimen

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caspopfungin were used as alternative agents. Patients with concurrent infections of CMV, bacteria or *aspergillus* were administered ganciclovir, cephalosporin, and itraconazole or voriconazole at the same time. Adjunctive corticosteroid was used in 58 (84%) of these patients. During the treatment period, a total of 23 (33%) patients developed shock. Of these patients, 20 (29%) required mechanical ventilation (MV), and 5 (7%) patients presented with pneumothorax. The 30-day in-hospital mortality rate was 26%, and the overall in hospital mortality rate was 32% (Fig 1). The deaths of 21 patients were attributed to PCP. One patient died from gastrointestinal bleeding.

Prognostic factors related to death

The univariate comparisons of the factors between survivors and non-survivors were shown in Table 3. There were no significant differences between age, gender, comorbidities, previous usage of immunosuppressive regimens, duration and dose of glucocorticoids, immunoglobulin (Ig) levels, and lactate dehydrogenase (LDH) levels. The lymphocyte count (575 \pm 766 cell//

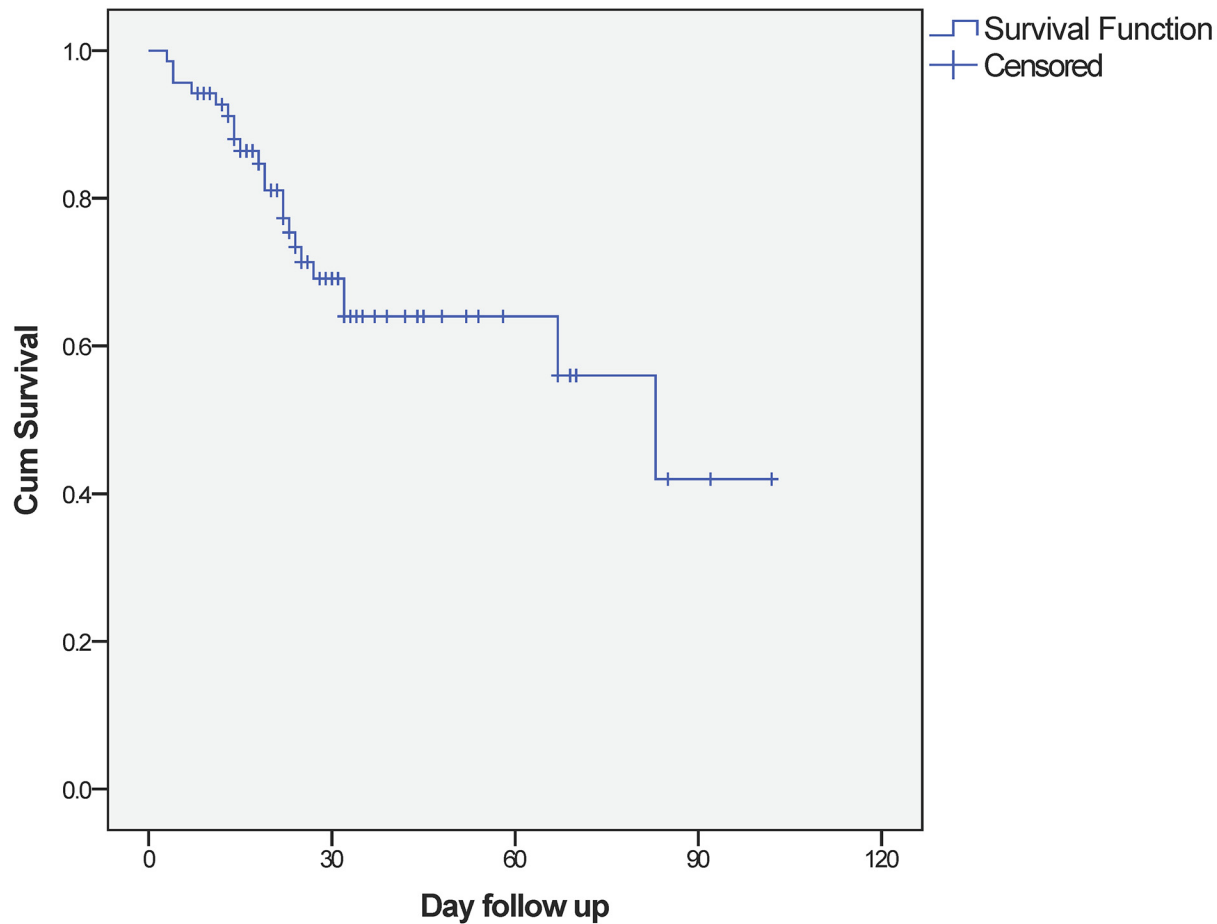


Fig 1. Kaplan-Meier survival curve for the patients with pneumocystis pneumonia.

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μl vs. 423 ± 304 cell/ μl among survivals and non-survivals, respectively, $P = 0.481$) and CD4+ T cell count (182 ± 209 cell/ μl vs. 122 ± 145 cell/ μl , $P = 0.236$) were higher in the surviving patients, but the results were not statistically significant. The rates of initial TMPco treatment failure in the survival and non-survival group were also not significantly different. A low PaO₂/FiO₂ ratio at diagnosis (248 ± 97 vs. 152 ± 62 , $P = 0.000$), shock during treatment (19% vs. 64%, $P = 0.001$), and use of MV (13% vs. 64%, $P = 0.000$) was related to poor prognosis. In addition, the low blood albumin level (32 ± 4 g/L vs. 27 ± 4 g/L, $P = 0.030$) and co-infection with *aspergillus* (4% vs. 27%, $P = 0.011$) were also associated with poor prognosis.

In multivariate analysis, all covariates with a P value < 0.05 (PaO₂/FiO₂ ratios, incidence of shock, MV, ALB level, *aspergillus* co-infection) were included in the regression analysis. The result showed that the PaO₂/FiO₂ ratios, mechanical ventilation, and ALB level were dependent predictors of mortality (Table 4).

Subgroup analysis of patients with SLE

Thirty-nine PCP patients with underlying SLE were analysed as a subgroup. The mortality rate in this group was 25.6% (10/39). Similar to the other CTD patients, a lower PaO₂/FiO₂ ratios

Table 3. Univariate analyses of risk factors among PCP patients determining survival rates.

Variables	Patients No. (%)		P value
	Survivors (n = 47)	Non-survivors (n = 22)	
Age, median years (range)	39(13–75)	41(14–78)	0.499
Male gender	18(38%)	7(32%)	0.498
Comorbidity			
Chronic renal disease	19(40%)	7(32%)	0.374
Idiopathic pulmonary fibrosis	3(6%)	4(18%)	0.140
Heart failure	2(4%)	0	1.000
Diabetes mellitus	5(11%)	3(14%)	0.703
Immunosuppressive agents	45(96%)	18(81%)	0.077
Previous days	319±751	196±422	0.731
Biological agents	4(9%)	1(5%)	1.000
Glucocorticoids dose (mg/d), mean ±SD*	49±22	49±34	0.779
Previous days	391±815	312±453	0.960
Severity of illness			
PaO ₂ /FiO ₂ ratios, mean ±SD	248±97	152±62	0.000
Shock	9(19%)	14(64%)	0.001
Mechanical ventilation	6(13%)	14(64%)	0.000
Pneumothorax	1(2%)	4(18%)	0.033
Laboratory findings			
WBC count, cell/ul, mean ±SD	6753±4043	8670±5380	0.338
Lymphocytes, cell/ul, mean ±SD	575±766	423±304	0.481
CD4+ T cell, cell/ul	182±209	122±145	0.236
Albumin, g/L, mean ±SD	32±4	27±4	0.003
Serum IgG, g/L, mean ±SD	9.66±5.66	8.82±4.14	0.909
Serum IgA, g/L, mean ±SD	1.76±0.66	2.36±0.90	0.153
Serum IgM, g/L, mean ±SD	1.07±0.45	0.88±0.37	0.354
LDH, U/L	488±148	563±213	0.264
Co-infections			
CMV	9(19%)	4(18%)	1.000
Bacterium	3(6%)	2(9%)	0.651
<i>Aspergillus</i>	2(4%)	6(27%)	0.011
Treatment			
Adjunctive steroid therapy	39(85%)	19(86%)	1.000
Use of TMPco	42(89%)	17(77%)	0.271
Not respond to TMPco	3(6%)	4(18%)	0.191

* Corticosteroids doses were expressed as the prednisolone equivalent dose

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Table 4. Multivariate analysis for Prognostic factors.

Variables	Adjusted OR	95%CI	P value
PaO ₂ /FiO ₂ ratios	1.005	1.002–1.009	0.006
Incidence of shock	0.589	0.240–1.444	0.247
Mechanical ventilation	1.676	0.642–4.380	0.292
Albumin	1.089	1.002–1.183	0.044
<i>Aspergillus</i> co-infection	0.679	0.342–5.199	0.679

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(234 ± 127 vs. 150 ± 73 , $P = 0.014$), incidence of shock (14% vs. 60%, $P = 0.009$), and usage of mechanical ventilation (7% vs. 80%, $P = 0.000$) were associated with poor prognosis in these patients. Interestingly, higher CD4+ T cell counts were found in the survival group (175 ± 128 vs. 75 ± 33 , $P = 0.024$). Higher proportions of non-survivors were female, but no statistical significance was found (51% vs. 90%, $P = 0.057$).

Discussion

PCP is an uncommon but fatal disease in patients with autoimmune disease. The clinical manifestations in these patients differ from those with HIV infection and the mortality rate is significantly higher (between 32% to 44% in previous reports) [8,14,15]. We accumulated the largest possible number of patients with CTD in China who developed PCP and analysed the symptoms, clinical courses, and prognosis factors in these patients. To our knowledge, this is the largest study performed on HIV-negative PCP patients with underlying autoimmune diseases.

In our study, all patients were under glucocorticoid therapy and most of them combined this with immunosuppressive agents. The most common patient radiologic features identified through CT scanning were ground glass opacity and reticulation [16]. The laboratory result showed that the lymphocyte count, CD4+ T cell count, and serum ALB were lower than normal and that the LDH level was elevated. WBC count and Ig levels were normal in these patients. These findings were similar with the previous report [12]. The co-infection rate in our study was higher than that in previous studies performed in HIV negative patients [10]. We thought the disturbance of the immune system in these patients and the usage of immunosuppressive agents might have caused these patients to become more susceptible to infections.

The overall mortality rate in our study was 32%, which is similar to that found in previously published studies [14,8]. Previous studies have demonstrated that high $D(A-a)O_2$ was associated with poor prognosis in non-HIV PCP patients [10,11]. In our study, PaO_2/FiO_2 ratios in the survival and non-survival groups were 248 ± 97 vs. 152 ± 62 respectively. The lower PaO_2/FiO_2 ratio, which was also a representative of ventilation-perfusion abnormality, was related to poor prognosis in CTD patients.

Hypoalbuminemia decreased plasma colloid osmotic pressure, which increased pulmonary vascular permeability and compromised the intravascular volume, placing the patient at risk for inadequate blood flow to vital organs. Previous studies showed that hypoalbuminemia had a positive correlation to increased lung injury and could be a significant indicator of mortality and morbidity in critically ill patients [17,18,19]. Lower ALB level is also reported to be associated with poor prognosis in PCP patients with CTD and HIV-negative patients [8,20]. But in a recent research performed by Kim et al., hypoalbuminemia was not considered as an independent predictor of mortality. In our study, the mean ALB level was higher in survivors than non-survivors. The difference was significant in both univariate and multivariate analysis. These findings suggest that ALB levels might be a predicting factor to be used in the diagnosis of PCP patients with CTD.

Incidence of shock and MV were reported to correlate with worse prognosis in non-HIV PCP patients [11]. In our study, the incidence of shock and MV rate also differed significantly in the survival and non-survival groups. These factors reflect the seriousness of diseases including organ failure, poor general condition and the need of intensive care, which made the patients more vulnerable to in hospital infections and other complications. Although these events were not identified as independent risk factors in multivariate analysis, more attention should be paid to patients with these conditions.

Aspergillus infection is one of the most common invasive fungal infections in SLE [21]. Patients with active disease and under high doses of corticosteroids (>60 mg/day) are more

vulnerable to *aspergillus* infection [21]. Because of the state of immunosuppression, mixed infection was common in these patients and the mortality rate was high. In our study, we also found that *aspergillus* co-infection was associated with higher mortality in univariate analysis. Due to the low incidence of combined *aspergillus* infection, the relationship was found to be not significant in multivariate analysis.

Despite presenting intolerance and adverse events, TMPco is still the first-line therapeutic regimen for *P. jirovecii* pneumonia [22,23,24]. The failure of initial TMPco treatment has been reported as a poor prognosis factor in HIV-negative patients. In our study, 86% of patients were initially treated with TMPco, and first-line therapy failure was observed in 12% of the patients. However, this study did not show any differences in the TMPco failure rate between the survival and non-survival groups. We hypothesized that this finding may be due to the limited sample size, intensive monitoring of the patient's symptoms and a prompt change to the second-line regimen (clindamycin, primaquine and/or caspofungin).

As a retrospective study, our study has several limitations. First, the incidence of SLE in our study is much higher than the incidence in other CTDs in China. SLE was thus the most common underlying autoimmune disease in our study. Second, some patients were diagnosed by PCR based assay. This diagnostic procedure is more sensitive than microscopic examination but cannot distinguish between infection and colonization. However, we used only bronchoalveolar lavage fluids specimens and performed the test in patients with clinical symptoms and radiologically compatible manifestations of PCP, which may have elevated the positive predictive value and diagnostic accuracy of this procedure [25,26]. Third, because it was a retrospective study, some factors that may have influenced the outcome of PCP were unavailable, such as LDH and C-reactive protein. The pneumonia severity index, the treatment delay and disease activity of underlying CTD could not be evaluated well. Furthermore, our study was carried out in a single institution. As the largest rheumatic autoimmune treatment centre in China, our hospital may have CTD patients with diseases that are more serious and with more comorbidities. The selection bias may have influenced the significance of our findings. A future prospective study of Chinese patients with a larger sample size is still required.

Conclusion

In summary, this study found that the mortality rate is high in PCP patients with underlying autoimmune disease in China. A lower PaO₂/FiO₂ ratios and ALB level were dependent predictors of mortality.

Supporting Information

S1 Table. Clinical characters and diagnostic procedure.

(XLSX)

S2 Table. Radiologic characters in CT scanning.

(XLSX)

S3 Table. Comorbidities, treatment and outcome.

(XLSX)

S4 Table. Laboratory findings and co-infections.

(XLSX)

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Author Contributions

Conceived and designed the experiments: MC XT K-FX MW. Analyzed the data: JZ MC. Contributed reagents/materials/analysis tools: JZ MC XTFQ. Wrote the paper: MC XT. Made the confirmation of all patients’ CTD diagnosis: JL.

References

1. Sepkowitz KA. Opportunistic infections in patients with and patients without Acquired Immunodeficiency Syndrome. *Clin Infect Dis*. 2002; 34(8):1098–107.
2. Mansharamani NG, Garland R, Delaney D, Koziel H. Management and outcome patterns for adult *Pneumocystis carinii* pneumonia, 1985 to 1995: comparison of HIV-associated cases to other immunocompromised states. *Chest*. 2000; 118(3):704–11. PMID: [10988192](#)
3. Wang XL, Wang XL, Wei W, An CL. Retrospective study of *Pneumocystis pneumonia* over half a century in mainland China. *J Med Microbiol*. 2011; 60(Pt 5):631–8.
4. Ainoda Y, Hirai Y, Fujita T, Isoda N, Totsuka K. Analysis of clinical features of non-HIV *Pneumocystis jirovecii* pneumonia. *J Infect Chemother*. 2012; 18(5):722–8. doi: [10.1007/s10156-012-0408-5](#) PMID: [22460829](#)
5. Takeuchi T, Kameda H. The Japanese experience with biologic therapies for rheumatoid arthritis. *Nat Rev Rheumatol*. 2010 Nov; 6(11):644–52. doi: [10.1038/nrrheum.2010.154](#) PMID: [20877307](#)
6. Li J, Huang XM, Fang WG, Zeng XJ. *Pneumocystis carinii* pneumonia in patients with connective tissue disease. *J Clin Rheumatol*. 2006; 12(3):114–7.
7. Tasaka S, Tokuda H. *Pneumocystis jirovecii* pneumonia in non-HIV-infected patients in the era of novel immunosuppressive therapies. *J Infect Chemother*. 2012; 18(6):793–806. doi: [10.1007/s10156-012-0453-0](#) PMID: [22864454](#)
8. Aoki Y, Iwamoto M, Kamata Y, Nagashima T, Yoshio T, Okazaki H, et al. Prognostic indicators related to death in patients with *Pneumocystis pneumonia* associated with collagen vascular diseases. *Rheumatol Int*. 2009; 29(11):1327–30. doi: [10.1007/s00296-009-0857-z](#) PMID: [19142640](#)
9. Vananuvat P, Suwannalai P, Sungkanuparph S, Limsuwan T, Ngamjanyaporn P, Janwityanujit S. Primary prophylaxis for *Pneumocystis jirovecii* pneumonia in patients with connective tissue diseases. *Semin Arthritis Rheum*. 2011; 41(3):497–502. doi: [10.1016/j.semarthrit.2011.05.004](#) PMID: [21959291](#)
10. Kim SJ, Lee J, Cho YJ, Park YS, Lee CH, Yoon HI, et al. Prognostic factors of *Pneumocystis jirovecii* pneumonia in patients without HIV infection. *J Infect*. 2014; 69(1):88–95. doi: [10.1016/j.jinf.2014.02.015](#) PMID: [24607411](#)
11. Ko Y, Jeong BH, Park HY, Koh WJ, Suh GY, Chung MP, et al. Outcomes of *Pneumocystis pneumonia* with respiratory failure in HIV-negative patients. *J Crit Care*. 2014; 29(3):356–61. doi: [10.1016/j.jcrc.2013.12.005](#) PMID: [24440053](#)
12. Hardak E, Neuberger A, Yigla M, Berger G, Finkelstein R, Sprecher H, et al. Outcome of *Pneumocystis jirovecii* pneumonia diagnosed by polymerase chain reaction in patients without human immunodeficiency virus infection. *Respirology*. 2012; 17(4):681–6. doi: [10.1111/j.1440-1843.2012.02158.x](#) PMID: [22390188](#)
13. Guo F, Chen Y, Yang S-L, Xia H, Li X-W, Tong Z-H. *Pneumocystis Pneumonia* in HIV-Infected and Immunocompromised Non-HIV Infected Patients: A Retrospective Study of Two Centers in China. *PLoS ONE*. 2014; 9(7):e101943. doi: [10.1371/journal.pone.0101943](#) PMID: [25029342](#)
14. Roblot F, Godet C, Le Moal G, Garo B, FaouziSouala M, Dary M, et al. Analysis of underlying diseases and prognosis factors associated with *Pneumocystis carinii* pneumonia in immunocompromised HIV-negative patients. *Eur J Clin Microbiol Infect Dis*. 2002; 21(7):523–31.
15. Godeau B, Coutant-Perronne V, Le ThiHuong D, Guillevin L, Magadur G, De Bandt M, et al. *Pneumocystis carinii* pneumonia in the course of connective tissue disease: report of 34 cases. *J Rheumatol*. 1994; 21(2):246–51. PMID: [8182632](#)
16. Vogel MN, Vatlach M, Weissgerber P, Goepfert B, Claussen CD, Hetzel J, et al. HRCT-features of *Pneumocystis jirovecii* pneumonia and their evolution before and after treatment in non-HIV immunocompromised patients. *Eur J Radiol*. 2012; 81(6):1315–20. doi: [10.1016/j.ejrad.2011.02.052](#) PMID: [21420818](#)

17. Aman J, van der Heijden M, van Lingen A, Girbes AR, van NieuwAmerongen GP, van Hinsbergh VW, et al. Plasma protein levels are markers of pulmonary vascular permeability and degree of lung injury in critically ill patients with or at risk for acute lung injury/acute respiratory distress syndrome. *Crit Care Med*. 2011; 39(1):89–97. doi: [10.1097/CCM.0b013e3181feb46a](https://doi.org/10.1097/CCM.0b013e3181feb46a) PMID: [21057316](https://pubmed.ncbi.nlm.nih.gov/21057316/)
18. Dubois MJ, Orellana-Jimenez C, Melot C, De Backer D, Berre J, Leeman M, et al. Albumin administration improves organ function in critically ill hypoalbuminemic patients: A prospective, randomized, controlled, pilot study. *Crit Care Med*. 2006; 34(10):2536–40. PMID: [16915107](https://pubmed.ncbi.nlm.nih.gov/16915107/)
19. Tiwari LK, Singhi S, Jayashree M, Baranwal AK, Bansal A. Hypoalbuminemia in critically sick children. *Indian J Crit Care Med*. 2014; 18(9):565–9. doi: [10.4103/0972-5229.140143](https://doi.org/10.4103/0972-5229.140143) PMID: [25249740](https://pubmed.ncbi.nlm.nih.gov/25249740/)
20. Ewig S, Bauer T, Schneider C, Pickenhain A, Pizzulli L, Loos U, et al. Clinical characteristics and outcome of *Pneumocystis carinii* pneumonia in HIV-infected and otherwise immunosuppressed patients. *Eur Respir J*. 1995; 8(9):1548–53.
21. Wang LR, Barber CE, Johnson AS, Barnabe C. Invasive fungal disease in systemic lupus erythematosus: a systematic review of disease characteristics, risk factors, and prognosis. *Semin Arthritis Rheum*. 2014; 44(3):325–30. doi: [10.1016/j.semarthrit.2014.06.001](https://doi.org/10.1016/j.semarthrit.2014.06.001) PMID: [25129259](https://pubmed.ncbi.nlm.nih.gov/25129259/)
22. Thomas CF Jr, Limper AH. *Pneumocystis pneumonia*. *N Engl J Med*. 2004; 350(24):2487–98. PMID: [15190141](https://pubmed.ncbi.nlm.nih.gov/15190141/)
23. Catherinot E, Lanternier F, Bougnoux ME, Lecuit M, Couderc LJ, Lortholary O. *Pneumocystis jirovecii* Pneumonia. *Infect Dis Clin North Am*. 2010; 24(1):107–38. doi: [10.1016/j.idc.2009.10.010](https://doi.org/10.1016/j.idc.2009.10.010) PMID: [20171548](https://pubmed.ncbi.nlm.nih.gov/20171548/)
24. Gilroy SA, Bennett NJ. *Pneumocystis pneumonia*. *Semin Respir Crit Care Med*. 2011; 32(6):775–82.
25. Orsi CF, Bettua C, Pini P, Venturelli C, La Regina A, Morace G, et al. Detection of *Pneumocystis jirovecii* and *Aspergillus* spp. DNA in bronchoalveolar lavage fluids by commercial real-time PCR assays: comparison with conventional diagnostic tests. *New Microbiol*. 2015; 38(1):75–84. PMID: [25742150](https://pubmed.ncbi.nlm.nih.gov/25742150/)
26. Robert-Gangneux F, Belaz S, Revest M, Tattevin P, Jouneau S, Decaux O, et al. Diagnosis of *Pneumocystis jirovecii* pneumonia in immunocompromised patients by real-time PCR: a 4-year prospective study. *J Clin Microbiol*. 2014; 52(9):3370–6.