

Eco-evolutionary dynamics modulate plant responses to global change depending on plant diversity and species identity

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Abstract Global change has dramatic impacts on grassland diversity. However, little is known about how fast species can adapt to diversity loss and how this affects their responses to global change. Here, we performed a common garden experiment testing whether plant responses to global change are influenced by their selection history and the conditioning history of soil at different plant diversity levels. Using seeds of four grass species and soil samples from a 14-year-old biodiversity experiment, we grew the offspring of the plants either in their own soil or in soil of a different community, and exposed them either to drought, increased nitrogen input, or a combination of both. Under nitrogen addition, offspring of plants selected at high diversity produced more biomass than those selected at low diversity, while drought neutralized differences in biomass production. Moreover, under the influence of global change drivers, soil history, and to a lesser extent plant history, had species-specific effects on trait expression. Our results show that plant diversity modulates plant-soil interactions and growth strategies of plants, which in turn affects plant eco-evolutionary pathways. How this change affects species' response to global change and whether this can cause a feedback loop should be investigated in more detail in future studies.

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Editor's evaluation

Dietrich et al. aimed to test the hypothesis that a decline in species richness due to various global change drivers selects for traits that will make species more vulnerable to the further effects of these drivers, amplifying thus the initial diversity decline. This research is of prime importance to botanists, plant ecologists, and ecosystem ecologists wanting to model the effects of global climate change on plant diversity and productivity.

Introduction

Human activities, such as the combustion of fossil fuels and the intensification of agriculture, are leading to global environmental changes, causing increased air temperatures, altered precipitation patterns, and rising amounts of nitrogen to ecosystems (IPCC, **Pörtner et al., 2021**). The consequences are more frequent extreme weather events such as droughts (**Dai et al., 2018**) and a growing accumulation of nitrogen in the soils (**Holland et al., 2005**). Both, drought and increased nitrogen

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eLife digest Over the last hundred years, human activities including burning of fossil fuels, clearing of forests, and fertilizer use have caused environmental changes that have resulted in many species of plants, animals and other forms of life becoming extinct.

Loss of plant species can change the local environment by, for example, altering the availability of nutrients and local communities of microbes in the soil. This may, in turn, cause remaining plant species to develop differently: they may take up fewer resources or become more prone to pathogens, both of which may alter their physical appearance. However, little is known about whether this happens and, if so, how rapidly such changes occur.

Since 2002, researchers in Germany have been running a long-term project known as the Jena Experiment to study how plants behave when they grow in communities with different numbers of other plant species. For the experiment, various species of grass and other plants commonly found in grasslands were grown together in different combinations. Some plots contained many species (referred to as "high diversity") and others contained only a few ("low diversity").

Here, Dietrich et al. collected seeds from four grasses grown for 12 years in Jena Experiment plots with two or six plant species. The seeds were then transferred to pots and grown in a greenhouse using soil either from the plot where the seeds originated or from another plot with a different diversity level. To simulate human-made changes in the environment, the team added nitrogen fertilizer or decreased how much they watered some of the plants.

The greenhouse experiment showed that after receiving nitrogen fertilizer, the seeds from the high diversity Jena Experiment plots grew into larger plants than the seeds from the low diversity plots. But there was no difference in size when the plants were watered less. Moreover, both fertilizer and watering treatment had different effects on the plants' physical appearance (root and leaf architecture) depending on the soil in which they were growing in.

The findings of Dietrich et al. suggest that plants may respond differently to changes in their environment based on their origins and the soil they are growing in. This study provides the first indication that species loss could accelerate a further loss of species due to changes in how the plants develop and the communities of organisms living in the soil.

input, in turn, further influence ecosystems and climatic conditions; hence, they are known as major global change drivers (*Sage*, *2020*).

Some of the most tremendous negative effects of global change are changes in ecological communities (Dornelas et al., 2014) and the extinction of species (Sage, 2020), whereby plant species are particularly affected due to their low mobility, with drastic consequences for the functioning of ecosystems. Studies in grassland biodiversity experiment have shown that low- and high-diversity plant communities significantly differ in their productivity and stability (Isbell et al., 2015; Marguard et al., 2009; Tilman et al., 2006). Low-diversity communities were shown to lose productivity over time, while high-diversity communities are more stable, so that plant diversity-productivity relationships become more positive over time (Cardinale et al., 2007; Meyer et al., 2016; Reich et al., 2012). A different development of plant-soil and plant-plant interactions at low and high diversity are assumed to be the important drivers of these strengthening biodiversity effects (Eisenhauer et al., 2019; Thakur et al., 2021). At low plant diversity, an accumulation of soil-borne pathogens might be responsible for lower plant community productivity (Mommer et al., 2018; Thakur et al., 2021), while in high-diversity communities, complementarity effects among plants and an accumulation of soilborne mutualists inhibit such negative processes, causing a higher productivity of these plant communities (Barry et al., 2019; Cardinale et al., 2007; Reich et al., 2012). Next to these biotic drivers, also diversity-dependent differences in abiotic conditions can influence plant community productivity (Barry et al., 2019). Previous studies demonstrated that a decrease of species richness alters the vegetation structure and density (Lorentzen et al., 2008), which in turn can have strong impacts on the availability of light, water, and nutrients for plants (Bachmann et al., 2018; Fischer et al., 2018; Lange et al., 2019).

Consequently, these findings raise the question, whether populations of the same plant species develop differently over time when growing at high or low diversity due to differences in

eco-evolutionary feedbacks (Bailey et al., 2006; Linhart, 1988; Post and Palkovacs, 2009; terHorst et al., 2016). There is empirical evidence that plant individuals at high diversity are selected for greater niche complementarity among species leading to a more complete use of available resources (Zuppinger-Dingley et al., 2014). At low plant diversity, in contrast, the accumulation of soil-borne pathogens may cause persistent species to adapt to this increase by producing more defense compounds. Thus, over time selection could favor individuals that invest more in defense and less in growth, which could be measurable as lower biomass production and altered values of traits related to growth (Dietrich et al., 2020b; Lau and Lennon, 2011; Mraja et al., 2011). For example, plants could decrease in height, shift along the leaf economic spectrum, and/or change their root architecture so that they can allocate more resources into defense. Indeed, several studies in the Jena Experiment demonstrated that plants not only show phenotypic plasticity in response to altered growth conditions in low- and high-diversity communities, but provided first evidence for micro-evolutionary processes. For example, only four years after sowing, the grass species Lolium perenne L. showed genetic differentiation from the source population, which was probably due to genetic drift as well as genotype-specific interactions with other species in plant communities of different diversity (Nestmann et al., 2011). Moreover, a recent study by van Moorsel et al., 2019 in the Jena Experiment demonstrated genetic and epigenetic divergence among plants originated either from monocultures or mixtures in three out of five studied species, suggesting rapid emergence of low-diversity and highdiversity genotypes.

Taken together, low and high plant diversity may differently affect eco-evolutionary feedbacks and thus the microevolution of plants, which could have the consequence that plants selected at different diversity respond differently to global change drivers. In a previous transplant experiment in the field, it was shown that some of the studied grassland species showed differences in their phenotype depending on plant history (monoculture or mixture) and soil environment (*Lipowsky et al., 2011*). Several greenhouse or common garden studies came to similar conclusions, that is, that plants selected at low or high diversity or grown with 'own' or different soil biota vary in their productivity and trait expression (*Hahl et al., 2020; Rottstock et al., 2017; van Moorsel et al., 2018b*; *Zuppinger-Dingley et al., 2014*). Such diversity-induced differences in the phenotype could lead to different responses of plants to global change drivers. For example, it is possible that, plants selected at low diversity have a phenotype which is related to a lower resistance against drought than plants selected at high diversity. Such changes would contribute to a faster extinction of species, which makes research into these processes an essential frontier.

In summary, differences in plant-plant and plant-soil interactions at low and high diversity may lead to differences in eco-evolutionary feedbacks; however, little is known about how rapidly and pervasively these differences occur (terHorst et al., 2016). Moreover, it is not known whether these differences affect the response of plants to global change drivers (Pugnaire et al., 2019), such as drought, nitrogen input, or a combination of both, which is assumed to be a common scenario in the future (Craven et al., 2016; Sage, 2020). To address these knowledge gaps, we performed a common garden experiment using plant and soil material from a long-running biodiversity experiment (Jena Experiment). For our study, we collected seeds of four grass species and took samples of soil biota (soil samples), which both had been selected for 14 years, either at low or high diversity (communities with two or six plant species and different plant species composition). The selection of species was based on a sufficient seed production of plants, whereby the four grass species show a spectrum of different growth strategies ranging from very dominant to subordinate. Plants were grown either in soil inoculated with their home soil biota, that is, soil biota of the community, where the seeds had been collected, or in soil inoculated with away soil biota, that is, with soil biota of a different plant community (differing in plant diversity or composition). The aim of the study was to test whether plant history (origin of plants: two- or six-plant species communities), soil history (origin of soil biota: twoor six-plant species communities), and soil treatment (home or away) influence the response of the plants to global change. Therefore, plants were either non-treated (control), or exposed to drought, increased nitrogen input, or a combination of both, drought and nitrogen input, in a full factorial design. We hypothesized that.

(I) plant and soil communities in the field developed differently at low and high diversity over time, that is, plants at low diversity were selected to invest more resources into defense due to higher accumulation of soil-borne pathogens, while plants at high diversity were selected to invest into growth

via complementarity effects and higher number of soil-borne mutualists. Therefore, we expected that productivity and trait expression of control plants (without global change drivers) in our common garden experiment differ depending on plant history, soil history, and soil treatment; for example, offspring of plants selected at low diversity has a lower biomass production and altered values of traits related to growth, because more resources are invested into defense, while the opposite is true for offspring of plants selected at high diversity.

(II) global change drivers have a strong impact on biomass production and trait expression of plants. We expected that drought reduces, and nitrogen input increases, plant biomass, and that single global change drivers alter the expression of traits related to growth. The combination of both global change drivers, however, has no net effect, because drought and nitrogen input compensate each other's impacts.

(III) because of different development of plants and soil communities at low and high diversity, offspring of plants selected at different diversity and grown in different soil (home vs. away soil, soil from low- vs. high-diversity communities) respond differently to global change drivers regarding performance and trait expression.

Results

Hypothesis 1: Offspring of plants selected at different diversity and grown in different soil (high vs. low diversity, home vs. away) show differences in productivity and trait expression

Biomass production

Overall, legacy treatments had a low impact on biomass production, and explain a small portion of variance (**Table 1**). Plants grown in soil of six-species communities tended to produce more root biomass than plants in soil of two-species communities in the control (**Table 2**; **Figure 1**). At species-level, Arrhenatherum elatius produced more root biomass and had higher root-shoot ratio, and Dactylis glomerata produced more shoot and total biomass in soil of six-species than two-species communities (**Figures 1 and 2a**; **Appendix 1—Tables 1 and 3**). The other two species, *Poa trivialis* and Alopecurus pratensis did not differ significantly in biomass production dependent on soil or plant history (**Figures 1 and 2a**; **Appendix 1—Tables 2 and 4**). Initial shoot number showed no influence on later biomass production except for shoot biomass of *D. glomerata* and root biomass of *A. elatius*, which, however, did not change the general patterns.

Plant traits and pathogen infestation

Legacy treatments had no consistent effects across the four species on the expression of shoot, leaf, or root traits in the control (**Appendix 1—table 5**). At species-level, legacy treatments did not affect trait expression in *A. elatius* (*Figure 2a*; *Appendix 1—table 6*). Plants of *A. pratensis* were taller in home than in away-different soil and had thicker roots (higher root diameter) in six- than in two-species soil (*Figure 2a*; *Appendix 1—table 7*). Plants of *D. glomerata* had higher leaf greenness and stomatal conductance, when seeds originated from two-species communities (*Figure 2a*; *Appendix 1—table 8*). Plants of *P. trivialis* had lower shoot nitrogen concentration and root diameter, and higher SRL in home soil than in away soil (*Figure 2a*; *Appendix 1—table 9*).

We found a low pathogen infestation of A. elatius and A. pratensis ($0.8\% \pm 1.9\%$ (SD) and $0.1\% \pm 0.5\%$, respectively), mainly by the rust species *Puccinia graminis* Pers. and *Puccinia coronata* Corda. Plants of D. glomerata and P. trivialis, in contrast, were strongly infested by the mildew *Blumeria graminis* (DC.) Speer ($3.1\% \pm 4.2\%$ and $8.6\% \pm 16.5\%$, respectively). Regarding legacy treatments, D. glomerata plants had a lower infestation when grown in home soil than in away soil, while mildew infestation of P. trivialis plants did not differ between legacy treatments (*Figure 2a; Appendix 1—table 10*).

given in bold, marginally significant	, כווי , ש-א t effects (p	alues (p) allo v < 0.1) in ita	u expranteu v ilics.		י, כמוכעומ				1. /o/. JUJI	ווורמוור בוובר	ט.ט / ט) גו) מוב
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Species identity (ID)	З	73.25	< 0.001	19.54	ю	80.17	< 0.001	17.36	S	121.30	< 0.001	63.65
Plant history	1	3.48	0.062	0.34	1	1.36	0.244	0.05	1	3.40	0.065	0.23
Soil history	1	0.01	0.915	< 0.01	-	0.04	0.851	< 0.01	-	0.49	0.484	< 0.01
Soil treatment	2	2.17	0.338	0.02	2	1.20	0.548	< 0.01	2	3.66	0.161	0.08
Drought (D)	1	83.05	< 0.001	10.30	1	110.26	< 0.001	14.11	-	2.81	0.094	0.09
Nitrogen input (N)	1	257.26	< 0.001	22.83	1	425.93	< 0.001	34.17	-	15.89	< 0.001	0.78
Species ID × Plant history	ĸ	0.71	0.872	< 0.01	ε	1.77	0.621	< 0.01	ε	0.63	0.890	< 0.01
Species ID × Soil history	ю	1.68	0.642	< 0.01	e	0.18	0.980	< 0.01	ε	3.64	0.303	0.02
Species ID × Soil treatment	9	4.29	0.638	< 0.01	9	6.64	0.355	0.01	9	2.30	0.891	< 0.01
Species ID × D	З	52.00	< 0.001	3.08	З	43.11	< 0.001	1.92	S	98.61	< 0.001	4.45
Species ID × N	ĸ	30.46	< 0.001	1.55	ε	33.73	< 0.001	1.35	ε	18.28	< 0.001	0.62
D×N	-	35.27	< 0.001	1.82	-	27.47	< 0.001	1.10	-	10.90	0.001	0.40
Species ID × Plant history × D	4	0.92	0.922	< 0.01	4	4.42	0.353	0.02	4	0.72	0.948	< 0.01
Species ID × Soil history × D	4	1.17	0.883	< 0.01	4	5.33	0.255	0.05	4	0.54	0.969	< 0.01
Species ID x Soil treatment x D	8	2.81	0.946	< 0.01	8	4.78	0.781	< 0.01	8	3.30	0.914	< 0.01
Species ID × Plant history × N	4	2.66	0.617	< 0.01	4	5.75	0.219	0.05	4	1.69	0.792	< 0.01
Species ID × Soil history × N	4	6.59	0.159	0.13	4	3.47	0.482	< 0.01	4	5.26	0.262	0.04
Species ID x Soil treatment x N	Ø	9.35	0.314	0.03	8	4.62	0.797	< 0.01	Ø	15.48	0.050	0.25
Species ID × Plant history × D × N	4	14.85	0.005	0.50	4	27.25	< 0.001	0.87	4	12.61	0.013	0.32
Species ID x Soil history x D x N	4	13.14	0.011	0.43	4	14.39	0.006	0.37	4	11.81	0.019	0.28
Species ID x Soil treatment x D x N	ω	6.19	0.626	< 0.01	Ø	7.91	0.442	< 0.01	8	4.81	0.778	< 0.01

Table 1. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment), global

change treatments (drought, nitrogen input), and their interactions on plant performance (total biomass, shoot biomass, root biomass).

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Species ID × ST 6 7.55 0.273 6 5.43 0.490 6 3.05 0.803 6 9.25 (Species ID × SH	e	3.68	0	1.299	e	3.00	0.391	c	3.11	0.375	3	4.02	0.259
	Species ID x ST	9	7.55	0	1.273	9	5.43	0.490	9	3.05	0.803	9	9.25	0.160

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Table 2. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment), and

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Hypothesis 2: Global change drivers have a strong impact on biomass production and trait expression

Biomass production

Overall, global change drivers had a strong impact on almost all response variables, and explained a large portion of variance, together with species ID (**Table 1**; **Figure 3b–d**; **Appendix 2—Tables 1–9**). Compared to control plants, drought reduced shoot biomass production, which was found across all study species and at species-level (**Figure 3a and d**). In contrast, drought did not have consistent effects on root biomass (**Figure 3a and d**). Drought had positive impact on root biomass of *A. elatius* and *D. glomerata*, while root biomass of *A. pratensis* decreased under drought and did not change significantly in *P. trivialis* (**Figure 3a and d**). Total biomass production was decreased, when plants were exposed to drought (**Figure 3a and d**) except for *D. glomerata*, where it was not different from the control (**Figure 3d**). Root-shoot ratios increased under drought (**Figure 3a and d**), which was found for all species except for *P. trivialis* (no significant change; **Figure 3d**).

Nitrogen input increased shoot, root, and thus also total biomass across the four species (*Figure 3b*) as well as in separate analyses of *A. elatius* and *A. pratensis* (*Figure 3e*). Plants of *D. glomerata* and *P. trivialis* did not change root biomass when fertilized (*Figure 3e*). Nitrogen input caused a decrease in root-shoot ratio in all species (*Figure 3a and e*).

When plants were treated with both global change drivers in combination, the negative impact of drought on shoot biomass was cancelled out by the positive impact of nitrogen input leading to an overall slight increase of shoot biomass (compared to control plants) that was also significant at



Figure 2. Schematic overview of the results of the common garden experiment testing how plants with a different origin (plant history) or grown in different soil (soil history, soil treatment) differ in performance and trait expression (**a**), under the influence of global change drivers drought (**b**), nitrogen input (**c**), and the combination of both (**d**). Illustrated is the impact of legacy treatments (= "legacy effect") and global change treatments (= "global change effect") on shoot and root biomass production as well as on plant traits (growth height ("Height"), shoot nitrogen concentration ("N_{shoot}"), leaf greenness ("Greenness"), leaf dry matter content ("LDMC"), specific leaf area ("SLA"), stomatal conductance ("g_s"), mildew infestation ("Mildew"), root diameter ("Dia"), specific root length ("SRL"), root length density ("RLD")) of the four study species. For legacy effects, schematic illustrations of plants indicate differences in shoot and/or root biomass, when originated from two-species (= 2) or six-species (= 6) communities (= plant history (PH); green color), when grown in two-species (=2) or six-species (= 6) community soil (= soil history (SH), blue color), or when grown in away (= a) or home (= h) soil (= soil treatment; a_s = away same soil; orange color). Arrows behind traits (for legacy effects) indicate, in which treatment group the value was significantly higher (arrow up) or lower (arrow down), e.g. "- SH: SLA 16" indicate that SLA in plants grown in six-species soil was higher than in two-species soil and "- LDMC 1h" indicate that LDMC was higher in plants increased (big arrow up) or decreased (big arrow down) due to the impact of the respective global change effects) indicate and increase (arrow up) or decrease (arrow up) or the trait value due to the impact of the respective global change effects) indicate and increase (arrow up) or decrease (arrow down) of the trait value due to the impact of the respective global change effects) indicate and increase (arrow up) or decrease (a

the species-level except for A. elatius (**Figure 3c and f**). Consistent with this, the positive impact of nitrogen input on root biomass was also cancelled out by drought when plants were treated with both global change drivers, i.e., control plants and plants treated with both global change drivers did not differ in root biomass production, across all study species (**Figure 3c**). At species-level, the combination of both global change drivers had an additive effect on root biomass production of *A. elatius* and *D. glomerata*, that is, plants of both species showed highest root biomass when treated with both global change drivers (**Figure 3f**). In *A. pratensis* and *P. trivialis*, both global change drivers in combination decreased root biomass production (**Figure 3f**). Taken together, the combination of both global change drivers (**Figure 3f**). In C. pratensis production, across all study species and for the high-productive species *A. elatius* and *D. glomerata*, while plants of the low-productive species *A. pratensis* and *P. trivialis* of the low-productive species *A. pratensis* and *P. trivialis* had a similar total biomass production as in the control (**Figure 3c and f**).



Figure 3. Response of plants treated with drought, nitrogen input, or a combination of both relative to non-treated plants (control) for total biomass, shoot biomass, root biomass, and root-shoot ratio across four study species (**a**–**c**) and separately for each species (**d**–**f**). Points are means and error bars are standard deviation. No symbol indicates significant differences between plants treated with global change driver and control plants, n.s. indicate no significant difference.

Root-shoot ratios were as low as in fertilized plants, across all species and in *P. trivialis* (*Figure 3c and f*). Plants of *A. elatius* and *D. glomerata* increased root-shoot ratios, similar to plants under drought (*Figure 3f*). In contrast, *A. pratensis* strongly decreased root-shoot ratios resulting in the lowest values compared to the other treatments (*Figure 3f*).

Plant traits and pathogen infestation

Across all study species, drought did not significantly alter growth height, but nitrogen input increased height (*Figure 3b and c*). When treated with both global change drivers, drought canceled out the positive nitrogen input effect, leading to similar height of plants treated with both global change drivers and control plants (*Figure 3d*). Further, drought and nitrogen input increased shoot nitrogen concentrations and leaf greenness, with additive effects when both global change drivers were applied together (*Figure 3b-d*). Drought did not influence LDMC and SLA, while nitrogen input decreased LDMC and increased SLA (*Figure 3b and c*). When treated with both global change drivers, drought mitigated the decrease of LDMC under nitrogen input, while the increase of SLA under nitrogen input did not change with drought (*Figure 3d*). Stomatal conductance was increased, when plants were treated with drought, but did not change when fertilized irrespective of the drought treatment (*Figure 3b-d*). In terms of root traits, we found a decrease of RLD under drought (irrespective of fertilization) and an increase in root diameter under nitrogen input (irrespective of drought; *Figure 3b-d*). Results of species-specific changes in trait expression under global change drivers can be found in *Figure 3b-d* and Appendix 2.

In *D. glomerata*, mildew infestation remained unchanged when treated with drought, but increased with nitrogen input. When treated with both global change drivers, mildew infestation was as high as in fertilized plants (*Figure 3b–d*). In *P. trivialis*, mildew infestation was increased under drought and when fertilized, while the combination of both global change drivers led to the highest mildew infestation (*Figure 3b–d*).



Figure 4. Total biomass (**a**–**c**), shoot biomass (**d**–**f**), and root biomass (**g**–**i**) of plants (across all four study species) originated from two- or six-species communities (plant history; **a**, **d**, **g**); grown in soil originated from two-species or six-species communities (soil history; **b**, **e**, **h**); or grown in home, away-same or away-different soil (soil treatment; **c**, **f**, **i**) and were either non-treated (control) or treated with drought, nitrogen input (N input) or a combination of both (D + N). Bars show mean values (\pm 1 SE); stars above bars indicate significant differences (p < 0.05), stars in brackets indicate marginally significant differences (p < 0.1).

Hypothesis 3: Offspring of plants selected at different diversity and grown in different soil (high vs. low diversity, home vs. away) respond differently to global change drivers

Biomass production

Plants from two- and six-species communities did not differ in shoot biomass production when treated with drought, but plants from six-species communities treated with drought tended to produce more root biomass than plants from two-species communities across all study species (**Table 2**; **Figure 4a**, **d and g**). At species-level, we found no significant effects of legacy treatments under drought (**Figure 2b**; **Appendix 1—Tables 1–4**).

When plants were fertilized, we found an impact of plant history across all study species: fertilized plants originated from six-species communities had a higher root and total biomass production than plants from two-species communities (**Table 2**; **Figure 4a and g**). This was also found in *D. glomerata*

plants, which tended to produce more shoot and total biomass when originated from six-species communities (*Figure 2c; Appendix 1—table 3*). In *A. elatius*, total biomass production of fertilized plants was significantly higher (and shoot biomass marginally significantly higher), when plants were grown in home and away-same than in away-different soil (*Figure 2c; Appendix 1—table 1*). In *A. pratensis*, fertilized plants grown in two-species community soil tended to produce more total biomass than in six-species community soil (*Figure 2c; Appendix 1—table 2*), while fertilized *P. trivialis* showed no significant differences (*Figure 2c; Appendix 1—table 4*).

When plants were treated with both global change drivers, the effects of nitrogen input were cancelled out or changed by drought, i.e., there was no significant impact of legacy treatments on biomass production across all study species and for A. *elatius* (**Table 2**; **Figure 2d**; **Appendix 1**—**table 1**). In D. glomerata, the significant influence of plant history disappeared, but plants in home and away-different soil showed higher root-shoot ratios than plants in away-same soil (**Figure 2d**; **Appendix 1**—**table 3**). Plants of *P. trivialis* treated with both global change drivers tended to have higher root biomass and root-shoot ratios when grown in home than in away-same soil (**Figure 2d**; **Appendix 1**—**table 4**). In contrast to the overall trend, A. *pratensis* was the only species which showed a similar response to nitrogen input and treatment with both global change drivers: the biomass production was higher in two- than in six-species community soil (for both global change drivers: significant higher root biomass and root-shoot ratios; **Figure 2d**; **Appendix 1**—**table 2**).

Plant traits and pathogen infestation

Shoot nitrogen concentration was not influenced by plant or soil history when treated with drought, but fertilized plants in six-species soil had higher shoot nitrogen concentrations than in two-species soil (soil history effect). Moreover, fertilized plants had lower shoot nitrogen concentrations in home than in away-different soil (soil treatment effect). When plants were treated with both global change drivers, the nitrogen input effect on soil history was cancelled out by drought, while the impact of soil treatment did not: plants in home soil still had lower shoot nitrogen concentration than plants in away soil (**Appendix 1—table 5**). Other plant traits (growth height, leaf greenness, LDMC, SLA, stomatal conductance, root traits) did not significantly differ depending on legacy treatments when plants were treated with nitrogen or drought (**Appendix 1—table 5**). At species level, we found a large number of different responses depending on legacy treatments and type of global change driver, which can be found in **Figure 3b-d** and Appendix 1.

Mildew infestation of *D. glomerata* plants exposed to drought was higher in home than in away soil, while this drought effect was cancelled out by nitrogen input (**Appendix 1—table 10**). Mildew infestation of *P. trivialis* plants was not significantly influenced by legacy treatments, neither with nor without global change drivers (**Appendix 1—table 10**).

Discussion

Hypothesis 1: Offspring of plants selected at different diversity and grown in different soil (high vs. low diversity, home vs. away) show differences in productivity and trait expression

Our findings that A. elatius and D. glomerata plants in soil of high-diversity communities produce more biomass than in soil of low-diversity communities are in line with several greenhouse studies showing that soil conditioned by multiple plant species has a more positive impact on plant growth than soil conditioned by only one or two plant species (**Guerrero-Ramírez et al., 2019**; **Yang et al., 2015**). Plants probably suffered more from pathogens when grown in soil of low-diversity communities and/or benefitted more from interactions with soil mutualists in soil of high-diversity communities (**Dietrich et al., 2021**; **Guerrero-Ramírez et al., 2019**; **Schnitzer et al., 2011**). Interestingly, this soil legacy effect was only found in A. elatius and D. glomerata, which were both highly productive species in the long-term field experiment. In contrast, the low-productive species A. pratensis and P. trivialis showed no significant difference when grown in differently conditioned soils. We can only speculate about the underlying reasons. It is possible that soil of low-diversity communities containing A. elatius and/or D. glomerata had a higher number of (species-specific) pathogens than plots containing A. pratensis and/or P. trivialis, due the higher productivity of A. elatius and D. glomerata in the field and thus more resources for pathogens. This accumulation of species-specific pathogens could lead to reduced productivity of *A. elatius* and *D. glomerata* offspring grown in low-diversity soil. However, it is also possible that *A. elatius* and *D. glomerata* benefit more, and *A. pratensis* and *P. trivialis* less, from soil mutualists, which can be more abundant in soil from high-diversity than in soil from low-diversity plant communities.

In contrast to biomass production, we did not find any significant influence of soil history on plant trait expression of *A. elatius* and *D. glomerata*. Nevertheless, we detected some other legacy treatment effects on plant trait expression, which was also found in related studies (*van Moorsel et al., 2018a*; *van Moorsel et al., 2018b*). The impact of soil history on root diameter of *A. pratensis*, and the impact of soil treatment (home/away) on the growth height of *A. pratensis*, on shoot nitrogen concentrations and root traits of *P. trivialis*, and on mildew infestation of *D. glomerata* indicate that plant-soil interactions influence growth, defense, and resource use strategies of plants (*Xi et al., 2021*), while this impact is species-specific. Moreover, *D. glomerata* plants had higher leaf greenness and stomatal conductance, when originated from low-diversity than from high-diversity plant communities. This could be an adaptation to higher light availability and lower soil moisture in low-diversity communities due to lower shading (*Bachmann et al., 2018; Fischer et al., 2018; Lorentzen et al., 2008*).

Hypothesis 2: Global change drivers have a strong impact on the productivity and trait expression of plants

In accordance with our second hypothesis, we found that drought reduced total biomass production. This was mainly caused by a loss of shoot biomass, while drought differently affected root biomass production of the studied grass species. Individuals of *A. elatius* and *D. glomerata* increased in root biomass at the expense of shoot biomass, leading to higher root-shoot ratio under drought. This is a commonly observed strategy to avoid dehydration, which enables plants to tap water from deeper soil layers (in the field) and at the same time minimizes the water loss caused by transpiration (*Eziz et al., 2017*). In contrast, the low-productive species either decreased instead of increased root biomass (*A. pratensis*) or did not change root biomass production (*P. trivialis*) under drought. Interestingly, the low-productive species had a three times higher loss of total biomass under drought (*A. pratensis*: -17.1%; *P. trivialis*: -15.3%) than the highly-productive species (*A. elatius*: -6.4%; *D. glomerata*: -5.7%, no significant loss of total biomass in *D. glomerata*). Presumably, the drought resistance strategy of *A. elatius* and *D. glomerata* is more effective, which is possibly a competitive advantage under the field conditions of the Jena Experiment, explaining the dominance of these species.

The influence of drought on the expression of plant traits was plant species-specific, except for shoot nitrogen concentrations and leaf greenness, which increased under drought in three species (except *P. trivialis*). Similar results were found in previous studies (*Kocoń and Staniak, 2014; Rolando et al., 2015*) and indicate a general strategy against drought stress: plants decrease the cell density of shoot tissues, in line with the reduction of shoot biomass to minimize the water loss, leading to an increase in the concentration of nitrogen compounds and chlorophyll (strong correlation between leaf greenness and chlorophyll concentration were found in *Bachmann et al., 2018*). At species-level, the low-productive species showed trait expression changes similar to biomass loss under drought, while the highly-productive species *D. glomerata* decreased in LDMC and increased in stomatal conductance, which is contrary to recent studies showing the opposite strategy to resist drought (high LDMC, low stomatal conductance; *Bristiel et al., 2018; Jaballah et al., 2008; Lozano et al., 2020*). The results may differ because *D. glomerata* in our study was infested by the mildew *Blumeria graminis*, which may have changed the leaf structure, and thus also trait expression changes under drought.

Furthermore, our second hypothesis was confirmed by showing that nitrogen input increased biomass production. At species-level, shoot biomass was increased in all four species, while root biomass was enhanced only in *A. elatius* and *A. pratensis*. In *D. glomerata* and *P. trivialis*, there was also a slight, but non-significant increase in root biomass. Both species showed a strong increase in mildew infestation when fertilized. This confirms the nitrogen-disease hypothesis indicating that nitrogen supply increases infection severity by altering leaf properties and resources for pathogens (*Dordas, 2008*). In *D. glomerata* and *P. trivialis*, severe infestation by powdery mildew *Blumeria graminis* may have led to a decrease in rates of net photosynthesis (*Hibberd et al., 1996; Mandal et al., 2009*), so that the reduced amount of energy was mainly invested in shoot biomass, e.g., for a higher leaf turnover, and less in root biomass.

We found consistent changes in plant trait expression over all four species in response to nitrogen input: growth height (except *A. elatius*), shoot nitrogen concentrations, and leaf greenness increased in all four species when fertilized, confirming an earlier study by **Siebenkäs et al., 2015**. Further nitrogen-induced changes in trait expression were likely affected by mildew infestation: the highly-infested species (*D. glomerata, P. trivialis*) showed lower LDMC and higher SLA, while LDMC and SLA of non-infested species did not change. Probably, *D. glomerata* and *P. trivialis* plants responded to the increase in mildew infestation with a change in the leaf architecture (**Cappelli et al., 2020**), which could enable plants to turn over their leaves more quickly and thus produce constantly new and unaffected leaves. With regard to root traits, the non-infested species decreased in specific root length (and *A. pratensis* also in root diameter), while root traits remained unchanged in the highly-infested species. The decrease in SRL and increase in diameter (i.e. thicker and shorter roots) in combination with the increase in root biomass of the fertilized *A. elatius* and *A. pratensis* plants indicate that these plants changed the root architecture building fewer fine roots when nutrient availability is enhanced, which is in line with similar research (**Siebenkäs et al., 2015**).

Finally, we hypothesized that global change drivers cancel out each other's impact when applied together. This was true for the low-productive species *A. pratensis* and *P. trivialis*, which did not change in total biomass production compared to control plants as also found in other research (*Carlsson et al., 2017*). However, the strong decrease in root-shoot ratios indicates that *A. pratensis* and *P. trivialis* plants changed their growth strategies. Interestingly, the high-productive species *A. elatius* and *D. glomerata* slightly increased in total biomass, which is mainly explainable by the additive positive impact of drought and nitrogen input on root biomass, resulting in increased root-shoot ratios. Obviously, dominant (or highly-productive) species in our study benefitted more strongly from the combined application of the global change drivers in comparison to subordinate (or low-productive) species. Assuming that dry periods are becoming more frequent (*Ruosteenoja et al., 2017*) and nitrogen deposition may steadily rise (*Reay et al., 2008*), our results suggest that competitive interactions change under the impact of multiple global change drivers, and subordinate species may become more severely threatened by extinction (*Pugnaire et al., 2019*).

Moreover, our results show that the combined effects of the two global change drivers on plant trait expression may differ from the effect of drought or nitrogen input alone, with strong negative effects for some plant species (e.g. highest mildew infestation of *P. trivialis* under combined impact of global change drivers). This suggests that plants change in physiology and morphology and thus in their response to global change, when a combined impact becomes more frequent, with an unknown influence on community composition and ecosystem functioning in the long term. This finding underlines the need for studies investigating multiple, interacting global change drivers (*Rillig et al., 2019*; *Thakur et al., 2018*).

Hypothesis 3: Offspring of plants selected at different diversity and grown in different soil (high vs. low diversity, home vs. away) respond differently to global change drivers

The soil history effect, that is, the beneficial effect of soil biota from high-diversity plant communities on biomass production of control plants, disappeared in treatments with global change drivers, which may be explainable by a change in soil community structure under drought (*Kaisermann et al., 2017*; *Pugnaire et al., 2019*) and/or nitrogen input (*Wei et al., 2018*). In line with our result, similar studies have shown that drought (*Fry et al., 2018*; *Wilschut and van Kleunen, 2021*) and nitrogen input (*in 't Zandt, 2019*) can interrupt or change plant-soil interactions.

Next to soil history, we also found altered plant responses to global change drivers when plants originated from low- or high-diversity communities (plant history). When treated with drought, there was no significant difference, but nitrogen input had a more positive impact on plants originated from high-diversity than from low-diversity communities. Possibly plants at high diversity were selected for greater niche complementarity (*Zuppinger-Dingley et al., 2014*), while plants at low diversity were selected for increased defense against species-specific pathogens (*Eisenhauer et al., 2019*), that accumulate in low-diversity environments (*Dietrich et al., 2021*; *van Ruijven et al., 2020*). Consequently, the offspring of individuals originated from high-diversity communities may be more efficient in allocating additional resources in increased growth, explaining our results. Interestingly, we did not find any significant plant history effect in plants treated with both global change drivers, indicating

that drought had a strong impact on the growth strategy of the plants and can counteract positive diversity effects.

Finally, we found that plants in home and away soil may respond differently to global change drivers; however, this was only true for the high-productive species *A. elatius*: plants benefitted more from fertilization in home and away-same than in away-different soil. The home advantage supports the idea that a decrease of plant diversity can lead to changes in plant-soil interactions and thus to differences in eco-evolutionary feedbacks at low and high diversity (*terHorst et al., 2016*). With our data in hand, we cannot determine the exact reason why we found the home advantage under fertilization but not under control conditions; however, our results show that plants may respond differently to global change drivers depending on the soil community with which they interact, which is consistent with previous findings (*Puy et al., 2021*).

Similar to the biomass production results, almost all differences in trait expression found in control plants disappeared when treated with global change drivers. Instead, many other changes in trait expression occurred depending on the type of global change driver treatment and plant species identity. Taken together, these results indicate that mainly soil biota (soil history and soil treatment) and only to a lesser extent plant history play an important role in the expression of traits under the influence of global change drivers, which is in line with previous findings (*Puy et al., 2021*). This suggests that the soil biota composition is strongly associated with the physiology and morphology of the plants. Therefore, shifts in soil biota composition due to plant species loss and/or global change driver impact can have strong effects on the response of plants to global change, which could further accelerate plant community change and species loss (*Pugnaire et al., 2019; Yang et al., 2021*).

Conclusion

In the present study, we showed for the first time that offspring of plants selected at low and high plant diversity differently respond to global change and that plant-soil interactions play a significant role in this process. These differences were mainly related to changes in trait expression, while changes in biomass production were minor, and they were strongly dependent on plant species identity and their competitiveness in the field, as well as the type of global change driver (drought, nitrogen input, or both). Although we did not find clear evidence that plants selected at low diversity generally suffer more under global change than plants selected at high diversity, it is possible that the species-specific responses alter species interactions and accelerate global change effects in the long run. To better assess the risk of such a potential feedback loop, future research is urgently necessary, especially, studies that test the long-term influence of global change drivers on plants and soil biota selected at different diversity under more realistic conditions, for example as plants growing in communities under field conditions.

Materials and methods

The Jena Experiment

Seed and soil material for our common garden experiment was collected from a long-term biodiversity experiment, the Jena Experiment, which is located in the floodplain of the Saale river near Jena (Thuringia, Germany, 50° 55'N, 11° 35'E, 130 m a.s.l.) (**Roscher et al., 2004**; **Weisser et al., 2017**). Before the establishment of the Jena Experiment in 2002, the site was a highly fertilized arable field, which had been used for growing wheat and vegetables from the early 1960s until 2000. Mean annual air temperature recorded from 2007 to 2016 at the experimental site (Weather Station Jena-Saaleaue, Max-Planck-Institute for Biogeochemistry Jena, https://www.bgc-jena.mpg.de/wetter/) was 9.7 °C, and mean annual precipitation was 587 mm. The soil of the study site is a Eutric Fluvisol, whereas soil texture changes from sandy loam to silty clay with increasing distance from the river Saale. Thus, four blocks were arranged parallel to the riverside (**Roscher et al., 2004**).

Material for our study was collected in a sub-experiment of the Jena Experiment, the so-called Dominance Experiment. The species pool of this experiment included nine species, which often reach dominance in Central European mesophilic grasslands of the Arrhenatherion type (*Ellenberg,* **1988**): five grasses, two legumes, and two herbs. Sown plant species richness levels were 1, 2, 3, 4, 6, and 9 species. Each species occurred eight times in the different compositions of each species-richness level. Moreover, each possible two-species combination was present with equal frequency

at each species-richness level of the mixtures (i.e., 2–9 species; more information about the design can be found in **Roscher et al., 2004**). In May 2002, seeds were sown with a density of 1000 viable seeds per m². Seeds from all species were purchased from a commercial supplier (Rieger-Hofmann GmBH, Blaufelden-Raboldshause, Germany). From 2002 to 2009, plants were grown in plots of 3.5 \times 3.5 m; from 2010 onwards, plot size was reduced to 1 \times 1 m. Plots were mown every year in June and September and mown plant material was removed. All plots were regularly weeded and never fertilized.

Seed collection, selection of study species, and experimental plots

In summer 2016, we collected seed material from the nine species in all Dominance Experiment plots (as bulk sample per species and plot) and stored them in a freezer (at -20 °C) until further use. We chose four grass species (Alopecurus pratensis L., Arrhenatherum elatius (L.) P. Beauv. ex J. Presl et C. Presl, Dactylis glomerata L., Poa trivialis L.) as study species. Furthermore, we selected 12 plots per species, six two-species and six six-species plots, that is, 48 plots in total, where sufficient seed material was available. It should be noted that from here on we refer to plots with two plant species as 'low-diversity communities' and plots with six species as 'high-diversity communities', although the species richness of high-diversity communities can be much greater in nature depending on the considered scale. We chose six-species plots as 'high-diversity communities', because they were available with different community compositions and could better represent the effects of species richness as a measure of diversity than the replicates of the 9-species plots with the same species composition. Moreover, previous studies have shown that plant productivity and soil biota communities already differ between 2- and 6-species communities (Dietrich et al., 2021; Roscher et al., 2007). The selected plots were evenly distributed in the four blocks of the experiment (Roscher et al., 2004). The study species differed strongly in their biomass production in the Dominance Experiment plots. In the two-species plots, all four species showed a high biomass production; however, in the six-species plots, only A. elatius and D. glomerata were highly-productive, while A. pratensis and P. trivialis showed intermediate levels and decreased in biomass production over the years (Clark et al., 2019; Roscher et al., 2007). For simplification, from here onwards, A. elatius and D. glomerata are referred to as 'highly-productive' species, while A. pratensis and P. trivialis are referred to as 'lowproductive' species.

Preparation of background substrate and study plants

For the pot substrate, we used a sterilized sand-soil mix (= background substrate), which was then inoculated with fresh living soil (5% of the total substrate by weight) from the selected plots. This inoculation method is a common procedure to investigate plant-soil interactions and has the advantage that only low amounts of living soil are needed and that potential abiotic feedbacks are eliminated (*Pernilla Brinkman et al., 2010*). To produce sterile background substrate, we collected 1.6 m³ soil substrate from the Jena Experiment in May 2017. This soil substrate was a mix of excavated soil material from different experimental plots, which was stored for several years at the experimental area. The soil substrate was sieved to 10 mm, homogenized, and mixed with 0.4 m³ quartz sand (WF 33, Quarzwerke GmbH, Walbeck, Germany). Afterwards, the soil-sand mix was steam-sterilized twice for 150 minutes at ~80 °C. More information about the steam-sterilization method and changes of abiotic and biotic soil properties can be found in *Dietrich et al., 2020a*.

For the preparation of study plants, QuickPot trays of 20 cm³ volume (Hermann Meyer KG, Rellingen, Germany) were sterilized with a potassium hypochlorite solution (Eau de Javel: 2.6 g KClO to 100 ml water; 1:1) and filled with an autoclaved mixture of sand and soil from the Jena Experiment (1:1; sterilized twice for 40 min at 121 °C) in June 2017. Each species and origin (i.e., plot) was sown with two or three seeds per pot plate cell. QuickPot trays were placed in an open greenhouse (Research Station Bad Lauchstädt, UFZ) to promote germination by natural daily temperature fluctuations. Trays were regularly watered (with demineralized water). On 29 June 2017, *A. pratensis* seeds were reseeded because of low germination rate. For the other three species, one seedling per pot plate cell was removed if more than two seeds were germinated. It should be noted that we cannot exclude possible maternal effects in our common garden experiment, because we used seed material collected in different plots of the field experiment; however, differences in maternal effects can also

be important drivers of eco-evolutionary feedbacks and can significantly influence plant responses to global change (**Puy et al., 2021; Rottstock et al., 2017**).

Common garden experiment

In July 2017, 12 soil cores (5 cm diameter, 10 cm depth) were taken in a grid of 20×20 cm in each Dominance Experiment plot selected for the study and stored in a cooling chamber (4 °C). Soil cores were pooled per plot and sieved through a sieve with 5 mm mesh size to remove stones and coarse roots. Then, 2800 cm³ steam-sterilized background substrate was thoroughly mixed with 150 cm³ fresh-sieved living soil and filled in a heat-cleaned pot (3 L, diameter 14.9 cm, height 18 cm) with 12 replicates per plot. Seedlings per pot plate cell were separated, and two seedlings per species with same plot origin were transplanted into one pot (Figure 5). In four pots per plot, we transplanted plants, which had the same plot origin as the inoculated soil (home soil treatment); in the other eight pots, plant and soil origin were different (away soil treatment). In four of these away pots, species richness of plant and soil origin was the same, but plant species composition was different (away-same soil treatment), and in the other four away pots, species richness of plant and soil origin was different (away-different soil treatment; Figure 5). Seedlings of D. glomerata were transplanted on 18 July 2017, followed by A. elatius (20 July 2017), P. trivialis (20 and 24 July 2017), and A. pratensis (26-28 July 2017). Seedlings were immediately watered with 200 ml demineralized water after transplantation, and the initial number of shoots was counted. In total, the experiment consisted of 576 pots, each with two plants. The pots were placed in an open greenhouse with a roof, which automatically closes at rain, and ambient temperatures (Research Station Bad Lauchstädt, UFZ). Pots were distributed in six blocks placing the 12 pots filled with soil from one plot in one block, that is, in each block, there were 12 pots with soil of one two-species and one six-species plot per species. The position of the pots within the blocks was randomly chosen and changed once a month to avoid potential side effects by neighboring pots and edge effects of the tables.

During the first week after planting, plants were watered every day with 200 ml demineralized water. From week two to four, all pots were watered every other day with 380 ml demineralized water without further treatments to allow the establishment of plants and soil biota in the pots (380 ml were used to achieve a water saturation of the soil of 60%; calculation can be found in Appendix 3). On 23 August 2017, treatments with the global change drivers were started. For every treatment (control, drought, nitrogen input, combination of drought and nitrogen input), we used three of the 12 pots per plot (one home, one away-same, and one away-different pot, respectively; *Figure 5*).

- I. For control, pots were watered as before (380 ml; every other day) and were not fertilized.
- II. Drought was simulated by reduced water saturation (= 30% water saturation = 225 ml; calculation can be found in Appendix 3). Pots were still watered every other day but with 225 ml instead of 380 ml demineralized water.
- III. Nitrogen input was applied once a week with 95 mg NH₄NO₃ (33.125 mg nitrogen) resulting in a total nitrogen amount of 265 mg after eight fertilization events, which is equivalent to a nitrogen input of 150 kg ha⁻¹ year⁻¹ nitrogen (medium value for managed grasslands in Germany; Häußermann et al., 2019). Fertilized plants were watered as before (380 ml; every other day).
- IV. For the combination of drought and nitrogen input, pots were watered with a reduced amount (225 ml) and were fertilized once a week (in the same way as for the nitrogen input treatment alone).

Once a month, all pots were weighted before watering. The measured weight per pot was subtracted from dry soil weight plus the assigned amount of water (380 or 225 ml). The difference revealed the amount of water which was then used to water the pot to keep the anticipated levels of water saturation for the drought and control treatment.

Data collection

After 11 weeks of growth with global change driver treatments, plants were harvested block-wise (between 16 October and 8 November 2018). Before harvest, aboveground traits and leaf fungal pathogen infestation were measured (**Table 3**). For growth height (in cm), we measured the stretched shoot length of the longest vegetative shoot per plant. Only 15% of the plants had flowered, which was neglected due to the small case number. For leaf greenness (unitless estimate of foliar chlorophyll content), three fully expanded leaves from vegetative shoots of each plant were measured with a



Figure 5. Overview of experimental design. In 2016, ripe seeds of four grass species were collected in two- and six-species plots of the Dominance Experiment (Jena Experiment), stored in a freezer and allowed to germinate in spring 2017. After germination, soil samples were collected from the plots and mixed with sterilized background soil (5% + 95%), filled in pots and planted with two seedlings (12 pot replicates per plot). In four pots per plot, plant and soil had the same plot origin (home soil); in four pots, species richness of plant and soil origin were the same, but plant species composition was different (away-same soil) and in four pots, species richness of plant and soil origin were different (= different origin of plant and soil; away-different soil; total Nr_{pots} = 576). Plants were exposed to global change drivers: drought, nitrogen input, or the combination of drought and nitrogen input, or were not treated (control).

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 Table 3. Summary list of response variables and experimental factors of the common garden experiment.

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Variable	Abbreviation	Unit	Description
Response variables			
Biomass production			
Total biomass	Total Bm	$g_{\rm total}$	Shoot and root biomass per pot
Shoot mass	Shoot Bm	g_{shoot}	Shoot biomass per pot
Root mass	Root Bm	g _{root}	Root biomass per pot
Root-shoot ratio	-	$g_{root} g_{shoot}^{-1}$	Root biomass divided by shoot biomass per pot
Aboveground traits			
Growth height	-	cm	Stretched shoot length of longest vegetative shoot*
Shoot nitrogen concentration	N _{Shoot}	mg N g_{shoot}^{-1}	Nitrogen mass per dry shoot mass
Leaf greenness	-	-	Unitless estimate of leaf chlorophyll concentration*
Specific leaf area	SLA	${mm_{leaf}}^2 {mg_{leaf}}^{-1}$	Leaf area per dry leaf mass*
Leaf dry matter content	LDMC	$mg_{leaf} g_{leaf}^{-1}$	Dry leaf mass per water-saturated fresh leaf mass*
Stomatal conductance	g₅	mmol m ⁻² s ⁻¹	Stomatal conductance per leaf area"
Belowground traits			
Root diameter	Dia	mm	Average root diameter of the root subsample
Specific root length	SRL	$m_{root} g_{root}^{-1}$	Root length per dry root biomass (subsample)
Root length density	RLD	${\rm cm_{root}} {\rm cm_{soil}}^{-3}$	Root length (extrapolated) per soil volume (pot)
Pathogen infestation	-	%	Percentage of infested leaf area (estimated)*
Experimental factors			
Species identity	Species ID	-	Study species
Legacy treatments			
Plant history	PH	-	Species richness of the plant community, where the seeds were collected – two or six plant species
Soil history	SH	-	Species richness of plant community, where the soil for inoculation was taken – two or six plant species
			Origin of seed and soil in one pot:
Soil treatment	ST	-	 same plot origin = home soil treatment different plot origin, but same species richness = away-same soil treatment different plot origin, different species richness = away-different soil treatment
Global change driver treatments	global change driver / global change / GC		
Drought treatment	Drought / D	_	30% instead of 60% water saturation
Nitrogen input treatment	Nitrogen input / N	_	Fertilization with NH₄NO₃ (150 kg ha¹ year¹ nitrogen)

*averaged per pot.

SPAD 502 Plus Chlorophyll Meter (Spectrum Technologies, Inc) and values were averaged per plant. Stomatal conductance (g_s ; mmol m⁻² s⁻¹) was measured at one fully expanded leaf per plant (i.e., two leaves per pot) with a SC-1 Leaf Porometer (Decagon Devices Inc). This was done block-wise and always one day after watering, between 10 a.m. and 3 p.m. Shortly before harvest, the percentage of total leaf area, which was infested by fungal pathogens was estimated for each plant. A subsample of leaves per species was taken to identify pathogens morphologically at the species level under a light microscope. Moreover, three fully expanded leaves per individual were cut, packed in wet paper towels to achieve water saturation, and stored overnight in a cooling chamber at 4 °C. On the next day, leaves were weighed as bulk sample per pot (i.e. six leaves) after removing water droplets with

tissue paper. Total leaf area was measured with a leaf area meter (LI-3000C Area Meter equipped with LI3050C transparent conveyer belt accessory, LICOR, USA) Afterwards, leaf samples were dried at 70 °C for 48 hr and weighed. LDMC was calculated as the ratio of dry weight to fresh weight (mg_{leaf}^{-1}) and SLA as the ratio of leaf area to dry weight (mm_{leaf}^{-1}).

For biomass harvest, plants were cut at ground level, and roots were cleaned by rinsing off all soil over a 0.5 mm sieve. The fresh root biomass was weighed and a subsample of around 1–2 g fresh weight was stored at –20 °C. At a later point, roots were thawed and scanned on a flatbed scanner at 800 dpi (Epson Expression 10000 XL scanner, Regent Instruments, Quebec, Canada), and root diameter and root length of the subsample were measured with an image analysis software (WinRHIZO; Regent Instruments, Quebec City, Canada). Specific root length (SRL) was calculated as the ratio of root length to root dry biomass (of the subsample; $m_{root} g_{root}^{-1}$) and root length density (RLD) as the ratio of root length to soil volume in the pot (root length was extrapolated from the ratio of dry root biomass of the measured subsample to total dry root mass per pot; $cm_{root} cm_{soil}^{-3}$).

All biomass samples were dried at 70 °C for 48 hr and then weighed. To calculate total shoot biomass per pot (each with two individuals), dry shoot biomass and dry leaf mass of the sample used for leaf area measurements were added. To calculate total root biomass, dry biomass of the scanned subsample was extrapolated from the ratio of fresh root biomass to dry root biomass per pot and added to the weighed dry root biomass per pot.

For chemical analysis, shoot biomass of each pot was chopped, and a subsample was ground with a ball mill. Then, 10 mg milled material was used to determine shoot nitrogen concentration with near-infrared spectroscopy (MPA Multi Purpose FT-NIR Analyzer, Bruker GmbH, Ettlingen, Germany). The calibration models used to predict shoot nitrogen concentrations were derived from laboratory data generated from previous samples of grass species. The accuracy of the predictions was verified by a repeated nitrogen concentration analysis of 45 randomly selected samples with an elemental analyzer (Vario EL Element Analyzer, Elementar, Hanau, Germany). Significant positive correlation (p < 0.001, r = 0.97, N = 45) between concentrations resulted from near-infrared spectroscopy and analysis with the elemental analyzer demonstrate high accuracy of our predictions.

Data analysis

To test whether the plants performed differently depending on legacy treatments (plant history, soil history, soil treatment [home/away]), or type of global change treatment, linear mixed-effects models were fitted for all measured response variables per pot as summarized in **Table 3**. Furthermore, some variables were transformed to meet the assumptions of normality and variance homogeneity: if necessary, root biomass and RLD were square root-transformed and root-shoot ratio, SLA, stomatal conductance, SRL, and pathogen infestation were log-transformed. Furthermore, outlier values of LDMC of three *P. trivialis* pots (extremely low values), and LDMC and SLA of one *A. elatius* pot (extremely low LDMC, high SLA) were excluded from the analysis.

For mixed-effect model analysis, we started with a null model with the random effects only (fitted with maximum likelihood). We used seed plot identity (plot, where the seeds had been collected) and soil plot identity (plot, from which the inoculation soil had been taken) as random effects. Then, we successively added the fixed effects with species identity first, followed by the legacy treatments: plant history (species richness of the plant community, where the seeds had been collected: two or six), soil history (species richness of the plant community, where the soil for inoculation had been taken: two or six), and soil treatment (home, away-same, away-different), followed by the global change driver treatments: drought (control or drought) and nitrogen input (control or nitrogen), and finally all interactions between species identity and the other fixed effects to check whether species differ in their responses. For analysis of stomatal conductance, we used daytime and air temperature as covariates, which were entered before adding the experimental factors to account for possible effects of the measurement time. Moreover, to decompose the variability attributable to model terms, mixed-effect models (for biomass production) were fitted, but with the restricted maximum likelihood method. Then, the share of explained variability was estimated as the difference between total variability attributed to random effects in models not including, and models including, the respective fixed effect (= variance decomposition; Siebenkäs et al., 2015). Because of multiple significant interactions between species identity and other fixed effects (Table 1), we further analyzed the response variables separately per species. Therefore,

we used the same fixed effect structure as explained above, but without species identity and additionally with the interactions between legacy treatments and global change driver treatments (which was not done in the first model, because otherwise, it would have become too complex). For pathogen infestation, we only analyzed data of *D. glomerata* and *P. trivialis*, because of very low infestation rates of *A. elatius* and *A. pratensis* plants. To test whether initial size influenced the performance of the phytometers later in the experiment, we added initial shoot number as a fixed effect before the other fixed effects in separate models for analysis of shoot and root biomass production.

Because of multiple significant interactions between legacy treatments and global change driver treatments (**Appendix 2—Tables 1–10**), we further analyzed the data for each global change driver treatment separately. We used plant history, soil history, and soil treatment as fixed effects for species-specific analysis, and for analyses across all four species, we extended the models by fitting species identity first and all possible interactions between species identity and legacy treatments in the end.

All models were fitted with maximum likelihood (ML), and likelihood ratio tests were used to decide on the significance of the fixed effects. Tukey's HSD test was used to test differences among soil treatment groups. All calculations and statistical analyses were done in R (version 3.6.1, R Development Core Team, http://www.R-project.org) including the package *Ime4* (glmer and Imer; **Bates et al., 2015**) and *multcomp* (Tukey HSD; **Hothorn et al., 2008**) for mixed-effects model analysis.

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Author contributions

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Additional files

Supplementary files

• Transparent reporting form

Data availability

The data reported in this paper have been deposited in Dryad, which can be publicly accessed at https://doi.org/10.5061/dryad.gmsbcc2p7.

The following dataset was generated:

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References

- Bachmann D, Roscher C, Buchmann N. 2018. How do leaf trait values change spatially and temporally with light availability in a grassland diversity experiment? *Oikos (Copenhagen, Denmark)* **127**:935–948. DOI: https://doi.org/10.1111/oik.04533
- Bailey JK, Wooley SC, Lindroth RL, Whitham TG. 2006. Importance of species interactions to community heritability: a genetic basis to trophic-level interactions. *Ecology Letters* **9**:78–85. DOI: https://doi.org/10. 1111/j.1461-0248.2005.00844.x, PMID: 16958871
- Barry KE, Mommer L, van Ruijven J, Wirth C, Wright AJ, Bai Y, Connolly J, De Deyn GB, de Kroon H, Isbell F, Milcu A, Roscher C, Scherer-Lorenzen M, Schmid B, Weigelt A. 2019. The future of complementarity: disentangling causes from consequences. *Trends in Ecology & Evolution* **34**:167–180. DOI: https://doi.org/10.1016/j.tree.2018.10. 013
- Bates D, Maechler M, Bolker B, Walker S, Christensen RHB, Singmann H, Dai B, Grothendieck G, Green P, Bolker MB. 2015. Package 'Ime4.' Convergence 12:2.
- Bristiel P, Gillespie L, Østrem L, Balachowski J, Violle C, Volaire F. 2018. Experimental evaluation of the robustness of the growth–stress tolerance trade-off within the perennial grass Dactylis glomerata. *Functional Ecology* 32:1944–1958. DOI: https://doi.org/10.1111/1365-2435.13112
- **Cappelli SL**, Pichon NA, Kempel A, Allan E. 2020. Sick plants in grassland communities: a growth-defense trade-off is the main driver of fungal pathogen abundance. *Ecology Letters* **23**:1349–1359. DOI: https://doi.org/10.1111/ele.13537
- Cardinale BJ, Wright JP, Cadotte MW, Carroll IT, Hector A, Srivastava DS, Loreau M, Weis JJ. 2007. Impacts of plant diversity on biomass production increase through time because of species complementarity. PNAS 104:18123– 18128. DOI: https://doi.org/10.1073/pnas.0709069104, PMID: 17991772
- Carlsson M, Merten M, Kayser M, Isselstein J, Wrage-Mönnig N. 2017. Drought stress resistance and resilience of permanent grasslands are shaped by functional group composition and N fertilization. Agriculture, Ecosystems & Environment 236:52–60. DOI: https://doi.org/10.1016/j.agee.2016.11.009
- Clark AT, Ann Turnbull L, Tredennick A, Allan E, Harpole WS, Mayfield MM, Soliveres S, Barry K, Eisenhauer N, Kroon H, Rosenbaum B, Wagg C, Weigelt A, Feng Y, Roscher C, Schmid B, Brophy C. 2019. Predicting species abundances in a grassland biodiversity experiment: Trade-offs between model complexity and generality. *Journal of Ecology* **108**:774–787. DOI: https://doi.org/10.1111/1365-2745.13316
- **Craven D**, Isbell F, Manning P, Connolly J, Bruelheide H, Ebeling A, Roscher C, Van Ruijven J, Weigelt A, Wilsey B. 2016. Plant diversity effects on grassland productivity are robust to both nutrient enrichment and drought. *Philosophical Transactions of the Royal Society B Biological Sciences* **371**:20150277. DOI: https://doi.org/10. 1098/rstb.2015.0277
- Dai AG, Zhao TB, Chen J. 2018. Climate Change and Drought: a Precipitation and Evaporation Perspective. Current Climate Change Reports 4:301–312. DOI: https://doi.org/10.1007/s40641-018-0101-6
- Dietrich P, Cesarz S, Eisenhauer N, Roscher C. 2020a. Effects of steam sterilization on soil abiotic and biotic properties. *Soil Organisms* 92:99–108. DOI: https://doi.org/10.25674/so92iss2pp99
- Dietrich P, Roeder A, Cesarz S, Eisenhauer N, Ebeling A, Schmid B, Schulze ED, Wagg C, Weigelt A, Roscher C. 2020b. Nematode communities, plant nutrient economy and life-cycle characteristics jointly determine plant

monoculture performance over 12 years. *Oikos (Copenhagen, Denmark)* **129**:466–479. DOI: https://doi.org/ 10.1111/oik.06989

- Dietrich P, Cesarz S, Liu T, Roscher C, Eisenhauer N. 2021. Effects of plant species diversity on nematode community composition and diversity in a long-term biodiversity experiment. *Oecologia* **197**:297–311. DOI: https://doi.org/10.1007/s00442-021-04956-1, PMID: 34091787
- Dordas C. 2008. Role of nutrients in controlling plant diseases in sustainable agriculture. A review. Agronomy for Sustainable Development 28:33–46. DOI: https://doi.org/10.1051/agro:2007051
- **Dornelas M**, Gotelli NJ, McGill B, Shimadzu H, Moyes F, Sievers C, Magurran AE. 2014. Assemblage time series reveal biodiversity change but not systematic loss. *Science (New York, N.Y.)* **344**:296–299. DOI: https://doi.org/10.1126/science.1248484, PMID: 24744374
- **Eisenhauer N**, Bonkowski M, Brose U, Buscot F, Durka W, Ebeling A, Fischer M, Gleixner G, Heintz-Buschart A, Hines J, Jesch A, Lange M, Meyer S, Roscher C, Scheu S, Schielzeth H, Schloter M, Schulz S, Unsicker S, van Dam N, et al. 2019. Biotic interactions, community assembly, and eco-evolutionary dynamics as drivers of long-term biodiversity–ecosystem functioning relationships. *Research Ideas and Outcomes* **5**:e47042. DOI: https://doi.org/10.3897/rio.5.e47042

Ellenberg H. 1988. Vegetation Ecology of Central Europe. Cambridge University Press.

- Eziz A, Yan Z, Tian D, Han W, Tang Z, Fang J. 2017. Drought effect on plant biomass allocation: A meta-analysis. Ecology and Evolution **7**:11002–11010. DOI: https://doi.org/10.1002/ece3.3630, PMID: 29299276
- Fischer C, Leimer S, Roscher C, Ravenek J, de Kroon H, Kreutziger Y, Baade J, Beßler H, Eisenhauer N, Weigelt A, Mommer L, Lange M, Gleixner G, Wilcke W, Schröder B, Hildebrandt A, Bardgett R. 2018. Plant species richness and functional groups have different effects on soil water content in a decade-long grassland experiment. *Journal of Ecology* **107**:127–141. DOI: https://doi.org/10.1111/1365-2745.13046
- Fry EL, Johnson GN, Hall AL, Pritchard WJ, Bullock JM, Bardgett RD. 2018. Drought neutralises plant-soil feedback of two mesic grassland forbs. *Oecologia* 186:1113–1125. DOI: https://doi.org/10.1007/s00442-018-4082-x, PMID: 29399737
- **Guerrero-Ramírez NR**, Reich PB, Wagg C, Ciobanu M, Eisenhauer N. 2019. Diversity-dependent plant–soil feedbacks underlie long-term plant diversity effects on primary productivity. *Ecosphere (Washington, D.C)* **10**:e02704.
- Hahl T, Moorsel SJ, Schmid MW, Zuppinger-Dingley D, Schmid B, Wagg C, Thakur M. 2020. Plant responses to diversity-driven selection and associated rhizosphere microbial communities. *Functional Ecology* 34:707–722. DOI: https://doi.org/10.1111/1365-2435.13511
- HäußermannU, BachM, KlementL, BreuerL. 2019. Stickstoff-Flächenbilanzen für Deutschland mit Regionalgliederung Bundesländer und Kreise—Jahre 1995 bis 2017; Methodik, Ergebnisse und Minderungsmaßnahmen. Abschlussbericht TEXTE 131:2019. .
- Hibberd JM, Richardson P, Whitbread R, Farrar JF. 1996. Effects of leaf age, basal meristem and infection with powdery mildew on photosynthesis in barley grown in 700 mumol mol-1CO2. *New Phytologist* **134**:317–325. DOI: https://doi.org/10.1111/j.1469-8137.1996.tb04636.x
- Holland EA, Braswell BH, Sulzman J, Lamarque JF. 2005. NITROGEN DEPOSITION ONTO THE UNITED STATES AND WESTERN EUROPE: SYNTHESIS OF OBSERVATIONS AND MODELS. *Ecological Applications* **15**:38–57. DOI: https://doi.org/10.1890/03-5162
- Hothorn T, Bretz F, Westfall P. 2008. Simultaneous inference in general parametric models. Biometrical Journal. Biometrische Zeitschrift 50:346–363. DOI: https://doi.org/10.1002/bimj.200810425, PMID: 18481363
- in 't Zandt D. 2019. Plant-soil feedback is shut down when nutrients come to town. *Plant and Soil* **439**:541–551. DOI: https://doi.org/10.1007/s11104-019-04050-9
- Isbell F, Craven D, Connolly J, Loreau M, Schmid B, Beierkuhnlein C, Bezemer TM, Bonin C, Bruelheide H, de Luca E, Ebeling A, Griffin JN, Guo Q, Hautier Y, Hector A, Jentsch A, Kreyling J, Lanta V, Manning P, Meyer ST, et al. 2015. Biodiversity increases the resistance of ecosystem productivity to climate extremes. *Nature* **526**:574–577. DOI: https://doi.org/10.1038/nature15374, PMID: 26466564
- Jaballah S, Gribaa A, Volaire F, Ferchichi A. 2008. Ecophysiological responses of perennial grasses Stipa lagascae and Dactylis glomerata under soil water deficit. *Options Méditerranéennes Serie A* **79**:303–307.
- Kaisermann A, de Vries FT, Griffiths RI, Bardgett RD. 2017. Legacy effects of drought on plant–soil feedbacks and plant–plant interactions. New Phytologist **215**:1413–1424. DOI: https://doi.org/10.1111/nph.14661
- Kocoń A, Staniak M. 2014. Productivity and gas exchange parameters of selected pasture grasses under drought stress. Journal of Food, Agriculture and Environment **12**:131–135.
- Lange M, Koller-France E, Hildebrandt A, Oelmann Y, Wilcke W, Gleixner G. 2019. How plant diversity impacts the coupled water, nutrient and carbon cycles. Advances in Ecological Research 61:185–219. DOI: https://doi.org/ 10.1016/bs.aecr.2019.06.005
- Lau JA, Lennon JT. 2011. Evolutionary ecology of plant-microbe interactions: soil microbial structure alters selection on plant traits. *The New Phytologist* **192**:215–224. DOI: https://doi.org/10.1111/j.1469-8137.2011. 03790.x, PMID: 21658184
- Linhart YB. 1988. Intrapopulation differentiation in annual plants III The Contrasting Effects of Intra-and Interspecific Competition. Evolution; International Journal of Organic Evolution 42:1047–1064. DOI: https:// doi.org/10.1111/j.1558-5646.1988.tb02523.x, PMID: 28581179
- Lipowsky A, Schmid B, Roscher C. 2011. Selection for monoculture and mixture genotypes in a biodiversity experiment. *Basic and Applied Ecology* **12**:360–371. DOI: https://doi.org/10.1016/j.baae.2011.03.005

- Lorentzen S, Roscher C, Schumacher J, Schulze ED, Schmid B. 2008. Species richness and identity affect the use of aboveground space in experimental grasslands. *Perspectives in Plant Ecology, Evolution and Systematics* 10:73–87. DOI: https://doi.org/10.1016/j.ppees.2007.12.001
- Lozano YM, Aguilar-Trigueros CA, Flaig IC, Rillig MC. 2020. Root trait responses to drought are more heterogeneous than leaf trait responses. *Functional Ecology* **34**:2224–2235. DOI: https://doi.org/10.1111/ 1365-2435.13656
- Mandal K, Saravanan R, Maiti S, Kothari IL. 2009. Effect of downy mildew disease on photosynthesis and chlorophyll fluorescence in Plantago ovata Forsk. *Journal of Plant Diseases and Protection* **116**:164–168. DOI: https://doi.org/10.1007/BF03356305
- Marquard E, Weigelt A, Temperton VM, Roscher C, Schumacher J, Buchmann N, Fischer M, Weisser WW, Schmid B. 2009. Plant species richness and functional composition drive overyielding in a six-year grassland experiment. *Ecology* **90**:3290–3302. DOI: https://doi.org/10.1890/09-0069.1, PMID: 20120799
- Meyer ST, Ebeling A, Eisenhauer N, Hertzog L, Hillebrand H, Milcu A, Pompe S, Abbas M, Bessler H, Buchmann N, De Luca E, Engels C, Fischer M, Gleixner G, Hudewenz A, Klein AM, de Kroon H, Leimer S, Loranger H, Mommer L, et al. 2016. Effects of biodiversity strengthen over time as ecosystem functioning declines at low and increases at high biodiversity. *Ecosphere (Washington, D.C)* 7:e01619. DOI: https://doi.org/10.1002/ecs2.1619
- Mommer L, Cotton TEA, Raaijmakers JM, Termorshuizen AJ, van Ruijven J, Hendriks M, van Rijssel SQ, van de Mortel JE, van der Paauw JW, Schijlen EGWM, Smit-Tiekstra AE, Berendse F, de Kroon H, Dumbrell AJ. 2018. Lost in diversity: the interactions between soil-borne fungi, biodiversity and plant productivity. *The New Phytologist* 218:542–553. DOI: https://doi.org/10.1111/nph.15036, PMID: 29468690
- Mraja A, Unsicker SB, Reichelt M, Gershenzon J, Roscher C. 2011. Plant community diversity influences allocation to direct chemical defence in Plantago lanceolata. PLOS ONE 6:e28055. DOI: https://doi.org/10.1371/journal. pone.0028055
- Nestmann S, Sretenovic Rajicic T, Dehmer K, Fischer M, Schumacher J, Roscher C. 2011. Plant species diversity and composition of experimental grasslands affect genetic differentiation of Lolium perenne populations. *Molecular Ecology* 20:2188–2203. DOI: https://doi.org/10.1111/j.1365-294X.2011.05027.x
- Pernilla Brinkman E, Van der Putten WH, Bakker EJ, Verhoeven KJ. 2010. Plant-soil feedback: experimental approaches, statistical analyses and ecological interpretations. *Journal of Ecology* **98**:1063–1073. DOI: https://doi.org/10.1111/j.1365-2745.2010.01695.x
- Pörtner HO, Scholes RJ, Agard J, Archer E, Arneth A, Bai X, Barnes D, Burrows M, Chan L, Cheung WL, Diamond S, Donatti C, Duarte C, Eisenhauer N, Foden W, Gasalla MA, Handa C, Hickler T, Hoegh-Guldberg O, Ichii K, et al. 2021. IPBES-IPCC co-sponsored workshop report on biodiversity and climate change. Zenodo. https://zenodo.org/
- **Post DM**, Palkovacs EP. 2009. Eco-evolutionary feedbacks in community and ecosystem ecology: interactions between the ecological theatre and the evolutionary play. *Philosophical Transactions of the Royal Society B Biological Sciences* **364**:1629–1640. DOI: https://doi.org/10.1098/rstb.2009.0012
- Pugnaire FI, Morillo JA, Penuelas J, Reich PB, Bardgett RD, Gaxiola A, Wardle DA, van der Putten WH. 2019. Climate change effects on plant-soil feedbacks and consequences for biodiversity and functioning of terrestrial ecosystems. *Science Advances* 5:eaaz1834. DOI: https://doi.org/10.1126/sciadv.aaz1834
- Puy J, Carmona CP, Hiiesalu I, Öpik M, Bello F, Moora M. 2021. Mycorrhizal symbiosis alleviates plant water deficit within and across generations via phenotypic plasticity. *Journal of Ecology* **110**:262–276. DOI: https://doi.org/ 10.1111/1365-2745.13810
- Reay DS, Dentener F, Smith P, Grace J, Feely RA. 2008. Global nitrogen deposition and carbon sinks. *Nature Geoscience* **1**:430–437. DOI: https://doi.org/10.1038/ngeo230
- Reich PB, Tilman D, Isbell F, Mueller K, Hobbie SE, Flynn DFB, Eisenhauer N. 2012. Impacts of biodiversity loss escalate through time as redundancy fades. *Science (New York, N.Y.)* **336**:589–592. DOI: https://doi.org/10. 1126/science.1217909, PMID: 22556253
- Rillig MC, Ryo M, Lehmann A, Aguilar-Trigueros CA, Buchert S, Wulf A, Iwasaki A, Roy J, Yang G. 2019. The role of multiple global change factors in driving soil functions and microbial biodiversity. *Science (New York, N.Y.)* 366:886–890. DOI: https://doi.org/10.1126/science.aay2832
- Rolando JL, Ramirez DA, Yactayo W, Monneveux P, Quiroz R. 2015. Leaf greenness as a drought tolerance related trait in potato (Solanum tuberosum L. *Environmental and Experimental Botany* **110**:27–35. DOI: https://doi.org/10.1016/j.envexpbot.2014.09.006
- Roscher C, Schumacher J, Baade J, Wilcke W, Gleixner G, Weisser WW, Schmid B, Schulze ED. 2004. The role of biodiversity for element cycling and trophic interactions: an experimental approach in a grassland community. *Basic and Applied Ecology* 5:107–121. DOI: https://doi.org/10.1078/1439-1791-00216
- Roscher C, Schumacher J, Weisser WW, Schmid B, Schulze ED. 2007. Detecting the role of individual species for overyielding in experimental grassland communities composed of potentially dominant species. *Oecologia* 154:535–549. DOI: https://doi.org/10.1007/s00442-007-0846-4, PMID: 17851699
- Rottstock T, Kummer V, Fischer M, Joshi J. 2017. Rapid transgenerational effects in Knautia arvensis in response to plant community diversity. *Journal of Ecology* **105**:714–725. DOI: https://doi.org/10.1111/1365-2745.12689
- Ruosteenoja K, Markkanen T, Venäläinen A, Räisänen P, Peltola H. 2017. Seasonal soil moisture and drought occurrence in Europe in CMIP5 projections for the 21st century. *Climate Dynamics* **50**:1177–1192. DOI: https://doi.org/10.1007/s00382-017-3671-4
- Sage RF. 2020. Global change biology: A primer. *Global Change Biology* **26**:3–30. DOI: https://doi.org/10. 1111/gcb.14893, PMID: 31663217

- Schnitzer SA, Klironomos JN, Hillerislambers J, Kinkel LL, Reich PB, Xiao K, Rillig MC, Sikes BA, Callaway RM, Mangan SA, van Nes EH, Scheffer M. 2011. Soil microbes drive the classic plant diversity-productivity pattern. *Ecology* 92:296–303. DOI: https://doi.org/10.1890/10-0773.1, PMID: 21618909
- Siebenkäs A, Schumacher J, Roscher C. 2015. Phenotypic plasticity to light and nutrient availability alters functional trait ranking across eight perennial grassland species. *AoB PLANTS* 7:plv029. DOI: https://doi.org/10.1093/aobpla/plv029
- terHorst CP, Zee PC, Bailey JK. 2016. Eco-evolutionary dynamics in plant–soil feedbacks. *Functional Ecology* **30**:1062–1072. DOI: https://doi.org/10.1111/1365-2435.12671
- Thakur MP, Reich PB, Hobbie SE, Stefanski A, Rich R, Rice KE, Eddy WC, Eisenhauer N. 2018. Reduced feeding activity of soil detritivores under warmer and drier conditions. *Nature Climate Change* 8:75–78. DOI: https:// doi.org/10.1038/s41558-017-0032-6, PMID: 29375673
- Thakur MP, van der Putten WH, Wilschut RA, Veen GFC, Kardol P, van Ruijven J, Allan E, Roscher C, van Kleunen M, Bezemer TM. 2021. Plant-Soil Feedbacks and Temporal Dynamics of Plant Diversity-Productivity Relationships. *Trends in Ecology & Evolution* **36**:651–661. DOI: https://doi.org/10.1016/j.tree.2021.03.011, PMID: 33888322
- Tilman D, Reich PB, Knops JMH. 2006. Biodiversity and ecosystem stability in a decade-long grassland experiment. *Nature* **441**:629–632. DOI: https://doi.org/10.1038/nature04742, PMID: 16738658
- van Moorsel SJ, Hahl T, Wagg C, De Deyn GB, Flynn DFB, Zuppinger-Dingley D, Schmid B. 2018a. Community evolution increases plant productivity at low diversity. *Ecology Letters* 21:128–137. DOI: https://doi.org/10. 1111/ele.12879, PMID: 29148170
- van Moorsel SJ, Schmid MW, Hahl T, Zuppinger-Dingley D, Schmid B. 2018b. Selection in response to community diversity alters plant performance and functional traits. *Perspectives in Plant Ecology, Evolution and Systematics* 33:51–61. DOI: https://doi.org/10.1016/j.ppees.2018.05.002
- van Moorsel SJ, Schmid MW, Wagemaker NCAM, van Gurp T, Schmid B, Vergeer P. 2019. Evidence for rapid evolution in a grassland biodiversity experiment. *Molecular Ecology* 28:4097–4117. DOI: https://doi.org/10. 1111/mec.15191, PMID: 31336411
- van Ruijven J, Ampt E, Francioli D, Mommer L. 2020. Do soil-borne fungal pathogens mediate plant diversity– productivity relationships? Evidence and future opportunities. *Journal of Ecology* **108**:1810–1821. DOI: https://doi.org/10.1111/1365-2745.13388
- Wei W, Yang M, Liu Y, Huang H, Ye C, Zheng J, Guo C, Hao M, He X, Zhu S. 2018. Fertilizer N application rate impacts plant-soil feedback in a sanqi production system. *Science of the Total Environment* 633:796–807. DOI: https://doi.org/10.1016/j.scitotenv.2018.03.219Get
- Weisser WW, Roscher C, Meyer ST, Ebeling A, Luo GJ, Allan E, Besser H, Barnard RL, Buchmann N, Buscot F, Engels C, Fischer C, Fischer M, Gessler A, Gleixner G, Halle S, Hildebrandt A, Hillebrandt H, de Kroon H, Lange M, et al. 2017. Biodiversity effects on ecosystem functioning in a 15-year grassland experiment: Patterns, mechanisms, and open questions. *Basic and Applied Ecology* 23:1–73. DOI: https://doi.org/10.1016/j.baae. 2017.06.002
- Wilschut RA, van Kleunen M. 2021. Drought alters plant-soil feedback effects on biomass allocation but not on plant performance. *Plant and Soil* **462**:285–296. DOI: https://doi.org/10.1007/s11104-021-04861-9
- Xi N, Adler PB, Chen D, Wu H, Catford JA, Bodegom PM, Bahn M, Crawford KM, Chu C. 2021. Relationships between plant-soil feedbacks and functional traits. *Journal of Ecology* **109**:3411–3423. DOI: https://doi.org/ 10.1111/1365-2745.13731
- Yang L, Maron JL, Callaway RM. 2015. Inhibitory effects of soil biota are ameliorated by high plant diversity. Oecologia 179:519–525. DOI: https://doi.org/10.1007/s00442-015-3351-1
- Yang G, Roy J, Veresoglou SD, Rillig MC. 2021. Soil biodiversity enhances the persistence of legumes under climate change. New Phytologist **229**:2945–2956. DOI: https://doi.org/10.1111/nph.17065
- Zuppinger-Dingley D, Schmid B, Petermann JS, Yadav V, De Deyn GB, Flynn DF. 2014. Selection for niche differentiation in plant communities increases biodiversity effects. *Nature* 515:108–111. DOI: https://doi.org/ 10.1038/nature13869

Appendix 1

Hypothesis 3: offspring of plants selected at different diversity and grown in different soil (high vs. low diversity, home vs. away) respond differently to global change drivers

Plant traits and pathogen infestation (across species and for each species) Growth height did not differ depending on soil or plant history when plants were treated with global change drivers across all study species and for *D. glomerata* (Table S1, S4). Plants of *A. elatius* in home soil were smaller than plants in away-same soil (Table S2). Nitrogen input had no influence, while plants were tallest in home soil and smallest in away-different soil when treated with both global change drivers (Table S2). Plants of *A. pratensis* exposed to drought were taller when grown in home than in away-different soil; however, this positive home effect was also only found in control plants (marginal significant; Table S3). When fertilized, the positive home effect on growth height disappeared (Table S3). Plants of *P. trivialis* were taller in two- than in six-species community soil when treated with both global change drivers, but they were not different when treated separately with drought or nitrogen input (Table S5).

Leaf greenness and shoot nitrogen concentrations were not influenced by legacy treatments when exposed to drought. When fertilized, plants still did not differ in leaf greenness but had higher shoot nitrogen concentrations in six-species than in two-species soil, found across all study species and for *D. glomerata* (Table S1, S4). Moreover, fertilized plants had a lower shoot nitrogen concentration in home than in away-different soil, found across all species and for *A. pratensis* (Table S1, S3). When plants were treated with both global change drivers, the nitrogen input effect on soil history was cancelled out by drought (across all species and for *D. glomerata*), while the impact of soil treatment did not: plants in home soil still had lower shoot nitrogen concentration than plants in away soil (across all species and for *A. pratensis*).

Plants treated with global change drivers did not differ significantly in LDMC or SLA dependent on legacy treatments, across all study species and in *A. elatius* (Table S1, S2). Drought resulted in higher LDMC of *A. pratensis* plants grown in six-species soil, and the combined application of drought and nitrogen input resulted in lower SLA in home than in away soil (Table S3). Fertilized *D. glomerata* plants had higher SLA in six- than in two-species community soil (Table S4). Plants of *P. trivialis* treated with both global change drivers had lower LDMC in two- than in six-species community soil (Table S5).

Stomatal conductance (g_s) did not differ significantly depending on legacy treatments when plants were treated with global change drivers across all study species and for *A. elatius* and *P. trivialis* (Table S1, S2, S5). In *A. pratensis*, fertilized plants showed a lower g_s when grown in home than in away soil. This effect was cancelled out by drought, when treated with both global change drivers (Table S3). In *D. glomerata*, plants had higher g_s when originated from six-species communities and treated with both global change drivers; however, this was also found in control plants (Table S4).

Across all study species, root diameter, SRL and RLD were not influenced by legacy treatments when treated with global change drivers (Table S1). In *A. elatius*, root traits also did not differ, when treated with single global change drivers, but under the combined influence of both global change drivers, plants grown in away-different soil showed the highest SRL, and plants in away-same soil had the lowest SRL (Table S2). In *A. pratensis*, plants exposed to drought had higher SRL and RLD in two- than in six-species soil. When fertilized, we did not find an effect of legacy treatment, but the combination of both global change drivers led to higher SRL and lower root diameter when plants were grown in away-same than in away-different or home soil (Table S3). In *D. glomerata*, RLD of plants exposed to drought was higher when originated from six-species than from two-species communities. This positive diversity impact disappeared when fertilized (Table S4). In *P. trivialis*, SRL were lower in plants grown in six-species community soil, when exposed to drought. When fertilized, this difference disappeared (Table S5).

Mildew infestation of *D. glomerata* plants exposed to drought was higher in home than in away soil, while this drought effect was cancelled out by nitrogen input (Table S6). Mildew infestation of *P. trivialis* plants was not significantly influenced by plant or soil history, neither with nor without global change drivers (Table S6).

Appendix 1—table 1. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio) of *A. elatius*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

							Total	biomass	5				
			Cont	rol		Drou	ght		Nitrog	en		D x N	
A. elatius		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.35	0.557	1	0.82	0.364	1	0.71	0.401	1	0.26	0.613
Soil history		1	1.08	0.298	1	0.76	0.383	1	0.06	0.811	1	0.47	0.494
Soil treatment		2	0.10	0.949	2	2.91	0.233	2	6.44	0.040	2	0.99	0.610
	_						Shoot	biomass	5				
			Contr	ol		Droug	Jht		Nitroge	en		D x N	
		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.12	0.726	1	3.36	0.067	1	1.27	0.260	1	0.01	0.904
Soil history		1	0.35	0.556	1	0.24	0.621	1	0.55	0.460	1	0.63	0.428
Soil treatment		2	2.08	0.354	2	2.89	0.236	2	5.24	0.073	2	0.98	0.613
							Root I	biomass					
			Contr	ol		Droug	Jht		Nitroge	en		D x N	
		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.03	0.860	1	0.15	0.701	1	0.01	0.916	1	0.36	0.551
Soil history		1	3.81	0.051	1	0.62	0.433	1	0.17	0.676	1	0.22	0.636
Soil treatment		2	2.05	0.359	2	2.38	0.304	2	2.25	0.325	2	1.68	0.432
							Root-shoo	t ratio					
		Con	trol			Drought	:		Nitroge	en		D x N	
	Df	Chi ²		р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.07		0.797	1	1.62	0.203	1	0.16	0.691	1	0.31	0.576
Soil history	1	4.86		0.027	1	0.24	0.626	1	0.50	0.479	1	0.07	0.787
Soil treatment	2	3.11		0.211	2	2.39	0.302	2	0.18	0.915	2	1.88	0.391
Soil treatment Plant history Soil history Soil treatment	Df 1 1 2	2 Con Chi ² 0.07 4.86 3.11	2.05	0.359 P 0.797 0.027 0.211	2 Df 1 1 2	2.38 Drought Chi ² 1.62 0.24 2.39	0.304 Root-shood D D D D D D D D D D D D D	2 t ratio Df 1 1 2	2.25 Nitroge Chi ² 0.16 0.50 0.18	0.325 P 0.691 0.479 0.915	2 Df 1 1 2	1.68 D x N Chi ² 0.31 0.07 1.88	0.432 P 0.57 0.78 0.35

Appendix 1—table 2. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio) of *A. pratensis*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

						Total I	biomass					
		Contro	bl		Droug	ht		Nitrog	en		D x N	I
A. pratensis	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.05	0.820	1	0.27	0.603	1	1.63	0.202	1	1.44	0.230
Soil history	1	0.02	0.879	1	1.05	0.306	1	2.97	0.085	1	2.07	0.151
Soil treatment	2	3.43	0.180	2	0.17	0.917	2	1.29	0.525	2	2.80	0.247

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									-				
			Contro	bl		Droug	ht		Nitroge	en		D x N	
		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.11	0.741	1	0.29	0.590	1	0.65	0.421	1	2.23	0.135
Soil history		1	0.14	0.710	1	0.33	0.564	1	0.86	0.354	1	<0.01	0.971
Soil treatment		2	0.15	0.927	2	1.84	0.398	2	1.03	0.596	2	1.35	0.509
							Root	biomas	s				
			Contr	ol		Drou	ght		Nitrog	jen		D x N	
		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.13	0.719	1	0.23	0.629	1	0.97	0.324	1	0.47	0.495
Soil history		1	0.15	0.703	1	1.16	0.281	1	1.83	0.176	1	3.98	0.046
Soil treatment		2	2.78	0.250	2	1.38	0.501	2	0.47	0.789	2	3.16	0.206
							Root-shoo	t ratio					
		Cor	ntrol			Drought			Nitrog	en		D x N	
	Df	Chi	2	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.13	0	.719	1	0.01	0.920	1	0.30	0.584	1	0.90	0.342
Soil history	1	0.20	0 0	.654	1	0.31	0.579	1	0.42	0.517	1	4.57	0.033
Soil treatment	2	1.33	0	.514	2	4.94	0.084	2	0.04	0.982	2	0.37	0.832

Shoot biomass

Appendix 1—table 3. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio) of *D. glomerata*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

						Total	biomass	5				
		Contro	ol		Drough	nt		Nitrog	en		D x N	
D. glomerata	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.56	0.456	1	2.22	0.136	1	3.09	0.079	1	0.13	0.715
Soil history	1	6.28	0.012	1	0.76	0.384	1	0.73	0.394	1	<0.01	0.978
Soil treatment	2	1.52	0.467	2	0.94	0.626	2	1.26	0.533	2	0.73	0.693
						Shoot	: biomas	55				
		Cont	rol		Droug	ght		Nitro	gen		DxN	1
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.02	0.885	1	1.28	0.259	1	3.18	0.075	1	0.22	0.640

						Root I	piomass					
		Contro	bl		Droug	nt		Nitroge	en		D x N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.40	0.236	1	2.55	0.111	1	1.98	0.160	1	0.04	0.848
Soil history	1	0.90	0.343	1	0.45	0.501	1	0.99	0.319	1	0.21	0.644
Soil treatment	2	2.49	0.288	2	2.06	0.358	2	0.02	0.992	2	3.16	0.206

0.81

0.44

1

2

0.369

0.801

0.33

3.34

1

2

0.567

0.188

0.004

0.216

8.27

3.06

1

2

Soil history

Soil treatment

0.700

0.932

0.15

0.14

1

2

		Control			Drough	t		Nitroge	n		D x N	
	Df	Chi ²	р									
Plant history	1	1.65	0.199	1	1.71	0.191	1	0.93	0.335	1	0.01	0.936
Soil history	1	<0.01	0.983	1	0.44	0.505	1	0.43	0.514	1	0.75	0.387
Soil treatment	2	3.14	0.208	2	2.84	0.242	2	0.20	0.906	2	7.72	0.021

Root-shoot ratio

Appendix 1—table 4. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio) of *P. trivialis*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

						Total b	ioma	55				
		Contro	ol		Droug	ht		Nitrog	en		D x ľ	N
P. trivialis	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.12	0.732	1	1.25	0.264	1	0.28	0.599	1	0.43	0.513
Soil history	1	0.12	0.731	1	0.14	0.704	1	0.07	0.796	1	0.05	0.826
Soil treatment	2	0.01	0.995	2	1.82	0.404	2	1.69	0.430	2	4.06	0.131
						Shoot l	bioma	155				
		Cont	rol		Drou	ght		Nitrog	gen		D x	N
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.01	0.920	1	1.91	0.167	1	0.39	0.532	1	0.01	0.943
Soil history	1	<0.01	0.973	1	0.47	0.492	1	0.46	0.499	1	0.19	0.663

	Root biomass													
		Contr	ol		Droug	ıht		Nitrog	en		Dxl	N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р		
Plant history	1	0.21	0.647	1	0.66	0.417	1	1.48	0.224	1	1.45	0.229		
Soil history	1	0.33	0.566	1	1.24	0.266	1	0.74	0.389	1	0.03	0.870		
Soil treatment	2	1.36	0.506	2	1.10	0.577	2	1.99	0.370	2	5.03	0.081		

0.81

0.667

2

1.22

0.545

2

2.96

0.227

2

1.34

Soil treatment

0.511

2

		Root-shoot ratio												
		Contro	ol		Droug	nt		Nitroge	en		D x N	1		
	Df	Chi ²	р											
Plant history	1	0.23	0.630	1	0.14	0.708	1	2.00	0.158	1	2.25	0.134		
Soil history	1	0.23	0.630	1	3.19	0.074	1	1.57	0.211	1	0.15	0.697		
Soil treatment	2	3.61	0.164	2	0.68	0.711	2	2.16	0.340	2	5.12	0.077		

Appendix 1—table 5. Summary of mixed-effect model analyses testing the effects of species identity, legacy treatments (plant history, soil history, soil treatment) and their interactions on plant trait expressions, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

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Growth	height
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	Contr	Control		Drou	ıght		Nitro	ogen		D x N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Species ID	3	36.51	<0.001	3	46.47	<0.001	3	26.45	<0.001	3	53.85	<0.001
Plant history (PH)	1	1.76	0.185	1	1.08	0.299	1	0.06	0.812	1	0.75	0.387
Soil history (SH)	1	0.48	0.488	1	0.86	0.354	1	1.52	0.217	1	1.40	0.237
Soil treatment (ST)	2	3.99	0.136	2	5.49	0.064	2	2.68	0.262	2	4.37	0.113
Species ID x PH	3	4.12	0.249	3	4.53	0.210	3	2.62	0.455	3	0.17	0.982
Species ID x SH	3	3.65	0.301	3	1.16	0.762	3	1.14	0.766	3	6.66	0.084
Species ID x ST	6	8.19	0.224	6	13.52	0.035	6	6.01	0.423	6	7.18	0.305

	Shoot nitrogen concentration													
	Conti	rol		Dro	ught		Nitr	ogen		DxI	N			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р		
Species ID	3	49.63	<0.001	3	23.08	<0.001	3	73.52	<0.001	3	30.02	<0.001		
Plant history (PH)	1	0.94	0.333	1	0.08	0.775	1	0.50	0.480	1	0.03	0.871		
Soil history (SH)	1	<0.01	0.963	1	1.50	0.221	1	4.67	0.031	1	<0.01	0.953		
Soil treatment (ST)	2	2.94	0.230	2	1.32	0.517	2	7.52	0.023	2	8.53	0.014		
Species ID x PH	3	2.80	0.424	3	5.03	0.170	3	4.00	0.262	3	2.20	0.533		
Species ID x SH	3	1.14	0.767	3	2.99	0.392	3	7.02	0.071	3	0.31	0.958		
Species ID x ST	6	12.36	0.054	6	6.88	0.332	6	6.13	0.409	6	4.73	0.579		

	Leaf greenness												
	Cont	rol		Drou	ıght		Nitro	gen		Dx	J		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Species ID	3	45.88	<0.001	3	44.96	<0.001	3	54.85	<0.001	3	71.04	<0.001	
Plant history (PH)	1	1.61	0.204	1	0.11	0.740	1	0.43	0.514	1	0.02	0.876	
Soil history (SH)	1	0.18	0.675	1	1.84	0.175	1	1.04	0.308	1	0.11	0.738	
Soil treatment (ST)	2	2.10	0.350	2	1.62	0.444	2	0.41	0.813	2	1.62	0.445	
Species ID x PH	3	4.39	0.222	3	3.98	0.264	3	1.88	0.600	3	2.78	0.427	
Species ID x SH	3	4.45	0.216	3	3.44	0.329	3	0.89	0.829	3	0.35	0.950	
Species ID x ST	6	3.54	0.739	6	3.92	0.688	6	8.79	0.186	6	3.38	0.759	

	LDMC											
	Control			Dro	ught		Niti	ogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Species ID	3	32.76	<0.001	3	22.47	<0.001	3	78.30	<0.001	3	43.04	<0.001
Plant history (PH)	1	0.33	0.565	1	2.01	0.156	1	0.03	0.861	1	0.03	0.870
Soil history (SH)	1	0.02	0.887	1	0.56	0.456	1	0.06	0.808	1	0.17	0.680
Soil treatment (ST)	2	2.83	0.243	2	1.27	0.529	2	1.34	0.511	2	0.80	0.670
Species ID x PH	3	1.71	0.635	3	0.26	0.967	3	1.00	0.802	3	4.79	0.188
Species ID x SH	3	1.69	0.638	3	4.04	0.257	3	5.48	0.140	3	2.91	0.405
Species ID x ST	6	3.52	0.742	6	1.10	0.981	6	5.73	0.454	6	11.22	0.082

	SLA												
	Control			Dro	ught		Nitr	ogen		Dх	N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Species ID	3	86.36	<0.001	3	57.20	<0.001	3	101.71	<0.001	3	73.53	<0.001	
Plant history (PH)	1	0.19	0.661	1	0.39	0.530	1	1.55	0.214	1	0.33	0.567	

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	SLA											
	Control			Dro	ught		Nitr	rogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Soil history (SH)	1	0.64	0.425	1	0.01	0.926	1	3.35	0.067	1	0.26	0.607
Soil treatment (ST)	2	4.38	0.112	2	1.43	0.488	2	2.32	0.313	2	1.50	0.472
Species ID x PH	3	1.58	0.663	3	1.26	0.738	3	0.96	0.810	3	4.38	0.223
Species ID x SH	3	2.26	0.521	3	1.47	0.690	3	3.69	0.297	3	1.90	0.592
Species ID x ST	6	2.38	0.882	6	2.88	0.824	6	4.08	0.666	6	14.22	0.027

	Stomatal co	nductance											
	Control			Dro	ught		Nitr	ogen		Dх	N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Temperature	1	0.75	0.388	1	1.40	0.237	1	3.18	0.074	1	0.18	0.670	
Daytime	1	18.95	<0.001	1	13.20	<0.001	1	5.72	<0.001	1	16.06	<0.001	
Species ID	3	45.36	<0.001	3	24.61	<0.001	3	42.88	<0.001	3	21.71	<0.001	
Plant history (PH)	1	0.60	0.438	1	0.01	0.910	1	0.48	0.490	1	2.95	0.086	
Soil history (SH)	1	0.10	0.757	1	0.05	0.818	1	1.15	0.283	1	0.07	0.797	
Soil treatment (ST)	2	0.08	0.963	2	2.67	0.263	2	4.85	0.088	2	0.20	0.905	
Species ID x PH	3	4.59	0.204	3	3.18	0.365	3	4.89	0.180	3	4.89	0.180	
Species ID x SH	3	2.60	0.457	3	3.53	0.317	3	3.23	0.358	3	3.36	0.340	
Species ID x ST	6	8.36	0.213	6	4.47	0.614	6	3.82	0.701	6	4.76	0.575	

Root diameter

	Cont	Control		Dro	ught		Nitro	ogen		Dxl	N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Species ID	3	97.02	<0.001	3	103.81	<0.001	3	93.37	<0.001	3	106.66	<0.001	
Plant history (PH)	1	0.87	0.352	1	0.02	0.883	1	<0.01	0.951	1	0.08	0.775	
Soil history (SH)	1	0.17	0.680	1	0.22	0.643	1	1.41	0.235	1	0.03	0.873	
Soil treatment (ST)	2	2.42	0.298	2	0.93	0.629	2	1.28	0.528	2	0.46	0.793	
Species ID x PH	3	0.79	0.852	3	0.19	0.979	3	4.53	0.291	3	3.28	0.350	
Species ID x SH	3	6.10	0.107	3	3.40	0.334	3	5.40	0.145	3	0.31	0.959	
Species ID x ST	6	9.36	0.155	6	2.06	0.914	6	1.41	0.965	6	13.49	0.036	

	SKL											
	Cont	rol		Dro	ught		Nit	rogen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Species ID	3	125.58	<0.001	3	123.96	<0.001	3	117.21	<0.001	3	144.90	<0.001
Plant history (PH)	1	0.31	0.579	1	0.04	0.833	1	2.81	0.094	1	0.05	0.830
Soil history (SH)	1	<0.01	0.986	1	1.17	0.279	1	1.37	0.242	1	1.48	0.224
Soil treatment (ST)	2	1.46	0.483	2	0.67	0.717	2	4.01	0.135	2	0.28	0.869
Species ID x PH	3	5.15	0.161	3	2.11	0.550	3	2.96	0.397	3	2.31	0.510
Species ID x SH	3	3.89	0.274	3	6.14	0.105	3	3.40	0.334	3	1.93	0.586
Species ID x ST	6	13.23	0.040	6	2.92	0.819	6	2.90	0.821	6	14.70	0.023

	RLD											
	Cont	rol		Dro	ught		Nitr	ogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Species ID	3	99.14	<0.001	3	101.33	<0.001	3	91.27	<0.001	3	75.25	<0.001

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	RLD											
	Con	trol		Drou	ught		Nitr	ogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history (PH)	1	0.00	0.956	1	3.36	0.067	1	0.11	0.742	1	0.98	0.323
Soil history (SH)	1	2.93	0.087	1	0.14	0.710	1	0.67	0.413	1	0.55	0.460
Soil treatment (ST)	2	2.50	0.286	2	2.56	0.279	2	0.03	0.983	2	4.98	0.083
Species ID x PH	3	1.35	0.716	3	5.11	0.164	3	2.59	0.459	3	0.59	0.900
Species ID x SH	3	5.42	0.144	3	2.89	0.409	3	0.45	0.929	3	0.49	0.921
Species ID x ST	6	2.77	0.838	6	4.44	0.617	6	0.91	0.989	6	6.27	0.393

Appendix 1—table 6. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant trait expressions of *A. elatius*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

A. elatius	Grov	vth height										
	Cont	rol		Droug	ght		Nit	rogen		DxI	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.32	0.569	1	2.94	0.087	1	1.01	0.314	1	0.13	0.719
Soil history	1	1.50	0.221	1	0.07	0.787	1	0.14	0.706	1	0.29	0.593
Soil treatment	2	2.67	0.263	2	10.64	0.005	2	1.55	0.461	2	7.58	0.023
	Sho	ot nitroge	n concentra	tion								
	Con	trol		Drou	ght		Nitro	gen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.52	0.472	1	3.46	0.063	1	<0.01	0.974	1	0.06	0.802
Soil history	1	0.89	0.347	1	0.04	0.843	1	1.64	0.200	1	0.06	0.803
Soil treatment	2	1.40	0.497	2	1.54	0.462	2	1.99	0.369	2	2.07	0.354
	Leaf	greennes	s									
	Con	trol		Dro	ught		Nitr	ogen		D x I	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.19	0.275	1	0.60	0.438	1	1.13	0.288	1	0.22	0.636
Soil history	1	1.50	0.221	1	0.99	0.321	1	0.03	0.862	1	0.15	0.699
Soil treatment	2	5.20	0.074	2	0.44	0.801	2	3.64	0.162	2	0.84	0.656
	LDN	IC										
	Con	trol		Drou	ght		Nitro	gen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.01	0.942	1	1.15	0.284	1	<0.01	0.987	1	1.02	0.313
Soil history	1	0.07	0.798	1	0.13	0.718	1	0.04	0.837	1	0.31	0.580
Soil treatment	2	0.03	0.985	2	0.34	0.844	2	2.00	0.369	2	2.44	0.295
	SLA											
	Con	trol		Dro	ught		Nitr	ogen		D x I	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.44	0.507	1	0.61	0.435	1	0.48	0.488	1	1.63	0.202
Soil history	1	0.04	0.836	1	0.22	0.638	1	0.88	0.348	1	1.08	0.300
Soil treatment	2	0.59	0.744	2	0.13	0.936	2	2.74	0.254	2	3.10	0.212

	Cont	rol	Drought Nitrogen D x N									
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Temperature	1	0.05	0.827	1	0.53	0.465	1	0.91	0.340	1	0.09	0.763
Daytime	1	6.15	0.013	1	3.92	0.048	1	0.68	0.408	1	0.37	0.544
Plant history	1	0.49	0.484	1	0.05	0.824	1	1.23	0.267	1	0.18	0.670
Soil history	1	0.83	0.361	1	0.13	0.718	1	0.92	0.336	1	<0.01	0.998
Soil treatment	2	0.96	0.618	2	1.69	0.429	2	2.99	0.224	2	0.33	0.846
	Roo	t diamete	r									
	Con	trol		Drou	ight		Ni	trogen		D>	< N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.24	0.627	1	<0.01	0.972	1	0.46	0.497	1	0.45	0.503
Soil history	1	1.37	0.242	1	0.53	0.467	1	2.59	0.108	1	0.10	0.754
Soil treatment	2	4.85	0.089	2	0.52	0.770	2	1.00	0.605	2	3.86	0.145
	SR	L										
	Co	ntrol		Dro	ought		Nit	rogen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.80	0.371	1	0.16	0.686	1	2.32	0.128	1	0.54	0.462
Soil history	1	0.06	0.807	1	0.02	0.884	1	2.66	0.103	1	0.21	0.649
Soil treatment	2	2.94	0.230	2	1.81	0.404	2	4.63	0.099	2	9.49	0.009
	RLD)										
	Con	trol		Dro	ught		Nitr	ogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.02	0.313	1	0.03	0.859	1	2.42	0.120	1	1.44	0.230
Soil history	1	2.51	0.113	1	1.14	0.286	1	1.03	0.310	1	0.46	0.500
Soil treatment	2	4.52	0.104	2	1.24	0.539	2	0.26	0.878	2	1.40	0.497

Stomatal conductance

Appendix 1—table 7. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant trait expressions of A. pratensis, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

A. pratensis	Gro	wth heigh	t									
	Con	trol		Dro	ought		Nitr	ogen		Dх	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.50	0.221	1	0.56	0.454	1	0.03	0.868	1	0.94	0.332
Soil history	1	0.44	0.508	1	0.15	0.700	1	0.03	0.874	1	0.82	0.365
Soil treatment	2	5.77	0.056	2	6.56	0.038	2	3.00	0.223	2	0.26	0.879
	Shoot	nitrogen	concentra	tion								
	Contr	ol		Drou	ght		Nitro	gen		D x N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.75	0.186	1	0.17	0.680	1	0.10	0.755	1	0.84	0.358
Soil history	1	0.37	0.544	1	0.96	0.328	1	0.01	0.939	1	0.00	0.966
Soil treatment	2	4.61	0.100	2	1.74	0.419	2	9.05	0.011	2	6.83	0.033

ratensis Growth height

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	Leaf	greennes	5											
	Con	trol		D	rought			Nitro	ogen			DxN	I	
	Df	Chi ²	р	D	of C	hi² p	b	Df	Cł	hi²	р	Df	Chi ²	р
Plant history	1	0.07	0.786	1	1.	03 0).311	1	0.5	58	0.445	1	0.18	0.673
Soil history	1	0.03	0.869	1	1.	85 C).174	1	0.9	90	0.343	1	0.19	0.661
Soil treatment	2	1.16	0.560	2	0.	60 ().743	2	0.0	61	0.737	2	2.21	0.332
	LDM	с												
	Cont	rol		D	rought			Nitro	ogen			DxI	J	
	Df	Chi ²	р	D	f C	ni² p)	Df	C	hi²	р	Df	Chi ²	р
Plant history	1	0.34	0.561	1	0.	38 0	.538	1	0.	40	0.527	1	2.17	0.140
Soil history	1	0.11	0.736	1	3.	62 0	.057	1	2.	32	0.128	1	0.05	0.821
Soil treatment	2	0.36	0.835	2	1.	42 0	.492	2	1.	.18	0.555	2	3.91	0.141
	SLA													
	Contro	ol			Drought			N	litrog	gen		DxI	J	
	Df	Chi ²	р		Df	Chi ²	р	D	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.07	0.78	6	1	0.32	0.572	1		0.00	0.984	1	1.28	0.259
Soil history	1	0.20	0.65	4	1	2.81	0.094	1		0.23	0.632	1	0.05	0.828
Soil treatment	2	2.21	0.33	1	2	0.70	0.704	2		1.18	0.555	2	8.59	0.014
	Stomat	al conduct	ance											
	Contro	1		Drou	ght		Nit	rogen			Dx	N		
	Df	Chi ²	р	Df	Chi ²	р	Df		Chi ²	р	Df	Ch	i²	р
Temperature	1	1.17	0.279	1	0.22	0.642	1		0.44	0.507	7 1	0.1	7	0.678
Daytime	1	0.77	0.379	1	0.07	0.786	1		1.13	0.289	7 1	8.3	8	0.004
Plant history	1	0.05	0.824	1	0.16	0.690	1		0.66	0.41	5 1	0.6	1	0.436
Soil history	1	1.30	0.255	1	0.14	0.706	1		0.79	0.373	3 1	0.5	3	0.466
Soil treatment	2	2.35	0.308	2	4.41	0.110	2		2.55	0.00	2 2	1.5	9	0.452
	Root o	liameter												
	Contro	ol		Drou	ght		Nitrog	en			D x N			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi	2	р	Df	Ch	i²	р
Plant history	1	0.28	0.595	1	0.18	0.673	1	0.20)	0.653	1	0.0	19	0.770
Soil history	1	5.61	0.018	1	0.95	0.331	1	1.34	1	0.246	1	0.0	1	0.942
Soil treatment	2	1.02	0.602	2	0.29	0.865	2	1.25	5	0.535	2	6.0	6	0.048
	SRL													
	Contr	ol		Drou	ıght		Nitrog	en			D x	N		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²		р	Df	C	hi²	р
Plant history	1	0.42	0.515	1	0.24	0.623	1	0.61		0.435	1	C	.01	0.916
Soil history	1	0.33	0.567	1	7.10	0.008	1	0.17		0.677	1	2	.73	0.098
Soil treatment	2	5.24	0.073	2	0.88	0.644	2	0.11		0.945	2	6	.03	0.049
	RLD													
	Contr	ol		Droug	ght		Nitrog	jen			DxN	1		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²		р	Df	Ch	i²	р
Plant history	1	0.28	0.595	1	0.12	0.729	1	0.08		0.781	1	0.0	19	0.763
Soil history	1	0.75	0.387	1	4.79	0.029	1	0.13		0.716	1	0.0	3	0.861

Appendix 1—table 8. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant trait expressions of *D. glomerata*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (P < 0.05) are given in bold, marginally significant effects (P < 0.1) in italics.

$\begin{tabular}{ c c c c c c c c c c } \hline P $ $Orought $$ $Nitrogen $$ $Orought $$ $Nitrogen $$ $Orought $$ Or	 P 0.912 0.675 0.581 hi² P 56 0.453 05 0.818
Df Chi ² p Df Chi ² 1 0.01 Soil history 1 0.69 0.405 1 0.91 0.340 1 1.25 0.263 1 0.18 Soil treatment 2 1.66 0.436 2 1.06 0.589 2 2.37 0.306 2 1.09 Shoot nitrogen concentration Df Chi ² p Df	 P 0.912 0.675 0.581 hi² P 56 0.453 0.5 0.818
Plant history 1 0.73 0.394 1 0.11 0.741 1 0.06 0.802 1 0.01 Soil history 1 0.69 0.405 1 0.91 0.340 1 1.25 0.263 1 0.18 Soil history 1 0.69 0.436 2 1.06 0.589 2 2.37 0.306 2 1.09 Shoot nitrogen concentration Drought Nitrogen D x N Df Chi ² p	0.912 0.675 0.581 hi ² p 56 0.453 05 0.818
Soil history 1 0.69 0.405 1 0.91 0.340 1 1.25 0.263 1 0.18 Soil treatment 2 1.66 0.436 2 1.06 0.589 2 2.37 0.306 2 1.09 Shoot nitrogen concentration Drought Nitrogen D x N Df Chi ² p Df Chi ² Qi and	 0.675 0.581 hi² p 56 0.453 05 0.818
Soil treatment 2 1.66 0.436 2 1.06 0.589 2 2.37 0.306 2 1.09 Shoot nitrogen concentration Control Drought Nitrogen D x N Df Chi ² p Df Chi ² Di Chi ² Di Chi ² Di Chi ² <	 0.581 hi² p 56 0.453 05 0.818
Shoot nitrogen concentration Control Drought Nitrogen D x N Df Chi ² p Df Chi ² Qi Q	hi ² p 56 0.453 05 0.818
Control Drought Nitrogen D x N Df Chi ² p Df Chi ² N O.5 O.55 1 2.38 O.123 1 O.5 O.5 Soil history 1 <0.01 0.952 1 2.18 O.140 1 8.44 O.004 1 0.02 Dif Dif Dif Dif Dif	hi ² p 56 0.453 05 0.818
Df Chi ² p Df Chi ² Di	hi ² p 56 0.453 05 0.818
Plant history 1 1.13 0.289 1 0.56 0.455 1 2.38 0.123 1 0.4 Soil history 1 <0.01	560.453050.818
Soil history 1 <0.01 0.952 1 2.18 0.140 1 8.44 0.004 1 0.0 Soil treatment 2 2.72 0.257 2 2.46 0.293 2 3.07 0.215 2 0.7 Leaf greenness	05 0.818
Soil treatment 2 2.72 0.257 2 2.46 0.293 2 3.07 0.215 2 0.7	
Leaf greenness	71 0.701
Control Drought Nitrogen D x N	
Df Chi ² p Df Chi ² p Df Chi ² p Df C	:hi² p
Plant history 1 4.93 0.026 1 0.02 0.886 1 0.17 0.680 1 0.	.13 0.723
Soil history 1 1.23 0.267 1 1.17 0.279 1 0.15 0.703 1 0.	.01 0.908
Soil treatment 2 2.33 0.313 2 3.58 0.167 2 1.16 0.560 2 0.	.68 0.713
LDMC	
Control Drought Nitrogen D x N	
Df Chi ² p Df Chi ² p Df Chi ² p Df Ch	hi² p
Plant history 1 0.86 0.353 1 1.18 0.278 1 0.37 0.540 1 0.0	64 0.423
Soil history 1 2.03 0.154 1 0.12 0.727 1 2.21 0.137 1 0.	28 0.594
Soil treatment 2 2.36 0.307 2 0.20 0.905 2 1.74 0.418 2 3.0	05 0.218
SLA	
Control Drought Nitrogen D x N	
Df Chi ² p Df Chi ² p Df Chi ² p Df C	:hi² p
Plant history 1 1.41 0.235 1 0.01 0.904 1 1.50 0.220 1 0.	.14 0.706
Soil history 1 2.29 0.130 1 0.28 0.595 1 3.86 0.050 1 0.	.02 0.888
Soil treatment 2 2.60 0.272 2 1.88 0.392 2 0.09 0.956 2 0.	.89 0.641
Stomatal conductance	
Control Drought Nitrogen D x N	
Df Chi ² p Df Chi ² p Df Chi ² p Df Chi ²	р
Temperature 1 1.12 0.289 1 <0.01 0.951 1 0.04 0.843 1 0.08	0.782
Daytime 1 24.06 <0.001 1 12.16 <0.001 1 4.04 0.044 1 4.37	0.037
Plant history 1 3.77 0.052 1 1.05 0.304 1 1.79 0.181 1 4.89	0.027
Soil history 1 1.44 0.231 1 1.55 0.214 1 0.47 0.493 1 2.34	0.126

	Root	diameter											
	Cont	rol		Droug	ght		Nitr	ogen			D	k N	
	Df	Chi ²	р	Df	Chi ²	р	Df		Chi ²	р	Df	Chi ²	р
Plant history	1	0.64	0.422	1	0.02	0.876	0 1		1.83	0.176	1	2.43	0.119
Soil history	1	0.33	0.567	1	2.50	0.114	1		0.34	0.559	1	0.16	0.691
Soil treatment	2	0.60	0.741	2	3.21	0.201	2		0.16	0.924	2	2.03	0.363
	SRL												
	Cont	rol		Drou	ught		Ni	troge	n		Dx	1	
	Df	Chi ²	р	Df	Chi ²	р	Df		Chi ²	р	Df	Chi ²	р
Plant history	1	2.55	0.111	1	0.54	0.462	1		0.08	0.777	1	0.36	0.548
Soil history	1	1.73	0.188	1	1.42	0.233	1		0.32	0.570	1	0.22	0.643
Soil treatment	2	2.23	0.329	2	0.24	0.888	2		2.28	0.320	2	2.38	0.304
	RLD												
	Cont	rol			Drought			Nit	rogen		DxN	1	
	Df	Chi ²	р		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.01	0.923	3	1	7.58	0.006	1	0.77	0.380	1	0.03	0.862
Soil history	1	0.27	0.602	2	1	0.02	0.901	1	0.54	0.464	1	0.18	0.673
Soil treatment	2	0.36	0.83	5	2	4.51	0.105	2	0.96	0.619	2	5.25	0.073

Appendix 1—table 9. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on plant trait expressions of *P. trivialis*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

P. trivialis	Grow	th height										
	Contr	ol		Droug	ght		Nitro	ogen		D x N	1	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	2.81	0.094	1	0.32	0.571	1	0.98	0.323	1	0.16	0.688
Soil history	1	0.62	0.429	1	0.92	0.338	1	1.12	0.289	1	5.02	0.025
Soil treatment	2	4.77	0.092	2	1.59	0.452	2	2.99	0.224	2	1.14	0.566
	Shoot	nitrogen co	oncentration									
	Contro	ol		Drou	ıght		Nit	rogen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	<0.01	0.986	1	0.01	0.934	1	0.07	0.785	1	0.01	0.915
Soil history	1	0.15	0.695	1	0.57	0.452	1	0.06	0.802	1	0.45	0.503

	Leaf g	reenness										
	Contro	bl		Droug	ht		Nitro	ogen		Dх	N	
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	Р
Plant history	1	0.14	0.708	1	2.41	0.120	1	0.04	0.845	1	2.35	0.126
Soil history	1	1.41	0.236	1	1.10	0.295	1	0.38	0.537	1	0.09	0.769
Soil treatment	2	0.13	0.936	2	0.37	0.833	2	5.22	0.074	2	0.97	0.616

0.313

2

1.18

0.554

2

3.86

0.145

2.33

Soil treatment

2

9.66

0.008

2

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	LDMC											
	Contr	ol			Drought			Nitrogen		Dх	N	
	Df	Chi ²	Р		Df Chi ²	Р		Df Cł	ni ² P	Df	Chi ²	Р
Plant history	1	0.81	0.369		1 0.24	0.627		1 0.0	0.826	1	1.34	0.247
Soil history	1	0.08	0.776		1 0.01	0.927		1 0.4	17 0.492	1	4.25	0.039
Soil treatment	2	3.34	0.188		2 0.72	0.696		2 3.0	0.222	2	2.64	0.268
	SLA											
	Contro	1		Dro	ought			Nitrogen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р		Df Ch	i² p	Df	Chi ²	р
Plant history	1	0.26	0.611	1	0.21	0.643		1 1.4	4 0.231	1	0.80	0.372
Soil history	1	0.41	0.522	1	0.40	0.528		1 1.4	7 0.226	1	0.33	0.565
Soil treatment	2	2.29	0.319	2	0.53	0.769		2 3.3	5 0.187	2	4.08	0.130
	Stomata	al conductar	ice									
	Control			D	rought			Nitroger	1	Dx	N	
	Df	Chi ²	р	D	f Chi ²	р		Df C	hi² p	Df	Chi ²	р
Temperature	1	10.96	0.001	1	8.08	0.004	ļ.	1 7.	25 0.007	1	4.31	0.038
Daytime	1	3.93	0.047	1	1.12	0.289		1 1.	22 0.270	1	6.35	0.012
Plant history	1	<0.01	0.949	1	0.60	0.439		1 2.	96 0.085	1	0.29	0.589
Soil history	1	0.68	0.410	1	0.95	0.330		1 2.	72 0.099	1	0.14	0.704
Soil treatment	2	2.46	0.293	2	0.54	0.763		2 0.	95 0.622	2	1.49	0.474
	Root dia	meter										
	Control			Droug	ht		Nitro	gen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.16	0.686	1	0.07	0.794	1	0.55	0.458	1	2.91	0.088
Soil history	1	3.06	0.080	1	0.31	0.579	1	0.95	0.329	1	0.06	0.800
Soil treatment	2	7.48	0.024	2	0.28	0.870	2	0.07	0.967	2	2.00	0.369
	SRL											
	Control			Drou	ght		Ni	trogen		D x	N	
	Df	Chi ²	р	Df	Chi ²	р	Df	Ch	i² p	Df	Chi ²	р
Plant history	1	2.10	0.147	1	0.94	0.332	1	1.8	2 0.178	1	1.04	0.308
Soil history	1	1.83	0.177	1	3.68	0.055	1	2.2	0.133	1	0.19	0.660
Soil treatment	2	5.73	0.057	2	0.56	0.755	2	1.9	0.374	2	1.83	0.401
	RLD											
	Control			Drough	nt		Nit	rogen		D x N	i	
	Df	Chi ²	р	Df	Chi ²	р	Df	Ch	i² p	Df	Chi ²	р
Plant history	1	0.23	0.632	1	0.01	0.904	1	0.0	0.920	1	0.54	0.463
Soil history	1	3.38	0.066	1	0.07	0.792	1	0.0	0.926	1	0.16	0.685
Soil treatment	2	0.63	0.731	2	0.61	0.739	2	0.1	6 0.924	2	3.25	0.197

Appendix 1—table 10. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment) on mildew infestation of *D. glomerata* and *P. trivialis*, when non-treated (control) or treated with GC drivers (drought, nitrogen input, drought and nitrogen input (D x N)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

Mildew infestation							D. g	lomerat	а				
			Contro	l		Droug	Jht		Nitrog	jen		DxN	1
		Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history		1	0.58	0.447	1	1.18	0.277	1	0.88	0.348	1	0.26	0.613
Soil history		1	0.41	0.522	1	2.63	0.105	1	<0.01	0.946	1	0.11	0.746
Soil treatment		2	6.01	0.049	2	7.65	0.022	2	0.93	0.628	2	0.09	0.958
							P. triviali	s					
		Control			Dro	ught			Nitrogen			D x N	
	Df	Chi ²	р	Df	Chi	2	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	<0.01	0.996	1	<0.0	1 ().973	1	0.03	0.860	1	0.21	0.647
Soil history	1	1.20	0.274	1	2.66	. (0.103	1	1.68	0.195	1	0.05	0.817
Soil treatment	2	3.94	0.139	2	1.78	3 ().412	2	0.16	0.921	2	2.10	0.350

Appendix 1—table 11. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment) and their interactions on root-shoot ratio, when non-treated (control) or treated with global change drivers (drought, nitrogen input, drought and nitrogen input ($D \times N$)).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

		Root-Shoot ratio												
		Contro	ol		Droug	ht		Nitrog	en	D x N				
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р		
Species ID	3	115.37	<0.001	3	116.36	<0.001	3	101.12	<0.001	3	108.37	<0.001		
Plant history (PH)	1	0.02	0.880	1	1.48	0.225	1	1.64	0.200	1	0.46	0.496		
Soil history (SH)	1	1.81	0.178	1	1.60	0.206	1	0.24	0.622	1	<0.01	0.992		
Soil treatment (ST)	2	0.46	0.793	2	1.96	0.376	2	1.19	0.551	2	3.54	0.170		
Species ID x PH	3	3.88	0.275	3	1.47	0.690	3	0.86	0.836	3	2.77	0.428		
Species ID x SH	3	5.98	0.113	3	3.99	0.263	3	2.53	0.471	3	3.71	0.295		
Species ID x ST	6	10.54	0.104	6	6.76	0.344	6	1.85	0.933	6	14.79	0.022		

Appendix 2

Hypothesis 2: global change drivers have a strong impact on biomass production and trait expression

Plant traits (separately for each species)

The grass *P. trivialis* was the only species which growth height decreased with drought, all other species showed no significant change under drought. Under nitrogen input, the species *P. trivialis, A. pratensis,* and *D. glomerata* (marginally significant) increased in growth height, while under the combined impact of both global change drivers, no species significantly changed in growth height (*D. glomerata* marginally significantly increased in height).

The species A.elatius, D. glomerata, and A. pratensis increased in shoot nitrogen concentration and leaf greenness under the impact of drought and/or nitrogen input (similar to analysis across all species). In P. trivialis, drought did not affect shoot nitrogen concentration or leaf greenness, and there was no additive impact of both global change drivers on leaf greenness (leaf greenness was as high as in fertilized plants).

Global change drivers had no significant influence on LDMC or SLA of *A. elatius* and *A. pratensis* except for LDMC decrease and SLA increase of *A. elatius* plants when treated with both global change drivers. Plants of *D. glomerata* decreased in LDMC and increased in SLA when treated with single global change drivers, while nitrogen input had a stronger impact than drought. When treated with both global change drivers, *D. glomerata* plants had still a significantly lower LDMC and higher SLA compared to control plants. In *P. trivialis*, drought had no significant influence on LDMC and SLA, while nitrogen input decreased LDMC and increased SLA. When treated with both global change drivers as high as in fertilized plants.

In *D. glomerata*, stomatal conductance was increased, when plants were treated with drought, and in *A. pratensis* decreased, when treated with both global change drivers. Stomatal conductance in *A. elatius* and *P. trivialis* did not change with global change treatments.

In A. elatius, SRL decreased when fertilized, irrespective of drought, while other root traits did not change significantly. In A. pratensis, drought, nitrogen input, and both global change drivers together had similar negative impacts on SRL and RLD (except for RLD under nitrogen input, which did not change). Root diameter of A. pratensis plants increased under single global change drivers with additive effects under the combined application. In D. glomerata, RLD increased and in P. trivialis RLD decreased and root diameter increased, when treated with both global change drivers.

Appendix 2—table 1. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on root-shoot ratio.

	ĸ	ot-shoot ratio	
	Df	Chi ²	р
Species identity (ID)	3	133.41	<0.001
Plant history	1	1.11	0.292
Soil history	1	1.08	0.300
Soil treatment	2	1.81	0.404
Drought (D)	1	60.01	<0.001
Nitrogen input (N)	1	89.83	<0.001
Species ID x Plant history	3	0.87	0.832
Species ID x Soil history	3	4.07	0.254
Species ID x Soil treatment	6	2.79	0.835
Species ID x D	3	95.53	<0.001

Shown are degrees of freedom (Df), Chi^2 and P-values (P). Significant effects (P < 0.05) are given in bold, marginally significant effects (P < 0.1) in italics.

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Appendix 2-table 1 Continued on next page

Appendix 2—table 1 Continued

Ro	ot-shoot ratio	
Df	Chi ²	р
3	9.31	0.025
1	2.19	0.139
4	2.02	0.733
4	1.58	0.812
8	4.97	0.760
4	2.91	0.573
4	3.18	0.528
8	18.18	0.020
4	10.42	0.034
4	11.14	0.025
8	5.20	0.736
	Df 3 1 4 4 4 4 4 8 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 8 4 4 8 4 8 4 8 4 8 4 8 4 8 4 8 4 8 4 8 8 8	Root-shoot ratio Df Chi ² 3 9.31 1 2.19 4 2.02 4 1.58 8 4.97 4 2.91 4 3.18 8 18.18 4 10.42 4 11.14 8 5.20

Appendix 2—table 2. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio).

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

	Total biomass Shoot			oot biomas	s	Roo	Root biomass			Root-shoot ratio			
	Df	Chi	р	Df	Chi	р	Df	Chi	р	Df	Chi	р	
Species ID	3	73.25	<0.001	3	80.17	<0.001	3	121.30	<0.001	3	133.41	<0.001	
Plant history	1	3.48	0.062	1	1.36	0.244	1	3.40	0.065	1	1.11	0.292	
Soil history	1	0.01	0.915	1	0.04	0.851	1	0.49	0.484	1	1.08	0.300	
Soil treatment	2	2.17	0.338	2	1.20	0.548	2	3.66	0.161	2	1.81	0.404	
Drought (D)	1	83.05	<0.001	1	110.26	<0.001	1	2.81	0.094	1	60.01	<0.001	
Nitrogen input (N)	1	257.26	<0.001	1	425.93	<0.001	1	15.89	<0.001	1	89.83	<0.001	
D×N	1	29.23	<0.001	1	23.02	<0.001	1	8.50	0.004	1	1.75	0.185	
Plant history x D	1	0.22	0.639	1	0.21	0.643	1	0.01	0.916	1	<0.01	0.977	
Soil history x D	1	<0.01	0.944	1	0.07	0.786	1	0.10	0.746	1	0.23	0.635	
Soil treatment x D	2	1.79	0.409	2	0.77	0.681	2	1.37	0.503	2	1.29	0.526	
Plant history x N	1	1.48	0.224	1	1.59	0.207	1	0.60	0.437	1	0.35	0.553	
Soil history x N	1	3.44	0.064	1	1.33	0.249	1	2.46	0.116	1	0.83	0.363	
Soil treatment x N	2	1.43	0.489	2	1.40	0.496	2	0.43	0.806	2	0.49	0.782	
Plant history x D x N	1	2.12	0.146	1	0.84	0.358	1	1.78	0.183	1	1.27	0.260	
Soil history x D x N	1	0.95	0.330	1	2.78	0.095	1	0.08	0.780	1	0.03	0.864	
Soil treatment x D x N	2	1.37	0.504	2	1.93	0.381	2	0.91	0.635	2	0.73	0.693	

Appendix 2—table 3. Summary of mixed-effect model analyses testing the effects of species identity (N = 4), legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant trait expression.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

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	Gro	wth heigh	t	Shoo	ot nitrogen co	onc.	Leaf	greenness	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Species ID	3	71.45	<0.001	3	57.20	<0.001	3	79.55	<0.001
Plant history	1	0.15	0.694	1	<0.01	0.960	1	0.05	0.830
Soil history	1	1.60	0.207	1	0.64	0.425	1	0.17	0.683
Soil treatment	2	3.98	0.137	2	2.27	0.321	2	0.60	0.742
Drought (D)	1	18.71	<0.001	1	65.46	<0.001	1	66.15	<0.001
Nitrogen input (N)	1	32.93	<0.001	1	772.20	<0.001	1	523.86	<0.001
D x N	1	1.10	0.294	1	48.85	<0.001	1	<0.01	0.997
Plant history x D	1	2.99	0.084	1	0.06	0.806	1	<0.01	0.950
Soil history x D	1	0.51	0.477	1	0.11	0.735	1	1.57	0.210
Soil treatment x D	2	3.54	0.171	2	0.02	0.990	2	0.69	0.707
Plant history x N	1	0.50	0.478	1	1.34	0.246	1	0.91	0.341
Soil history x N	1	1.41	0.235	1	0.19	0.666	1	1.54	0.215

Continued on next page

	Gro	wth height		Shoot nitrogen conc.				Leaf greenness			
	Df	Chi ²	р	Df	Chi ²	р	D	f Chi ²	р		
Soil treatment x N	2	1.87	0.392	2	3.30	0.192	2	2.42	0.299		
Plant history x D x N	1	0.83	0.364	1	0.21	0.645	1	0.79	0.373		
Soil history x D x N	1	0.69	0.407	1	3.06	0.080	1	<0.01	0.977		
Soil treatment x D x N	2	4.94	0.085	2	1.56	0.458	2	0.04	0.983		
	LDM	с		SLA			Stoma	atal conducta	nce		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р		
Air temperature	-	-	-	-	-	-	1	5.34	0.021		
Daytime	-	-	-	-	-	-	1	38.25	<0.001		
Species ID	3	80.52	<0.001	3	124.00	<0.001	3	47.15	<0.001		
Plant history	1	0.80	0.373	1	0.06	0.805	1	1.25	0.264		
Soil history	1	0.10	0.750	1	1.22	0.270	1	0.37	0.543		
Soil treatment	2	1.13	0.570	2	1.64	0.441	2	3.38	0.185		
Drought (D)	1	0.94	0.333	1	0.11	0.743	1	0.90	0.343		
Nitrogen input (N)	1	62.84	<0.001	1	61.63	<0.001	1	8.16	0.004		
D x N	1	6.69	0.010	1	0.01	0.904	1	9.33	0.002		
Plant history x D	1	0.04	0.841	1	0.34	0.559	1	0.06	0.806		
Soil history x D	1	0.49	0.484	1	0.02	0.883	1	0.65	0.420		
Soil treatment x D	2	0.24	0.887	2	0.23	0.889	2	0.18	0.914		
Plant history x N	1	0.65	0.421	1	0.16	0.688	1	0.69	0.406		
Soil history x N	1	0.12	0.734	1	1.07	0.300	1	0.63	0.428		
Soil treatment x N	2	0.66	0.719	2	2.92	0.232	2	0.08	0.960		
Plant history x D x N	1	0.16	0.687	1	1.77	0.183	1	0.87	0.351		
Soil history x D x N	1	<0.01	0.962	1	0.95	0.331	1	0.02	0.887		
Soil treatment x D x N	2	2.27	0.322	2	1.33	0.514	2	3.73	0.155		
	Roo	t diameter		SR	۲L.		R	LD			
	Df	Chi ²	р	Df	Chi ²	р	D	f Chi ²	р		
Species ID	3	165.58	<0.001	3	174.84	<0.001	3	125.84	<0.001		
Plant history	1	0.03	0.872	1	0.32	0.569	1	1.14	0.286		
Soil history	1	0.37	0.544	1	0.36	0.546	1	0.25	0.617		
Soil treatment	2	1.50	0.473	2	2.80	0.246	2	4.97	0.083		
Drought (D)	1	11.19	0.001	1	7.67	0.006	1	16.09	<0.001		
Nitrogen input (N)	1	19.83	<0.001	1	6.68	0.010	1	1.29	0.257		
D×N	1	0.25	0.619	1	1.27	0.261	1	2.14	0.144		
Plant history x D	1	0.37	0.544	1	0.34	0.559	1	0.67	0.414		
Soil history x D	1	0.12	0.725	1	0.48	0.491	1	0.07	0.798		
Soil treatment x D	2	1.67	0.434	2	0.65	0.723	2	0.44	0.802		
Plant history x N	1	0.40	0.528	1	1.91	0.167	1	<0.01	0.944		
Soil history x N	1	0.42	0.515	1	0.15	0.703	1	0.86	0.353		
Soil treatment x N	2	0.27	0.872	2	1.69	0.430	2	0.08	0.959		
Plant history x D x N	1	0.20	0.652	1	1.22	0.270	1	0.12	0.734		
Soil history x D x N	1	1.48	0.224	1	3.47	0.063	1	3.94	0.047		
Soil treatment x D x N	2	0.75	0.686	2	0.84	0.659	2	1.02	0.600		

Appendix 2—table 4. Summary of mixed-effect model analyses testing the effects legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant performance (total biomass, shoot biomass, root biomass and root-shoot ratio) of *A. elatius* and *A. pratensis*.

Shown are degrees of freedom (Df), Chi² and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

	A. elatius														
	Total	biomass		Shoo	ot biomass		Roo	ot biomas	is	Root	t-shoot rat	io			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р			
Plant history	1	0.26	0.609	1	0.05	0.827	1	0.10	0.747	1	0.11	0.738			
Soil history	1	0.39	0.533	1	0.03	0.865	1	1.02	0.312	1	1.28	0.258			
Soil treatment	2	2.06	0.357	2	1.59	0.452	2	1.31	0.520	2	0.30	0.861			
Drought (D)	1	21.54	<0.001	1	50.79	<0.001	1	6.13	0.013	1	67.84	<0.001			
Nitrogen input (N)	1	125.48	<0.001	1	128.72	<0.001	1	31.68	<0.001	1	13.70	<0.001			
D×N	1	36.23	<0.001	1	45.06	<0.001	1	1.86	0.173	1	0.13	0.715			
Plant history x D	1	1.01	0.315	1	2.37	0.123	1	0.05	0.823	1	1.28	0.258			
Soil history x D	1	0.27	0.606	1	2.01	0.156	1	0.71	0.399	1	2.11	0.146			
Soil treatment x D	2	1.21	0.545	2	3.22	0.200	2	0.13	0.939	2	1.21	0.545			
Plant history x N	1	0.92	0.337	1	2.00	0.157	1	0.02	0.879	1	0.46	0.497			
Soil history x N	1	0.87	0.352	1	0.05	0.832	1	1.37	0.242	1	2.29	0.130			
Soil treatment x N	2	3.07	0.215	2	0.80	0.669	2	6.25	0.044	2	5.64	0.060			
Plant history x D x N	1	0.07	0.792	1	<0.01	0.980	1	0.15	0.696	1	0.02	0.884			
Soil history x D x N	1	0.61	0.434	1	0.05	0.822	1	0.89	0.344	1	1.17	0.279			
Soil treatment x D x N	2	3.61	0.165	2	2.25	0.326	2	1.33	0.515	2	0.56	0.757			

	А. р	oratensis										
	Tota	al biomas	5	Sho	ot biomass		Root	biomass		Root	-shoot ratio	
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.57	0.452	1	0.42	0.518	1	0.43	0.512	1	<0.01	0.985
Soil history	1	0.68	0.408	1	<0.01	0.945	1	1.47	0.225	1	0.80	0.371
Soil treatment	2	0.34	0.845	2	0.29	0.865	2	0.23	0.892	2	0.07	0.967
Drought (D)	1	71.43	<0.001	1	38.06	<0.001	1	60.92	<0.001	1	0.15	0.696
Nitrogen input (N)	1	74.74	<0.001	1	162.92	<0.001	1	9.71	0.002	1	55.50	<0.001
D×N	1	26.47	<0.001	1	3.98	0.046	1	24.94	<0.001	1	16.49	<0.001
Plant history x D	1	0.08	0.772	1	0.51	0.477	1	0.48	0.488	1	1.07	0.301
Soil history x D	1	0.43	0.512	1	0.37	0.546	1	0.20	0.653	1	0.01	0.912
Soil treatment x D	2	1.17	0.557	2	0.19	0.911	2	2.12	0.346	2	3.60	0.165
Plant history x N	1	0.40	0.529	1	1.26	0.261	1	0.02	0.875	1	0.14	0.709
Soil history x N	1	5.45	0.020	1	1.19	0.275	1	4.53	0.033	1	1.24	0.265
Soil treatment x N	2	2.78	0.249	2	2.50	0.287	2	1.21	0.547	2	0.13	0.938
Plant history x D x N	1	0.55	0.458	1	0.02	0.881	1	0.59	0.442	1	0.08	0.771
Soil history x D x N	1	0.28	0.595	1	0.30	0.585	1	0.78	0.376	1	1.44	0.230
Soil treatment x D x N	2	0.91	0.634	2	0.05	0.975	2	1.45	0.485	2	2.41	0.300

Appendix 2—table 5. Summary of mixed-effect model analyses testing the effects legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant performance (total biomass, shoot biomass, root biomass and

root-shoot ratio) of *D. glomerata* and *P. trivialis*.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

	D. glomerata											
	Total	biomass		Sho	ot biomass		Roo	t biomass		Root	t-shoot rat	io
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	1.51	0.219	1	1.32	0.251	1	1.12	0.289	1	0.19	0.662
Soil history	1	0.00	0.957	1	0.01	0.912	1	0.07	0.787	1	0.05	0.829
Soil treatment	2	0.79	0.673	2	0.11	0.948	2	2.65	0.266	2	2.94	0.230
Drought (D)	1	0.98	0.323	1	12.71	<0.001	1	20.48	<0.001	1	58.54	<0.001
Nitrogen input (N)	1	82.06	<0.001	1	124.42	<0.001	1	8.87	0.003	1	16.79	<0.001
D x N	1	0.07	0.790	1	0.04	0.843	1	0.61	0.434	1	0.53	0.467
Plant history x D	1	0.05	0.821	1	0.55	0.458	1	0.24	0.623	1	1.40	0.236
Soil history x D	1	0.56	0.453	1	2.20	0.138	1	0.14	0.706	1	0.27	0.601
Soil treatment x D	2	0.09	0.955	2	0.55	0.758	2	1.09	0.579	2	3.01	0.222
Plant history x N	1	1.55	0.213	1	1.85	0.174	1	0.62	0.432	1	0.29	0.592
Soil history x N	1	1.42	0.234	1	2.24	0.135	1	0.26	0.612	1	0.25	0.618
Soil treatment x N	2	0.05	0.976	2	0.72	0.699	2	1.94	0.378	2	3.83	0.147
Plant history x D x N	1	4.64	0.031	1	3.35	0.067	1	4.09	0.043	1	3.81	0.051
Soil history x D x N	1	4.21	0.040	1	3.68	0.055	1	2.87	0.090	1	1.64	0.200
Soil treatment x D x N	2	1.70	0.428	2	3.03	0.220	2	0.66	0.718	2	0.32	0.853

	P. tri	ivialis										
	Tota	l biomass		Shoc	ot biomasp		Roo	Root biomass			shoot ratio	
	Df	Chi ²	р	Df	Chi ²	Р	Df	Chi ²	р	Df	Chi ²	р
Plant history	1	0.91	0.340	1	0.03	0.870	1	1.49	0.222	1	1.36	0.244
Soil history	1	0.26	0.611	1	0.08	0.781	1	2.43	0.119	1	4.29	0.038
Soil treatment	2	1.23	0.540	2	1.18	0.556	2	0.62	0.732	2	0.09	0.956
Drought (D)	1	23.05	<0.001	1	22.42	<0.001	1	8.93	0.003	1	0.00	0.988
Nitrogen input (N)	1	27.28	<0.001	1	87.31	<0.001	1	1.12	0.290	1	45.86	<0.001
D×N	1	3.81	0.051	1	2.16	0.141	1	2.81	0.094	1	2.10	0.147
Plant history x D	1	0.08	0.775	1	1.03	0.311	1	0.03	0.874	1	0.20	0.656
Soil history x D	1	<0.01	0.969	1	0.21	0.649	1	0.15	0.696	1	0.21	0.646
Soil treatment x D	2	0.80	0.670	2	0.69	0.708	2	0.38	0.828	2	1.04	0.594
Plant history x N	1	<0.01	0.972	1	0.87	0.350	1	0.32	0.569	1	0.73	0.391
Soil history x N	1	<0.01	0.984	1	0.01	0.936	1	0.01	0.920	1	0.03	0.857
Soil treatment x N	2	4.20	0.123	2	1.87	0.392	2	6.33	0.042	2	7.28	0.026
Plant history x D x N	1	0.25	0.614	1	<0.01	0.978	1	0.17	0.680	1	0.00	0.972
Soil history x D x N	1	0.02	0.890	1	1.11	0.292	1	1.09	0.296	1	2.88	0.089
Soil treatment x D x N	2	0.35	0.838	2	0.49	0.782	2	1.16	0.559	2	1.97	0.373

Appendix 2—table 6. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant trait expressions of *A. elatius*.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

A. elatius	Gro	wth heigl	nt	Shoo	ot nitrogen co	onc.	Leaf	Leaf greenness			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р		
Plant history	1	0.24	0.625	1	0.12	0.725	1	0.07	0.795		
Soil history	1	0.61	0.436	1	0.36	0.547	1	0.67	0.413		
Soil treatment	2	2.01	0.365	2	0.80	0.670	2	0.19	0.907		
Drought (D)	1	2.11	0.146	1	36.64	<0.001	1	30.19	<0.001		
Nitrogen input (N)	1	5.35	0.021	1	142.97	<0.001	1	153.54	<0.001		
D×N	1	0.02	0.881	1	32.71	<0.001	1	0.27	0.604		
Plant history x D	1	4.68	0.030	1	1.41	0.236	1	0.48	0.487		
Soil history x D	1	0.01	0.904	1	0.26	0.612	1	0.06	0.813		
Soil treatment x D	2	3.10	0.212	2	0.38	0.827	2	1.58	0.453		

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A. elatius	elatius Growth height Shoot nitrogen conc. Leaf greenness									
	Df	Chi ²	р	Df	Chi ²	р	I	Df Cl	hi²	р
Plant history x N	1	1.15	0.284	1	1.08	0.300		1	3.76	0.053
Soil history x N	1	0.61	0.434	1	0.20	0.656		1	1.09	0.295
Soil treatment x N	2	3.03	0.220	2	0.27	0.874	4	2	2.37	0.305
Plant history x D x N	1	0.59	0.443	1	1.85	0.174		1	0.37	0.545
Soil history x D x N	1	0.93	0.334	1	0.03	0.854		1	0.06	0.813
Soil treatment x D x N	2	7.64	0.022	2	0.26	0.877	2	2	1.95	0.377
	LDM	с		SLA			Stom	atal con	ductance	
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²		Р
Air temperature	-	-	-	-	-	-	1	< 0.0	1	0.948
Daytime	-	-	-	-	-	-	1	8.0	5	0.005
Plant history	1	0.46	0.500	5	1.69	0.194	1	0.4	9	0.486
Soil history	1	0.19	0.666	6	1.83	0.176	1	0.0	5	0.823
Soil treatment	2	1.37	0.504	8	1.14	0.565	2	3.3	8	0.184
Drought (D)	1	7.57	0.006	9	12.37	<0.001	1	4.5	8	0.032
Nitrogen input (N)	1	1.05	0.307	10	0.05	0.832	1	2.0	0	0.158
D x N	1	0.02	0.889	11	1.87	0.171	1	0.1	7	0.681
Plant history x D	1	1.48	0.224	12	1.94	0.164	1	1.0	8	0.298
Soil history x D	1	0.36	0.549	13	0.79	0.373	1	0.0	5	0.830
Soil treatment x D	2	< 0.01	0.998	15	1.73	0.420	2	0.7	3	0.693
Plant history x N	1	0.01	0.904	16	0.08	0.782	1	0.0	4	0.836
Soil history x N	1	0.01	0.936	17	1.69	0.193	1	0.3	6	0.549
Soil treatment x N	2	2.16	0.339	19	2.01	0.367	2	0.2	4	0.886
Plant history x D x N	1	<0.01	0.999	20	1.96	0.162	1	0.4	2	0.518
Soil history x D x N	1	0.10	0.752	21	0.15	0.696	1	1.4	8	0.224
Soil treatment x D x N	2	0.35	0.840	23	0.50	0.781	2	1.9	9	0.369
	F	loot diame	ter		SRL			RLD		
	0	Of Chi ²	Р		Df Chi ²	Р		Df	Chi ²	Р
Plant history	1	0.08	0.783		1 0.31	0.576		1	0.09	0.767
Soil history	1	0.23	0.629		1 0.22	0.639		1	0.82	0.364
Soil treatment	2	2.89	0.236		2 5.30	0.071		2	3.35	0.187
Drought (D)	1	0.32	0.572		1 5.25	0.022		1	0.04	0.851
Nitrogen input (N)	1	3.46	0.063		1 13.72	<0.001		1	0.13	0.723
D×N	1	0.01	0.932		1 1.62	0.204		1	<0.01	0.989
Plant history x D	1	0.39	0.531		1 0.11	0.740		1	0.77	0.380
Soil history x D	1	0.01	0.938		1 0.95	0.329		1	0.29	0.590
Soil treatment x D	2	2.11	0.349		2 0.51	0.775		2	0.45	0.797
Plant history x N	1	0.09	0.764		1 1.41	0.235		1	1.29	0.256
Soil history x N	1	1.35	0.246		1 0.32	0.573		1	3.53	0.060
Soil treatment x N	2	0.68	0.711		2 1.06	0.590		2	1.76	0.416
Plant history x D x N	1	1.68	0.194		1 2.73	0.099		1	3.70	0.054
Soil history x D x N	1	4.45	0.035		1 0.52	0.469		1	1.46	0.227

2.75

0.253

2

2.26

2

Soil treatment x D x N $\,$

2

2.00

0.369

0.324

Appendix 2—table 7. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant trait expressions of *A. pratensis*.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

A. pratensis		Growth height			oot nitrogen	conc.	Lea	Leaf greenness		
		Df Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Plant history		1 1.35	0.246	1	0.16	0.687	1	0.49	0.485	
Soil history		1 0.71	0.400	1	<0.01	0.967	1	0.11	0.745	
Soil treatment		2 8.50	0.014	2	1.38	0.501	2	0.20	0.903	
Drought (D)		1 1.07	0.300	1	15.42	<0.001	1	16.09	<0.001	
Nitrogen input (N)		1 10.63	0.001	1	246.65	<0.001	1	143.35	<0.001	
D x N		1 1.40	0.236	1	17.58	<0.001	1	0.86	0.353	
Plant history x D		1 0.16	0.692	1	<0.01	0.979	1	0.58	0.446	
Soil history x D		1 0.31	0.577	1	0.52	0.471	1	3.04	0.081	
Soil treatment x D		2 1.11	0.575	2	0.50	0.778	2	3.39	0.183	
Plant history x N		1 0.28	0.597	1	0.17	0.681	1	<0.01	0.994	
Soil history x N		1 0.01	0.919	1	0.10	0.747	1	1.10	0.293	
Soil treatment x N		2 2.42	0.299	2	6.58	0.037	2	0.19	0.911	
Plant history x D x N		1 0.18	0.672	1	0.87	0.352	1	1.06	0.304	
Soil history x D x N		1 0.45	0.501	1	0.49	0.485	1	0.03	0.863	
Soil treatment x D x N		2 0.85	0.654	2	2.08	0.353	2	0.32	0.854	
	IDM	<u> </u>		51.0			Stom	atal conductance		
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	P	
Air temperature	_	_	_	-	-	_	1	0.16	0.685	
Daytime	_	_	_	_	_	_	1	1.78	0.182	
Plant history	1	2.82	0.093	1	0.19	0.665	1	0.43	0.513	
Soil history	1	1.80	0.180	1	0.94	0.332	1	0.41	0.520	
Soil treatment	2	3.57	0.168	2	5.69	0.058	2	3.67	0.159	
Drought (D)	1	4.02	0.045	1	1.29	0.255	1	6.17	0.013	
Nitrogen input (N)	1	0.75	0.388	1	2.93	0.087	1	3.64	0.056	
D×N	1	0.33	0.566	1	0.41	0.524	1	3.45	0.063	
Plant history x D	1	0.13	0.715	1	0.27	0.604	1	0.03	0.862	
Soil history x D	1	0.16	0.685	1	<0.01	0.980	1	0.64	0.423	
Soil treatment x D	2	1.40	0.497	2	1.39	0.499	2	0.01	0.993	
Plant history x N	1	1.03	0.311	1	1.02	0.313	1	0.58	0.447	
Soil history x N	1	<0.01	0.950	1	0.78	0.377	1	0.18	0.669	
Soil treatment x N	2	0.64	0.726	2	2.56	0.278	2	0.27	0.874	
Plant history x D x N	1	0.80	0.372	1	1.67	0.197	1	2.57	0.109	
Soil history x D x N	1	4.17	0.041	1	1.01	0.315	1	0.23	0.634	
Soil treatment x D x N	2	0.18	0.912	2	1.09	0.581	2	15.71	<0.001	
	D- · "			CD'			DID			
	Koot di	Chi ²	P	5RL Df	Chi ²	P		Chi ²	P	
Plant history	1	0.01	0.025	1	0.28	0.597	1	0.04	L 000	
Coil bistony	1	0.19	0.735	1	0.01	0.07/	1	0.00	0.007	
SUILINISTORY	1	U. 18	0.076	I	0.01	0.734	1	0.92	0.337	

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	Root diameter			SRL			RLD	RLD			
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	Р		
Soil treatment	2	0.54	0.763	2	0.97	0.615	2	0.12	0.940		
Drought (D)	1	39.31	<0.001	1	5.25	0.022	1	82.01	<0.001		
Nitrogen input (N)	1	51.80	<0.001	1	5.33	0.021	1	0.34	0.560		
D×N	1	0.09	0.767	1	5.57	0.018	1	4.32	0.038		
Plant history x D	1	0.01	0.906	1	0.30	0.587	1	0.26	0.611		
Soil history x D	1	0.09	0.769	1	0.01	0.910	1	0.02	0.877		
Soil treatment x D	2	2.58	0.276	2	4.88	0.087	2	0.11	0.948		
Plant history x N	1	0.03	0.869	1	0.19	0.660	1	0.17	0.682		
Soil history x N	1	6.39	0.011	1	8.14	0.004	1	0.63	0.426		
Soil treatment x N	2	1.82	0.402	2	3.27	0.195	2	1.24	0.539		
Plant history x D x N	1	0.54	0.461	1	0.15	0.700	1	0.28	0.594		
Soil history x D x N	1	1.82	0.178	1	1.87	0.172	1	0.27	0.605		
Soil treatment x D x N	2	3.23	0.199	2	1.63	0.443	2	0.70	0.703		

Appendix 2—table 8. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant trait expressions of *D. glomerata*.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

D. glomerata	Grow	rth height		Shoot r	nitrogen con	c.	Lea	Leaf greenness		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Plant history	1	0.05	0.831	1	0.58	0.444	1	0.22	0.640	
Soil history	1	1.56	0.212	1	1.35	0.245	1	0.27	0.606	
Soil treatment	2	5.25	0.073	2	0.75	0.687	2	0.55	0.760	
Drought (D)	1	<0.01	0.976	1	19.10	<0.001	1	29.41	<0.001	
Nitrogen input (N)	1	11.51	0.001	1	183.85	<0.001	1	172.91	<0.001	
D x N	1	<0.01	0.949	1	3.72	0.054	1	0.08	0.781	
Plant history x D	1	0.01	0.920	1	0.05	0.828	1	2.75	0.097	
Soil history x D	1	0.82	0.366	1	0.08	0.08 0.774		0.22	0.639	
Soil treatment x D	2	0.48	0.785	2	0.25	0.880	2	0.21	0.899	
Plant history x N	1	0.91	0.341	1	2.96	0.086	1	0.61	0.437	
Soil history x N	1	0.23	0.633	1	0.32	0.571	1	1.75	0.186	
Soil treatment x N	2	0.35	0.840	2	0.29	0.866	2	4.92	0.085	
Plant history x D x N	1	0.12	0.733	1	1.62	0.204	1	0.54	0.462	
Soil history x D x N	1	0.71	0.400	1	5.07	0.024	1	<0.01	0.998	
Soil treatment x D x N	2	0.06	0.969	2	2.15	0.341	2	0.33	0.846	
	LDM	с		SLA			Stomata	omatal conductance		
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Air temperature	-	-	-	-	-	-	1	0.39	0.531	
Daytime	-	-	-	-	-	-	1	20.31	<0.001	
Plant history	1	0.12	0.727	1	0.80	0.371	1	5.08	0.024	
Soil history	1	0.58	0.445	1	0.32	0.573	1	<0.01	0.944	
Soil treatment	2	0.58	0.749	2	1.20	0.548	2	0.54	0.765	

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	LDMO	C		SLA			Stomatal conductance			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Drought (D)	1	0.07	0.798	1	0.54	0.461	1	9.01	0.003	
Nitrogen input (N)	1	55.57	<0.001	1	57.43	<0.001	1	2.72	0.099	
D×N	1	20.69	<0.001	1	6.61	0.010	1	6.34	0.012	
Plant history x D	1	0.04	0.842	1	0.46	0.498	1	0.07	0.793	
Soil history x D	1	0.01	0.926	1	0.09	0.762	1	<0.01	0.991	
Soil treatment x D	2	1.43	0.490	2	0.09	0.958	2	0.19	0.907	
Plant history x N	1	0.99	0.320	1	0.02	0.893	1	0.32	0.571	
Soil history x N	1	2.48	0.115	1	2.19	0.139	1	0.69	0.406	
Soil treatment x N	2	0.13	0.938	2	1.56	0.459	2	0.09	0.958	
Plant history x D x N	1	2.00	0.157	1	0.09	0.768	1	0.33	0.566	
Soil history x D x N	1	1.30	0.254	1	4.99	0.026	1	5.98	0.014	
Soil treatment x D x N	2	3.56	0.169	2	1.09	0.579	2	1.57	0.456	
	Root d	liameter		SRL			RLD			
	Df	Chi ²	р	Df	Chi ²	р	Df	Chi ²	р	
Plant history	1	0.60	0.438	1	0.96	0.326	1	2.61	0.107	
Soil history	1	0.06	0.805	1	0.07	0.791	1	0.01	0.933	
Soil treatment	2	0.07	0.967	2	0.58	0.749	2	2.44	0.296	
Drought (D)	1	0.93	0.335	1	1.16	0.281	1	9.45	0.002	
Nitrogen input (N)	1	1.22	0.270	1	0.37	0.545	1	7.05	0.008	
D x N	1	0.80	0.370	1	1.73	0.189	1	0.08	0.773	
Plant history x D	1	3.60	0.058	1	0.64	0.425	1	0.25	0.614	
Soil history x D	1	1.41	0.235	1	0.62	0.430	1	0.23	0.632	
Soil treatment x D	2	0.65	0.721	2	1.95	0.377	2	2.43	0.297	
Plant history x N	1	0.19	0.667	1	2.05	0.152	1	0.03	0.854	
Soil history x N	1	0.60	0.437	1	<0.01	0.994	1	0.21	0.646	
Soil treatment x N	2	0.85	0.653	2	0.97	0.616	2	1.76	0.414	
Plant history x D x N	1	1.49	0.222	1	0.14	0.712	1	3.11	0.078	
Soil history x D x N	1	0.49	0.483	1	3.54	0.060	1	1.07	0.301	
Soil treatment x D x N	2	1.65	0.438	2	1.16	0.559	2	0.20	0.907	

Appendix 2—table 9. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on plant trait expressions of *P. trivialis*.

Shown are degrees of freedom (Df), Chi^2 and P-values (P). Significant effects (P < 0.05) are given in bold, marginally significant effects (P < 0.1) in italics.

P. trivialis	Grow	rth height		Shoot	nitrogen conc.		Leaf greenness		
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	Р
Plant history	1	0.06	0.800	1	0.00	0.997	1	0.93	0.334
Soil history	1	2.29	0.131	1	0.05	0.824	1	1.10	0.294
Soil treatment	2	1.66	0.435	2	0.51	0.776	2	1.15	0.563
Drought (D)	1	30.17	<0.001	1	5.46	0.019	1	1.42	0.233
Nitrogen input (N)	1	12.16	<0.001	1	297.03	<0.001	1	108.82	<0.001

Appendix 2—table 9 Continued on next page

Appendix 2—table 9 Continued

P. trivialis	Growth height		SI	Shoot nitrogen conc.				Leaf g	Leaf greenness		
	Df	Chi ²	Р	D	f	C	hi²	Ρ	Df	Chi ²	Р
D×N	1	1.72	0.	190 1			17.06	<0.001	1	1.09	0.296
Plant history x D	1	0.22	0.	637 1			0.11	0.736	1	3.08	0.079
Soil history x D	1	2.28	0.	131 1			0.53	0.469	1	0.06	0.806
Soil treatment x D	2	3.11	0.	211 2			1.03	0.598	2	0.18	0.916
Plant history x N	1	5.16	0.	023 1			0.05	0.821	1	0.13	0.719
Soil history x N	1	3.49	0.	062 1			0.04	0.842	1	0.36	0.549
Soil treatment x N	2	2.08	0.	354 2			1.04	0.594	2	1.98	0.371
Plant history x D x N	1	0.92	0.	336 1			0.03	0.865	1	0.11	0.738
Soil history x D x N	1	0.13	0.	718 1			0.18	0.669	1	0.00	0.967
Soil treatment x D x N $$	2	2.11	0.	348 2			5.57	0.062	2	1.74	0.418
	LD	MC			S	SLA			Stoma	tal conducta	ıce
	Df	:	Chi ²	Р	0	Df	Chi ²	Р	Df	Chi ²	Р
Air temperature	-		-	-	-		-	-	1	38.70	<0.001
Daytime	-		-	-	-		-	-	1	18.64	<0.001
Plant history	1		<0.01	0.965	1	I	0.62	0.431	1	0.18	0.675
Soil history	1		0.08	0.777	1		0.49	0.485	1	0.71	0.399
Soil treatment	2		1.64	0.441	2	2	2.12	0.346	2	3.25	0.197
Drought (D)	1		2.85	0.091	1		2.75	0.097	1	0.22	0.636
Nitrogen input (N)	1		57.72	<0.001	1		41.44	<0.001	1	0.06	0.800
D x N	1		0.39	0.534	1		0.62	0.431	1	2.87	0.090
Plant history x D	1		1.09	0.296	1		0.38	0.540	1	2.86	0.091
Soil history x D	1		2.26	0.133	1		0.45	0.502	1	0.01	0.908
Soil treatment x D	2		0.19	0.908	2	2	1.33	0.515	2	0.40	0.819
Plant history x N	1		3.37	0.066	1		1.56	0.212	1	0.35	0.554
Soil history x N	1		0.54	0.461	1		0.21	0.645	1	2.45	0.118
Soil treatment x N	2		1.89	0.388	2	2	3.10	0.213	2	1.36	0.508
Plant history x D x N	1		0.13	0.720	1		0.58	0.446	1	0.14	0.704
Soil history x D x N	1		1.15	0.283	1		1.01	0.315	1	7.44	0.006
Soil treatment x D x N $$	2		3.30	0.192	2	2	0.99	0.610	2	2.20	0.333
	Root d	iameter			SRL				RLD		
	Df	Chi ²	Р)	Df		Chi ²	Р	Df	Chi ²	Р
Plant history	1	2.10		0.147	1		2.38	0.123	1	0.04	0.840
Soil history	1	0.08		0.781	1		0.30	0.581	1	1.31	0.253
Soil treatment	2	0.13		0.938	2		0.31	0.856	2	1.13	0.568
Drought (D)	1	14.18	<	0.001	1		0.89	0.347	1	18.25	<0.001
Nitrogen input (N)	1	0.17		0.677	1		3.49	0.062	1	0.03	0.872
D×N	1	0.88		0.349	1		0.25	0.618	1	1.16	0.282
Plant history x D	1	0.40		0.525	1		0.27	0.602	1	0.16	0.692
Soil history x D	1	0.48		0.487	1		0.20	0.655	1	1.36	0.244
Soil treatment x D	2	5.85		0.054	2		0.50	0.777	2	0.43	0.808
Plant history x N	1	1.28		0.258	1		0.07	0.795	1	0.28	0.594
Soil history x N	1	1.21		0.271	1		0.36	0.549	1	0.65	0.418

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	Root diameter			SRL			RLD	RLD		
	Df	Chi ²	Р	Df	Chi ²	Р	Df	Chi ²	Р	
Soil treatment x N	2	2.99	0.225	2	9.11	0.011	2	0.33	0.846	
Plant history x D x N	1	0.33	0.566	1	0.05	0.821	1	0.02	0.878	
Soil history x D x N	1	4.11	0.043	1	9.74	0.002	1	2.06	0.151	
Soil treatment x D x N	2	0.52	0.772	2	1.40	0.495	2	1.43	0.488	

Appendix 2—table 10. Summary of mixed-effect model analyses testing the effects of legacy treatments (plant history, soil history, soil treatment), global change treatments (drought, nitrogen input) and their interactions on mildew infestation of *D. glomerata* and *P. trivialis*.

Shown are degrees of freedom (Df), Chi^2 and p-values (p). Significant effects (p < 0.05) are given in bold, marginally significant effects (p < 0.1) in italics.

Mildew infestation	D. glomerata	1		P. trivialis					
	Df	Chi ²	р	Df	Chi ²	р			
Plant history	1	0.29	0.588	1	0.01	0.939			
Soil history	1	0.24	0.622	1	4.16	0.041			
Soil treatment	2	0.22	0.896	2	3.36	0.187			
Drought (D)	1	2.44	0.119	1	10.69	0.001			
Nitrogen input (N)	1	42.75	<0.001	1	38.76	<0.001			
D×N	1	1.05	0.305	1	0.98	0.321			
Plant history x D	1	0.03	0.855	1	0.02	0.889			
Soil history x D	1	2.25	0.134	1	0.07	0.788			
Soil treatment x D	2	5.79	0.055	2	0.25	0.884			
Plant history x N	1	<0.01	0.953	1	0.25	0.614			
Soil history x N	1	0.21	0.643	1	0.50	0.477			
Soil treatment x N	2	0.32	0.854	2	1.22	0.544			
Plant history x D x N	1	3.00	0.083	1	0.09	0.770			
Soil history x D x N	1	1.69	0.193	1	0.93	0.335			
Soil treatment x D x N	2	7.15	0.028	2	0.62	0.734			

Appendix 3

Calculation of irrigation water quantity per pot

1) After one week of growing, pots were watered until 100% saturation and then weighted (= Weight $_{\rm wet\,soil}).$

2) To determine the amount of water, which is needed to get 60% water saturation (control value), we used the following equations:

(I) $\frac{22\% (\text{water holding capacity}) \times 60\% \text{ saturation}}{100\% \text{ saturation}} = 13.2\%$

(II) Weight_{wet soil} - $\frac{\text{Weight}_{\text{wet soil}} \times 100\% \text{ saturation}}{13.2\% + 100}$ = Weight_{water control}

First, we multiplied the water holding capacity of the Jena Experiment soil-sand mix (22%) times 60% saturation and then divided the result by 100% saturation. Second, Weight _{wet soil} was multiplied with 100 and then divided by 113.2. Third, the calculated weight for a 60% saturation was subtracted from Weight _{wet soil} per pot and averaged over all pots, which resulted in 380 ml water.

3) Drought was simulated by 50% lower water saturation (30% saturation), while the amount of water was calculated as followed:

(I) $\frac{22\%$ (water holding capacity) $\times 30\%$ saturation = 6.6%

(II) Weight_{wet soil} $-\frac{\text{Weight}_{\text{wet soil}} \times 100\% \text{ saturation}}{6.6\% + 100} = \text{Weight}_{\text{water drought}}$