

RESEARCH

Open Access

# CO<sub>2</sub> flagging - an improved method for the collection of questing ticks

Călin M Gherman<sup>1</sup>, Andrei D Mihalca<sup>1\*</sup>, Mirabela O Dumitrache<sup>1</sup>, Adriana Györke<sup>1</sup>, Ioan Oroian<sup>2</sup>, Mignon Sandor<sup>2</sup> and Vasile Cozma<sup>1</sup>

## Abstract

**Background:** Most epidemiological studies on tick-borne pathogens involve collection of ticks from the environment. An efficient collection method is essential for large sample pools. Our main aim was to evaluate the efficacy of a new method, where traditional flagging was enhanced by the use of CO<sub>2</sub> dispersed into the white flannel. The CO<sub>2</sub> was spread through a rubber hose network inserted into the flag blanket. The research was conducted in spring, in March-April 2011 in two locations from Cluj County, Romania.

**Methods:** The research was conducted in March-April 2011 in two locations from Cluj County, Romania. The flag to be tested contained a fine silicone rubber hose network which dispersed the CO<sub>2</sub> in the shaft. On each collection site n=30 samplings were performed. Each sampling consisted in the simultaneous use of both flags (with and without CO<sub>2</sub>) by two persons. The CO<sub>2</sub> concentration level on the flag canvas surface was measured. The efficacy of the method was determined by counting comparatively the total number of ticks and separate developmental stage count.

**Results:** Using the CO<sub>2</sub> improved flag, 2411 (59%) *Ixodes ricinus* and 100 (53.8%) *Dermacentor marginatus* ticks were captured, while the CO<sub>2</sub>-free flag accounted for the collection of 1670 *I. ricinus* (41%) and 86 (46.2%) *D. marginatus* ticks. The addition of CO<sub>2</sub> prompted a concentration difference on the surface of the flag ranging between 756.5 and 1135.0 ppm with a mean value of 848.9 ppm.

**Conclusion:** The study showed that the CO<sub>2</sub> enhanced sweep flag increased the ability of *I. ricinus* ( $p < 0.001$ ) but not of *D. marginatus* to be attracted to the flag blanket.

**Keywords:** Flagging, Carbon dioxide, Questing ticks, *Ixodes ricinus*

## Background

Ticks (suborder Ixodida) are obligate blood-sucking acarines attacking a wide variety of hosts from all tetrapod vertebrate classes [1,2]. Around 700 species of hard ticks are currently recognized as valid species [1]. Most of these species are three-host ticks (i.e. each stage detaches after engorgement) [3,4]. Regardless of the number of hosts, each tick must find a suitable host. In three-host ticks, most of their multiannual life is not spent attached to the host but as free-living organisms. Thus, newly hatched larvae, unfed nymphs and unfed

adults are in a permanent host finding state. Host detection and attachment in Ixodidae is achieved through three main alternative behavioral patterns: questing, hunting and tick-host cohabitation (nidicolous ticks) [4].

Most epidemiological studies on tick-borne pathogens involve collection of ticks from the environment [5]. Thus, an efficient collection method is essential for large datasets. Tick collection methods had been reviewed by Gray [6]. He divided these methods into four major categories: (1) flagging or dragging methods; (2) trapping using carbon dioxide baits; (3) collecting from hosts and (4) walking (i.e. on the clothes of the collectors). Despite all these methods are relative and do not estimate density (number per unit area) or absolute size (total number as measured in mark-release-recapture methods) [7], each of these has a variable efficacy depending on several

\* Correspondence: amihalca@usamvcluj.ro

<sup>1</sup>Department of Parasitology and Parasitic Diseases, University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, Calea Mănăștur 3-5, Cluj-Napoca 400372, Romania

Full list of author information is available at the end of the article

factors (i.e. habitat type, tick species, developmental stage etc.). Nevertheless, all methods have been improved over the time in order to increase their efficacy [8].

One of the most important ticks species (regarding its range, abundance, and vectorial importance) in the Palearctic region, with tendency to expand its spread in Northern Europe [9,10], is *Ixodes ricinus* [11]. The host seeking strategy of all developmental stages in *I. ricinus* is questing, when ticks are typically positioned on the vegetation with their legs extended, waiting for a moving host to which they attach [12]. Though, questing is a complex behavioral process, which involves responses to stimuli like host movement, concentration of environmental carbon dioxide and increase of temperature [4]. The most commonly used method for collection of questing ticks is flagging. However, flagging stimulates only the tick sensor for movement, and leaves the other two sensorial components of questing (i.e. carbon dioxide and temperature) unexploited.

The vast majority of ecological and epidemiological studies of tick-borne pathogens involve collection of unfed ticks from the environment. In this view, our main aim was to evaluate the efficacy of a new method, where traditional flagging was enhanced by the use of dispersed CO<sub>2</sub> into the white flannel.

## Methods

### Sweep and flag design

The sweep consists of a shaft and a flag. The shaft is constructed from a hollow aluminum tube, and the flag from white technical flannel. A JBL 500 g CO<sub>2</sub> bottle with a CO<sub>2</sub> solenoid regulator attached (both aquarium use, JBL Aquarium®, Germany) were fixed with plastic lock seals on the shaft. A silicone rubber hose was attached to the CO<sub>2</sub> solenoid regulator; the hose was introduced through the aluminum shaft and connected to the flag. The rubber hose was pierced (to release CO<sub>2</sub>) and attached to the flag by sewing forming a network structure (Figure 1). The hose was made of bending-resistant silicone rubber with the inner diameter of 1 mm and the outer diameter of 2 mm with a wall thickness of 0.5 mm.

The flag surface area was 0.48 m<sup>2</sup> (80 x 60 cm) to allow unrestricted passage across all types of vegetation. Two identical flags were made; one of them with CO<sub>2</sub> and the other without CO<sub>2</sub> (control).

### Study area

Two hilly areas were chosen: Vultureni and Faget, both in Cluj county (Figure 2), according to preliminary results of sampling for the evaluation of the presence of tick-borne pathogens in Romania (manuscript under preparation). The habitats consisted in herbaceous

vegetation alternating with small shrubs, located at the edge of woods, specific areas for ticks. The climate is moderate continental, influenced by the vicinity of the Apuseni Mountains and Atlantic influences from west of the country, in autumn and winter [13]. The study was conducted in spring, between the end of March and the end of April 2011, as tick abundance is higher in North-western Romania in this season [14]. The GPS was used to measure the distance.

### Sampling procedure

On each collection site n = 30 samplings were performed (total 60 samplings in the two sites). Each sampling consisted in the simultaneous use of both flags, (with and without CO<sub>2</sub>), on 2 m wide adjacent areas, by two persons who had interchanged the sweep every fifty meters for the homogeneity of results. After each 5 m the flags were checked for ticks. All ticks were collected regardless their species and fixed in pure ethanol. Specific identification was performed using morphological keys [15] under a binocular microscope.

### Determination of carbon dioxide

The CO<sub>2</sub> concentration level on the flag canvas surface was measured using a portable CIRAS-2 Photosynthesis System equipped with a SRC-1 Soil Respiration Chamber (PP Systems International Inc®, USA). CO<sub>2</sub> level was determined at a single time, but from several points of the flag (n = 20) according to instruction manual and recommendations [16]. The results were expressed in ppm as compared to the standard CO<sub>2</sub> concentration of 393.71 ppm (reference data for 24.04.2011) [16].

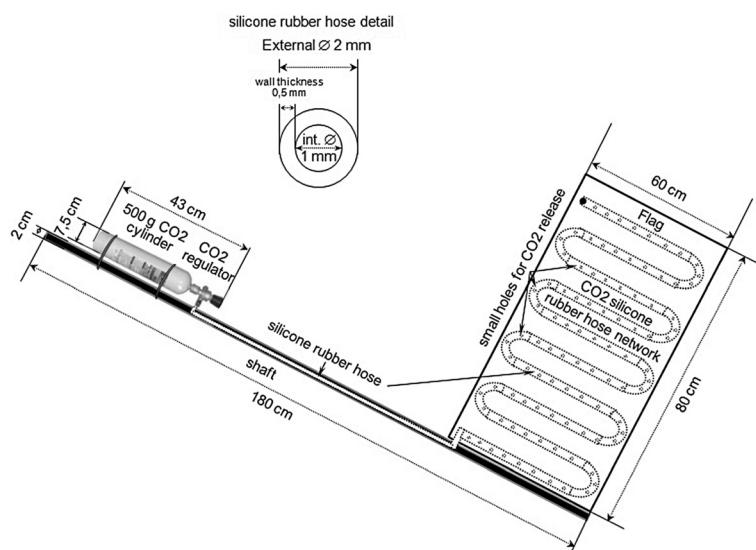
### Statistical analysis

The efficacy of the method was determined by counting the total number of ticks and separate developmental stage count, attached to the CO<sub>2</sub> flag compared with the control (CO<sub>2</sub>-free flag). For statistical analysis, the values were compared in CHI-SQUARE TEST [17]. A p value of <0.05 was considered statistically significant. The relative risk (RR) is a ratio of the probability of the event occurring in the exposed group versus a non-exposed group [18].

## Results

A total number of 4267 of ticks belonging to two species were collected in the 60 samplings: *I. ricinus* (n = 4081) and *Dermacentor marginatus* (n = 186).

*I. ricinus* accounted for 4081 tick captures (adults, nymphs and larvae) were collected in the 60 samplings (Table 1). Of these, 2617 (64.1%) were adults, 1422 (34.9%) nymphs and 42 (1%) larvae. Using CO<sub>2</sub> improved flag were captured 2411 (59%) ticks and 1670 (41%) without CO<sub>2</sub>.



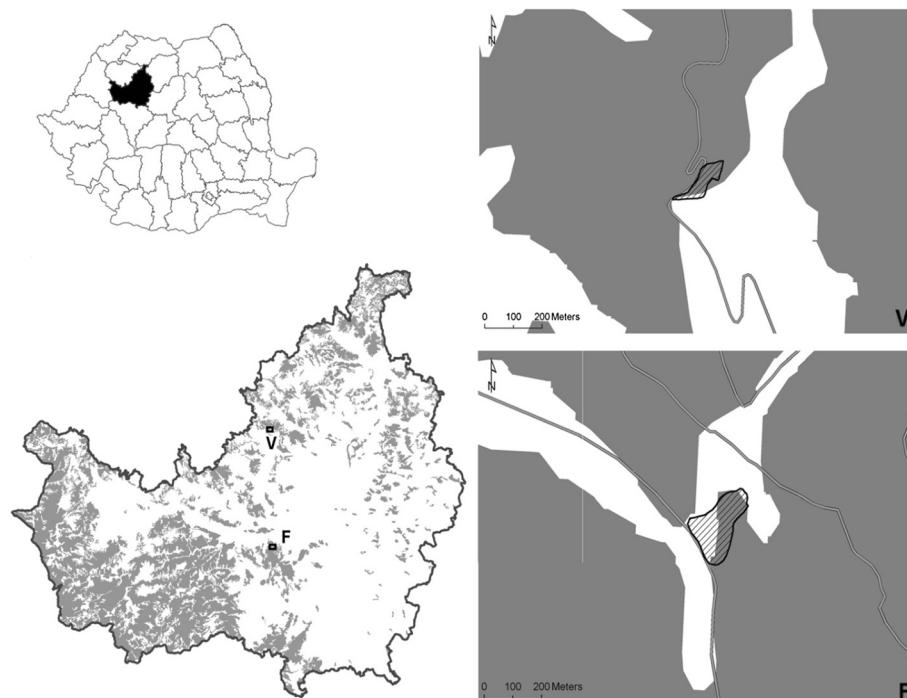
**Figure 1** Sweep CO<sub>2</sub> flag diagram.

The statistical analysis revealed highly statistically significant ( $p < 0.001$ ) difference between the two variables in adults and nymphs, in both locations and overall; for larvae, the recorded statistical differences were not significant (Table 2). Carbon dioxide flagging was more effective than CO<sub>2</sub>-free flagging, with average values of RR ranging between 1.4 and 1.6 for adults and nymphs.

The addition of CO<sub>2</sub> prompted a difference on the surface of the flag ranging between 756.5 and 1135.0 ppm with a mean value of 848.9 ppm.

## Discussion

Flagging is the most widespread tick collection method. Over time, several improvements to this method were



**Figure 2** Field studies location (shaded area shows the flagging surface).

**Table 1 Number of questing *Ixodes ricinus* ticks collected by flagging with and without CO<sub>2</sub>**

Location/stage/method	Faget		Vultureni		Total (both locations)		
	IR	DM	IR	DM	IR	DM	
<b>Males</b>	CO <sub>2</sub> +	456	27	211	22	667	49
	CO <sub>2</sub> -	325	21	160	20	485	41
<b>Females</b>	CO <sub>2</sub> +	587	26	281	25	868	51
	CO <sub>2</sub> -	407	23	190	22	597	45
<b>Nymphs</b>	CO <sub>2</sub> +	547	0	304	0	851	0
	CO <sub>2</sub> -	353	0	218	0	571	0
<b>Larvae</b>	CO <sub>2</sub> +	22	0	3	0	25	0
	CO <sub>2</sub> -	16	0	1	0	17	0
<b>TOTAL</b>	CO <sub>2</sub> +	1612	53	799	47	2411	100
	CO <sub>2</sub> -	1101	44	569	42	1670	86
	Per location	2713	97	1368	89	4081	186

proposed [8] and now there is many types used: double walking flagging and double walking with baited flagging [19], walking flagging with a loose-fitting and white cotton flannel garment worn [20] and strip-flag method [21].

The successful hosts attack in *Ixodes* species expressed as the ability to adhere to a flannel flag is influenced by many factors: light and/or shadow, radiation heat (temperature), mechanical vibration of questing substrate, host odor, and CO<sub>2</sub> concentration [22].

Chemical mediators also known as semiochemicals are as well important for behavioral patterns in ticks. These information-bearing compounds are secreted by animals into the external environment, and when recognized they trigger a specific behavioral response such as food location, sexual partner location or escape [23]. Semiochemicals are categorized into four major categories: (1) pheromones; (2) allomones; (3) kairomones; and (4) synomones [24].

Carbon dioxide acts like an attractant kairomone for ticks [25]. Experimentally, it acted as an attracting agent causing almost immediate activation in the soft ticks *Ornithodoros coriaceus* quiescent ticks [25]. Adults of *Dermacentor andersoni* respond also very well to stimulation with carbon dioxide. Garcia, 1965, described a system based on release of CO<sub>2</sub> for the collection of *D. andersoni*. The system involved a piece of dry ice placed on a wire mesh platform in the desired area. The results indicate that the CO<sub>2</sub> method is more sensitive for detection of adult ticks than is the conventional flagging technique [26]. Carbon dioxide (dry ice) trapping method was demonstrated to be effective in the collection of some tick species: *I. ricinus* [5], *Amblyomma americanum* [27-29] and *A. hebraeum* [30]. Our method is important in Europe as it enhances the capture of *I. ricinus*, the main vector of *Borrelia burgdorferi* s.l. and the tick-borne encephalitis (TBE) virus [31].

Our results are consistent with those cited above. The number of adults and nymphs of *I. ricinus* collected was significantly increased using CO<sub>2</sub> enhanced blanket comparing with flagging without CO<sub>2</sub>. The lower number of larvae collected can be explained by the months of sampling, March-April, when larvae may be not fully active and by the quality of blanket, made by technical flannel, material which may not have reached the lower levels of the vegetation where larvae sit to quest.

These data show that the responsiveness to CO<sub>2</sub> is enhanced during host-seeking periods of the life cycle and reduced at other times [32]. However, others [29] did not establish significant differences between flagging method with a strip blanket and CO<sub>2</sub> traps or rabbits scent baits. The response time of ticks to host attachment was shown to be dependent on the tick species and CO<sub>2</sub> concentration in the environment [33]. In *Amblyomma maculatum*, *A. americanum* and *Dermacentor variabilis*,

**Table 2 Statistical significance of tick collection efficacy using flagging with and without CO<sub>2</sub>**

Location	Faget		Vultureni		Total				
	<i>I. ricinus</i>	<i>D. marginatus</i>		<i>I. ricinus</i>	<i>D. marginatus</i>		<i>I. ricinus</i>	<i>D. marginatus</i>	
		P	RR		P	RR		P	RR
<b>Males</b>	<b>CO<sub>2</sub>+</b>	0.0001	0.31	0.0002	0.83	0.0001	0.30		
	<b>CO<sub>2</sub>-</b>	1.40 (1.27–1.56)	1.29 (0.86–1.91)	1.32 (1.14–1.53)	1.1 (0.72–1.69)	1.38 (1.27–1.50)	1.20 (0.89–1.61)		
<b>Females</b>	<b>CO<sub>2</sub>+</b>	0.0001	0.69	0.0001	0.68	0.0001	0.47		
	<b>CO<sub>2</sub>-</b>	1.44 (1.32–1.58)	1.13(0.76–1.68)	1.48(1.30–1.67)	1.14(0.76–1.71)	1.45(1.35–1.57)	1.13(0.85–1.51)		
<b>Nymphs</b>	<b>CO<sub>2</sub>+</b>	0.0001		0.0001		0.0001			
	<b>CO<sub>2</sub>-</b>	1.6 (1.4–1.7)		1.4 (1.2–1.6)		1.5 (1.4–1.6)			
<b>Larvae</b>	<b>CO<sub>2</sub>+</b>	0.25		0.48		0.13			
	<b>CO<sub>2</sub>-</b>	1.4 (0.9–2.2)		3 (0.5–18)		1.5 (0.9–2.3)			

the groups preconditioned with low ambient CO<sub>2</sub> (422 ppm) always produced response times of longer duration than ticks preconditioned to high ambient CO<sub>2</sub> (956 ppm) [34].

The lack of statistical significance for *D. marginatus* between the two collecting methods compared in the present study might be caused by the major differences in the sample size. The small total number of *D. marginatus* collected (regardless the method used) can be explained by the typical area of this species which prefers biotopes characterized by xerophilic plant communities: dry pasture shrub communities, grazing black locust forests (*Robinietum*), forest-steppe or grikis (*Quercetum pubescens* and *Cometo-Quercetum*), margins of oak forests and bushy ridges between the fields and field paths [35]. Our study area is characterized by deforested hills and covered with low vegetation; predominant species in forest areas are hornbeam, birch, poplar, hazel, elm, ash and maple. Although *D. marginatus* is almost as widespread in Romania as *I. ricinus*, [36], the population density is significantly lower in most of the sampled localities [37]. Regarding seasonality, in Eastern Europe, *D. marginatus* is most numerous in February and March [38] and most of our sampling was done in April.

Flagging technique seems to work better for other species: *Ixodes dammini* [27], *I. pacificus*, *D. occidentalis* and *D. variabilis* [32], *I. rubicundus* [39] or *I. ricinus* [40]. Four different methods of surveying were tested for *D. variabilis* and *I. banksi* and it was shown that the most successful was flagging, compared with carbon dioxide trapping, nest boxes and collection from hosts [41].

Concerning the temperature, *I. persulcatus* is a more cold-resistant tick than *I. ricinus* and it is more successful both in adhering to the flag and in remaining attached to it at two ranges: 6–10°C and 17–22°C [42]. In general a greater percentage of *I. rubicundus* displayed an appetence response at lower (12, 17 and 21°C) than at high (30°C) temperatures [39]. It is known that all life stages of *I. ricinus* are equipped to sense shifts in light intensity. This allows *I. ricinus* to use onset of darkness to trigger mobility when desiccation risk is reduced in nature [43]. The nymphs are stimulated to walk horizontally by humidity and host scent. When the atmosphere is sufficiently wet they are likely to walk towards odor secreted by host skin [44,45]. The larvae of *I. hirsti* seem to be more sensitive to shade and heat, while they were unresponsive to CO<sub>2</sub> concentration and host odor [46]. Radiation heat and shadowing caused the greatest percentage of *I. rubicundus* to display an appetence response; shadowing and radiation heat had the least effect on *R. punctatus* [22]. A single mechanical perturbation of the substratum caused a mean of 50% of *I. rubicundus*

to display an appetence response. Constant mechanical perturbation resulted in a progressive decrease in the proportion of ticks reacting [22]. Host scent is known to initiate questing behavior in *I. persulcatus*, *I. ricinus* and *I. crenulatus*. Both *I. ricinus* and *I. crenulatus* respond strongly to sheep wool [47,48].

Entire-blanket flagging is a better sampling method for *I. ricinus* comparing with others variants of flagging or dragging. Significantly more nymphs and adults were caught by the entire-blanket versus strip-blanket flagging [49]. Flagging was 1.5–1.7 times as effective as dragging; impregnation of the cloths with different substances, like host odor, increased the efficacy by 2.4 (dragging) to 2.8 (flagging) times [39]. From several types of material used (i.e. cotton, woolen flannel, "molleton" - soft thick cotton, and toweling - spongecloth), the last was the best cloth type to optimize the number of ticks collected [36]. However, dry-ice-baited tick-traps is more effective for ticks with increased mobility, like *A. americanum* [27].

Our enhanced technique that combines, for the first time, classical flagging and carbon dioxide trapping methods improved significantly the ability of *I. ricinus* to adhere to flag blanket. By introducing CO<sub>2</sub> in the white blanket we tried to simulate the time approach of a host to the tick. Sudden increase of the CO<sub>2</sub> concentration in the air stimulates its vivacity and questing position, increasing the chance of attachment of ticks to the flag.

## Conclusion

The study showed that the CO<sub>2</sub> enhanced sweep flag increased the ability of *I. ricinus* ( $p < 0.001$ ) but not of *D. marginatus* to be attracted to the flag blanket.

## Competing interests

The authors declare that they have no competing interests.

## Acknowledgements

This research was financed by project PCCE 7/2010 by CNCSIS.

## Author details

<sup>1</sup>Department of Parasitology and Parasitic Diseases, University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, Calea Măñăstur 3-5, Cluj-Napoca 400372, Romania. <sup>2</sup>Department of Plant and Environmental Protection, University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, Calea Măñăstur 3-5, Cluj-Napoca 400372, Romania.

## Authors' contributions

GCM wrote the manuscript, performed flagging. MAD concept idea, performed the flagging. DMO identified the ticks. GA statistical analysis. OI measurement of CO<sub>2</sub>. SM measurement of CO<sub>2</sub>. CV team coordinator. All authors read and approved the final manuscript.

Received: 28 December 2011 Accepted: 21 June 2012

Published: 21 June 2012

## References

1. Guglielmone AA, Robbins RG, Apanaskevich DA, Petney TN, Estrada-Pena A, Horak IG, Shao RF, Barker SC: The Argasidae, Ixodidae and Nuttalliellidae (Acaria: Ixodida) of the world: a list of valid species name. Zootaxa 2010, 2528:1–28.

2. Keirans JE: Systematics of the Ixodida (Argasidae, Ixodidae, Nuttalliellidae): An overview and some problems. In *Tick Vector Biology, Medical and Veterinary Aspects*. Edited by Fivaz B, Petney T, Horak I. Berlin: Springer Verlag; 1992:1–21.
3. Oliver JH Jr: Biology and systematics of ticks (Acar: Ixodida). *Annu Rev Ecol Syst* 1989, 20:397–430.
4. Petney NT, Robbins RG, Guglielmone AA, Apanaskevich DA, Estrada-Peña A, Horak IG, Shao R: *A Look at the World of Ticks*. In *Progress in Parasitology*. Edited by Mehlhorn H. Berlin: Springer Verlag; 2011:283–296.
5. Capelli G, Ravagnan S, Montarsi F, Ciocchetta S, Cazzin S, Porcellato E, Babiker AM, Cassini R, Salvato A, Cattoli G, Otranto D: Occurrence and identification of risk areas of *Ixodes ricinus*-borne pathogens: a cost-effectiveness analysis in north-eastern Italy. *Parasite Vector* 2012, 5:61.
6. Gray JS: A carbon dioxide trap for prolonged sampling of *Ixodes ricinus* L. populations. *Exp Appl Acarol* 1985, 1:35–44.
7. Braks M, Van der Giessen J, Kretzschmar M, Van Pelt W, Scholte EJ, Reusken C, Zeller H, Van Bortel W, Sprong H: Towards an integrated approach in surveillance of vector-borne diseases in Europe. *Parasite Vector* 2011, 4:192.
8. Carroll JF, Schmidtmann ET: Tick sweep: modification of the tick drag-flag method for sampling nymphs of the deer tick (Acar: Ixodidae). *J Med Entomol* 1992, 29(2):352–355.
9. Jore S, Viljujgrein H, Hofshagen M, Brun-Hansen H, Kristoffersen AB, Nygård K, Brun E, Ottesen P, Sævik BK, Ytrehus B: Multi-source analysis reveals latitudinal and altitudinal shifts in range of *Ixodes ricinus* at its northern distribution limit. *Parasite Vector* 2011, 4:84.
10. Jaenson TG, Jaenson DG, Eisen L, Petersson E, Lindgren E: Changes in the geographical distribution and abundance of the tick *Ixodes ricinus* during the past 30 years in Sweden. *Parasite Vector* 2012, 5:8.
11. Estrada-Peña A: Geostatistics as predictive tools to estimate *Ixodes ricinus* (Acar: Ixodidae) habitat suitability in the western Palearctic from AVHRR satellite imagery. *Exp Appl Acarol* 1999, 23:337–349.
12. Randolph SE: Tick ecology: processes and patterns behind the epidemiological risk posed by ixodid ticks as vectors. *Parasitol* 2004, 129: S37–S65.
13. Trusca V, Alecu M: Romania General Facts. In *Romania's Third National Communication on Climate Change under the United Nations Framework Convention on Climate Change*. Edited by Ministry of Environment and Water Management. Holstebro: Grue & Hornstrup; 2005:21–65.
14. Chițimia L: [Ecology of Ixodidae in Southwestern Romania] [in Romanian]. PhD Thesis. Timișoara: Universitatea de Științe Agricole și Medicină Veterinară a Banatului; 2006:283.
15. Feider Z: *[Fauna of the Popular Republic of Romania. Volume 5/2. Acaromorpha, Suprafamily Ixodoidea]* [in Romanian]. București: Editura Academiei Republicii Populare Române; 1965.
16. Conway T, Tans P: NOAA/ESRL. www.esrl.noaa.gov/gmd/ccgg/trends/. Accessed 30 november 2011.
17. Snedecor GW, Cochran WG: *Statistical Methods*. 8th edition. Ames: Iowa State University Press; 1989.
18. Localio AR, Margolis DJ, Berlin JA: Relative risks and confidence intervals were easily computed indirectly from multivariable logistic regression. *J Clin Epidemiol* 2007, 60(9):874–882.
19. Fernández-Ruvalcaba M, Jesus F, De La Torre P, Cordoba-Juarez G, García-Vazquez Z, Rosario-Cruz R, Saltijeral-Oaxaca J: Animal bait effect on the recovery of *Boophilus microplus* larvae from experimentally infested grass in Morelos, Mexico. *Parasitol Latinoam* 2003, 58:54–58.
20. Chapman ES, Siegle TE: A novel method for tick retrieval reveals northern migration of *Amblyomma americanum* into Pike County, IL, USA. *Int J Acarol* 2000, 26(4):391–393.
21. Gray JS, Lohan G: The development of a sampling method for the tick *Ixodes ricinus* and its use in a redwater fever area. *Ann Appl Biol* 1982, 101(3):421–427.
22. Fourie LJ, Snyman A, Kok DJ, Horak IG, Van Zyl JM: The appetence behaviour of two South African paralysis-inducing ixodid ticks. *Exp Appl Acarol* 1993, 17:921–930.
23. Sonenshine DE: Pheromones and other semiochemicals of ticks and their use in tick control. *Parasitology* 2004, 129:S405–S425.
24. Evenden ML, Judd GJR, Borden JH: A synomone imparting distinct sex pheromone communication channels for *Choristoneura rosaceana* (Harris) and *Pandemis limitata* (Robinson) (Lepidoptera: Tortricidae). *Chemoecology* 1999, 9:73–80.
25. Garcia R: Carbon dioxide as an attractant for certain ticks (Acarina: Argasidae and Ixodidae). *Ann Entomol Soc Am* 1962, 55(5):605–606.
26. Garcia R: Collection of *Dermacentor andersoni* (Stiles) with carbon dioxide and its application in studies of Colorado tick fever virus. *Am J Trop Med Hyg* 1965, 14(6):1090–1093.
27. Ginsberg HS, Ewing CP: Comparison of flagging, walking, trapping, and collecting from hosts as sampling methods for northern deer ticks, *Ixodes dammini*, and lone-star ticks, *Amblyomma americanum* (Acar: Ixodidae). *Exp Appl Acarol* 1989, 7:313–322.
28. Kinzer DR, Presley SM, Hair JA: Comparative efficiency of flagging and carbon dioxide-baited sticky traps for collecting the lone star tick, *Amblyomma americanum* (Acarina: Ixodidae). *J Med Entomol* 1990, 27(5):750–755.
29. Schulze TL, Jordan RA, Hung RW: Biases associated with several sampling methods used to estimate abundance of *Ixodes scapularis* and *Amblyomma americanum* (Acar: Ixodidae). *J Med Entomol* 1997, 34(6):615–623.
30. Anderson RB, Scrimgeour GJ, Kaufman WR: Responses of the tick, *Amblyomma hebraicum* (Acar: Ixodidae), to carbon dioxide. *Exp Appl Acarol* 1998, 22:667–681.
31. Dautel H, Kahl O: Ticks (Acar: Ixodoidea) and their medical importance in the urban environment. In *Proceedings of the Third International Conference on Urban Pests*: 19–22 July 1999. Edited by Robinson WH, Rettich F, Rambo GW. Prague; 1999:73–82.
32. Li X, Dunley JE: Optimal sampling and spatial distribution of *Ixodes pacificus*, *Dermacentor occidentalis* and *Dermacentor variabilis* ticks (Acar: Ixodidae). *Exp Appl Acarol* 1998, 22:233–248.
33. Gray JS: Studies on the larval activity of the tick *Ixodes ricinus* L. in Co. Wicklow, Ireland. *Exp Appl Acarol* 1985, 1:307–316.
34. Holscher KH, Gearhart HL, Barker RW: Electrophysiological responses of three tick species to carbon dioxide in the laboratory and field. *Ann Entomol Soc Am* 1980, 73(3):288–292.
35. Nosek J: The ecology, bionomics, behaviour and public health importance of *Dermacentor marginatus* and *D. reticulatus* ticks. *Wiad Parazytol* 1972, XVIII(4–5):721–725.
36. Mihalca AD, Dumitrache MO, Magdaș C, Gherman CM, Domşa C, Mircean V, Ghira IV, Pocora V, Ionescu DT, Sikó Barabási S, Cozma V, Sándor AD: Synopsis of the hard ticks (Acar: Ixodidae) of Romania with update on host associations and geographical distribution. *Exp Appl Acarol* 2012, doi:10.1007/s10493-012-9566-5.
37. Mihalca AD, Gherman CM, Magdas V, Dumitrache MO, Györke A, Sándor AD, Domşa C, Oltean M, Mircean V, Mărăcăianu DL, D'Amico G, Păduraru AO, Cozma V: *Ixodes ricinus* is the dominant questing tick in forest habitats in Romania: the results from a countrywide dragging campaign. *Exp Appl Acarol* 2012, doi:10.1007/s10493-012-9568-3.
38. Hornok S: Allochronic seasonal peak activities of *Dermacentor* and *Haemaphysalis* spp. under continental climate in Hungary. *Vet Parasitol* 2009, 163:366–369.
39. Fourie LJ, Van Der Lingen F, Kok DJ: Improvement of field sampling methods for adult Karoo paralysis ticks, *Ixodes rubicundus* (Acar: Ixodidae), through addition of host odour. *Exp Appl Acarol* 1995, 19:93–101.
40. Vassallo M, Pichon B, Cabaret J, Figureau C, Perez-Eid C: Methodology for sampling questing nymphs of *Ixodes Ricinus* (Acar: Ixodidae), the principal vector of lyme disease in Europe. *J Med Entomol* 2000, 37(3):335–339.
41. Keaney P: *The Effectiveness Of Various Trapping Techniques In Collecting Ixodes scapularis and Other Species of Ticks*, Bios 569 - Practicum in Aquatic Biology, 1651 Turtle Creek Ct. Advisor: Dr. Craig. 1994.
42. Uspensky I: Ability of successful attack in two species of ixodid ticks (Acar: Ixodidae) as a manifestation of their aggressiveness. *Exp Appl Acarol* 1993, 17:673–683.
43. Perret JL, Guerin PM, Diehl PA, Vlimant M, Gern L: Darkness induces mobility, and saturation deficit limits questing duration, in the tick *Ixodes ricinus*. *J Exp Biol* 2003, 206:1809–1815.
44. Crooks E, Randolph SE: Walking by *Ixodes ricinus* ticks: intrinsic and extrinsic factors determine the attraction of moisture or host odour. *J Exp Biol* 2006, 209:2138–2142.
45. Greenfield BPJ: Environmental parameters affecting tick (*Ixodes ricinus*) distribution during the summer season in Richmond Park, London. *Bioscience Horizons* 2011, 0(0):1–9.

46. Oorebeek M, Sharrad T, Kleindorfer S: What attracts larval *Ixodes hirsti* (Acaria: Ixodidae) to their host? *Parasitol Res* 2009, **104**:623–628.
47. Lees AD: The behaviour and physiology of ticks. *Acarol* 1969, **11**(3):397–410.
48. Elizarov YA, Vasewta AA: A distant orientation of blood-sucking ixodid ticks to the host's attractant factors. (in Russian). *Parazit Sb* 1976, **10**:136–141.
49. Tack W, Madder M, De Frenne P, Vanhellemont M, Gruwez R, Verheyen K: The effects of sampling method and vegetation type on the estimated abundance of *Ixodes ricinus* ticks in forests. *Exp Appl Acarol* 2011, **54**:285–292.

doi:10.1186/1756-3305-5-125

**Cite this article as:** Gherman et al.: CO<sub>2</sub> flagging - an improved method for the collection of questing ticks. *Parasites & Vectors* 2012 **5**:125.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

