



Review Article

Unraveling the role of long non-coding RNAs in therapeutic resistance in acute myeloid leukemia: New prospects & challenges

Siddhant Sharma

Department of Chemical and Biological Engineering, University of British Columbia, Vancouver, British Columbia, V6T 1Z3, Canada



ARTICLE INFO

Keywords:

Acute myeloid leukemia
Chemotherapy
lncRNAs
Chemoresistance
Self-renewal
Apoptosis

ABSTRACT

Acute Myeloid Leukemia (AML) is a fatal hematological disease characterized by the unchecked proliferation of immature myeloid blasts in different tissues developed by various mutations in hematopoiesis. Despite intense chemotherapeutic regimens, patients often experience poor outcomes, leading to substandard remission rates. In recent years, long non-coding RNAs (lncRNAs) have increasingly become important prognostic and therapeutic hotspots, due to their contributions to dysregulating many functional epigenetic, transcriptional, and post-translational mechanisms leading to alterations in cell expressions, resulting in increased chemoresistance and reduced apoptosis in leukemic cells. Through this review, I highlight and discuss the latest advances in understanding the major mechanisms through which lncRNAs confer therapy resistance in AML. In addition, I also provide perspective on the current strategies to target lncRNA expressions. A better knowledge of the critical role that lncRNAs play in controlling treatment outcomes in AML will help improve existing medications and devise new ones.

1. Introduction

Hematopoiesis is a highly regulated process of developing various blood cell lineages from hematopoietic stem cells (HSCs) that undergo multiple, intermediate stages of maturation [1]. Leukemia is a diverse family of life-threatening hematologic malignancies that manifest from an oligoclonal, unregulated proliferation of failed differentiated HSCs. According to the Canadian Cancer Society, cancer is the prime factor responsible for deaths in Canada, and of the 124,200 reported cases of cancer in Canadian males in 2023, approximately 3.14 % of cases accounted for leukemia while for the 114,900 female cancer cases, leukemia accounted for 2.17 % of all cases. The National Cancer Institute SEER predicted an estimated 59,610 new cases of leukemia in the general US population for 2023 with the predicted mortality rate at a staggering estimate of 39.7 % (23,710 deaths). While there exists a wide heterogeneity in the genetic abnormalities and risk factors that lead to leukemogenesis, with treatment options varying on a case-by-case basis, the general factors that increase the risk of leukemia include high/fatal doses of radiation; long exposure to carcinogenic species like Benzene, and workplace accidents/hazards [2]. Based on the diseases' rate of progression and cell lineage affected, the World Health Organization (WHO) has broadly classified leukemia into 4 unique subtypes namely acute myeloid leukemia (AML), chronic myeloid leukemia (CML), acute

lymphoblastic leukemia (ALL), and chronic lymphoblastic leukemia (CLL) [3,4]. AML is a rare clonal malignancy stemming specifically from the abnormal development and uncontrolled proliferation of immature blood cells called myeloid blasts which subsequently infiltrate the bone marrow (BM), and other tissues, interfering with other healthy cells and preventing them from performing their biological functions [5]. The diagnosis of AML is based on a greater than or equal to 20 % threshold limit of the presence of immature, malignant myeloid blasts in the blood and other tissues [6,7]. While rarely diagnosed in children and youth, AML is the most common form of acute leukemia occurring in adults in the USA. The estimated general US population diagnosed with acute myeloid leukemia was 20,380 cases for 2023 with deaths estimated to be at a ghastly approximation of 11,310 (55.4 % for all diagnosed cases) [8]. Growing advances in hematological cancer research have dramatically improved therapy outcomes with most patients achieving complete remission (CR). Induction chemotherapy is the first treatment regimen to combat cancer with a single or a concoction of different chemotherapeutic reagents. An intense 7-day cytarabine (Cyt) + 3-day anthracycline regimen remains unchallenged as induction chemotherapy to treat AML for over 40 years [9]; but despite achieving a promising 60–80 % remission rate in the younger generation and 40–60 % in adults greater than 65 years of age [6,10], approximately 10–20 % of all patients fail to respond to induction chemotherapy and more than

E-mail address: sid0507@student.ubc.ca.<https://doi.org/10.1016/j.ncrna.2024.05.009>

Received 12 February 2024; Received in revised form 19 May 2024; Accepted 20 May 2024

Available online 20 May 2024

2468-0540/© 2024 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co. Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

85 % patients experience relapse which diminishes the probability of survival after further treatment regimens [11]. The genomic, and molecular heterogeneity, of AML, is diverse and growing evidence suggests that strong therapeutic resistance in leukemic cells is strongly correlated to the observed high relapse and poor prognosis in patients [12,13]. The underlying mechanisms influencing leukemic cells to become more resistant to therapeutic effects have not yet been fully elucidated. Therefore, a greater mechanistic understanding of pathways mediating therapeutic resistance in AML is an urgent, indispensable need that will help open novel avenues for creating more effective and safe treatment procedures. lncRNAs are the group of non-coding RNAs (ncRNAs)

greater than 200 nucleotides in length but lack appreciable protein translation [14–16]. Several studies have documented the significant contributions of lncRNAs in enhancing therapeutic resistance in various cancers including lung cancer [17–21], breast cancer [22–26], colorectal cancer [27–30], gastric cancer [31–34], and ovarian cancer [35–37]. In this review, I offer thorough insights into the numerous ways through which lncRNAs induce and support chemoresistance, self-renewal, and immortalization against programmed death in AML (Fig. 1). Knowing the critical roles played by different lncRNAs in AML will improve our awareness of the interactions between different underlying processes that result in heightened chemo refractory and

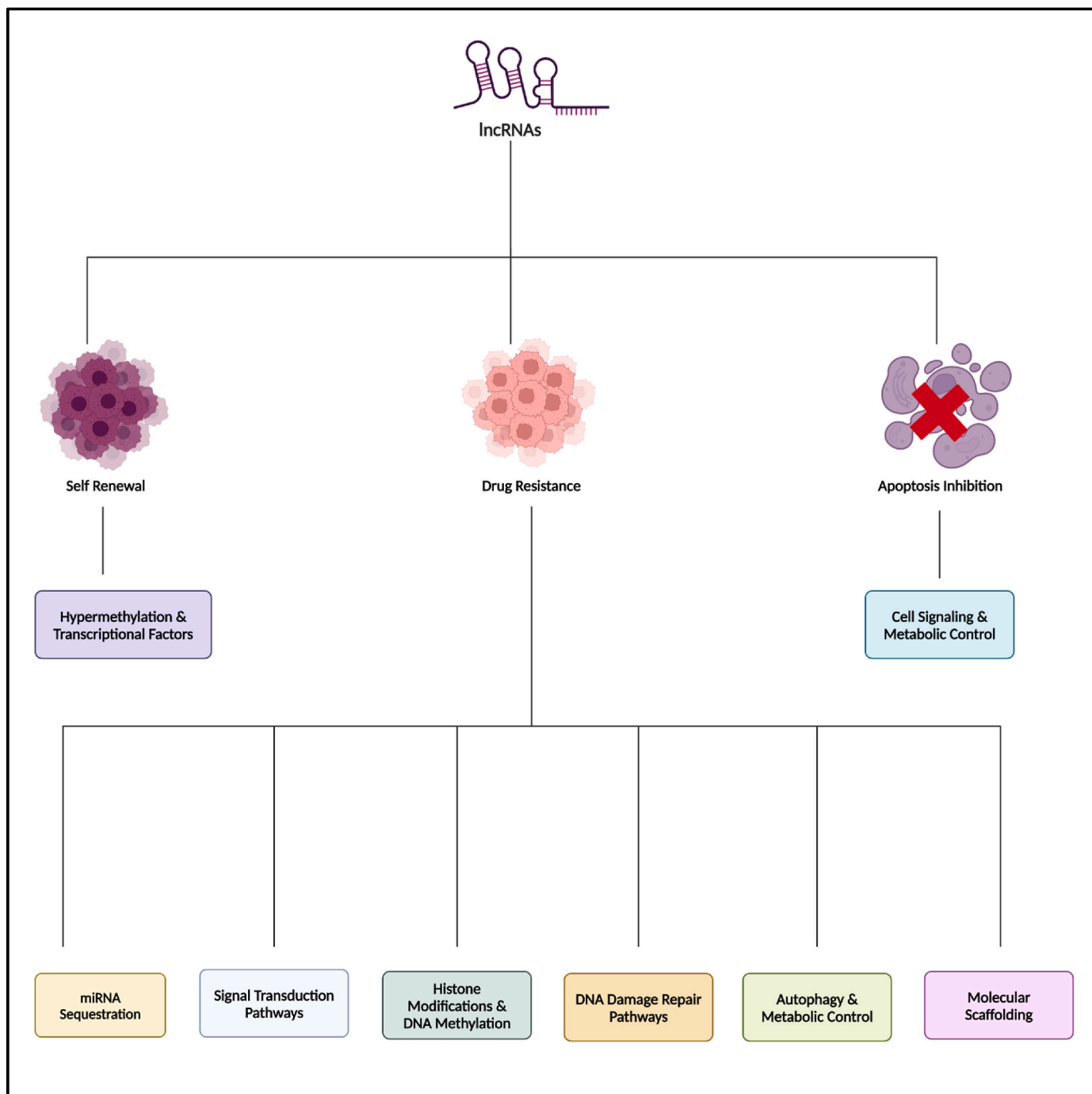


Fig. 1. Graphical Abstract

An illustration summarizing the different mechanisms through which lncRNAs regulate drug resistance, promote apoptosis inhibition, and enhance the self-renewal capacity of LSCs.

ultimately lead to the development of novel medicines, creating new avenues for the treatment of AML. Finally, I hope that the studies discussed herein will encourage the creation of brand-new questions and lines of inquiry that will have a significant influence on clinical practice.

2. Significance of lncRNA expressions in AML

With great strides being accomplished in cancer research, it has been found that the dysregulation of various lncRNA expressions is an important factor responsible for leukemogenesis. Indeed, a diverse spectrum of evidence suggests that lncRNAs can be exploited as important prognostic and therapeutic targets [15,38–41]. lncRNAs are involved in regulating cellular proliferation, myeloid differentiation, drug resistance, leukemic cell metabolism, RNA modifications, and inhibition of apoptosis in AML [42]. Illustriously, in addition to enhancing m⁶A methylation, the lncRNA UCA1 also increases the stability of CXCR4 and CYP1B1 by binding to and then upregulating the expression of the METTL14 protein. METTL14 is substantially overexpressed in AML patients with t(11q23), t(15; 17), and t(8; 21) cytogenetic abnormalities [43], and studies also suggest that both CXCR4 and CYP1B1 play an important role in AML pathogenesis and represent promising therapeutic targets [44–46]. Given that METTL14 is a crucial component of UCA1-modified m⁶A methylation and that m⁶A methylation levels are likewise considerably expressed in AML, the significance of the UCA1-mediated METTL14/CXCR4/CYP1B1 axis becomes important [47]. Various studies highlight how lncRNAs upregulate various signal transductions to facilitate AML metastasis and restrain apoptosis. A prominent example is the lncRNA SNHG16 which is substantially expressed in several cancers and is essential for tumorigenesis as it is involved in the growth and proliferation of cancer cells, as well as enhancing chemoresistance, metastasis, and preventing cell apoptosis [48]. Little is known about this lncRNA's function, specifically in AML. Recently, it was shown that SNHG16 regulates oligoclonal proliferation and advocates aggressive cell migration in AML. SNHG16 directly binds to the CELF2 protein and overexpression of SNHG16 down-regulates CELF2 expression levels since their interaction gives SNHG16 an authoritative control over the regulation of CELF2 protein to its mRNA target, inherently reducing the stability of the CELF2 mRNA. Since PTEN's main function is to negatively regulate the PI3K/AKT signal pathway, whose overexpression is a major contributor to carcinogenesis, enhanced SNHG16 expression decreases PTEN activity, leading to an increase in PI3K/AKT signaling [49]. The role of lncRNAs as competing endogenous RNAs (ceRNAs) has garnered increasing attention in recent times. Patients with M5 subtype AML are reported to have significant levels of HOTTIP expression. The microRNA-608 (miRNA-608) gene is the main target of HOTTIP, and M5, U-837, and THP-1 AML cell lines with increased HOTTIP expression exhibited significantly lower levels of miRNA-608. As a ceRNA, HOTTIP traps miRNA-608 inside of itself. CCK-8 and flow cytometry demonstrated that this encapsulation upregulates DDA1 expression, which is normally tightly downregulated by miRNA-608. This increase in DDA1 is responsible for increasing the proliferative potential of AML. As a result, the HOTTIP/miRNA-608/DDA1 axis is discovered to hasten AML pathogenesis [50]. The lncRNA SBF2-AS1 is overexpressed in AML. The active tumor suppressor miR-188-5p is sequestered by SBF2-AS1, which acts as a ceRNA to promote ZFP91 expression and boost AML proliferation. ZFP91 is an active tumorigenesis promoter, and miR-188-5p inhibits ZFP91 mRNA expression by binding to its 3'-UTR. Since high levels of ZFP91 are linked to poor prognosis in AML patients, its elevation by SBF2-AS1 through the/miR-188-5p/ZFP91 axis is clinically valuable since evidence further suggests G0/G1 cell cycle arrest and greater apoptosis in AML cells on SBF2-AS1 knockdown [51]. Conspicuously, in the case of AML, lncRNAs are known for being significantly involved in metabolic reprogramming [52]. For instance, ANRIL accelerates AML advancement by modulating glucose metabolism. The Adiponectin receptor 1 (AdipoR1) expression level is found to be positively correlated

with ANRIL, evident from knockdown studies demonstrating that silencing AdipoR1 boosts cell apoptosis and lessens proliferation like the results observed on ANRIL knockout. Aberrant glucose metabolism is an important component of leukemogenesis [53]. Mechanistically, the expressions of pAMPK, SIRT1, GLUT1, and LDHA are increased when ANRIL expression is increased in AML cells. Taken in tandem, these results highlight that ANRIL functions as an important marker in AML by mediating uncontrolled proliferation through the AdipoR1/-AMPK/SIRT1 axis [54].

3. Significance of lncRNAs in regulating chemoresistance

The development of both intrinsic and acquired forms of chemoresistance continues to be the biggest impediment to improving the lives of cancer patients [55]. Intrinsic chemoresistance refers to the cancer cells' natural capacity to fend off the effects of anti-cancer medications even before the patient is administered them. Causes for the emergence of intrinsic multidrug resistance in cancers are explained by ingrained genetic mutations in tumor microenvironments (TMEs), decreased responsiveness brought on by cancer stem cells, and the development of defense mechanisms to counteract the cytotoxic effects of chemotherapeutic agents. A progressive decrease in the medicine's effects during or after the completion of the chemotherapeutic regimen, on the other hand, indicates acquired drug resistance. Similar to its intrinsic counterpart, acquired chemoresistance is caused by many circumstances, including the activation of multiple oncogenes, physiochemical changes in the drug target, and finally, advancements in the TMEs post-treatment [56]. Cancer cells fight drug effects by a variety of strategies, which can be broadly categorized in a dichotomic sense as physical and metabolic responses. Physical features such as extracellular matrix (ECM) architecture, TME matrix stiffness, and cell adhesion-mediated drug resistance (CAM-DR) all contribute to chemoresistance in various carcinomas. The biochemical pathways that contribute to the enhancement of drug-resistant characteristics in cancers include effects of tumor-associated macrophages (TAMs), hypoxia, glycolysis, intercellular communication, cytokine-cancer cell communication, and secretion of exosomes by cancer cells [57]. Other mechanisms include overexpression of specific ABC transporter proteins [58,59], increased autophagy [60,61], and lysosomal sponging in cellular alcoves [15,62,63]. While many miRNAs, transcription factors, and histone complexes have been well known to modulate chemoresistance in cancers [64–71], how lncRNAs contribute to and control chemoresistance in malignancies is an ongoing research subject of significant importance. To rephrase this question, it would be more useful to consider what precise pathways lncRNAs use to increase drug resistance in cancer cells. Many attempts have been made to address this question. For instance [26], showed that the lncRNA DDX11-AS1 promoted doxorubicin (DOX) resistance in breast cancer cells by interacting with and upregulating levels of the LIN28A protein. Consequently, this resulted in enhanced stability of mRNA expressions of ATG7 and ATG12 proteins leading to DOX resistance. In another study by [34] it was found that the lncRNA EIF3J-DT stabilized the mRNA expression of the ATG14 protein by interacting with it directly while also simultaneously sponging the miRNA MIR188-3p to prevent ATG14 degradation to induce chemoresistance in gastric cancer. Besides breast and gastric cancer, lncRNAs play an active role in mediating drug resistance in various other malignancies like lung cancer [72,73], ovarian cancer [74,75], osteosarcoma [76,77], lymphoma [78], and other hematological malignancies [79,80]. The processes by which lncRNAs regulate chemoresistance include but are not restricted to, post-translational histone modifications, mRNA stabilization, miRNA sequestration, anti-apoptotic signaling, and interactions with transcription factors [81,82]. I would like to point out that despite an abundance of evidence pointing to the multifaceted role of lncRNAs in AML [83–87], the specific actions taken by lncRNAs for mediating drug resistance in AML have not been clearly explained. Moreover, while scientific research over the past five years has answered some

questions, it has also opened doors to new potential research frontiers, as I will discuss ahead.

3.1. Mechanism 1: lncRNA mediated miRNA sequestration

Since some lncRNAs have miRNA binding sites, they regulate gene expression by inhibiting the quantity of miRNA available to its corresponding downstream target's mRNA. This process is known as sequestration or molecular sponging. Interactions between miRNAs and lncRNAs play an important role in the development of chemoresistance in AML. For instance, the overexpression of the lncRNA SATB1-AS1 causes desensitization of AML cells towards Adriamycin (ADR or DOX) and Cytarabine (Cyt). Principally, miR-580 functions as a tumor suppressor in AML, and SATB1-AS1 sponges miR-580 to boost levels of the 2'-5'-oligoadenylate synthetase 2 (OAS2) protein. Furthermore, the SATB1-AS1/miR-580/OAS2 axis is itself regulated by the GSK3 β / β -catenin signaling pathway. This signal transduction is identified by the increased expressions of β -catenin in AML cells exhibiting greater resistance to ADR/Cyt chemotherapeutic treatment. SATB1-AS1 therefore has been identified as a novel lncRNA intricately involved in mediating drug resistance in AML cells and can be exploited as a therapeutic target [88]. BM samples extracted from patients receiving DOX treatment overexpress the lncRNA HOXA-AS2. In addition, miR-520c-3p levels are markedly substandard in both the THP-1/ADR and U937/ADR cell lines, and knocking out HOXA-AS2 enhances miR-520c-3p expression. Furthermore, miR-520c-3p binds to several putative locations on the 3'-UTR of S100A4 mRNA to inhibit its expression. Remarkably, S100A4 is an oncogene that is essential for the development of chemoresistance in a variety of malignancies [89]. Therefore, in tandem, the HOXA-AS2/miR-520c-3p/S100A4 axis is a lucrative target that can be targeted to overcome DOX resistance in AML [90]. lncRNA NEAT2, also known as MALAT1 is an 8.7 knt lncRNA transcript localized in the nucleus at the 11q13.1 chromosomal region in humans that exhibits extensive evolutionary conservation relative to most other lncRNAs [91]. Numerous studies have demonstrated how it affects chemoresistance in different cancers via miRNA sponging including gastric cancer [92–94], lung cancer [95], bladder cancer [96], and breast cancer [97,98]. MALAT1 enhances Ara-C resistance in AML cells by retarding miR-96 expression. It is observed that MALAT1 expression is significantly elevated in the HL60, and THP-1 cell lines exhibiting strong resistance towards Ara-C. On the other hand, there is a marked down-regulation of miR-96, suggesting that MALAT1 functions in opposition to miR-96. MALAT1 expression in AML cells is negated by miR-96 overexpression, and conversely, decreasing miR-96 expression eliminates these effects [99]. Lysine Demethylation 4C (KDM4C), an oncogene frequently discovered to be overexpressed in a variety of malignancies contributes to the course of these diseases as well as their resistance to radiation and medication [100–103]. Interestingly, KDM4C plays a critical role in regulating MALAT1 expression in AML cells. Mechanistically, KDM4C demethylates the MALAT1 promoter region to boost MALAT1 expression in the HL60/Ara-C cell line. Apart from the previously discussed miR-96, another miRNA target of MALAT1 is miR-328-3p. Downregulated miR-328-3p levels are the consequence of MALAT1 overexpression, indicating a negative correlation between the two. Cyclin D2 (CCND2) is a cyclin-dependent kinase that is involved in promoting chemoresistance in various cancers [104,105]. As CCND2 is a target of miR-328-3p in AML, upregulating miR-328-3p reduces CCND2 expression levels, helping AML cells become more sensitive to Ara-C treatment [106]. Therefore, KDM4C's epigenetic regulation of MALAT1 and its subsequent control over the miR-328-3p/CCND2 axis constitutes a unique mechanism for MALAT1-mediated AML chemoresistance. To mitigate Ara-C resistance in AML, therapeutics that oppose and stifle the expression of either or both MALAT1 and KDM4C can be developed. The activation of the myelocytomatosis (MYC or c-MYC) oncogene is considered an important hallmark of cancer due to its role in upregulating self-renewal capacity [107,108], and

chemoresistance [109,110]. Interestingly lncRNAs have been known to regulate c-MYC expression to moderate chemoresistance in different cancers [32,111,112]. The lncRNA XIST is involved in mediating drug resistance and metastasis in various cancers [113]. Compared to normal BM samples, XIST is substantially expressed in KG-1 BM cell lines derived from AML patients. Reduction in XIST expression in KG-1 cell lines positively correlates with enhanced susceptibility to DOX therapy. Furthermore, XIST specifically targets the tumor suppressor miR-29a to block its expression, which in turn increases the expression of the MYC oncogene, leading to heightened DOX resistance in AML cells. Thus, inhibiting XIST expression to limit MYC activity via the XIST/miR-29a/MYC axis may provide a useful therapeutic approach to combating DOX resistance [114]. Almost all eukaryotes contain a class of biological molecules called tetraspanins (Tspans), which are key players in the construction and regulation of the plasma membrane. Tspans act as molecular scaffolds that assemble and attract proteins to specific areas of the cell membrane to form functional units that emit signal pathways [115]. Despite evidence suggesting their importance in the regulation of various cellular processes, Tspans also play important roles in the proliferation and chemoresistance of various carcinomas [116–119]. Interestingly, HL60/ADR and K562/ADR resistant cell lines overexpress the lncRNA KCNQ1OT1, and miR-193a-3p remains repressed as a direct target of KCNQ1OT1. Tspan3 is a downstream target of miR-193a-3p and is shown to be raised in both the ADR-resistant cell lines. Together, KCNQ1OT1 sequesters miR-193a-3p to upregulate Tspan3 and mediate ADR resistance in AML cells. Thus, it is worthwhile to develop strategies to selectively target the KCNQ1OT1/miR-193a-3p/Tspan3 axis to diminish ADR resistance in AML [120]. Surprisingly, abnormal MEG3 lncRNA expressions are linked to a worse prognosis and higher ADR resistance in AML cells. ADR resistance in AML cells can be increased by mutating MEG3 to down-regulate the expression of miR-155 and increase the expression of mannosyltransferase ALG9 [121]. In azacitidine-resistant AML cell lines THP-1-/Aza, another lncRNA, NRF2-AS1, is overexpressed. Its expression is positively associated with that of insulin-like growth factor-1 (IGF1), which is also overproduced in the same cell line. Aza resistance results from NRF2-AS1's inhibition of miR-483-3p expression, which in turn adversely affects IGF1 expression in AML [122]. When looked at from a broader perspective, it is axiomatic to say that the important role played by lncRNAs in modulating drug resistance by inhibiting their miRNA targets is an important mechanism that holds significant therapeutic importance for clinicians.

3.2. Mechanism 2: Signal transduction pathways regulated by lncRNAs

Several signaling pathways play important roles in leukemogenesis and multidrug resistance [123–125]. Interestingly, a wide range of evidence suggests that lncRNAs are also capable of altering certain signaling pathways, resulting in more drug-resistant cells [126–128]. Illustriously, the overexpression of HOTAIR facilitates chemoresistance in AML cells by mediating the AKT/NOTCH1 signaling pathway. p21 and p53 are well-studied biomarkers known for their role in suppressing cancer metastasis [129,130]. HOTAIR is highly expressed in the K562 and A02 ADR drug-resistant cell lines. Selectively inhibiting HOTAIR expression sensitizes AML cells to ADR therapy. Mechanistically, HOTAIR upregulates NOTCH1 gene expression to induce chemoresistance in AML cells and functions as a negative regulator of p21. HOTAIR inhibition leads to an increase in p21 expression levels and a stark decrease in AKT phosphorylation that sensitizes K562/A02 cells to ADR [131]. The lncRNA CRNDE was discovered to be substantially expressed in AML cells with strong ADR resistance in a recent study by [132]. The expression of p-glycoprotein (P-gp) and multidrug resistance protein 1 (MDR1) was significantly elevated in ADR-resistant AML cells. Abolishing CRNDE expression significantly decreased MDR1/P-gp levels and HL60/ADR cells were re-sensitized to ADR. At the molecular level, β -catenin, c-MYC, and cyclin D1 were among the key components that

were inhibited because of downregulating CRNDE, which suppressed the Wnt/ β -catenin pathway. Cytoplasmic lncRNA UCA1 facilitates the desensitization of AML cells to daunorubicin (DNR) by stimulating the PI3K/AKT signaling pathway. The primary miRNA target of UCA1 in the HL60 and U937 AML cell lines is miR-613. Furthermore, in both AML cell lines UCA1 overexpression enhanced the levels of p-PI3K/PI3K and p-AKT/AKT. When combined, these findings suggest that the PI3K/AKT pathway was activated because of UCA1-mediated silencing of miR-613. Therefore, UCA1 increases DNR resistance in AML cells by blocking miR-613, which activates the PI3K/AKT signaling pathway [133]. LINC00239 is overexpressed in KG-1 and HL60 AML cell lines and inhibits the effects of DOX by upregulating the PI3K/AKT/mTOR signaling pathway. Elevated expression of LINC00239 caused greater phosphorylation of mTOR and AKT, resulting in higher resistance of AML cells to DOX in the HL60 AML cell line. This impact was confirmed by pre-treatment with their respective dual inhibitor, NVP-BEZ235, which suppressed the expression levels of mTOR and PI3K/AKT respectively leading to a considerable reduction in the phosphorylation levels enhanced by LINC00239 [134]. lncRNA HOTAIRM1 is highly expressed in AML cells that are resistant to glucocorticoids (GC). ChIP-seq data indicates that the acute myeloid leukemia-1 (AML1) oncogene directly binds to H3K4me3 and H3K27ac sites on HOTAIRM1's promoter region causing increased histone methylation and acetylation, heightening HOTAIRM1 expression. The AML1-HOTAIRM1 molecular scaffold facilitates the repression of the ARHGAP18 gene by sequestering AML1 from the accessible, open chromatin region of ARHGAP18. Reduced ARHGAP18 expression eventually triggers the RHOA/ROCK1 signaling pathway by producing greater amounts of the RhoA-GTP protein complex. This, in turn, causes the BCL-2 gene to be overexpressed, which increases GC chemoresistance in AML [135]. A remarkable work by [136] discovered that the lncRNA GAS6-AS2 plays a significant role in strengthening Ara-C resistance in AML. GAS6-AS2 is overexpressed in the K562/Ara-C line alongside AXL protein and reducing GAS6-AS2 expression declined AXL mRNA levels, effectively sensitizing K262 AML cells to Ara-C treatment. Mechanistically, GAS6-AS2 in-trans interacts with DNMT1 and DNMT3A at the promoter region of AXL to reduce methylation on its upstream CpG sites that consequently enhances AXL transcription. Additionally, the effects of GAS6-AS2 were confirmed in NSG mice *in-vivo* and the MOLM14 AML cell line *in-vitro*. It was found that GAS6-AS2 expedites the activation of the GAS6/TAM signaling axis, which in MOLM14 cells imparts the observed Ara-C resistance.

3.3. Mechanism 3: Regulation of histone modifications & DNA methylation

Previously, I discussed how HOTAIR participates in the Wnt/ β catenin pathway to enhance chemoresistance in AML. HOTAIR further imparts chemoresistance in AML through another pathway. The Phosphate and tensin homolog (PTEN) is a tumor suppressor gene that negatively regulates the expression of the PI3K/AKT/mTOR signaling pathway, a major pathway involved in the development of many cancers including AML [137]. Under-expressed or mutated forms of PTEN expression have been identified in various malignancies [138–140]. It was recently found that the HL60/ADM cell line showed heightened levels of HOTAIR while PTEN was starkly downregulated due to a heavily methylated promoter region. Silencing HOTAIR resulted in a marked downregulation of DNMT3b while rescuing its expression had the opposite effect. Therefore, the overexpression of HOTAIR enhances DNMT3b levels which diminishes PTEN and culminates in increased ADM resistance [141]. It is noteworthy that additional research is necessary to delve deeper into the function of HOTAIR in controlling AML chemoresistance, as this work did not identify any additional downstream target of PTEN or precisely annotate a mechanism that controlled HOTAIR expression in AML. The tumor suppressor gene located at the chr5q31.1 chromosomal region - Zinc finger CCHC-type

containing 10 (ZCCHC10) plays an active role in downregulating various cancers [142]. However, it is often under-expressed or mutated in various cancers causing carcinogenesis [143]. According to [144] the lncRNA SNHG1 epigenetically silences ZCCHC10 expression in AML cells allowing them to develop resistance towards venetoclax. Briefly, in a similar mechanism adopted by HOTAIR to suppress PTEN in ADM-resistant AML cells, lncRNA SNHG1 functions as a guide molecule to bind directly to both DNMT1 and DNMT3b and aid them to the promoter of ZCCHC10 to inhibit its expression. Venetoclax functions as a BCL-2 inhibitor [145] and enhanced ZCCHC10 mediated apoptosis of AML cells in ML2 and ALM3 cell lines and in mice incubated with MOLM-13-Vec and MOLM13-Zh10 xenograft models. Therefore, the SNHG1/ZCCHC10/p53 apoptosis pathway can serve as a valuable therapeutic target for AML therapy. Enhancer of zeste homolog 2 (EZH2), the enzymatic catalytic component of PRC2, increases the H3K27 methylation of its target genes to affect their expression [146]. Chemoresistance in malignancies, including leukemias, is caused by hypermethylated CpG islands and anomalous DNA methylation records at CpG-depleted areas [147,148]. Moreover, it's also thought that elevated H3K27me3 can facilitate chromatin remodeling, which drives the target genes' chromatin into a more docile state to minimize DNA-drug interactions [149] as a means to increase drug resistance in tumors. EZH2 positively regulates H3K27me3 expression to confer chemoresistance in neoplastic diseases. The role of EZH2-mediated H3K27me3 chemoresistance remains enigmatic in AML. Intriguingly, the lncRNA TUG1 directly binds to EZH2 to epigenetically limit the expression of miR-34a as observed in HL60 and HL60/ADR cell lines. TUG1 improves EZH2 recruitment on miR-34a to enrich H3K27me3 levels on the miR-34a promoter region which substantially reduces AML cells susceptibility to ADR treatment. Taken together, these findings imply that TUG1 promotes chemoresistance in AML by downregulating miR-34a through EZH2 recruitment [150]. The TUG1/miR-34a/EZH2 regulatory axis is a useful therapeutic target to improve ADR sensitization in AML cells. The antisense lncRNA USP30-AS1 is located on the chr12(q24,11) on the USP30 gene's opposite strand [151]. [40] discovered that the lncRNA USP30-AS1 regulates USP30 gene expression to enhance survival and inhibit apoptosis in HL60 AML cells. Mechanistically, USP30-AS1 enriched USP30's promoter region with both H3K4me3 and H3K27ac marks while the former histone mark was enhanced partially by binding to the protein complex ASH2L.

3.4. Mechanism 4: lncRNAs participate in DNA damage repair pathways

Chemotherapy and radiotherapy treatments currently offered to cancer patients can elicit various unwarranted immunogenic reactions and induce injuries in various tissues causing genomic instability. Irregularities in the human genome are also a direct cause of the development and recurrence of various cancers [152]. lncRNA-mediated DNA damage repair induces chemoresistance in various malignancies [153–155]. Therefore, a greater understanding of the pathways regulated by lncRNAs leading to DNA damage repair might reveal new insights about how AML cells become resistant to external drug and radiation influence. Recently [156], identified that the nuclear factor kappa B subunit (RELA), an important component of DNA repair pathways, modulates the transcription of lncRNA uc002jit. In their previous study, the authors proved that RELA also regulated the transcription of poly (ADP-ribose) polymerase 1 (PARP1) and dual silencing of both PARP1 and RELA resulted in elevated levels of DNA damage in AML cells [157]. Accumulating evidence suggests that PARP1 upregulation is associated with greater resistance to anti-cancer treatment [158–160]. Interestingly, blocking uc002jit directly lowers PARP1 mRNA stability and levels, making AML cells more susceptible to DNR therapy. Hence the RELA/uc002jit/PARP1 pathway holds significant value for researchers to manufacture drugs aimed at inhibiting uc002jit in cancer cells to activate and elevate levels of DNA repair. LINC00152 is a large intergenic lncRNA of length 828 bp located on the 2p11.2 chromosome.

Extensively studied, this specific lncRNA has been known to regulate chemoresistance in many malignancies [161–164]. Recently [165], identified that LINC00152 was highly expressed in CD34⁺CD38[−] leukemic stem cells (LSCs), and silencing LINC00152 resulted in substandard PARP1 expression which was also significantly elevated in CD34⁺CD38[−] cells which further sensitized LSCs to DOX treatment. Although a detailed mechanism is still unknown, nonetheless, since both LINC00152 and PARP1 were found to be highly expressed in signaling pathways associated with DNA damage repair, future research should be directed towards investigating the exact mechanistic action of LINC00152 in DNA damage repair in AML.

3.5. Mechanism 5: Autophagy regulation by lncRNAs

Autophagy is a self-destructive process in which defective cells and organelles are eliminated. Often, they are also recycled to preserve cellular functioning during endogenous or external nutritional stress. Changes in various autophagy processes play a significant role in the genesis of several deadly diseases, including cancer [166]. The mechanisms regulating autophagy in malignant neoplasms remain an active topic of scientific investigation. Interestingly, lncRNA-mediated autophagy-related pathways play an active role in promoting resistance to anti-cancer drugs [17,34,167,168]. The lncRNA DANCR is overexpressed in Ara-C-resistant AML cell lines. It imparts drug resistance by restricting miR-874-3p expression. Downregulating miR-874-3p decreases its inhibitory effect on ATG16L1, a significant autophagy-related protein that promotes autophagosome formation [169] whose upregulation is linked to enhanced drug resistance. As a result, DANCR-mediated autophagy serves as an anti-apoptotic route through which AML cells increase their survival in response to medication effects. The DNAJ family is a class of proteins associated with heat shock protein 70s (Hsp70s); that are essential matrons involved in various structural and protein activities like folding and degradation. DNAJB9 is a newly discovered member of this protein family; responsible for stimulating ATPase enterprises by interacting with binding immunoglobulin protein (BiP) in the endoplasmic reticulum (ER) lumen, where it is localized [170]. The lncRNA SNHG5 has miR-32 as its proximate target and inhibits its expression. DNAJB9 shares a positive correlation with SNHG5 and conversely, a negative correlation exists between miR-32 and DNAJB9. SNHG5 modulates the SNHG5/miR-32/DNAJB9 axis to directly target autophagy for enhancing drug resistance in AML cells [171].

3.6. Mechanism 6: Metabolic control - The Warburg effect & lncRNAs in AML

Changes in metabolic pathways in cancer cells are crucial distinguishing features that set them apart from other normal cells in the body. Cancer cells frequently modify their metabolic pathways to maintain survival and growth, and a better understanding of these pathways will surely contribute to the development of more effective treatments [172]. New evidence suggests that lncRNAs play an active role in regulating cancer metabolism through signaling pathways and post-transcriptional and epigenetic modifications that are associated with increased drug resistance and poor prognosis [173–177]. The alarmingly high rate of glucose uptake by cancer cells to enhance aerobic glycolysis and lactate generation is known as the Warburg effect [178]. A discussion on the mechanisms regulating the Warburg effect is herein not elucidated, however, for a further academic understanding, I direct the readers to a review by [179]. This effect plays an important role in imparting chemoresistance to various cancers [180–182] including AML. The overexpression of the lncRNA UCA1 is directly responsible for reversing the inhibition of hypoxia-inducible factor 1- α (HIF-1 α) reliant glycolysis in HL60 and HL60/ADR cell lines. Mechanistically, UCA1 binds to miR-125a, thereby restraining its functionality and upregulating its target gene, HK2 expression, responsible

for elevated glycolysis in AML. In this way, UCA1 increases the chemoresistance in pediatric AML, and the UCA1/miR-125a/HK2 axis commands therapeutic importance [183]. The lncRNA TUG1 is highly expressed in HL60/ADR AML cell lines and unsurprisingly silencing TUG1 enhances the apoptotic effects of ADR. Curiously, the Warburg effect is also upregulated; characterized by aberrant uptake of glucose due to upregulated mRNA expression levels of HK2 and PKM2 in the HL60/ADR cell line. The enhanced levels of the PI3K/AKT signaling pathway are involved in intricately regulating the Warburg effect to induce chemoresistance in various carcinomas [184–186]. Recently a study by [187] corroborates the results from previous studies since downregulating TUG1 reduced AKT levels through reduced phosphorylation whilst hyperactivating AKT signaling reversed the ADR cytotoxic effects on abrogating TUG1 expression. Therefore, TUG1 enhances ADR resistance in AML by elevating the Warburg effect via enhancing AKT signaling [188].

3.7. Mechanism 7: Transcriptional regulation by lncRNAs as scaffolding domains

lncRNAs can regulate gene expression by functioning as molecular scaffolds for the assembly of various ribonucleic complexes (RNPs) and transcription factors [189]. Thus, lncRNAs can exercise precise genetic control by directly binding to multiple effector molecules for regulating the spatiotemporal dynamics of complex cellular events. lncRNA scaffolding is gaining increased attention due to its important role in the development of chemoresistance in various cancers [27,190]. A better understanding of the role of such scaffolding effects in cancer will allow researchers to change cell fate by using strategies to boost or suppress the expression of various molecular species involved in the design and assembly of such domains, which are quite complex [191]. Recently, a novel lncRNA AC026150.8 has been reported to function as a molecular scaffold for regulating chemoresistance in AML. In KG-1 and K562 cells, heightened levels of AC026150.8 led to stronger desensitization to Ara-C, while dampening its expression eliminated the consequences of drug resistance. Western blotting and bioinformatics research verified that the splicing factor, poly(rC)-binding protein 1 (PCBP1), is recruited by AC026150.8. Therefore, it is proposed that the PCBP1 - AC026150.8 scaffold controls atypical splicing in AML cells to increase their resistance to Ara-C [192]. However, since the precise mechanism of AC026150.8 - PCBP1 scaffold is only suggested to be able to mediate splicing in AML cells, more confirmation is needed. If validated, it would be interesting to know if any additional effector molecules outside the PCBP1 factor can attach to AC026150.8 to control splicing in AML cells. Since it is evident that this interaction does promote Ara-C resistance in AML cells, it is also vital to ascertain the specific mechanism of the AC026150.8 - PCBP1 scaffold. 40–160 nm wide extracellular vesicles inscribed within a lipid bilayer in eukaryotic species are known as exosomes [193]. Exosomes harbor various important molecular species including but not limited to RNAs, metabolites, and proteins [194], and are now considered to be crucial components of various aspects of cell communication, and signaling for mediating biological processes like wound healing [195–198], cell differentiation [199], and immune responses [200–202]. Interestingly lncRNAs transmitted by exosomes (exosomal lncRNAs) also regulate chemoresistance in various cancers and are important therapeutic targets [177,203–207]. In a fantastic study recently by [208], for the first time, a relationship between exosomes and lncRNAs to promote chemoresistance in AML has been established. The m⁶A demethylase FTO was overexpressed in bone marrow-derived mesenchymal stem cell exosomes (BM-MSCs-Exo). AML cells treated with FTO-exosomes (FTO-exo) in high levels were particularly unresponsive to Ara-C treatment. Mechanistically, FTO-exo boosted lncRNA GLCC1 expression in THP-1 and Kasumi-1 AML cell lines by demethylating the promoter of lncRNA GLCC1 to improve GLCC1-HuR binding. In addition, GLCC1 served as a scaffold between the oncogene c-MYC and insulin-like growth factor 2 mRNA binding

protein (IGF2BP1) and FTO-exo strengthened GLCC1/c-MYC signaling axis, making AML cells more resistant to Ara-C. It's vital to remember that this study solely confirmed the findings in an *in-vitro* environment and *in-vivo* modeling is still warranted to truly comprehend how GLCC1 scaffolding influences AML chemoresistance. The findings nonetheless offer compelling new insights (Table 1) (Fig. 2).

4. lncRNAs control apoptosis resistance pathways in AML

Achieving high amounts of cellular apoptosis of cancer cells is the primary goal of cancer treatment. In short, apoptosis is a type of planned cell death brought on by the dissolution of the cell membrane caused by caspase 6 (CAS6), which shreds down DNA into primitive structures [209]. The human genome is vulnerable to endogenous stresses caused by chemotherapy medications. These stresses result in the recruitment of BCL-2 and Bak to the mitochondria and the release of cytochrome *c* into the cytosol. The apoptosome protein complex is then generated through

the interplay amongst procaspase-9, Apaf-1, and cytochrome *c*. Apoptosomes cause caspase-9 (CAS9) and caspase-3 (CAS3) to become activated, which leads to apoptosis and therapeutic resistance in AML cells [210]. In contrast, the extrinsic pathway is regulated by the cellular communications between ligands and multiple cell death receptors like FAS [211]. Interestingly, pro-apoptotic gene downregulation, self-renewal of cancer cells, and abnormal expression of genes like BCL-2 are all significant contributors to greater chemoresistance in malignancies that protect cancer stem cells (CSCs) from apoptosis-mediated death. lncRNAs are known to regulate anti-apoptosis pathways to confer drug resistance in various malignancies. For instance, lncRNA SNHG6 upregulated BCL-2 expression through sequestering miR-127 to induce cisplatin resistance in gastric cancer [212]. Another lncRNA H19 prevented apoptosis in multiple myeloma cells by sponging miR-29b-3p expression to mediate and upregulate the translation of anti-apoptotic myeloid leukemia cell differentiation protein (MCL-1) that subsequently enhanced bortezomib resistance [213]. Recently the role of

Table 1
lncRNAs Regulate Chemoresistance in AML.

lncRNA	Molecular Target	Mechanism of Action	Cell Line Used	Modeling Environment	Drugs Used	References
SATB1-AS1	miR-580	Sequesters miR-580 to upregulate OAS2 expression	HL-60/ADR OCI-AML5/Cyt	Both <i>in-vitro</i> and <i>in-vivo</i>	ADR and Cyt	[88]
HOXA-AS2	miR-52C-3p	Downregulates miR-52C-3p expression to upregulate S100A4	U937/ADR and THP-1/ADR	<i>In-vitro</i>	ADR	[90]
MALAT1	miR-96 and miR-328-3p	Suppresses both miR-328-3p and miR-96. Under expression of miR-328-3p results in upregulation of CCND2	HL60 and THP-1	<i>In-vitro</i>	Ara-C	[99,106]
XIST	miR-29a	Upregulates c-MYC expression by antagonizing miR-29a	KG-1 BM	Both <i>in-vitro</i> and <i>in-vivo</i>	DOX	[114]
KCNQ1OT1	miR-193a-3p	Promotes Tspan3 expression by repressing miR-193a-3p	HL60/ADR and K562/ADR	<i>In-vitro</i>	ADR	[120]
MEG3	miR-155	Reduces expression of miR-155 to increase ALG9 levels	U937/ADR and THP-1/ADR	<i>In-vitro</i>	ADR	[121]
NRF2-AS1	miR-483-3p	Silences miR-483-3p expression to enhance IGF1 mRNA levels	THP-1/Aza	<i>In-vitro</i>	Aza	[122]
HOTAIR	p21/NOTCH signaling	Suppresses p21 expression and enhances NOTCH1 expression levels	K562/ADR	<i>In-vitro</i>	ADR	[131]
CRNDE	β -catenin, c-Myc, cyclinD1/Wnt/ β -catenin pathway	Upregulates β -catenin, c-Myc, and cyclinD1 to enhance the Wnt/ β -catenin pathway	HL60/ADR	<i>In-vitro</i>	ADR	[132]
UCA1	miR-613/PI3K/AKT/mTOR pathway	Suppresses miR-613 to enhance levels of p-PI3K/PI3K and p-AKT/AKT	HL60 and U-937	<i>In-vitro</i>	DNR	[133]
LINC00239	mTORC1	Increased mTORC1 activity by upregulating phosphorylation of mTOR and AKT	HL60	<i>In-vitro</i>	DOX	[134]
HOTAIRM1	ARHGAP18	Sponges AML1 away from ARHGAP18 to activate RHOA/ROCK1 signaling	HL60/ADR and K562/ADR	<i>In-vitro</i>	ADR	[120]
HOTAIR	DNMT3b	Suppresses PTEN expression by enhancing DNMT3b expression levels	HL60/ADM	<i>In-vitro</i>	ADM	[141]
SNHG1	ZCCHC10	Recruits DNMT1 and DNMT3b to ZCCHC10 promoter to downregulate its expression	ML2 and ALM3 cell lines and MOLM-13-Vec and MOLM13-Zh10 xenograft models	<i>In-vitro</i> and <i>in-vivo</i>	Venetoclax	[144]
TUG1	miR-34a	Suppresses miR-34a by enhancing EZH2 recruitment on miR-34a to enrich H3K27me3 levels on its promoter region	HL60/ADR	<i>In-vitro</i>	ADR	[150]
USP30-AS1	USP30	Enriches promoter region of USP30 with both histone methylation (H3K4me3) and acetylation (H3K27ac) marks	HL60	<i>In-vitro</i>	N/A	[40]
uc002jit.1	PARP1	Enhances PARP1 expression levels and stability	KG1 α xenograft model and Kasumi-1, and HEK293 cell lines	Both <i>in-vitro</i> and <i>in-vivo</i>	DNR	[156]
LINC00152	PARP1	Potentially regulates PARP1 expression (no specific mechanism discovered)	CD34 ⁺ CD38 ⁻ LSCs	<i>In-vitro</i>	DOX	[165]
DANCR	miR-874-3p	Raises ATGL16 protein levels by repressing miR-874-3p	HL60, U937, and KG1 α cell lines	<i>In-vitro</i>	Ara-C	[296]
SNHG5	miR-32	Inhibits miR-32 expression to upregulate DNAJB9 levels	HL60, HL60/ADR	<i>In-vitro</i>	ADR	[171]
UCA1	miR-125a	Inhibits miR-125a expression to upregulate HK2 levels	HL60, HL60/ADR	<i>In-vitro</i>	ADR	[183]
TUG1	AKT	Enhances AKT signaling	HL60/ADR	<i>In-vitro</i>	ADR	[188]
AC026150.8	PCBP1	The AC026150.8 - PCBP1 scaffold mutates alternative splicing in AML cells	KG-1 and K562 cell lines	<i>In-vitro</i>	Ara-C	[192]
GLCC1	c-MYC and IGF2BP1	Functions as a bridge to assemble c-MYC and IGF2BP1 on its domain	THP-1 and Kasumi-1 cell lines	<i>In-vitro</i>	Ara-C	[208]

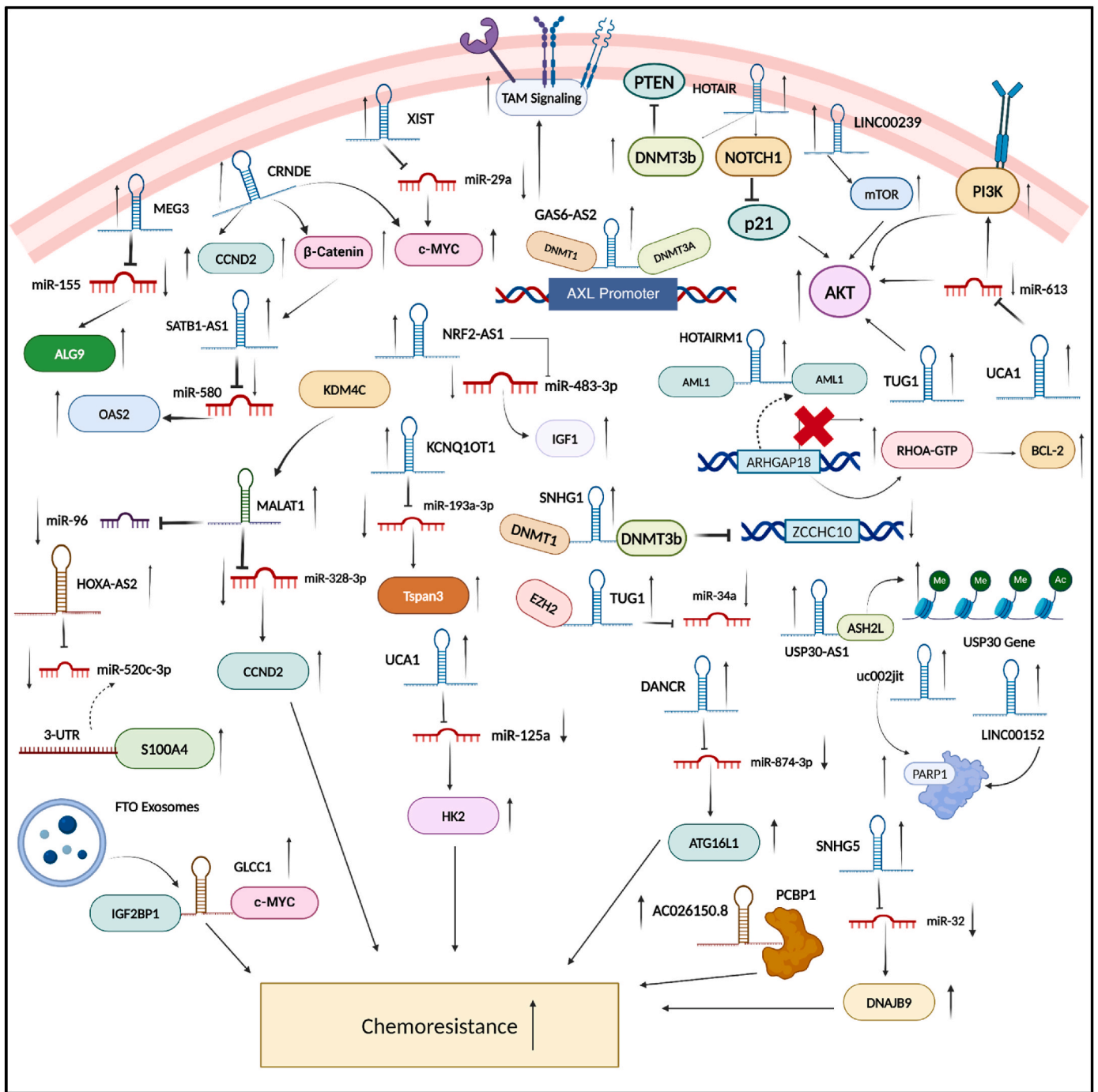


Fig. 2. lncRNAs regulate chemoresistance in AML

Schematic illustration summarizing the mechanisms by which lncRNAs regulate drug resistance in AML. The major molecular mechanisms through which lncRNAs enhance chemoresistance are A) miRNA sequestration, B) signal transduction pathways, C) histone modifications and DNA methylation, D) DNA damage repair pathways, E) autophagy and metabolic reprogramming, F) molecular scaffolding.

lncRNA HOTTIP in regulating FAS signaling to prevent apoptosis in various AML cell lines was examined. In detail, on knocking out HOTTIP in WT MOLM3 cells it was observed that the expression of FAS, CAS3, and CAS9 were significantly upregulated, and a similar effect was observed in CBS7/9^{+/-} and OCI-AML13 cell lines while the latter two cell lines also exhibited repressed HOXA gene expressions. It is proposed that at a transcriptional level, HOTTIP upregulates the expression of miR-196b that directly represses FAS expression as was observed with the increased apoptosis of AML cells on miR-196b downregulation in the MOLM13 cell line [214]. Although more research is necessary to fully

understand this mechanism, these findings demonstrate that HOTTIP controls the FAS signaling axis to prevent cellular apoptosis in AML cells. Boosting the lncRNA SLED1 expression in Kasumi and U937 AML cells resulted in diminished apoptosis when administered DOX treatment. In addition, flow cytometry studies confirmed increased apoptosis on abolishing SLED1, and it is postulated that SLED1 affects AML cell survival via two different mechanisms. First, as shown by KEGG pathway analysis, by participating in the PI3K/AKT signaling pathway; and second, by elevating BCL-2 expression. Although SLED1's mode of action is merely speculated, it is an intriguing point to consider with evident

therapeutic significance for subsequent research into investigating the potential of lncRNAs in mediating anti-apoptosis in AML [215]. Hepatocyte nuclear factor 4 α (HNF4 α) is an important transcription factor that plays an instrumental part in gene regulation by attracting cofactors to the promoter regions of its target genes [216] to facilitate metabolism and cellular development. Abnormal expressions of HNF4 α are associated with chemoresistance and metastasis in various cancers [217–219]. Recently it was demonstrated that LINP1 controlled HNF4 α expression to positively affect leukemic cells' capacity to survive. Moreover, a drop in the expression levels of lactate dehydrogenase A (LDHA) and glucose transporter 1 (GLUT1) was caused by silencing both LINP1 and HNF4 α , which reduced the uptake of glucose by AML cells. Lastly, over-expressing HNF4 α and LINP1 led to a considerable increase in pAMPK and WNT5A levels, suggesting that LINP1 regulates glucose metabolism in AML cells through the HNF4 α /AMPK/WNT5A metabolic pathway to enhance cell survival [52].

5. Mechanisms of leukemic stem cell self-renewal by lncRNAs

The ability of CSCs to self-renew remains one of the main obstacles to fighting chemoresistance. The ability of CSCs to differentiate and develop into various lineages is known as "self-renewal," and it is essentially limitless [220]. This unique ability of CSCs to protect themselves against apoptosis-mediated cell death is one of the main causes of patients' poor response to chemotherapy regimens and poor prognosis due to relapses [221–223]. A growing body of research indicates that lncRNAs actively mediate the ability of CSCs to undergo self-renewal and that they can both stimulate [224–227] and impede [41,226,228] this process. Pediatric AML patients with MLL-type leukemia, a very aggressive subtype of AML, have a dismal prognosis even with intensive use of cutting-edge chemotherapy treatments [229]. One of the main causes of these results is that the disease can reappear after drug exposure, which lowers the survival rate [230,231]. DOT1-like (DOT1L) methyltransferase protein uniquely interacts with the H3K79 histone site to regulate H3K79me/H3K79me₂/H3K79me₃ histone methylation in human genes [232] and its aberrant expression is associated with many facets of carcinogenesis [233,234]. Recently [235] investigated the role of lncRNA LAMP5-AS1 in regulating DOT1L in MLL leukemia. H3K79me₃/H3K29me₂ levels were dramatically reduced at loci of HOXA9, HOXA10, CDK6, and MEIS1 cancer stem genes situated in the HOXA cluster on LAMP5-AS1 knockdown in THP-1 and MOLM13 MLL patient cell lines. The methyltransferase activity of DOT1L was significantly affected by downregulating LAMP5-AS1, which markedly downregulated the expressions of cancer stemness genes preventing MLL recurrence. Consequently, LAMP5-AS1 might be a valuable therapeutic target to further abrogate self-renewal in AML. It is underestimated how transcription factors control the expression of lncRNAs, which in turn drives cancer development. Due to their interactions with many transcription factors, numerous studies have shown the carcinogenic relevance of lncRNAs as co-transcriptional regulators of gene expression [236–238]. Research indicates transcription factors actively contribute to chemoresistance induction in malignancies by affecting the expression of lncRNAs. For example, to impart oxaliplatin resistance in colorectal cancer cells, the transcription factor MYBL2 (B-MYB) stimulates the production of the lncRNA CCAT1, which downregulates the expression of the suppressor of cytokine signaling 3 (SOCS3) [67]. In urothelial cancer, another lncRNA TUG1, whose expression is boosted by the transcription factor nuclear factor erythroid 2 (Nrf2), increases DOX resistance [239]. Intriguingly [240], have reported that the lncRNA HOXB-AS3 plays a critical role in elevating the self-renewal capacity of LSCs. YTH domain-containing protein 1 (YTHDC1) is an important m⁶A reader that plays a crucial role in promoting chemoresistance in breast cancer [241,242], bladder cancer [140], colon cancer [243], ovarian cancer [244], and prostate cancer [245]. Inhibiting YTHDC1 expression in CD34⁺CD38⁻ LSCs accelerates the AML cell's demise. Mechanistically, YTHDC1 interacts with the m⁶A

alteration of the HOXB-AS3 RNA precursor to upregulate the expression of the spliceosome NR_033205.1. It appears that the YTHDC1/HOXB-AS3 circuit is a new and significant target that can be used to prevent LSCs from regenerating themselves, hence enhancing drug efficacy and patient health. The p15 tumor suppressor gene is frequently downregulated due to elevated methylation on its promoter region, and this facilitates leukemogenesis [246]. *In-vitro* studies using the U937 and Kasumi-1 AML lines, as well as *in-vivo* modeling using the U937 mouse xenograft model, demonstrated that HOTAIR increased methylation at the promoter of the p15 gene by enlisting the PRC2 complex to upregulate H3K27me₃ trimethylation, which continued the maintenance of LSCs [247] (Table 2) (Fig. 3).

6. Therapeutic relevance of tumor suppressor lncRNAs in AML

lncRNAs are often categorized as either oncogenic or tumor suppressors based on whether they play an important role in promoting carcinogenesis or inhibiting it. Often tumor suppressor expressions are significantly downregulated in cancers and upregulating their expressions can potentially counteract or abrogate altogether oncogenic effects like chemoresistance and self-renewal of CSCs. The importance of a lncRNA's fundamental nature in defining its activity is rather surprising, given that a lncRNA can retain oncogenic capabilities in one form of cancer while exhibiting tumor-suppressive features in another. Take for instance the case of the lncRNA NEAT1. It facilitates cancer cell proliferation, growth, and metastasis in various malignancies [248–251]. Unexpectedly, contrary to its oncogenic nature, lncRNA NEAT1 serves as a tumor suppressor in AML [252]. demonstrated that the expression of lncRNA NEAT1 was starkly downregulated in the K652 and THP-1 AML cell lines. Enhancing NEAT1 expression in both cell lines greatly sensitized AML cells to treatment by Alisertib and Bortezomib chemotherapeutic drugs. As previously mentioned, ABC proteins play a crucial role in desensitizing cancer cells to treatment, and overexpressing NEAT1 expression in both cell lines leads to a decrease in ABCG2 protein expression effectively reducing drug resistance. Furthermore, NEAT1 (specifically, NEAT1_1 isoform) is under-expressed in CD34⁺ LSCs. NEAT1_1 reduces the ability of LSCs to renew themselves by inhibiting the Wnt/ β signaling cascade and quickening the degradation of the DVL2 protein by interacting with the Trim56 E3 ubiquitin ligase [253]. Another unique lncRNA, MAGI2-AS3, possesses therapeutic significance since it reduces the ability of LSCs to reincarnate themselves by inducing the TET2 DNA methylase on the promoter of the LRIG1 protein that it targets. Additionally, it attaches to the LRIG1 protein directly and boosts demethylation on LRIG1's promoter [41]. Remarkably, a single lncRNA can function inside the same cancer as both an oncogene and a tumor suppressor. The lncRNA MEG3 is renowned evidence of this paradox. MEG3 can promote cellular death as well as the development of chemoresistance. Research carried out *in-vitro* on WT MOLM13 and U937 AML cell lines, as well as *in-vivo* on NSG mice, proved that overexpression of MEG3 inhibited the formation of tumors by ceasing the G0/G1 cell cycle phase. MEG3 has an intriguing dual mechanism of action as a tumor suppressor. Firstly, MEG3 stabilizes p53 to increase the level of RB protein, which subsequently lowers the expression of the DNMT3A DNA demethylase. Furthermore, TET2 and WT1 collaborate to control and improve MEG3 expression, which further suppresses tumor growth [254]. Additionally, it is noted that MEG3 expression is positively associated with miR-493-5p and that MEG3 is present in HL60/Ara-C AML at substandard levels. Raising MEG3 levels increases the production of miR-493-5p, which post-transcriptionally silences METTL3, preventing the activation of the MYC gene which consequently re-sensitizes AML cells to Ara-C therapy [255]. Nevertheless, because MEG3 sequesters miR-155 to raise ALG9 levels, which confers ADR resistance in AML cells, aberrant expression of MEG3 is also linked to the onset of chemoresistance [121]. Tumor suppressor lncRNAs typically work by inhibiting the expression of oncogenes, resulting in increased apoptosis. Illustriously, the lncRNA transcript H22954 proves to be an

Table 2
lncRNAs Modulate Apoptosis Resistance & Self Renewal in AML.

lncRNA	Molecular Target	Mechanism of Action	Cell Line Used	Modeling Environment	Drugs Used	References
HOTTIP	miR-196b	Potentially elevates miR-196b to control FAS signaling (more confirmation needed)	MOLM3, CBS7/9 ^{+/-} and OCI-AML13 cell lines	Both <i>in-vitro</i> and <i>in-vivo</i>	N/A	[214]
SLED1	Proteins in PI3K/AKT signaling and BCL-2	Potentially targets PI3K/AKT signaling and also elevates BCL-2 expression (more confirmation needed)	U937, Kasumi cell lines	<i>In-vitro</i>	DOX	[215]
LINP1	HN4F α	Elevates HN4F α Expression to further increase pAMPK and WNT5A levels	KG-1, THP-1, and NOD/SCID (NSG) mice models	<i>In-vitro</i> and <i>in-vivo</i>	N/A	[52]
LAMP5-AS1	DOT1L	Directly interacts with DOT1L to upregulate H3K79me3/H3K29me2 histone methylation at HOXA9, HOXA10, CDK6, and MEIS1 genes	THP-1, MV4-11, RS4-11, HEK293T, MOLM13 MLL cell lines and NOD/SCID mice models	Both <i>in-vitro</i> and <i>in-vivo</i>	N/A	[235]
HOXB-AS3	N/A	YTHDC1 regulates HOXB-AS3 expression by enhancing its spliceosome NR_033205.1 levels	CD34 ⁺ CD38 ⁻ LSCs (THP-1 cell line isolated) and NSG mice	Both <i>in-vitro</i> and <i>in-vivo</i>	N/A	[240]
HOTAIR	p15	Recruits PRC2 on the promoter of p15 to increase methylation on its promoter	U937 and Kasumi-1 AML lines and U937 mouse xenograft model	Both <i>in-vitro</i> and <i>in-vivo</i>	N/A	[247]

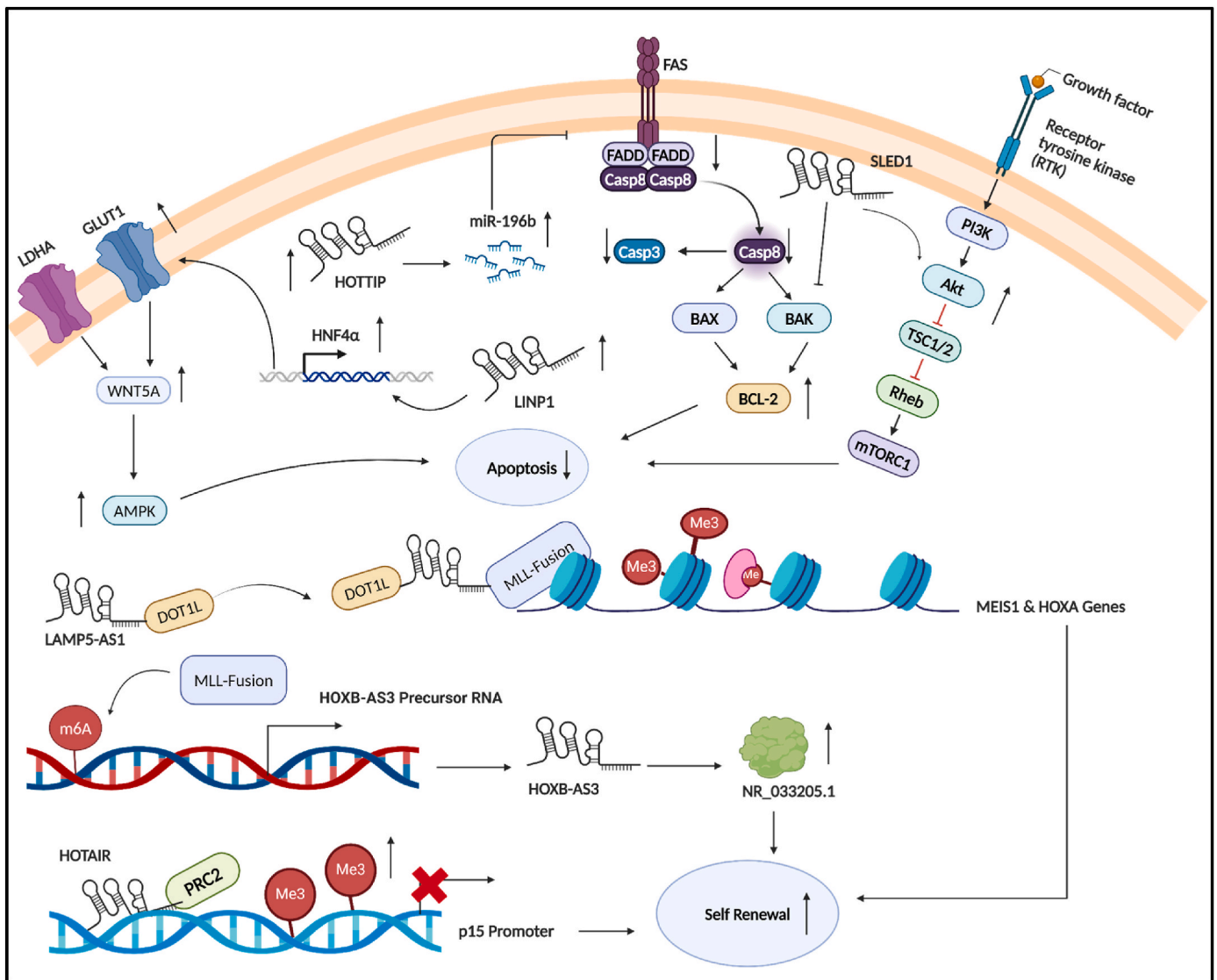


Fig. 3. lncRNAs Regulate Apoptosis Resistance & LSC Self Renewal Pathways in AML

Diagrammatic representation of the processes by which lncRNAs support the prevention of apoptosis and improve LSCs' ability to renew themselves. lncRNAs possess the ability to subvert pro-apoptotic gene expressions, rewire metabolism, and even aberrantly upregulate transduction pathways like the PI3K/AKT signaling to impede apoptosis. Furthermore, lncRNAs increase H3K79me3/H3K29me2/H3K79me3 histone methylation at their target gene's promoter region and interact directly with transcription factors, consequently enhancing LSC self-renewal potential.

important therapeutic target for overcoming BCL-2-mediated drug and extrinsic apoptosis resistance, as it lowered BCL-2 expression to enhance the expression of CAS9, CAS7, and PARP, while all effects of H22954 were reversed on rescuing BCL-2 expression [256]. In another case, the lncRNA LINC00998 has been found to increase the synthesis of the RNA-binding protein ZFP36 by binding directly to its promoter. As a result, ZFP36 binds to the 3' UTR of its target mTORC2 gene, compromising its expression by accelerating the process of mTORC2 mRNA degradation [257]. The data from the preceding research provides some significant insights and perspectives that further emphasize the need to create lncRNA-based treatments to counteract chemoresistance and enhance apoptosis levels in AML.

7. Perspectives & future directions

The largest barrier to improving AML patients' health remains chemoresistance. The resistance of AML cells to therapeutic effects is caused by a multitude of causes, but this review intends to highlight the role that lncRNAs play in controlling this process. It is important to understand that there is a great deal of variation in the chemotherapeutic medications employed, the pathways annotated, and the experimental modeling methodology across all the research reviewed. While most of the research examined in this review only contains *in-vitro* results, the absence of *in-vivo* verification makes it difficult to apply the results to valuable clinical settings. Therefore, using animal or patient-derived xenograft-based *in-vivo* modeling, future studies should examine and validate the mechanisms and effects revealed in the discussed publications. Validating the mechanistic actions of the lncRNAs GLCC1, AC026150.8, SLED1, TUG1, UCA1, SNHG5, DANCR, LINC00152, USP30-AS1, HOTAIR, HOTAIRM1, LINC00239, CRNDE, KCNQ1OT1, MEG3, NRF2-AS1, and MALAT1 is still warranted. To fully understand several of the above-discussed mechanisms, additional experimental validation is needed. For example, in the case of lncRNA AC026150.8, it is merely hypothesized that, in comparison to healthy control cells, splicing in AML cells may be altered by AC026150.8 due to the scaffold that exists between AC026150.8 and PCBP1. In another case, it was only speculated that LINC00152 would promote chemoresistance in AML through DNA repair pathways since both PARP1 and LINC00152 were significantly expressed in these pathways. It is evident that further research is necessary, and these are promising areas to investigate for the next studies. Briefly, more experimental work is required to fully understand the mechanisms of lncRNAs AC026150.8, LINC00152, SLED1, HOTTIP, and HOTAIR. The processes by which lncRNAs in AML enhance chemoresistance are remarkably intricate and go beyond straightforward interactions between lncRNAs and miRNAs. It's important to note whether any new downstream target genes and proteins exist in a mechanism where lncRNAs may have an impact on their expression in the processes previously mentioned. For instance, it's unclear if there are any further downstream targets for the lncRNA HOTAIR than PTEN that HOTAIR downregulates to regulate chemoresistance. Investigating these possibilities can improve our comprehension of lncRNAs' functioning mechanisms. Therapeutic resistance in AML is regulated by several other routes that have not yet been thoroughly investigated. Recent research has shed light on the impact of lncRNAs on mitochondrial dynamics [258]. It would be beneficial to investigate which lncRNAs influence mitochondrial processes to help AML cells escape therapeutic interference.

8. Challenges & strategies to target lncRNAs in AML

Tremendous developments in genomic technologies have afforded novel perspectives into the structural intricacies and functioning of the human genome. This has opened a vast expanse of fascinating and valuable information about lncRNAs that previously remained oblivious. Despite their inability to code for functional proteins, lncRNAs play a significant role in regulating gene expression and are the subject of

close attention and scrutiny. However, despite the swift technological advancement, our awareness of lncRNAs remains minimal. This is largely due to their low conservation compared to other mRNA species, which makes it challenging to investigate, comprehend, and compare their roles across multiple species. Furthermore, the lack of noticeable expression of lncRNAs makes it difficult to accurately measure their population in both cells and tissues. Lastly, the impact of a particular lncRNA's subcellular localization on its function is something we are just beginning to understand [259]. lncRNAs are often classified according to where they are in a cell and are heavily compartmentalized into subcellular domains including but not limited to the nucleus and cytoplasm. A great majority of lncRNAs, known as nuclear lncRNAs, are known to preferentially localize in the nucleus. Through chromatin remodeling and transcriptional mechanisms, they largely influence gene expression [260]. On the other hand, cytoplasmic lncRNAs play a crucial role in mediating post-transcriptional processes such as translation and mRNA degradation. They also operate as effector molecules in cell signaling pathways and interact with ribonucleic proteins (RBPs) to affect their gene expression [261]. It is evident that the subcellular habitat of lncRNAs directly influences the way they differ in function. Consequently, targeting lncRNAs in the human genome has proven to be difficult and represents a substantial obstacle for clinical studies. Interestingly, because lncRNAs do not code for well-conserved and functional proteins, they have long been regarded as "undruggable" macromolecules. This is one of the reasons why the present treatment regimen does not include any specific drugs aimed solely at regulating lncRNA expressions in AML patients. Nonetheless, fresh developments are paving the road to alter this paradigm as I discuss below.

8.1. Small interfering RNAs

Small interfering RNAs (siRNAs) have proven to be a game changer in the treatment of cancer patients. They work on the principle of RNA interference (RNAi), which is a method of gene silencing that uses a double-stranded RNA (dsRNA) to mandate the targeting of complementary mRNA to inhibit its expression through either transcriptional suppression or by inducing decay [262]. Illustriously, it was determined that siRNA (si-SNHG4) mediated silencing of the lncRNA SNHG4 in colorectal cancer cell line SW1116 appreciably reduced BCL-2 expression while also increasing CAS3 and BAX expression to enhance pro-apoptotic signaling [263]. Numerous other investigations have confirmed that siRNA-mediated lncRNA silencing is effective in boosting apoptosis and making cancer cells more susceptible to anti-cancer therapies [264–266]. Despite the progress, there are still a few issues that need to be resolved before siRNA therapies can be effectively applied in clinical settings. The biggest challenge is to deliver siRNA nanoparticles systemically to special target areas to initiate RNAi. To overcome this obstacle and increase the pharmacological effectiveness of siRNA treatments, lipid nanoparticle (LNP)-based delivery systems are currently being developed. These systems enable the integration of siRNAs into their supramolecular polymer matrix, optimizing the delivery mechanism by improving the drug encapsulation rate and boosting cellular uptake at the target site. Lipid nanoparticles embedded with siRNAs have been proposed to be viable therapies for an array of malignancies through rigorous testing both *in-vitro* and *in-vivo* models [267–270]. siRNAs are now being explored progressively more for applications in AML treatment. [271] investigated that in the t(8; 21) cytogenetic AML type, the lncRNA LINC01257 is significantly expressed. To block the expression of LINC01257, they created a Patisiran (FDA-validated) formulation containing an LNP-si-LINC01257 vessel. After being administered, the vessel was remarkably able to concentrate on and decrease LINC01257 expression, and cell count the Kasumi-1 AML cell line by 55 %. Whilst the study only validated the effects within *in-vitro* setting and *in-vivo* modeling in animal xenograft models is still warranted, nonetheless, this study indeed has set the wheels in motion for future studies to investigate the therapeutic application, safety, and

efficacy of using LNP-siRNA systems to target AML cells.

8.2. Antisense oligonucleotides

Antisense oligonucleotides (ASOs) are unit-stranded oligodeoxynucleotides that can influence protein expression through RNAi reminiscent of siRNAs [272]. Mechanistically, ASOs can manipulate the expression of genes, modify transcriptional and post-transcriptional processes, or sterically obstruct molecule-to-molecule interactions. ASO therapeutics are receiving more attention and are being evaluated in several clinical trials, which has aided in their entry into the pharmaceutical industry. While chemotherapeutic and radiotherapeutic intervention plays a major role in the current AML treatment regimen, novel possibilities for the development of more individualized treatment options, such as ASO therapies, can be launched as we keep discovering additional details about AML's molecular mechanisms. What's more assuring is that several ASO treatments have already been designed to treat AML, and a few of them are presently going through extensive clinical trials. Studies have primarily concentrated on developing strategies that directly target particular protein expressions that serve as surface biomarkers [273,274] or that play a role in anti-apoptotic pathways [275]. Developing and studying the sustainability of ASO treatments that target lncRNA expressions is crucial for achieving greater synergistic anti-cancer behaviors since lncRNAs are indispensable for AML cell self-renewal, chemoresistance, and prevention of apoptosis. While oligonucleotide-based therapies are beneficial, they have limitations that must be addressed before they can be further developed for clinical trials. For instance, incorporating foreign RNA molecules into the body might trigger undesirable immunogenic responses, which may reduce the quantity of medication endocytosed by the cells, further limiting the intended RNAi effects of such treatments [276,277]. Furthermore, such therapies need to minimize any harmful non-target effects to ensure that cells beyond the intended target ones remain in good health. Chemically conjugating glycoprotein molecules to siRNAs has proven effective in selectively targeting hepatocytes and is seen as a significant advancement in overcoming the body's natural biological barriers to nucleic acid medication delivery [278]. Future research can be devoted to potentially developing protein-conjugated siRNA delivery engines or investigating techniques to chemically modify oligonucleotide structures to enhance selectivity towards oncogenic lncRNA expressions in AML cells.

8.3. Plant-based compounds

Currently, compounds found naturally in plants are being significantly reviewed for potential anti-cancer properties. Numerous such compounds isolated from plants, such as Taxol, have served as the basis for the development of blockbuster chemotherapeutic medications, such as paclitaxel, which is extensively used to treat malignancies [279]. There is a growing body of research [280–284] investigating the extraction and engineering of plant-based molecules to create more effective anti-cancer drugs, and these studies may ultimately lead to effective treatments [285]. Accumulating evidence suggests that plant metabolites appear to have promising potential to progress the treatment of AML [286–289]. Studies published recently additionally indicate that such compounds could potentially target lncRNAs in AML and thus accelerate cellular death. For instance, it has been shown that Naringenin (Nar) increases programmed cell death by downregulating the expression of the lncRNA XIST, which was shown to be overexpressed in the HL60 AML cell line. Greater resistance to apoptosis resulted from the overexpression of miR-34a's downstream target HDAC1, which was caused by XIST suppressing miR-34a expression. Transfecting HL60 AML cells, with Nar, restrained XIST expression which facilitated apoptosis. This suggests that Nar-mediated control of the XIST/miR-34a/HDAC1 axis can be further clinically assessed using *in-vivo* models [290]. Matrine, an alkaloid derived from plants, was

shown to target the long noncoding RNA LINC01116 and impede its expression to elevate the levels of miR-592. This, in turn, eliminated the JAK/STAT3 signaling pathway and culminated in greater cellular apoptosis and lowered tumor growth [291]. Even though the aforementioned studies have demonstrated the potential for using plant-based compounds to specifically target lncRNAs in AML, more research is still needed to determine the precise mechanisms of action of these natural molecules. Additionally, greater *in-vivo* tests in animal xenograft models and clinical trials are necessary to effectively translate these findings to beneficial ends.

8.4. Future possibilities for targeting lncRNAs in AML

Transcriptional interference has made it feasible to target genes precisely and efficiently, thanks to the development of revolutionary genetic engineering technologies. However, significant difficulties arise when lncRNA expressions are ablated using ASO and siRNA treatments, such as low knockdown levels and a higher chance of off-target effects [292]. The more complicated a particular lncRNA transcriptional machinery is, the more difficult it is to target it. In such cases, CRISPR/CAS9 positions itself as a unique method with distinct benefits over other available therapeutics. While CRISPR/CAS9-mediated AML treatments have not yet been converted into clinical practice for treating patients, various studies have used the CRISPR/CAS9 system to selectively knock down lncRNAs, and this can potentially lead to novel therapeutic strategies in the future [214,293,294]. Furthermore, as I have detailed above, lncRNAs can directly interact with a variety of protein complexes to boost drug resistance and restrict apoptosis. Consequently, it may be possible to create synthetic morpholino oligomers that will deliberately obstruct and sterically impair such types of interactions between lncRNAs and proteins [295] (Fig. 4).

9. Conclusions

AML is a rare and deadly form of leukemia with a high death rate. Therefore, developing new therapies is crucial to helping those suffering from this difficult, uncommon cancer. I outlined the key mechanisms by which lncRNAs in AML bestow therapeutic resistance in this review. It follows naturally that more investigation is required to fully understand the immunological role of oncogenic lncRNAs. In addition, current research also needs to focus on how tumor suppressor lncRNAs can be programmed to target pathways that confer drug and apoptosis resistance in AML for the design of more efficient therapies. I support additional research into novel pathways and treatment targets to hasten the creation of more effective medications. Many novel strategies, such as oligonucleotide therapeutics, are being developed and expanding rapidly to selectively target and alter lncRNA expressions. While there are still many challenges to overcome before they become commonplace clinical treatments, they are exciting areas for future research due to their success in pre-clinical studies.

Outstanding questions

1. What are the fundamental processes by which lncRNAs control mitochondrial dynamics to evade therapeutic impact in AML patients?
2. How do lncRNAs regulate DNA damage repair pathways?
3. To comprehend the relationships between lncRNAs and other macromolecules, how can we effectively build novel genomic and proteomic approaches?
4. How can strategies be created to increase the expression of tumor suppressor lncRNAs?
5. How might the integration of human line xenograft models or patient tumor organoid models help us better understand how lncRNAs regulate AML?

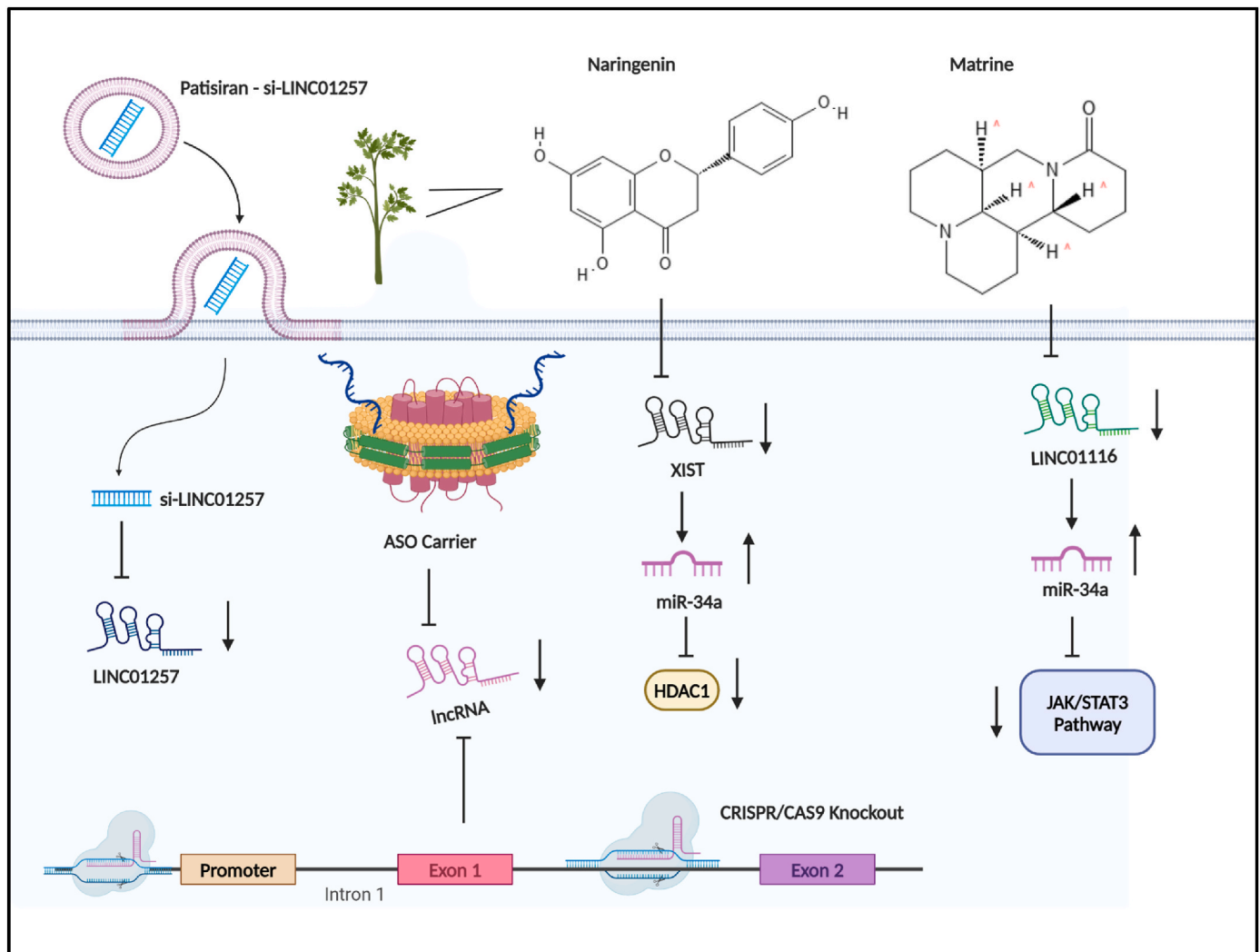


Fig. 4. Strategies to Target lncRNAs in AML

Figure highlighting strategies to target and inhibit the expression of oncogenic lncRNAs in AML. Currently, LNP-siRNA vehicles are the gold standard for targeting lncRNA expressions in various cancers and the LNP-si-LINC01257 vessel has been shown to inhibit the expression of the lncRNA LINC-1257 in AML. Plant metabolites like Nar and Matrine have proved to be clinically valuable as their heightened expression has been shown to decrease the expression of the lncRNAs XIST and LINC01116 leading to greater apoptosis levels in AML. Although CRISPR/CAS9 has been widely employed in several investigations to specifically knock down lncRNA expressions, it is yet unclear how this ground-breaking technology will be applied to actual clinical settings. Potential avenues for targeting lncRNAs in the future include the synthesis of ASO therapies and the use of morpholino oligomers to sterically obstruct lncRNA-protein complex interactions.

6. How can we enhance the systemic release of oligonucleotide therapies to target AML cells through the development of novel delivery systems?
7. How can we create delivery systems based on the CRISPR/CAS9 technology or morpholino oligomers to specifically knock down or block the interactions between oncogenic lncRNAs and proteins?
8. What plans of action can we build to reduce or eliminate the side effects of the new nucleic acid-based medicines and the present chemotherapeutic treatments?
9. How can we include more *in-vivo* modeling to better comprehend the mechanistic actions of oncogenic lncRNAs to create more potent treatments?
10. What methods do natural compounds use to specifically target and suppress the production of oncogenic lncRNA in AML? How may we apply that knowledge to create cutting-edge treatment plans that improve patient outcomes?

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data and material

Data sharing does not apply to this article as no new data was created or analyzed in this study.

Competing interests

There are no competing interests that I declare.

Funding

Not Applicable.

CRedit authorship contribution statement

Siddhant Sharma: Writing – review & editing, Writing – original draft, Visualization, Validation, Investigation, Formal analysis, Conceptualization.

Declaration of competing interest

I (Siddhant Sharma) declare that I have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgments

I would like to thank Aicha Asma Houfani and Dr. Leonard. J. Foster for their thoughtful discussions and support in developing the manuscript.

References

- [1] R. Elsaid, F. Soares-da-Silva, M. Peixoto, D. Amiri, N. Mackowski, P. Pereira, et al., Hematopoiesis: a Layered Organization across Chordate species, *Front. Cell Dev. Biol.* 8 (2020) 606642.
- [2] A.A. Bhat, S.N. Younes, S.S. Raza, L. Zarif, S. Nisar, I. Ahmed, et al., Role of non-coding RNA networks in leukemia progression, metastasis and drug resistance, *Mol. Cancer* 19 (2020) 57.
- [3] S. Saleem, J. Amin, M. Sharif, M.A. Anjum, M. Iqbal, S.-H. Wang, A deep network designed for segmentation and classification of leukemia using fusion of the transfer learning models, *Complex & Intelligent Systems* 8 (2022) 3105–3120.
- [4] P.K. Das, D.V. A. S. Meher, R. Panda, A. Abraham, A systematic review on recent advancements in deep and machine learning based Detection and classification of acute Lymphoblastic leukemia, *IEEE Access* 10 (2022) 81741–81763.
- [5] J. Zhang, Y. Gu, B. Chen, Mechanisms of drug resistance in acute myeloid leukemia, *OncoTargets Ther.* 12 (2019) 1937–1945.
- [6] P. Modarres, F. Mohamadi Farsani, A.A. Nekouie, S. Vallian, Meta-analysis of gene signatures and key pathways indicates suppression of JNK pathway as a regulator of chemo-resistance in AML, *Sci. Rep.* 11 (2021) 12485.
- [7] D.A. Arber, A. Orazi, R. Hasserjian, J. Thiele, M.J. Borowitz, M.M. Le Beau, et al., The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia, *Blood* 127 (2016) 2391–2405.
- [8] Leukemia - cancer stat facts. In: SEER [Internet]. [cited 13 Sep 2022]. Available: <https://seer.cancer.gov/statfacts/html/leuks.html>.
- [9] M. Luppi, F. Fabbiano, G. Visani, G. Martinelli, A. Venditti, Novel agents for acute myeloid leukemia, *Cancers* 10 (2018), <https://doi.org/10.3390/cancers10110429>.
- [10] C.A. Lachowicz, P.K. Reville, H. Kantarjian, E. Jabbour, G. Borthakur, N. Daver, et al., Venetoclax combined with induction chemotherapy in patients with newly diagnosed acute myeloid leukaemia: a post-hoc, propensity score-matched, cohort study, *Lancet Haematol* 9 (2022) e350–e360.
- [11] F. Guijarro, A. Bataller, M. Diaz-Beyá, A. Garrido, C. Coll-Ferrà, S. Vives, et al., Long-term outcomes in patients with relapsed/refractory acute myeloid leukemia and other high-risk myeloid malignancies after undergoing sequential conditioning regimen based on IDA-FLAG and high-dose melphalan, *Bone Marrow Transplant.* 57 (2022) 1304–1312.
- [12] G.R. Fajardo-Orduña, E. Ledesma-Martínez, I. Aguiñiga-Sánchez, M. de L. Mora-García, B. Weiss-Steider, E. Santiago-Osorio, Inhibitors of chemoresistance pathways in Combination with Ara-C to overcome multidrug resistance in AML. A Mini review, *Int. J. Mol. Sci.* 22 (2021), <https://doi.org/10.3390/ijms22094955>.
- [13] L. Cuesta-Casanovas, J. Delgado-Martínez, J.M. Cornet-Masana, J.M. Carbó, L. Clément-Demange, R.M. Riuseno, Lysosome-mediated chemoresistance in acute myeloid leukemia, *Cancer Drug Resist* 5 (2022) 233–244.
- [14] M. Ashrafzadeh, M.H. Gholami, S. Mirzaei, A. Zabolian, A. Haddadi, M. V. Farahani, et al., Dual relationship between long non-coding RNAs and STAT3 signaling in different cancers: new insight to proliferation and metastasis, *Life Sci.* 270 (2021) 119006.
- [15] M. Izadirad, L. Jafari, A.R. James, J.P. Unfried, Z.-X. Wu, Z.-S. Chen, Long noncoding RNAs have pivotal roles in chemoresistance of acute myeloid leukemia, *Drug Discov. Today* 26 (2021) 1735–1743.
- [16] C. Dieter, E.D. Lourenco, N.E. Lemos, Association of long non-coding RNA and leukemia: a systematic review, *Gene* 735 (2020) 144405.
- [17] F.-X. Huang, H.-J. Chen, F.-X. Zheng, Z.-Y. Gao, P.-F. Sun, Q. Peng, et al., LncRNA BLACAT1 is involved in chemoresistance of non-small cell lung cancer cells by regulating autophagy, *Int. J. Oncol.* 54 (2019) 339–347.
- [18] S. Fang, H. Gao, Y. Tong, J. Yang, R. Tang, Y. Niu, et al., Long noncoding RNA-HOTAIR affects chemoresistance by regulating HOXA1 methylation in small cell lung cancer cells, *Lab. Invest.* 96 (2016) 60–68.
- [19] Z. Fang, W. Chen, Z. Yuan, X. Liu, H. Jiang, LncRNA-MALAT1 contributes to the cisplatin-resistance of lung cancer by upregulating MRP1 and MDR1 via STAT3 activation, *Biomed. Pharmacother.* 101 (2018) 536–542.
- [20] Y. Cai, Z.-Y. Dong, J.-Y. Wang, LncRNA NNT-AS1 is a major mediator of cisplatin chemoresistance in non-small cell lung cancer through MAPK/Slug pathway, *Eur. Rev. Med. Pharmacol. Sci.* 22 (2018) 4879–4887.
- [21] L.-J. Tian, Y.-P. Wu, D. Wang, Z.-H. Zhou, S.-B. Xue, D.-Y. Zhang, et al., Upregulation of long noncoding RNA (lncRNA) X-Inactive specific transcript (XIST) is associated with cisplatin resistance in non-small cell lung cancer (NSCLC) by downregulating MicroRNA-144-3p, *Med Sci Monit* 25 (2019) 8095–8104.
- [22] Q. Chen, H. Shen, X. Zhu, Y. Liu, H. Yang, H. Chen, et al., A nuclear lncRNA Linc00839 as a Myc target to promote breast cancer chemoresistance via PI3K/AKT signaling pathway, *Cancer Sci.* 111 (2020) 3279–3291.
- [23] Q.-N. Zhu, G. Wang, Y. Guo, Y. Peng, R. Zhang, J.-L. Deng, et al., LncRNA H19 is a major mediator of doxorubicin chemoresistance in breast cancer cells through a cullin4A-MDR1 pathway, *Oncotarget* 8 (2017) 91990–92003.
- [24] X. Si, R. Zang, E. Zhang, Y. Liu, X. Shi, E. Zhang, et al., LncRNA H19 confers chemoresistance in ER α -positive breast cancer through epigenetic silencing of the pro-apoptotic gene BIK, *Oncotarget* 7 (2016) 81452–81462.
- [25] Z.-H. Li, N.-S. Yu, Q. Deng, Y. Zhang, Y.-Y. Hu, G. Liu, et al., LncRNA SNHG7 mediates the chemoresistance and stemness of breast cancer by sponging miR-34a, *Front. Oncol.* 10 (2020) 592757.
- [26] X. Si, G. Zhang, M. Li, M. Yao, X. Shi, Z. Dong, et al., LncRNA DDX11-AS1 promotes chemoresistance through LIN28A-mediated ATG12 mRNA stabilization in breast cancer, *Pharmacology* 108 (2023) 61–73.
- [27] S. Zhu, J. Mao, X. Zhang, P. Wang, Y. Zhou, J. Tong, et al., CAF-derived exosomal lncRNA FAL1 promotes chemoresistance to oxaliplatin by regulating autophagy in colorectal cancer, *Dig. Liver Dis.* (2023), <https://doi.org/10.1016/j.dld.2023.06.010>.
- [28] X. Zhang, D. Ma, B. Xuan, D. Shi, J. He, M. Yu, et al., LncRNA CAC1nc promotes chemoresistance of colorectal cancer by modulating alternative splicing of RAD51, *Oncogene* 42 (2023) 1374–1391.
- [29] B. Duan, H. Zhang, Z. Zhu, X. Yan, Z. Ji, J. Li, LncRNA LINC01871 sponging miR-142-3p to modulate ZYG11B promotes the chemoresistance of colorectal cancer cells by inducing autophagy, *Anti Cancer Drugs* 34 (2023) 827–836.
- [30] M. Li, S. Sun, Z. Bian, S. Yao, M. Liu, X. You, et al., SNHG15 promotes chemoresistance and glycolysis in colorectal cancer, *Pathol. Res. Pract.* 246 (2023) 154480.
- [31] J. Xu, Y. Xu, G. Ye, J. Qiu, LncRNA-SNHG1 promotes paclitaxel resistance of gastric cancer cells through modulating the miR-216b-5p-hexokinase 2 axis, *J. Chemother.* 35 (2023) 527–538.
- [32] Y. Zhu, B. Zhou, X. Hu, S. Ying, Q. Zhou, W. Xu, et al., LncRNA LINC00942 promotes chemoresistance in gastric cancer by suppressing MS12 degradation to enhance c-Myc mRNA stability, *Clin. Transl. Med.* 12 (2022) e703.
- [33] W. Zhu, L. Tan, T. Ma, Z. Yin, J. Gao, Long noncoding RNA SNHG8 promotes chemoresistance in gastric cancer via binding with hnRNP1 and stabilizing TROY expression, *Dig. Liver Dis.* 54 (2022) 1573–1582.
- [34] Y. Luo, S. Zheng, Q. Wu, J. Wu, R. Zhou, C. Wang, et al., Long noncoding RNA (lncRNA) EIF3J-DT induces chemoresistance of gastric cancer via autophagy activation, *Autophagy* 17 (2021) 4083–4101.
- [35] T.-L. Mao, M.-H. Fan, N. Dlamini, C.-L. Liu, LncRNA MALAT1 facilitates ovarian cancer progression through promoting chemoresistance and invasiveness in the tumor microenvironment, *Int. J. Mol. Sci.* 22 (2021), <https://doi.org/10.3390/ijms221910201>.
- [36] Q. Xu, Y.-B. Lin, L. Li, J. Liu, LncRNA TLR8-AS1 promotes metastasis and chemoresistance of ovarian cancer through enhancing TLR8 mRNA stability, *Biochem. Biophys. Res. Commun.* 526 (2020) 857–864.
- [37] Y. Wu, T. Wang, L. Xia, M. Zhang, Correction to: LncRNA WDFY3-AS2 promotes cisplatin resistance and the cancer stem cell in ovarian cancer by regulating hsa-miR-139-5p/SDC4 axis, *Cancer Cell Int.* 23 (2023) 176.
- [38] J. Zheng, Y. Song, Z. Li, A. Tang, Y. Fei, W. He, The implication of lncRNA expression pattern and potential function of lncRNA RP4-576H24.2 in acute myeloid leukemia, *Cancer Med.* 8 (2019) 7143–7160.
- [39] Y. Yuan, Q. Wang, S.L. Ma, L.Q. Xu, M.Y. Liu, B. Han, et al., lncRNA PCAT-1 interacting with FZD6 contributes to the malignancy of acute myeloid leukemia cells through activating Wnt/ β -catenin signaling pathway, *Am J Transl Res* 11 (2019) 7104–7114.
- [40] W. Zhou, S. Xu, T. Deng, R. Zhou, C. Wang, LncRNA USP30-AS1 promotes the survival of acute myeloid leukemia cells by cis-regulating USP30 and ANKRD13A, *Hum. Cell* 35 (2022) 360–378.
- [41] L. Chen, X. Fan, J. Zhu, X. Chen, Y. Liu, H. Zhou, LncRNA MAGI2-AS3 inhibits the self-renewal of leukaemic stem cells by promoting TET2-dependent DNA demethylation of the LRIG1 promoter in acute myeloid leukaemia, *RNA Biol.* 17 (2020) 784–793.
- [42] A. Kirtonia, M. Ashrafzadeh, A. Zarrabi, K. Hushmandi, A. Zabolian, A. K. Bejandi, et al., Long noncoding RNAs: a novel insight in the leukemogenesis and drug resistance in acute myeloid leukemia, *J. Cell. Physiol.* 237 (2022) 450–465.
- [43] H. Weng, H. Huang, H. Wu, X. Qin, B.S. Zhao, L. Dong, et al., METTL14 inhibits hematopoietic stem/progenitor differentiation and promotes leukemogenesis via mRNA m6A modification, *Cell Stem Cell* 22 (2018) 191–205.e9.

- [44] D. Cancilla, M.P. Rettig, J.F. DiPersio, Targeting CXCR4 in AML and ALL, *Front. Oncol.* 10 (2020) 1672.
- [45] Y. Xia, Q. Hong, X. Chen, H. Ye, L. Fang, A. Zhou, et al., APC2 and CYP1B1 methylation changes in the bone marrow of acute myeloid leukemia patients during chemotherapy, *Exp. Ther. Med.* 12 (2016) 3047–3052.
- [46] F. Nagai, Y. Hiyoshi, K. Sugimachi, H.-O. Tamura, Cytochrome P450 (CYP) expression in human myeloblastic and lymphoid cell lines, *Biol. Pharm. Bull.* 25 (2002) 383–385.
- [47] J. Li, Z. Li, X. Bai, X. Chen, M. Wang, Y. Wu, et al., LncRNA UCA1 promotes the progression of AML by upregulating the expression of CXCR4 and CYP1B1 by affecting the stability of METTL4, *J. Oncol.* 2022 (2022) 2756986.
- [48] Y. Xiao, T. Xiao, W. Ou, Z. Wu, J. Wu, J. Tang, et al., LncRNA SNHG16 as a potential biomarker and therapeutic target in human cancers, *Biomark. Res.* 8 (2020) 41.
- [49] M. Shi, R. Yang, J. Lin, Q.I. Wei, L. Chen, W. Gong, et al., LncRNA-SNHG16 promotes proliferation and migration of acute myeloid leukemia cells via PTEN/PI3K/AKT axis through suppressing CELF2 protein, *J. Biosci.* 46 (2021), <https://doi.org/10.1007/s12038-020-00127-1>.
- [50] M.-F. Zhuang, L.-J. Li, J.-B. Ma, LncRNA HOTTIP promotes proliferation and cell cycle progression of acute myeloid leukemia cells, *Eur. Rev. Med. Pharmacol. Sci.* 23 (2019) 2908–2915.
- [51] Y.-J. Tian, Y.-H. Wang, A.-J. Xiao, P.-L. Li, J. Guo, T.-J. Wang, et al., Long noncoding RNA SBF2-AS1 act as a ceRNA to modulate cell proliferation via binding with miR-188-5p in acute myeloid leukemia, *Artif. Cells, Nanomed. Biotechnol.* 47 (2019) 1730–1737.
- [52] J. Shi, R. Dai, Y. Chen, H. Guo, Y. Han, Y. Zhang, LncRNA LINC1 regulates acute myeloid leukemia progression via HNF4 α /AMPK/WNT5A signaling pathway, *Hematol. Oncol.* 37 (2019) 474–482.
- [53] A.J. Pulikkottil, S. Bamezai, T. Ammer, F. Mohr, K. Feder, N.M. Vegi, et al., TET3 promotes AML growth and epigenetically regulates glucose metabolism and leukemic stem cell associated pathways, *Leukemia* 36 (2022) 416–425.
- [54] L.-Y. Sun, X.-J. Li, Y.-M. Sun, W. Huang, K. Fang, C. Han, et al., LncRNA ANRIL regulates AML development through modulating the glucose metabolism pathway of AdipoR1/AMPK/SIRT1, *Mol. Cancer* 17 (2018) 127.
- [55] M. Levin, M. Stark, Y. Ofra, Y.G. Assaraf, Deciphering molecular mechanisms underlying chemoresistance in relapsed AML patients: towards precision medicine overcoming drug resistance, *Cancer Cell Int.* 21 (2021) 53.
- [56] X. Wang, H. Zhang, X. Chen, Drug resistance and combating drug resistance in cancer, *Cancer Drug Resist* 2 (2019) 141–160.
- [57] Y. Jo, N. Choi, K. Kim, H.-J. Koo, J. Choi, H.N. Kim, Chemoresistance of cancer cells: requirements of tumor microenvironment-mimicking in vitro models in anti-cancer drug development, *Theranostics* 8 (2018) 5259–5275.
- [58] H. Xiao, Y. Zheng, L. Ma, L. Tian, Q. Sun, Clinically-relevant ABC transporter for anti-cancer drug resistance, *Front. Pharmacol.* 12 (2021) 648407.
- [59] E.L. Giddings, D.P. Champagne, M.-H. Wu, J.M. Laffin, T.M. Thornton, F. Valenca-Pereira, et al., Mitochondrial ATP efflux ABC transporter-mediated drug efflux in cancer chemoresistance, *Nat. Commun.* 12 (2021) 2804.
- [60] X. Sui, R. Chen, Z. Wang, Z. Huang, N. Kong, M. Zhang, et al., Autophagy and chemotherapy resistance: a promising therapeutic target for cancer treatment, *Cell Death Dis.* 4 (2013) e838.
- [61] X. Li, Y. Zhou, Y. Li, L. Yang, Y. Ma, X. Peng, et al., Autophagy: a novel mechanism of chemoresistance in cancers, *Biomed. Pharmacother.* 119 (2019) 109415.
- [62] J. Hraběta, M. Belhajová, H. Subrtová, M.A. Merlos Rodrigo, Z. Heger, T. Eckschlager, Drug sequestration in lysosomes as one of the mechanisms of chemoresistance of cancer cells and the possibilities of its inhibition, *Int. J. Mol. Sci.* 21 (2020), <https://doi.org/10.3390/ijms21124392>.
- [63] R. Halaby, Influence of lysosomal sequestration on multidrug resistance in cancer cells, *Cancer Drug Resist* 2 (2019) 31–42.
- [64] W. Yang, W. Xiao, Z. Cai, S. Jin, T. Li, miR-1269b drives cisplatin resistance of human non-small cell lung cancer via modulating the PTEN/PI3K/AKT signaling pathway, *Oncotargets Ther.* 13 (2020) 109–118.
- [65] T. Yagi, H. Iinuma, T. Hayama, K. Matsuda, K. Nozawa, M. Tsukamoto, et al., Plasma exosomal microRNA-125b as a monitoring biomarker of resistance to mFOLFOX6-based chemotherapy in advanced and recurrent colorectal cancer patients, *Mol Clin Oncol* 11 (2019) 416–424.
- [66] T. Zhu, Z. Hu, Z. Wang, H. Ding, R. Li, J. Wang, et al., microRNA-301b-3p from mesenchymal stem cells-derived extracellular vesicles inhibits TXNIP to promote multidrug resistance of gastric cancer cells, *Cell Biol. Toxicol.* 39 (2023) 1923–1937.
- [67] F. Liu, Y. Wang, Y. Cao, Z. Wu, D. Ma, J. Cai, et al., Transcription factor B-MYB activates lncRNA CCAT1 and upregulates SOCS3 to promote chemoresistance in colorectal cancer, *Chem. Biol. Interact.* 374 (2023) 110412.
- [68] P. Zhao, J. Sun, X. Huang, X. Zhang, X. Liu, R. Liu, et al., Targeting the KLF5-EphA2 axis can restrain cancer stemness and overcome chemoresistance in basal-like breast cancer, *Int. J. Biol. Sci.* 19 (2023) 1861–1874.
- [69] L. Li, N. Wang, Y. Xiong, G. Guo, M. Zhu, Y. Gu, Transcription factor FOSL1 enhances drug resistance of breast cancer through DUSP7-mediated dephosphorylation of PEA15, *Mol. Cancer Res.* 20 (2022) 515–526.
- [70] J. Verigos, P. Karakaidos, D. Kordias, A. Papoudou-Bai, Z. Evangelou, H. V. Harissis, et al., The histone demethylase LSD1/kdm1a mediates chemoresistance in breast cancer via regulation of a stem cell program, *Cancers* 11 (2019), <https://doi.org/10.3390/cancers11101585>.
- [71] Y. Zhang, J.-G. Qiu, X.-Y. Jia, Y. Ke, M.-K. Zhang, D. Stieg, et al., METTL3-mediated N6-methyladenosine modification and HDAC5/YY1 promote IFFO1 downregulation in tumor development and chemo-resistance, *Cancer Lett.* 553 (2023) 215971.
- [72] X. Xu, X. Zhou, Z. Chen, C. Gao, L. Zhao, Y. Cui, Silencing of lncRNA XIST inhibits non-small cell lung cancer growth and promotes chemosensitivity to cisplatin, *Aging* 12 (2020) 4711–4726.
- [73] L. Cao, J. Chen, B. Ou, C. Liu, Y. Zou, Q. Chen, GAS5 knockdown reduces the chemo-sensitivity of non-small cell lung cancer (NSCLC) cell to cisplatin (DDP) through regulating miR-21/PTEN axis, *Biomed. Pharmacother.* 93 (2017) 570–579.
- [74] S. Liu, H. Lei, F. Luo, Y. Li, L. Xie, The effect of lncRNA HOTAIR on chemoresistance of ovarian cancer through regulation of HOXA7, *Biol. Chem.* 399 (2018) 485–497.
- [75] K. Huang, X. Chen, Z. Geng, X. Xiong, Y. Cong, X. Pan, et al., LncRNA SLC25A21-AS1 increases the chemosensitivity and inhibits the progression of ovarian cancer by upregulating the expression of KCNK4, *Funct. Integr. Genomics* 23 (2023) 110.
- [76] X. Huang, W. Zhang, F. Pu, Z. Zhang, LncRNA MEG3 promotes chemosensitivity of osteosarcoma by regulating antitumor immunity via miR-21-5p/p53 pathway and autophagy, *Genes Dis* 10 (2023) 531–541.
- [77] A.M. Lee, A. Ferdjallah, E. Moore, D.C. Kim, A. Nath, E. Greengard, et al., Long non-coding RNA ANRIL as a potential biomarker of chemosensitivity and clinical outcomes in osteosarcoma, *Int. J. Mol. Sci.* 22 (2021), <https://doi.org/10.3390/ijms222011168>.
- [78] C. Guo, M. Gong, Z. Li, Knockdown of lncRNA MCM3AP-AS1 attenuates chemoresistance of burkitt lymphoma to doxorubicin treatment via targeting the miR-15a/EIF4E Axis, *Cancer Manag. Res.* 12 (2020) 5845–5855.
- [79] L. Zhang, S. Zhou, T. Zhou, X. Li, J. Tang, Targeting the lncRNA DUXAP8/miR-29a/PIK3CA network restores doxorubicin chemosensitivity via PI3K-AKT-mTOR signaling and synergizes with inotuzumab ozogamicin in chemotherapy-resistant B-cell acute lymphoblastic leukemia, *Front. Oncol.* 12 (2022) 773601.
- [80] H. Dai, J. Wang, Z. Huang, H. Zhang, X. Wang, Q. Li, et al., LncRNA OIP5-AS1 promotes the autophagy-related imatinib resistance in chronic myeloid leukemia cells by regulating miR-30e-5p/ATG12 Axis, *Technol. Cancer Res. Treat.* 20 (2021) 15330338211052150.
- [81] D. Singh, Y.G. Assaraf, R.N. Gacche, Long non-coding RNA mediated drug resistance in breast cancer, *Drug Resist Updat* 63 (2022) 100851.
- [82] B. Chen, M.P. Dragomir, C. Yang, Q. Li, D. Horst, G.A. Calin, Targeting non-coding RNAs to overcome cancer therapy resistance, *Signal Transduct Target Ther* 7 (2022) 121.
- [83] Y. Tao, J. Zhang, L. Chen, X. Liu, M. Yao, H. Zhang, LncRNA CD27-AS1 promotes acute myeloid leukemia progression through the miR-224-5p/PBX3 signaling circuit, *Cell Death Dis.* 12 (2021) 510.
- [84] C.-Z. Wang, B.-B. Ma, Z.-J. Xu, J.-D. Zhou, T.-J. Zhang, Q. Chen, et al., Reduced expression of lncRNA DLEU7-AS1 is a novel favorable prognostic factor in acute myeloid leukemia, *Biosci. Rep.* 42 (2022), <https://doi.org/10.1042/BSR20212078>.
- [85] L. Peng, Y. Zhang, H. Xin, LncRNA SNHG3 facilitates acute myeloid leukemia cell growth via the regulation of miR-758-3p/SRGN axis, *J. Cell. Biochem.* 121 (2020) 1023–1031.
- [86] C. Li, Q. Gao, M. Wang, H. Xin, LncRNA SNHG1 contributes to the regulation of acute myeloid leukemia cell growth by modulating miR-489-3p/SOX12/Wnt/ β -catenin signaling, *J. Cell. Physiol.* 236 (2021) 653–663.
- [87] X. Ma, W. Zhang, M. Zhao, S. Li, W. Jin, K. Wang, Oncogenic role of lncRNA CRNDE in acute promyelocytic leukemia and NPM1-mutant acute myeloid leukemia, *Cell Death Discov.* 6 (2020) 121.
- [88] H. Zhou, X. Jia, F. Yang, P. Shi, Long noncoding RNA SATB1-AS1 contributes to the chemotherapy resistance through the microRNA-580/2'-5'-oligoadenylate synthetase 2 axis in acute myeloid leukemia, *Bioengineered* 12 (2021) 6403–6417.
- [89] N. Mencia, E. Selga, I. Rico, M.C. de Almagro, X. Villalobos, S. Ramirez, et al., Overexpression of S100A4 in human cancer cell lines resistant to methotrexate, *BMC Cancer* 10 (2010) 250.
- [90] X. Dong, Z. Fang, M. Yu, L. Zhang, R. Xiao, X. Li, et al., Knockdown of long noncoding RNA HOXA-AS2 suppresses chemoresistance of acute myeloid leukemia via the miR-520c-3p/S100A4 Axis, *Cell. Physiol. Biochem.* 51 (2018) 886–896.
- [91] G. Arun, D. Aggarwal, D.L. Spector, MALAT1 long non-coding RNA: functional implications, *Noncoding RNA* 6 (2020), <https://doi.org/10.3390/nrna6020022>.
- [92] H. YiRen, Y. YingCong, Y. Sunwu, L. Keqin, T. Xiaochun, C. Senrui, et al., Long noncoding RNA MALAT1 regulates autophagy associated chemoresistance via miR-23b-3p sequestration in gastric cancer, *Mol. Cancer* 16 (2017) 174.
- [93] Z. Zhang, M. Li, Z. Zhang, lncRNA MALAT1 modulates oxaliplatin resistance of gastric cancer via sponging miR-22-3p, *Oncotargets Ther.* 13 (2020) 1343–1354.
- [94] Z. Xi, J. Si, J. Nan, LncRNA MALAT1 potentiates autophagy-associated cisplatin resistance by regulating the microRNA-30b/autophagy-related gene 5 axis in gastric cancer, *Int. J. Oncol.* 54 (2019) 239–248.
- [95] T. Yang, H. Li, T. Chen, H. Ren, P. Shi, M. Chen, LncRNA MALAT1 depressed chemo-sensitivity of NSCLC cells through directly functioning on miR-197-3p/p120 catenin Axis, *Mol Cells* 42 (2019) 270–283.
- [96] P. Liu, X. Li, Y. Cui, J. Chen, C. Li, Q. Li, et al., LncRNA-MALAT1 mediates cisplatin resistance via miR-101-3p/VEGF-C pathway in bladder cancer, *Acta Biochim. Biophys. Sin.* 51 (2019) 1148–1157.
- [97] X. Yue, W.-Y. Wu, M. Dong, M. Guo, LncRNA MALAT1 promotes breast cancer progression and doxorubicin resistance via regulating miR-570-3p, *Biomed. J.* 44 (2021) S296–S304.

- [98] C. Shi, S. Ren, X. Zhao, Q. Li, lncRNA MALAT1 regulates the resistance of breast cancer cells to paclitaxel via the miR-497-5p/SHOC2 axis, *Pharmacogenomics* 23 (2022) 973–985.
- [99] N. Hu, L. Chen, C. Wang, H. Zhao, MALAT1 knockdown inhibits proliferation and enhances cytarabine chemosensitivity by upregulating miR-96 in acute myeloid leukemia cells, *Biomed. Pharmacother.* 112 (2019) 108720.
- [100] D.H. Lee, G.W. Kim, J. Yoo, S.W. Lee, Y.H. Jeon, S.Y. Kim, et al., Histone demethylase KDM4C controls tumorigenesis of glioblastoma by epigenetically regulating p53 and c-Myc, *Cell Death Dis.* 12 (2021) 89.
- [101] C.-Y. Lin, B.-J. Wang, B.-C. Chen, J.-C. Tseng, S.S. Jiang, K.K. Tsai, et al., Histone demethylase KDM4C stimulates the proliferation of prostate cancer cells via activation of AKT and c-myc, *Cancers* 11 (2019), <https://doi.org/10.3390/cancers11111785>.
- [102] C.-Y. Lin, B.-J. Wang, Y.-K. Fu, C. Huo, Y.-P. Wang, B.-C. Chen, et al., Inhibition of KDM4C/c-Myc/LDHA signalling axis suppresses prostate cancer metastasis via interference of glycolytic metabolism, *Clin. Transl. Med.* 12 (2023) e764.
- [103] X. Jie, W.P. Fong, R. Zhou, Y. Zhao, Y. Zhao, R. Meng, et al., USP9X-mediated KDM4C deubiquitination promotes lung cancer radioresistance by epigenetically inducing TGF- β 2 transcription, *Cell Death Differ.* 28 (2021) 2095–2111.
- [104] G. Yu, H. Zhou, W. Yao, L. Meng, B. Lang, lncRNA TUG1 promotes cisplatin resistance by regulating CCND2 via epigenetically silencing miR-194-5p in bladder cancer, *Mol. Ther. Nucleic Acids* 16 (2019) 257–271.
- [105] H. Hou, R. Yu, H. Zhao, H. Yang, Y. Hu, Y. Hu, et al., lncRNA OTUD6B-AS1 induces cisplatin resistance in cervical cancer cells through up-regulating cyclin D2 via miR-206, *Front. Oncol.* 11 (2021) 777220.
- [106] L. Xue, C. Li, J. Ren, Y. Wang, KDM4C contributes to cytarabine resistance in acute myeloid leukemia via regulating the miR-328-3p/CCND2 axis through MALAT1, *Ther. Adv. Chronic Dis* 12 (2021) 2040622321997259.
- [107] H.-L. Zhang, P. Wang, M.-Z. Lu, S.-D. Zhang, L. Zheng, c-Myc maintains the self-renewal and chemoresistance properties of colon cancer stem cells, *Oncol. Lett.* 17 (2019) 4487–4493.
- [108] W. Zhan, X. Liao, Y. Wang, L. Li, J. Li, Z. Chen, et al., circCTIC1 promotes the self-renewal of colon TICs through BPTF-dependent c-Myc expression, *Carcinogenesis* 40 (2019) 560–568.
- [109] Y. Di, X. Jing, K. Hu, X. Wen, L. Ye, X. Zhang, et al., The c-MYC-WDR43 signalling axis promotes chemoresistance and tumour growth in colorectal cancer by inhibiting p53 activity, *Drug Resist Updat* 66 (2023) 100909.
- [110] Y. Zeng, H. Jiang, X. Zhang, J. Xu, X. Wu, Q. Xu, et al., Canagliflozin reduces chemoresistance in hepatocellular carcinoma through PKM2-c-Myc complex-mediated glutamine starvation, *Free Radic. Biol. Med.* 208 (2023) 571–586.
- [111] X. Li, Y. Zhang, X. Wang, F. Lin, X. Cheng, Z. Wang, et al., Long non-coding RNA CTSLP8 mediates ovarian cancer progression and chemotherapy resistance by modulating cellular glycolysis and regulating c-Myc expression through PKM2, *Cell Biol. Toxicol.* 38 (2022) 1027–1045.
- [112] Y. Wang, M. Zhang, Z. Wang, W. Guo, D. Yang, MYC-binding lncRNA EPIC1 promotes AKT-mTORC1 signaling and rapamycin resistance in breast and ovarian cancer, *Mol. Carcinog.* 59 (2020) 1188–1198.
- [113] J. Yang, M. Qi, X. Fei, X. Wang, K. Wang, Long non-coding RNA XIST: a novel oncogene in multiple cancers, *Mol Med* 27 (2021) 159.
- [114] C. Wang, L. Li, M. Li, W. Wang, Y. Liu, S. Wang, Silencing long non-coding RNA XIST suppresses drug resistance in acute myeloid leukemia through down-regulation of MYC by elevating microRNA-29a expression, *Mol Med* 26 (2020) 114.
- [115] Y.-C. Li, Y. Wu, G. Chen, L.-Z. Zhu, X. Luo, Q.-Q. Nie, et al., Tetraspanins predict the prognosis and characterize the tumor immune microenvironment of glioblastoma, *Sci. Rep.* 13 (2023) 13317.
- [116] Y. Garcia-Mayea, C. Mir, L. Carballo, A. Sánchez-García, M. Bataller, M. E. Lleonart, TSPAN1, a novel tetraspanin member highly involved in carcinogenesis and chemoresistance, *Biochim. Biophys. Acta Rev. Canc* 1877 (2022) 188674.
- [117] X. Lu, L. An, G. Fan, L. Zang, W. Huang, J. Li, et al., EGFR signaling promotes nuclear translocation of plasma membrane protein TSPAN8 to enhance tumor progression via STAT3-mediated transcription, *Cell Res.* 32 (2022) 359–374.
- [118] J. Wang, Y. Zhou, D. Li, X. Sun, Y. Deng, Q. Zhao, TSPAN31 is a critical regulator on transduction of survival and apoptotic signals in hepatocellular carcinoma cells, *FEBS Lett.* 591 (2017) 2905–2918.
- [119] Y. Qi, W. Qi, S. Liu, L. Sun, A. Ding, G. Yu, et al., TSPAN9 suppresses the chemosensitivity of gastric cancer to 5-fluorouracil by promoting autophagy, *Cancer Cell Int.* 20 (2020) 4.
- [120] H. Sun, Y. Sun, Q. Chen, Z. Xu, lncRNA KCNQ1OT1 contributes to the progression and chemoresistance in acute myeloid leukemia by modulating Tspan3 through suppressing miR-193a-3p, *Life Sci.* 241 (2020) 117161.
- [121] Y. Yu, D. Kou, B. Liu, Y. Huang, S. Li, Y. Qi, et al., lncRNA MEG3 contributes to drug resistance in acute myeloid leukemia by positively regulating ALG9 through sponging miR-155, *Int J Lab Hematol.* 42 (2020) 464–472.
- [122] C. Hu, S. Li, X. Fu, X. Zhao, J. Peng, lncRNA NR2F1-AS1 was involved in azacitidine resistance of THP-1 cells by targeting IGF1 with miR-483-3p, *Cytokine* 162 (2023) 156105.
- [123] L. Sang, X. Wu, T. Yan, D. Naren, X. Liu, X. Zheng, et al., The m6A RNA methyltransferase METTL3/METTL14 promotes leukemogenesis through the mdm2/p53 pathway in acute myeloid leukemia, *J. Cancer* 13 (2022) 1019–1030.
- [124] I. Nepstad, K.J. Hatfield, I.S. Grønningseter, H. Reikvam, The PI3K-Akt-mTOR signaling pathway in human acute myeloid leukemia (AML) cells, *Int. J. Mol. Sci.* 21 (2020), <https://doi.org/10.3390/ijms21082907>.
- [125] H. Jiang, J. Tang, L. Qiu, Z. Zhang, S. Shi, L. Xue, et al., Semaphorin 4D is a potential biomarker in pediatric leukemia and promotes leukemogenesis by activating PI3K/AKT and ERK signaling pathways, *Oncol. Rep.* 45 (2021), <https://doi.org/10.3892/or.2021.7952>.
- [126] P. Han, J.-W. Li, B.-M. Zhang, J.-C. Lv, Y.-M. Li, X.-Y. Gu, et al., The lncRNA CRNDE promotes colorectal cancer cell proliferation and chemoresistance via miR-181a-5p-mediated regulation of Wnt/ β -catenin signaling, *Mol. Cancer* 16 (2017) 9.
- [127] S. Xing, Y. Qu, C. Li, A. Huang, S. Tong, C. Wu, et al., Deregulation of lncRNA-AC078883.3 and microRNA-19a is involved in the development of chemoresistance to cisplatin via modulating signaling pathway of PTEN/AKT, *J. Cell. Physiol.* 234 (2019) 22657–22665.
- [128] W. Zhang, Q. Wu, Y. Liu, X. Wang, C. Ma, W. Zhu, lncRNA HOTAIR promotes chemoresistance by facilitating epithelial to mesenchymal transition through miR-29b/PTEN/PI3K signaling in cervical cancer, *Cells Tissues Organs* 211 (2022) 16–29.
- [129] M. Michel, L. Kaps, A. Maderer, P.R. Galle, M. Moehler, The role of p53 dysfunction in colorectal cancer and its implication for therapy, *Cancers* 13 (2021), <https://doi.org/10.3390/cancers13102296>.
- [130] R.S. Moussa, K.C. Park, Z. Kovacevic, D.R. Richardson, Ironing out the role of the cyclin-dependent kinase inhibitor, p21 in cancer: novel iron chelating agents to target p21 expression and activity, *Free Radic. Biol. Med.* 133 (2019) 276–294.
- [131] M.-L. Li, Y. Wang, Y.-N. Xu, Q.-Y. Lu, Overexpression of lncRNA-HOTAIR promotes chemoresistance in acute leukemia cells, *Int. J. Clin. Exp. Pathol.* 13 (2020) 3044–3051.
- [132] Y. Kang, S. Zhang, W. Cao, D. Wan, L. Sun, Knockdown of lncRNA CRNDE suppresses proliferation and P-glycoprotein-mediated multidrug resistance in acute myelocytic leukemia through the Wnt/ β -catenin pathway, *Biosci. Rep.* 40 (2020), <https://doi.org/10.1042/BSR20193450>.
- [133] Q. Yao, L. Zhang, Y. Wang, J. Liu, L. Yang, Y. Wang, lncRNA UCA1 elevates the resistance of human leukemia cells to daunorubicin by the PI3K/AKT pathway via sponging miR-613, *Biosci. Rep.* 41 (2021), <https://doi.org/10.1042/BSR20201389>.
- [134] Y. Yang, W. Dai, Y. Sun, Z. Zhao, Long non-coding RNA linc00239 promotes malignant behaviors and chemoresistance against doxorubicin partially via activation of the PI3K/Akt/mTOR pathway in acute myeloid leukaemia cells, *Oncol. Rep.* 41 (2019) 2311–2320.
- [135] L. Liang, W. Gu, M. Li, R. Gao, X. Zhang, C. Guo, et al., The long noncoding RNA HOTAIRM1 controlled by AML1 enhances glucocorticoid resistance by activating RHOA/ROCK1 pathway through suppressing ARHGAP18, *Cell Death Dis.* 12 (2021) 702.
- [136] A.C. Bester, J.D. Lee, A. Chavez, Y.-R. Lee, D. Nachmani, S. Vora, et al., An integrated genome-wide CRISPRa approach to functionalize lncRNAs in drug resistance, *Cell* 173 (2018) 649–664.e20.
- [137] N. Fusco, E. Sajjadi, K. Venetis, G. Gaudioso, G. Lopez, C. Corti, et al., PTEN alterations and their role in cancer management: are we making headway on precision medicine? *Genes* 11 (2020) <https://doi.org/10.3390/genes11070719>.
- [138] L. Shi, W. Zhu, Y. Huang, L. Zhuo, S. Wang, S. Chen, et al., Cancer-associated fibroblast-derived exosomal microRNA-20a suppresses the PTEN/PI3K-AKT pathway to promote the progression and chemoresistance of non-small cell lung cancer, *Clin. Transl. Med.* 12 (2022) e989.
- [139] F. Zheng, J. Zhong, K. Chen, Y. Shi, F. Wang, S. Wang, et al., PINK1-PTEN axis promotes metastasis and chemoresistance in ovarian cancer via non-canonical pathway, *J. Exp. Clin. Cancer Res.* 42 (2023) 295.
- [140] Y. Su, B. Wang, J. Huang, M. Huang, T. Lin, YTHDC1 positively regulates PTEN expression and plays a critical role in cisplatin resistance of bladder cancer, *Cell Prolif.* 56 (2023) e13404.
- [141] W. Zhou, S. Xu, X. Chen, C. Wang, HOTAIR suppresses PTEN via DNMT3b and confers drug resistance in acute myeloid leukemia, *Hematology.* 26 (2021) 170–178.
- [142] Y. Ning, N. Hui, B. Qing, Y. Zhuo, W. Sun, Y. Du, et al., ZCCHC10 suppresses lung cancer progression and cisplatin resistance by attenuating MDM2-mediated p53 ubiquitination and degradation, *Cell Death Dis.* 10 (2019) 414.
- [143] Z.-H. Ma, P.-D. Shi, B.-S. Wan, MiR-410-3p activates the NF- κ B pathway by targeting ZCCHC10 to promote migration, invasion and EMT of colorectal cancer, *Cytokine* 140 (2021) 155433.
- [144] H. Zhou, Q. Zhang, W. Huang, C. He, C. Zhou, J. Zhou, et al., Epigenetic silencing of ZCCHC10 by the lncRNA SNHG1 promotes progression and venetoclax resistance of acute myeloid leukemia, *Int. J. Oncol.* 62 (2023), <https://doi.org/10.3892/ijo.2023.5512>.
- [145] P. Bose, V. Gandhi, M. Konopleva, Pathways and mechanisms of venetoclax resistance, *Leuk. Lymphoma* 58 (2017) 1–17.
- [146] R. Duan, W. Du, W. Guo, EZH2: a novel target for cancer treatment, *J. Hematol. Oncol.* 13 (2020) 104.
- [147] A. Magi, G. Mattei, A. Mingrino, C. Caprioli, C. Ronchini, G. Frigè, et al., High-resolution Nanopore methylome-maps reveal random hyper-methylation at CpG-poor regions as driver of chemoresistance in leukemias, *Commun. Biol.* 6 (2023) 382.
- [148] N. Gull, M.R. Jones, P.-C. Peng, S.G. Coetzee, T.C. Silva, J.T. Plummer, et al., DNA methylation and transcriptomic features are preserved throughout disease recurrence and chemoresistance in high grade serous ovarian cancers, *J. Exp. Clin. Cancer Res.* 41 (2022) 232.
- [149] K.A. Gelato, W. Fischle, Role of histone modifications in defining chromatin structure and function, *Biol. Chem.* 389 (2008) 353–363.
- [150] Q. Li, W. Song, J. Wang, TUG1 confers Adriamycin resistance in acute myeloid leukemia by epigenetically suppressing miR-34a expression via EZH2, *Biomed. Pharmacother.* 109 (2019) 1793–1801.

- [151] C. Li, X. Liang, Y. Liu, lncRNA USP30-AS1 sponges miR-765 and modulates the progression of colon cancer, *World J. Surg. Oncol.* 20 (2022) 73.
- [152] R. Huang, P.-K. Zhou, DNA damage repair: historical perspectives, mechanistic pathways and clinical translation for targeted cancer therapy, *Signal Transduct Target Ther* 6 (2021) 254.
- [153] H. Zhang, Y. Hua, Z. Jiang, J. Yue, M. Shi, X. Zhen, et al., Cancer-associated fibroblast-promoted lncRNA DNMT3OS confers radioresistance by regulating DNA damage response in esophageal squamous cell carcinoma, *Clin. Cancer Res.* 25 (2019) 1989–2000.
- [154] A. García-Venzor, E.A. Mandujano-Tinoco, A. Ruiz-Silvestre, J.M. Sánchez, F. Lizarraga, C. Zampedri, et al., lncMat2B regulated by severe hypoxia induces cisplatin resistance by increasing DNA damage repair and tumor-initiating population in breast cancer cells, *Carcinogenesis* 41 (2020) 1485–1497.
- [155] L. Ren, X. Qing, J. Wei, H. Mo, Y. Liu, Y. Zhi, et al., The DDUP protein encoded by the DNA damage-induced CTBP1-DT lncRNA confers cisplatin resistance in ovarian cancer, *Cell Death Dis.* 14 (2023) 568.
- [156] D. Li, Z. Yu, T. Wang, Y. Li, X. Chen, L. Wu, The role of the novel lincRNA uc002jit.1 in NF- κ B-mediated DNA damage repair in acute myeloid leukemia cells, *Exp. Cell Res.* 391 (2020) 111985.
- [157] D. Li, Y. Luo, X. Chen, L. Zhang, T. Wang, Y. Zhuang, et al., NF- κ B and poly (ADP-ribose) polymerase 1 form a positive feedback loop that regulates DNA repair in acute myeloid leukemia cells, *Mol. Cancer Res.* 17 (2019) 761–772.
- [158] F. Wang, O.G. Gouttiau, L. Wang, A. Peng, PARP1 upregulation in recurrent oral cancer and treatment resistance, *Front. Cell Dev. Biol.* 9 (2021) 804962.
- [159] F. Quinónero, C. Mesas, J.A. Muñoz-Gómez, C. Jiménez-Luna, G. Perazzoli, J. Prados, et al., PARP1 inhibition by Olaparib reduces the lethality of pancreatic cancer cells and increases their sensitivity to Gemcitabine, *Biomed. Pharmacother.* 155 (2022) 113669.
- [160] W.-W. Zuo, C.-F. Zhao, Y. Li, H.-Y. Sun, G.-M. Ma, Y.-P. Liu, et al., High expression of PARP1 in tumor and stroma cells predicts different prognosis and platinum resistance in patients with advanced epithelial ovarian cancer, *Front. Oncol.* 12 (2022) 931445.
- [161] Z. Bian, J. Zhang, M. Li, Y. Feng, S. Yao, M. Song, et al., Long non-coding RNA LINC00152 promotes cell proliferation, metastasis, and confers 5-FU resistance in colorectal cancer by inhibiting miR-139-5p, *Oncogenesis* 6 (2017) 395.
- [162] S. Zhang, W. Liao, Q. Wu, X. Huang, Z. Pan, W. Chen, et al., LINC00152 upregulates ZEB1 expression and enhances epithelial-mesenchymal transition and oxaliplatin resistance in esophageal cancer by interacting with EZH2, *Cancer Cell Int.* 20 (2020) 569.
- [163] X.-L. Hu, J. Wang, W. He, P. Zhao, W.-Q. Wu, Down-regulation of lncRNA Linc00152 suppressed cell viability, invasion, migration, and epithelial to mesenchymal transition, and reversed chemo-resistance in breast cancer cells, *Eur. Rev. Med. Pharmacol. Sci.* 22 (2018) 3074–3084.
- [164] W. Wang, F. Wu, Z. Zhao, K.-Y. Wang, R.-Y. Huang, H.-Y. Wang, et al., Long noncoding RNA LINC00152 is a potential prognostic biomarker in patients with high-grade glioma, *CNS Neurosci. Ther.* 24 (2018) 957–966.
- [165] C. Cui, Y. Wang, W. Gong, H. He, H. Zhang, W. Shi, et al., Long non-coding RNA LINC00152 regulates self-renewal of leukemia stem cells and induces chemo-resistance in acute myeloid leukemia, *Front. Oncol.* 11 (2021) 694021.
- [166] J.N.S. Vargas, M. Hamasaki, T. Kawabata, R.J. Youle, T. Yoshimori, The mechanisms and roles of selective autophagy in mammals, *Nat. Rev. Mol. Cell Biol.* 24 (2023) 167–185.
- [167] Q. Wu, J. Ma, J. Wei, W. Meng, Y. Wang, M. Shi, lncRNA SNHG11 promotes gastric cancer progression by activating the wnt/ β -catenin pathway and oncogenic autophagy, *Mol. Ther.* 29 (2021) 1258–1278.
- [168] P.-P. Li, R.-G. Li, Y.-Q. Huang, J.-P. Lu, W.-J. Zhang, Z.-Y. Wang, lncRNA OTUD6B-AS1 promotes paclitaxel resistance in triple negative breast cancer by regulation of miR-26a-5p/MTDH pathway-mediated autophagy and genomic instability, *Aging* 13 (2021) 24171–24191.
- [169] L. Sun, X. Wang, L. Chen, Z. Gao, S. Xu, C. Hu, et al., CPT1A mediates chemoresistance in human hypopharyngeal squamous cell carcinoma via ATG16L1-dependent cellular autophagy, *Cell Insight* 2 (2023) 100127.
- [170] S.H. Nasr, J.A. Vrana, S. Dasari, F. Bridoux, M.E. Fidler, S. Kaaki, et al., DNAJB9 is a specific immunohistochemical marker for fibrillary glomerulonephritis, *Kidney Int Rep* 3 (2018) 56–64.
- [171] D. Wang, T. Zeng, Z. Lin, L. Yan, F. Wang, L. Tang, et al., Long non-coding RNA SNHG5 regulates chemotherapy resistance through the miR-32/DNAJB9 axis in acute myeloid leukemia, *Biomed. Pharmacother.* 123 (2020) 109802.
- [172] I. Martínez-Reyes, N.S. Chandel, Cancer metabolism: looking forward, *Nat. Rev. Cancer* 21 (2021) 669–680.
- [173] H. Zheng, M. Zhang, X. Ke, X. Deng, D. Li, Q. Wang, et al., lncRNA XIST/miR-137 axis strengthens chemo-resistance and glycolysis of colorectal cancer cells by hindering transformation from PKM2 to PKM1, *Cancer Biomark* 30 (2021) 395–406.
- [174] Z.-Q. Zheng, Z.-X. Li, J.-L. Guan, X. Liu, J.-Y. Li, Y. Chen, et al., Long noncoding RNA TINCR-mediated regulation of acetyl-CoA metabolism promotes nasopharyngeal carcinoma progression and chemoresistance, *Cancer Res.* 80 (2020) 5174–5188.
- [175] H. Wu, B. Liu, Z. Chen, G. Li, Z. Zhang, MSC-induced lncRNA HCP5 drove fatty acid oxidation through miR-3619-5p/AMPK/PGC1 α /CEBPB axis to promote stemness and chemo-resistance of gastric cancer, *Cell Death Dis.* 11 (2020) 233.
- [176] W. He, B. Liang, C. Wang, S. Li, Y. Zhao, Q. Huang, et al., MSC-regulated lncRNA MACC1-AS1 promotes stemness and chemoresistance through fatty acid oxidation in gastric cancer, *Oncogene* 38 (2019) 4637–4654.
- [177] K. Domvri, S. Petanidis, D. Anastakis, K. Porpodis, C. Bai, P. Zarogoulidis, et al., Exosomal lncRNA PCAT-1 promotes Kras-associated chemoresistance via immunosuppressive miR-182/miR-217 signaling and p27/CDK6 regulation, *Oncotarget* 11 (2020) 2847–2862.
- [178] M.V. Libertì, J.W. Locasale, The Warburg effect: how does it benefit cancer cells? *Trends Biochem. Sci.* 41 (2016) 211–218.
- [179] P. Vaupel, G. Multhoff, Revisiting the Warburg effect: historical dogma versus current understanding, *J. Physiol.* 599 (2021) 1745–1757.
- [180] Y.-D. Chu, L.-C. Cheng, S.-N. Lim, M.-W. Lai, C.-T. Yeh, W.-R. Lin, Aldolase B-driven lactagenesis and CEACAM6 activation promote cell renewal and chemoresistance in colorectal cancer through the Warburg effect, *Cell Death Dis.* 14 (2023) 660.
- [181] J. Li, R. Gao, J. Zhang, USP22 contributes to chemoresistance, stemness, and EMT phenotype of triple-negative breast cancer cells by regulating the Warburg effect via c-myc deubiquitination, *Clin. Breast Cancer* 23 (2023) 162–175.
- [182] Y. Zhou, S. Huang, Y. Guo, M. Ran, W. Shan, W.-H. Chen, et al., Epigallocatechin gallate circumvents drug-induced resistance in non-small-cell lung cancer by modulating glucose metabolism and AMPK/AKT/MAPK axis, *Phytother Res.* 37 (2023) 5837–5853.
- [183] Y. Zhang, Y. Liu, X. Xu, Knockdown of lncRNA-UCA1 suppresses chemoresistance of pediatric AML by inhibiting glycolysis through the microRNA-125a/hexokinase 2 pathway, *J. Cell. Biochem.* 119 (2018) 6296–6308.
- [184] J. Cui, Y. Guo, H. Wu, J. Xiong, T. Peng, Everolimus regulates the activity of gemcitabine-resistant pancreatic cancer cells by targeting the Warburg effect via PI3K/AKT/mTOR signaling, *Mol Med* 27 (2021) 38.
- [185] X. Tian, M. Liu, X. Huang, Q. Zhu, W. Liu, W. Chen, et al., Noscapine induces apoptosis in human colon cancer cells by regulating mitochondrial damage and Warburg effect via PTEN/PI3K/mTOR signaling pathway, *OncoTargets Ther.* 13 (2020) 5419–5428.
- [186] H. Shao, J. Chen, A. Li, L. Ma, Y. Tang, H. Chen, et al., Salvigenin suppresses hepatocellular carcinoma glycolysis and chemoresistance through inactivating the PI3K/AKT/GSK-3 β pathway, *Appl. Biochem. Biotechnol.* 195 (2023) 5217–5237.
- [187] M.J. Ryu, J. Han, S.J. Kim, M.J. Lee, X. Ju, Y.L. Lee, et al., PTEN/AKT signaling mediates chemoresistance in refractory acute myeloid leukemia through enhanced glycolysis, *Oncol. Rep.* 42 (2019) 2149–2158.
- [188] L. Chen, H. Zhao, C. Wang, N. Hu, TUG1 knockdown enhances adriamycin cytotoxicity by inhibiting glycolysis in adriamycin-resistant acute myeloid leukemia HL60/ADR cells, *RSC Adv.* 9 (2019) 10897–10904.
- [189] D.M. Ribeiro, A. Zanzoni, A. Cipriano, R. Delli Ponti, L. Spinelli, M. Ballarino, et al., Protein complex scaffolding predicted as a prevalent function of long non-coding RNAs, *Nucleic Acids Res.* 46 (2018) 917–928.
- [190] X. Zhang, S. Zheng, C. Hu, G. Li, H. Lin, R. Xia, et al., Cancer-associated fibroblast-induced lncRNA UPK1A-AS1 confers platinum resistance in pancreatic cancer via efficient double-strand break repair, *Oncogene* 41 (2022) 2372–2389.
- [191] K.C. Wang, H.Y. Chang, Molecular mechanisms of long noncoding RNAs, *Mol. Cell.* 43 (2011) 904–914.
- [192] H. Zhang, Y. Zhao, X. Liu, Y. Liu, X. Wang, Y. Fu, et al., A novel upregulated lncRNA-AC026150.8 promotes chemo-resistance and predicts poor prognosis in acute myeloid leukemia, *Cancer Med.* 10 (2021) 8614–8629.
- [193] J. Dai, Y. Su, S. Zhong, L. Cong, B. Liu, J. Yang, et al., Exosomes: key players in cancer and potential therapeutic strategy, *Signal Transduct Target Ther* 5 (2020) 145.
- [194] R. Kalluri, V.S. LeBleu, The biology, function, and biomedical applications of exosomes, *Science* 367 (2020), <https://doi.org/10.1126/science.aau6977>.
- [195] M. Zhang, H. Liu, Q. Cui, P. Han, S. Yang, M. Shi, et al., Tendon stem cell-derived exosomes regulate inflammation and promote the high-quality healing of injured tendon, *Stem Cell Res. Ther.* 11 (2020) 402.
- [196] J. Wang, H. Wu, Y. Peng, Y. Zhao, Y. Qin, Y. Zhang, et al., Hypoxia adipose stem cell-derived exosomes promote high-quality healing of diabetic wound involves activation of PI3K/Akt pathways, *J Nanobiotechnology* 19 (2021) 202.
- [197] W. Liu, M. Yu, D. Xie, L. Wang, C. Ye, Q. Zhu, et al., Melatonin-stimulated MSC-derived exosomes improve diabetic wound healing through regulating macrophage M1 and M2 polarization by targeting the PTEN/AKT pathway, *Stem Cell Res. Ther.* 11 (2020) 259.
- [198] M. Duan, Y. Zhang, H. Zhang, Y. Meng, M. Qian, G. Zhang, Epidermal stem cell-derived exosomes promote skin regeneration by downregulating transforming growth factor- β 1 in wound healing, *Stem Cell Res. Ther.* 11 (2020) 452.
- [199] P. Yuan, L. Ding, H. Chen, Y. Wang, C. Li, S. Zhao, et al., Neural stem cell-derived exosomes regulate neural stem cell differentiation through miR-9-hes1 Axis, *Front. Cell Dev. Biol.* 9 (2021) 601600.
- [200] D. Shen, Z. He, Mesenchymal stem cell-derived exosomes regulate the polarization and inflammatory response of macrophages via miR-21-5p to promote repair after myocardial reperfusion injury, *Ann. Transl. Med.* 9 (2021) 1323.
- [201] M. Elashiry, M.M. Elashiry, R. Elsayed, M. Rajendran, C. Auersvald, R. Zeitoun, et al., Dendritic cell derived exosomes loaded with immunoregulatory cargo reprogram local immune responses and inhibit degenerative bone disease in vivo, *J. Extracell. Vesicles* 9 (2020) 1795362.
- [202] F. Yuan, W. Peng, Y. Yang, J. Xu, Y. Liu, Y. Xie, et al., Endothelial progenitor cell-derived exosomes promote anti-inflammatory macrophages via SOCS3/JAK2/STAT3 axis and improve the outcome of spinal cord injury, *J. Neuroinflammation* 20 (2023) 156.
- [203] Q. Gao, X. Fang, Y. Chen, Z. Li, M. Wang, Exosomal lncRNA UCA1 from cancer-associated fibroblasts enhances chemoresistance in vulvar squamous cell carcinoma cells, *J. Obstet. Gynaecol. Res.* 47 (2021) 73–87.

- [204] Y. Sun, G. Hao, M. Zhuang, H. Lv, C. Liu, K. Su, MEG3 lncRNA from exosomes released from cancer-associated fibroblasts enhances cisplatin chemoresistance in SCLC via a miR-15a-5p/CCNE1 Axis, *Yonsei Med. J.* 63 (2022) 229–240.
- [205] Y. Tong, L. Yang, C. Yu, W. Zhu, X. Zhou, Y. Xiong, et al., Tumor-secreted exosomal lncRNA POU3F3 promotes cisplatin resistance in ESCC by inducing fibroblast differentiation into CAFs, *Mol Ther Oncolytics* 18 (2020) 1–13.
- [206] Z. Zhang, J. Yin, C. Lu, Y. Wei, A. Zeng, Y. You, Exosomal transfer of long non-coding RNA SBF2-AS1 enhances chemoresistance to temozolomide in glioblastoma, *J. Exp. Clin. Cancer Res.* 38 (2019) 166.
- [207] R. Jan, G.-E.-S. Chaudhry, Understanding apoptosis and apoptotic pathways targeted cancer therapeutics, *Adv Pharm Bull* 9 (2019) 205–218.
- [208] R. Kou, T. Li, C. Fu, D. Jiang, Y. Wang, J. Meng, et al., Exosome-shuttled FTO from BM-MSCs contributes to cancer malignancy and chemoresistance in acute myeloid leukemia by inducing m6A-demethylation: a nano-based investigation, *Environ. Res.* (2023) 117783.
- [209] B.A. Carneiro, W.S. El-Deiry, Targeting apoptosis in cancer therapy, *Nat. Rev. Clin. Oncol.* 17 (2020) 395–417.
- [210] R. Jan, G.-E.-S. Chaudhry, Understanding apoptosis and apoptotic pathways targeted cancer therapeutics, *Adv Pharm Bull* 9 (2019) 205–218.
- [211] A. Ramos, S. Sadeghi, H. Tabatabaiean, Battling chemoresistance in cancer: root causes and strategies to uproot them, *Int. J. Mol. Sci.* 22 (2021), <https://doi.org/10.3390/ijms22179451>.
- [212] J. Mei, G. Liu, R. Li, P. Xiao, D. Yang, H. Bai, et al., lncRNA SNHG6 knockdown inhibits cisplatin resistance and progression of gastric cancer through miR-1297/BCL-2 axis, *Biosci. Rep.* 41 (2021), <https://doi.org/10.1042/BSR20211885>.
- [213] Y. Pan, Y. Zhang, W. Liu, Y. Huang, X. Shen, R. Jing, et al., lncRNA H19 overexpression induces bortezomib resistance in multiple myeloma by targeting MCL-1 via miR-29b-3p, *Cell Death Dis.* 10 (2019) 106.
- [214] A.P. Singh, H. Luo, M. Matur, M.A. Eshelman, K. Hamamoto, A. Sharma, et al., A coordinated function of lncRNA HOTTIP and miRNA-196b underpinning leukemogenesis by targeting FAS signaling, *Oncogene* 41 (2022) 718–731.
- [215] J. Jian, N. Wang, H. Hao, C. Yuan, Q. Liu, C. Ji, et al., SLED1 promoting cell proliferation and inhibiting apoptosis in acute myeloid leukemia: a study, *Appl. Biochem. Biotechnol.* 195 (2023) 6633–6652.
- [216] D.-D. Lv, L.-Y. Zhou, H. Tang, Hepatocyte nuclear factor 4 α and cancer-related cell signaling pathways: a promising insight into cancer treatment, *Exp. Mol. Med.* 53 (2021) 8–18.
- [217] Y. Huang, L. Xian, Z. Liu, L. Wei, L. Qin, Y. Xiong, et al., AMPK α 2/HNF4A/BORIS/GLUT4 pathway promotes hepatocellular carcinoma cell invasion and metastasis in low glucose microenvironment, *Biochem. Pharmacol.* 203 (2022) 115198.
- [218] B. Shokouhian, B. Negahdari, Z. Heydari, M. Totonchi, H. Aboulkheyr, E. A. Piryaei, et al., HNF4 α is possibly the missing link between epithelial-mesenchymal transition and Warburg effect during hepatocarcinogenesis, *Cancer Sci.* 114 (2023) 1337–1352.
- [219] L.-L. Li, Z. Peng, Q. Hu, L.-J. Xu, X. Zou, D.-M. Huang, et al., Berberine retarded the growth of gastric cancer xenograft tumors by targeting hepatocyte nuclear factor 4 α , *World J. Gastrointest. Oncol.* 14 (2022) 842–857.
- [220] L.T.H. Phi, I.N. Sari, Y.-G. Yang, S.-H. Lee, N. Jun, K.S. Kim, et al., Cancer stem cells (CSCs) in drug resistance and their therapeutic implications in cancer treatment, *Stem Cells Int.* 2018 (2018) 5416923.
- [221] C. Shen, Y. Sheng, A.C. Zhu, S. Robinson, X. Jiang, L. Dong, et al., RNA demethylase ALKBH5 selectively promotes tumorigenesis and cancer stem cell self-renewal in acute myeloid leukemia, *Cell Stem Cell* 27 (2020) 64–80.e9.
- [222] M. Marzagalli, F. Fontana, M. Raimondi, P. Limonta, Cancer stem cells-key players in tumor relapse, *Cancers* 13 (2021), <https://doi.org/10.3390/cancers13030376>.
- [223] M.L. De Angelis, F. Francescangeli, A. Zeuner, Breast cancer stem cells as drivers of tumor chemoresistance, dormancy and relapse: new challenges and therapeutic opportunities, *Cancers* 11 (2019), <https://doi.org/10.3390/cancers11101569>.
- [224] J. Shi, C. Guo, Y. Li, J. Ma, The long noncoding RNA TINCR promotes self-renewal of human liver cancer stem cells through autophagy activation, *Cell Death Dis.* 13 (2022) 961.
- [225] Y. Wang, P. Zhu, J. Luo, J. Wang, Z. Liu, W. Wu, et al., lncRNA HAND2-AS1 promotes liver cancer stem cell self-renewal via BMP signaling, *EMBO J.* 38 (2019) e101110.
- [226] P.-Y. Chen, P.-L. Hsieh, C.-Y. Peng, Y.-W. Liao, C.-H. Yu, C.-C. Yu, lncRNA MEG3 inhibits self-renewal and invasion abilities of oral cancer stem cells by sponging miR-421, *J. Formos. Med. Assoc.* 120 (2021) 1137–1142.
- [227] Z. Gao, Q. Wang, M. Ji, X. Guo, L. Li, X. Su, Exosomal lncRNA UCA1 modulates cervical cancer stem cell self-renewal and differentiation through microRNA-122-5p/SOX2 axis, *J. Transl. Med.* 19 (2021) 229.
- [228] X. Chen, R. Xie, P. Gu, M. Huang, J. Han, W. Dong, et al., Long noncoding RNA LBCS inhibits self-renewal and chemoresistance of bladder cancer stem cells through epigenetic silencing of SOX2, *Clin. Cancer Res.* 25 (2019) 1389–1403.
- [229] R.K. Slany, The molecular biology of mixed lineage leukemia, *Haematologica* 94 (2009) 984–993.
- [230] A.V. Krivtsov, D. Twomey, Z. Feng, M.C. Stubbs, Y. Wang, J. Faber, et al., Transformation from committed progenitor to leukaemia stem cell initiated by MLL-AF9, *Nature* 442 (2006) 818–822.
- [231] G. Basati, M. Khaksarian, S. Abbaszadeh, H.E. Lashgarian, A. Marzban, Cancer stem cells and nanotechnological approaches for eradication, *Stem Cell Investig* 6 (2019) 38.
- [232] M. Wong, P. Polly, T. Liu, The histone methyltransferase DOT1L: regulatory functions and a cancer therapy target, *Am. J. Cancer Res.* 5 (2015) 2823–2837.
- [233] R. Vatapalli, V. Sagar, Y. Rodriguez, J.C. Zhao, K. Unno, S. Pamarthy, et al., Histone methyltransferase DOT1L coordinates AR and MYC stability in prostate cancer, *Nat. Commun.* 11 (2020) 4153.
- [234] Z. Song, Z. Wei, Q. Wang, X. Zhang, X. Tao, N. Wu, et al., The role of DOT1L in the proliferation and prognosis of gastric cancer, *Biosci. Rep.* 40 (2020), <https://doi.org/10.1042/BSR20193515>.
- [235] W.-T. Wang, T.-Q. Chen, Z.-C. Zeng, Q. Pan, W. Huang, C. Han, et al., The lncRNA LAMP5-AS1 drives leukemia cell stemness by directly modulating DOT1L methyltransferase activity in MLL leukemia, *J. Hematol. Oncol.* 13 (2020) 78.
- [236] Y. Zhang, Y.-X. Huang, D.-L. Wang, B. Yang, H.-Y. Yan, L.-H. Lin, et al., lncRNA DSCAM-AS1 interacts with YBX1 to promote cancer progression by forming a positive feedback loop that activates FOXA1 transcription network, *Theranostics* 10 (2020) 10823–10837.
- [237] H. Xu, G. Zhao, Y. Zhang, H. Jiang, W. Wang, D. Zhao, et al., Long non-coding RNA PAXIP1-AS1 facilitates cell invasion and angiogenesis of glioma by recruiting transcription factor ETS1 to upregulate KIF14 expression, *J. Exp. Clin. Cancer Res.* 38 (2019) 486.
- [238] L. Tao, X. Wang, Q. Zhou, Long noncoding RNA SNHG16 promotes the tumorigenicity of cervical cancer cells by recruiting transcriptional factor SPI1 to upregulate PARP9, *Cell Biol. Int.* 44 (2020) 773–784.
- [239] Z. Sun, G. Huang, H. Cheng, Transcription factor Nrf2 induces the up-regulation of lncRNA TUG1 to promote progression and adriamycin resistance in urothelial carcinoma of the bladder, *Cancer Manag. Res.* 11 (2019) 6079–6090.
- [240] C. Wu, J. Cui, Y. Huo, L. Shi, C. Wang, Alternative splicing of HOXB-AS3 underlie the promoting effect of nuclear m6A reader YTHDC1 on the self-renewal of leukemic stem cells in acute myeloid leukemia, *Int. J. Biol. Macromol.* 237 (2023) 123990.
- [241] Y. Sun, D. Dong, Y. Xia, L. Hao, W. Wang, C. Zhao, YTHDF1 promotes breast cancer cell growth, DNA damage repair and chemoresistance, *Cell Death Dis.* 13 (2022) 230.
- [242] A. Wu, X. Wang, F. Zhang, X. Yang, Y. Quan, J. Dong, et al., YTHDF1 enhances stemness and chemoresistance in triple-negative breast cancer cells by upregulating SIAH2, *Mol. Carcinog.* (2023), <https://doi.org/10.1002/mc.23661>.
- [243] P. Chen, X.-Q. Liu, X. Lin, L.-Y. Gao, S. Zhang, X. Huang, Targeting YTHDF1 effectively re-sensitizes cisplatin-resistant colon cancer cells by modulating GLS-mediated glutamine metabolism, *Mol Ther Oncolytics* 20 (2021) 228–239.
- [244] L. Hao, J.-M. Wang, B.-Q. Liu, J. Yan, C. Li, J.-Y. Jiang, et al., m6A-YTHDF1-mediated TRIM29 upregulation facilitates the stem cell-like phenotype of cisplatin-resistant ovarian cancer cells, *Biochim. Biophys. Acta Mol. Cell Res.* 1868 (2021) 118878.
- [245] S. Yuan, S.-H. He, L.-Y. Li, S. Xi, H. Weng, J.-H. Zhang, et al., A potassium-chloride co-transporter promotes tumor progression and castration resistance of prostate cancer through m6A reader YTHDC1, *Cell Death Dis.* 14 (2023) 7.
- [246] S.X. Guo, T. Taki, H. Ohnishi, H.Y. Piao, K. Tabuchi, F. Bessho, et al., Hypermethylation of p16 and p15 genes and RB protein expression in acute leukemia, *Leuk. Res.* 24 (2000) 39–46.
- [247] S. Gao, B. Zhou, H. Li, X. Huang, Y. Wu, C. Xing, et al., Long noncoding RNA HOTAIR promotes the self-renewal of leukemia stem cells through epigenetic silencing of p15, *Exp. Hematol.* 67 (2018) 32–40, e3.
- [248] M. Zhang, W.-B. Wu, Z.-W. Wang, X.-H. Wang, lncRNA NEAT1 is closely related with progression of breast cancer via promoting proliferation and EMT, *Eur. Rev. Med. Pharmacol. Sci.* 21 (2017) 1020–1026.
- [249] W. Peng, Z. Wang, H. Fan, lncRNA NEAT1 impacts cell proliferation and apoptosis of colorectal cancer via regulation of akt signaling, *Pathol. Oncol. Res.* 23 (2017) 651–656.
- [250] X. Li, S. Wang, Z. Li, X. Long, Z. Guo, G. Zhang, et al., The lncRNA NEAT1 facilitates cell growth and invasion via the miR-211/HMG2 axis in breast cancer, *Int. J. Biol. Macromol.* 105 (2017) 346–353.
- [251] J.-T. Kou, J. Ma, J.-Q. Zhu, W.-L. Xu, Z. Liu, X.-X. Zhang, et al., lncRNA NEAT1 regulates proliferation, apoptosis and invasion of liver cancer, *Eur. Rev. Med. Pharmacol. Sci.* 24 (2020) 4152–4160.
- [252] C. Gao, J. Zhang, Q. Wang, C. Ren, Overexpression of lncRNA NEAT1 mitigates multidrug resistance by inhibiting ABCG2 in leukemia, *Oncol. Lett.* 12 (2016) 1051–1057.
- [253] H. Yan, Z. Wang, Y. Sun, L. Hu, P. Bu, Cytoplasmic NEAT1 suppresses AML stem cell self-renewal and leukemogenesis through inactivation of Wnt signaling, *Adv. Sci.* 8 (2021) e2100914.
- [254] Y. Lyu, J. Lou, Y. Yang, J. Feng, Y. Hao, S. Huang, et al., Dysfunction of the WT1-MEG3 signaling promotes AML leukemogenesis via p53-dependent and -independent pathways, *Leukemia* 31 (2017) 2543–2551.
- [255] A. Wang, Y. Chen, L. Shi, M. Li, L. Li, S. Wang, et al., Tumor-suppressive MEG3 induces microRNA-493-5p expression to reduce arabinocytosine chemoresistance of acute myeloid leukemia cells by downregulating the METTL3/MYC axis, *J. Transl. Med.* 20 (2022) 288.
- [256] X. Qi, Y. Jiao, C. Cheng, F. Qian, Z. Chen, Q. Wu, H22954, a novel long non-coding RNA down-regulated in AML, inhibits cancer growth in a BCL-2-dependent mechanism, *Cancer Lett.* 454 (2019) 26–36.
- [257] X. Fang, X. Pan, H. Mai, X. Yuan, S. Liu, F. Wen, LINC00998 functions as a novel tumor suppressor in acute myeloid leukemia via regulating the ZFP36 ring finger protein/mammalian target of rapamycin complex 2 axis, *Bioengineered* 12 (2021) 10363–10372.
- [258] E. Porcù, M. Benetton, V. Bisio, A. Da Ros, C. Tregnago, G. Borella, et al., The long non-coding RNA CDK6-AS1 overexpression impacts on acute myeloid leukemia differentiation and mitochondrial dynamics, *iScience* 24 (2021) 103350.
- [259] J.E. Freedman, J.M. Miano, National heart, lung, and blood Institute workshop participants*. Challenges and opportunities in linking long noncoding RNAs to

- cardiovascular, lung, and blood diseases, *Arterioscler. Thromb. Vasc. Biol.* 37 (2017) 21–25.
- [260] Q. Sun, Q. Hao, K.V. Prasanth, Nuclear long noncoding RNAs: key regulators of gene expression, *Trends Genet.* 34 (2018) 142–157.
- [261] J.H. Noh, K.M. Kim, W.G. McClusky, K. Abdelmohsen, M. Gorospe, Cytoplasmic functions of long noncoding RNAs, *Wiley Interdiscip Rev RNA* 9 (2018) e1471.
- [262] N. Agrawal, P.V.N. Dasaradhi, A. Mohammed, P. Malhotra, R.K. Bhatnagar, S. K. Mukherjee, RNA interference: biology, mechanism, and applications, *Microbiol. Mol. Biol. Rev.* 67 (2003) 657–685.
- [263] M. Khajehdehi, M. Khalaj-Kondori, B. Baradaran, The siRNA-mediated knockdown of SNHG4 efficiently induced pro-apoptotic signaling and suppressed metastasis in SW1116 colorectal cancer cell line, *Mol. Biol. Rep.* 50 (2023) 8995–9006.
- [264] T. Jiang, J. Zhu, S. Jiang, Z. Chen, P. Xu, R. Gong, et al., Targeting lncRNA DDIT4-AS1 sensitizes triple negative breast cancer to chemotherapy via suppressing of autophagy, *Adv. Sci.* 10 (2023) e2207257.
- [265] Y. Zhang, H. Ai, X. Fan, S. Chen, Y. Wang, L. Liu, Knockdown of long non-coding RNA HOTAIR reverses cisplatin resistance of ovarian cancer cells through inhibiting miR-138-5p-regulated EZH2 and SIRT1, *Biol. Res.* 53 (2020) 18.
- [266] J. Ma, L.-N. Han, J.-R. Song, X.-M. Bai, J.-Z. Wang, L.-F. Meng, et al., Long noncoding RNA LINC01234 silencing exerts an anti-oncogenic effect in esophageal cancer cells through microRNA-193a-5p-mediated CCNE1 downregulation, *Cell. Oncol.* 43 (2020) 377–394.
- [267] A.M. Vaidya, Z. Sun, N. Ayat, A. Schilb, X. Liu, H. Jiang, et al., Systemic delivery of tumor-targeting siRNA nanoparticles against an oncogenic lncRNA facilitates effective triple-negative breast cancer therapy, *Bioconjug Chem.* 30 (2019) 907–919.
- [268] C. Nicolescu, A. Vaidya, A. Schilb, Z.-R. Lu, Regulating oncogenic lncRNA DANCER with targeted ECO/siRNA nanoparticles for non-small cell lung cancer therapy, *ACS Omega* 7 (2022) 22743–22753.
- [269] Z. Bi, Q. Li, X. Dinglin, Y. Xu, K. You, H. Hong, et al., Nanoparticles (NPs)-Mediated lncRNA AFAP1-AS1 silencing to block wnt/ β -catenin signaling pathway for synergistic reversal of radioresistance and effective cancer radiotherapy, *Adv. Sci.* 7 (2020) 2000915.
- [270] K. Yang, L. Xu, Y. Xu, Q. Shen, T. Qin, Y. Yu, et al., Nanoparticles (NPs)-mediated lncBCMA silencing to promote eEF1A1 ubiquitination and suppress breast cancer growth and metastasis, *Acta Pharm. Sin.* B 13 (2023) 3489–3502.
- [271] P. Connerty, E. Moles, C.E. de Bock, N. Jayatilake, J.L. Smith, S. Meshinchi, et al., Development of siRNA-loaded lipid nanoparticles targeting long non-coding RNA LINC01257 as a novel and safe therapeutic approach for t(8;21) pediatric acute myeloid leukemia, *Pharmaceutics* 13 (2021), <https://doi.org/10.3390/pharmaceutics13101681>.
- [272] C. Rinaldi, M.J.A. Wood, Antisense oligonucleotides: the next frontier for treatment of neurological disorders, *Nat. Rev. Neurol.* 14 (2018) 9–21.
- [273] H. Chen, M.K. Jayasinghe, E.Y.M. Yeo, Z. Wu, M. Pirisinu, W.M. Usman, et al., CD33-targeting extracellular vesicles deliver antisense oligonucleotides against FLT3-ITD and miR-125b for specific treatment of acute myeloid leukaemia, *Cell Prolif.* 55 (2022) e13255.
- [274] C. Yan, J. Gu, Y. Zhang, K. Ma, R.J. Lee, Efficient delivery of the Bcl-2 antisense oligonucleotide G3139 via nucleus-targeted aCD33-NKSN nanoparticles, *Int J Pharm* 625 (2022) 122074.
- [275] B.Z. Carter, D.H. Mak, S.J. Morris, G. Borthakur, E. Estey, A.L. Byrd, et al., XIAP antisense oligonucleotide (AEG35156) achieves target knockdown and induces apoptosis preferentially in CD34+38- cells in a phase 1/2 study of patients with relapsed/refractory AML, *Apoptosis* 16 (2011) 67–74.
- [276] R.S. Geary, D. Norris, R. Yu, C.F. Bennett, Pharmacokinetics, biodistribution and cell uptake of antisense oligonucleotides, *Adv. Drug Deliv. Rev.* 87 (2015) 46–51.
- [277] S.T. Crooke, S. Wang, T.A. Vickers, W. Shen, X.-H. Liang, Cellular uptake and trafficking of antisense oligonucleotides, *Nat. Biotechnol.* 35 (2017) 230–237.
- [278] J.K. Nair, J.L.S. Willoughby, A. Chan, K. Charisse, M.R. Alam, Q. Wang, et al., Multivalent N-acetylgalactosamine-conjugated siRNA localizes in hepatocytes and elicits robust RNAi-mediated gene silencing, *J. Am. Chem. Soc.* 136 (2014) 16958–16961.
- [279] M. Fridlender, Y. Kapulnik, H. Koltai, Plant derived substances with anti-cancer activity: from folklore to practice, *Front. Plant Sci.* 6 (2015) 799.
- [280] N.H. Nguyen, Q.T.H. Ta, Q.T. Pham, T.N.H. Luong, V.T. Phung, T.-H. Duong, et al., Anticancer activity of novel plant extracts and compounds from adenoma bracteosum (bonati) in human lung and liver cancer cells, *Molecules* 25 (2020), <https://doi.org/10.3390/molecules25122912>.
- [281] P. Rajendran, U. Maheshwari, A. Muthukrishnan, R. Muthuswamy, K. Anand, B. Ravindran, et al., Myricetin: versatile plant based flavonoid for cancer treatment by inducing cell cycle arrest and ROS-reliant mitochondria-facilitated apoptosis in A549 lung cancer cells and in silico prediction, *Mol. Cell. Biochem.* 476 (2021) 57–68.
- [282] Z. Zhang, W. Sang, L. Xie, W. Li, B. Li, J. Li, et al., Polyphenol-based nanomedicine evokes immune activation for combination cancer treatment, *Angew Chem. Int. Ed. Engl.* 60 (2021) 1967–1975.
- [283] D. Sadava, S. Chen, Castasterone, a plant steroid hormone, affects human small-cell lung cancer cells and reverses multi-drug resistance, *Pharmaceutics* 16 (2023), <https://doi.org/10.3390/ph16020170>.
- [284] P. Furdak, N. Pieńkowska, G. Bartosz, I. Sadowska-Bartosz, Extracts of common vegetables inhibit the growth of ovary cancer cells, *Foods* 11 (2022), <https://doi.org/10.3390/foods11162518>.
- [285] M. Greenwell, P.K.S.M. Rahman, Medicinal plants: their use in anticancer treatment, *Int J Life Sci Pharma Res.* 6 (2015) 4103–4112.
- [286] B. Almajali, H.A.N. Al-Jamal, W.R. Wan Taib, I. Ismail, M.F. Johan, A. A. Doolaanea, et al., Thymoquinone suppresses cell proliferation and enhances apoptosis of HL60 leukemia cells through Re-expression of JAK/STAT negative regulators, *Asian Pac J Cancer Prev* 22 (2021) 879–885.
- [287] X. Lai, Y. Sun, X. Zhang, D. Wang, J. Wang, H. Wang, et al., Honokiol induces ferroptosis by upregulating HMOX1 in acute myeloid leukemia cells, *Front. Pharmacol.* 13 (2022) 897791.
- [288] X. Yu, H. Li, P. Hu, Y. Qing, X. Wang, M. Zhu, et al., Natural HDAC-1/8 inhibitor baicalein exerts therapeutic effect in CBF-AML, *Clin. Transl. Med.* 10 (2020) e154.
- [289] X. Gu, M. Guan, C. Jiang, Q. Song, X. Li, N. Sun, et al., Assessment of thiosemicarbazone-containing compounds as potential antileukemia agents against P-gp overexpressing drug resistant K562/A02 cells, *Chem. Biodivers.* 18 (2021) e2000775.
- [290] C. Wen, X. Lu, Y. Sun, Q. Li, J. Liao, L. Li, Naringenin induces the cell apoptosis of acute myeloid leukemia cells by regulating the lncRNA XIST/miR-34a/HDAC1 signaling, *Heliyon* 9 (2023) e15826.
- [291] P.-P. Zhang, F. Zhang, K. Zhu, J.-F. Zhu, Y. Yuan, Y.-L. Yang, et al., Matrine exerted an anti-tumor effect on acute myeloid leukemia via the lncRNA LINC01116/miR-592-mediated JAK/STAT pathway inactivation, *Neoplasma* 69 (2022) 123–135.
- [292] K. Zare, M. Shademan, M.M. Ghahramani Seno, H. Dehghani, CRISPR/Cas9 knockout strategies to ablate CCAT1 lncRNA gene in cancer cells, *Biol. Proced. Online* 20 (2018) 21.
- [293] T.R. Fernando, J.R. Contreras, M. Zampini, N.I. Rodriguez-Malave, M.O. Alberti, J. Anguiano, et al., The lncRNA CAS15 regulates SOX4 expression in RUNX1-rearranged acute leukemia, *Mol. Cancer* 16 (2017) 126.
- [294] H. Luo, G. Zhu, J. Xu, Q. Lai, B. Yan, Y. Guo, et al., HOTTIP lncRNA promotes hematopoietic stem cell self-renewal leading to AML-like disease in mice, *Cancer Cell* 36 (2019) 645–659.e8.
- [295] J.D. Moulton, Using morpholinos to control gene expression, *Curr Protoc Nucleic Acid Chem* 68 (4.30.1–4.30.29) (2017).
- [296] H. Zhang, L. Liu, L. Chen, H. Liu, S. Ren, Y. Tao, Long noncoding RNA DANCER confers cytarabine resistance in acute myeloid leukemia by activating autophagy via the miR-874-3P/ATG16L1 axis, *Mol. Oncol.* 15 (2021) 1203–1216.