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An Approach to Modify 14-Membered Lactone Macrolide Antibiotic Scaffolds

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acetal in acidic conditions. The structure of this atypical β -keto ketene acetal intermediate within the macrocyclic system has been determined by NMR and X-ray methods. The use of basic conditions at an elevated temperature yielded new, doubly $\alpha_{,\beta}$ -unsaturated ketone macrolide derivatives with (4*E*)-configuration as two conformational isomers of folded-in or folded-out conformations.



actone macrolides with different sizes of the macrocyclic rings, such as, clarithromycin (1), erythromycin, azithromycin, or leucomycins, are used worldwide as agents in antibacterial therapy.^{1,2} The resistance of different bacterial strains to these antibiotics prompted medicinal chemists to design novel modifications on the basis of their structural alterations mainly within the hydroxyl or ketone groups from aglycone and/or saccharide.³ Modifications and the total synthesis of 14-membered clarithromycin 1 and its congeners have been performed widely with the conserved stereochemistry at carbon C(12) and the ketone at the C(3)position, ketolide group antibiotics.⁴⁻⁷ Recently, it has been shown that contraction of the aglycone ring of classical lactone antibiotics contributed to the formation of effective antibacterial agents against multiresistant Gram-negative pathogens.⁸ It should be mentioned that the aglycone ring of 14membered lactone macrolides is much more strained compared to 15- and 16-membered ones and hence is more prone to intramolecular reactions.9 Thus, all the abovementioned transformations of 14-membered lactone aglycones or saccharide parts belonging to 1 or erythromycins, hindered often by intramolecular ketalizations, were performed for years via identical intermediates, contributing to the generation of a huge number of comparably substituted macrolide structures.^{3,10-14} Taking into account the above facts, here, we propose another type of approach to aglycone modifications of erythromycin-like antibiotics via novel intermediates, offering the opportunity for greater semisynthetic structural diversification of 14-membered macrolide scaffolds.

Clarithromycin (1) was transformed into 5 via multistep synthesis according to a previous report (Scheme 1).¹⁵ The synthesis of product 5 has been confirmed by X-ray crystallography (Figure S1). During attempts of an alternative preparation of 5, 2'-acetylated 2 was isolated and its X-ray structure was also determined (Figure S2). Structures of 5 and 2'-acetylated 2 show *E*-configuration of the double bond

Scheme 1. Synthetic Route to Clarithromycin Derivative 5



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© 2022 The Authors. Published by American Chemical Society C(10)=C(11). In the next step, **5** was treated with CDI (1,1'-Carbonyldiimidazole) in basic conditions yielding carbamate **6**, widely used in the syntheses of telithromycin-like ketolides.¹⁶ The isolated product **6** was converted into a novel-type structure for lactone macrolides, i.e., β -keto ketene acetal intermediate 7 (Scheme 2), at basic conditions and elevated

Scheme 2. Synthesis of Novel Clarithromycin Intermediates 7 and 8



temperature. The yield of this reaction was good (72%), and the structure of 7 was proven by NMR. This type of structure is unexpected and rare since, within 1,3-ketoester systems, usually, the ketone group is the favorable site of enolization. Similar types of ketene acetals, i.e., synthons used for heterocyclization, were obtained in a different way, not involving 1,3-ketoesters.¹⁸ To the best of our knowledge, only two examples of such ketene acetals within the 1,3ketoester moiety have been characterized by X-ray crystallog-raphy in the literature.^{19,20} After smooth and efficient methanolysis of the acetyl group of 7, derivative 8 was formed (Scheme 2). The X-ray structure of 8 and the spectral characteristics (Supporting Information) allowed us to unambiguously confirm the unique structure of the lactone macrolides, containing the protected lactone in a bicyclic system within a β -keto ketene acetal moiety (Figure 1) and the altered absolute configuration at C(12) when compared to 1. The newly formed ketene acetal moiety has Z-configuration and is conjugated with C(3)-ketone in an *s*-trans arrangement. This type of structure can be alternatively formed via



Figure 1. ORTEP diagram of 8. Displacement ellipsoids were drawn at the 50% probability level.

stereospecific S_N1 - or S_N2 -type mechanisms due to the nucleophilic attack of the anion localized at the oxygen of the ketene acetal on the electrophilic carbon atom C(12). On the one hand, when one takes into account steric crowding within the C(9)–C(13) portion and the macrocyclic ring strain, where tertiary carbocation at C(12) is being attacked by the enolate oxygen, the S_N1 mechanism should be favored. On the other hand, the basic conditions and the nature of the leaving group should favor the S_N2 -type mechanism.

Compound 7 is sensitive to acidic conditions as shown in Scheme 3. The treatment of 7 with an acidic methanol solution





afforded derivative 9, possessing an inverted absolute configuration (12R) relative to its epimer 5 (12S) as well as 1, erythromycin, azithromycin, telithromycin, or their many congeners. At these experimental conditions, protonation of the ketone group at C(3) enables the nucleophilic attack on the electrophilic carbon C(1), followed by the ketene acetal C(1)-OC(12) cleavage with the retention of (12R) absolute configuration relative to 7 (Scheme S1). The stereochemistry of 9 was evidenced by ¹H-¹H NOESY (Figure S3) and chemical shift differences found in ¹H and ¹³C NMR spectra compared to that of 5 (Tables S1 and S2). The altered stereochemistry at C(12) enables structure stabilization via the H-bond between C(12)-hydroxyl and the lactone group, as was evidenced by the shift of the $v(O_{12}-H)$ band toward lower wavenumbers (\sim 3100 cm⁻¹) and the reduced intensity of the band (Figure S4). This intramolecular H-bond impacts the conformation of the whole aglycone, which is well reflected in chemical shift differences of 5 and 9 in ¹H and ¹³C NMR spectra (Tables S1 and S2). The DFT calculated structure of 9 stabilized by an intramolecular H-bond is shown in Figure 2.

An attempt at shortening the synthetic path leading to 7 from 5 afforded an 80% yield of novel-type product 10, lacking desosamine saccharide (Scheme 3). The unexpected elimination of the saccharide portion from the C(5) position of 5 is realized via the E1cB mechanism, as noted earlier for 16membered lactones.²¹ Broadening of the ν (C=O) band at ~1670 cm⁻¹ in the FT-IR spectrum of 10 showed that, in addition to unsaturated ketone at C(9), another unsaturated one at C(3) also exists (Figure S5). However, this type of unsaturated derivative 10 is formed as the mixture of two compounds (10a and 10b) having an identical 4*E*-configuration of the conjugated double bond with the ketone at C(3), as proved by 1D and 2D NMR spectra (Tables S1 and



Figure 2. Calculated structure of **9** on the basis of assumed contacts found in the NOESY spectrum using an XC functional: BLYP-D3 with basis set TZ2P (ADF package; see the Supporting Information).

S2; Figure S6). Initial attempts to separate 10a and 10b by the HPLC method with C18 and chiral columns and different mobile phases were unsuccessful (Figure S7). Further efforts at HPLC separation of 10a and 10b on a C8 column with the reversed stationary phase were successful (Figure S8); hence, 10a and 10b can be described as the two different conformational isomers instead of "conformers". Moreover, the occurrence of these two conformational isomers was evidenced independently on the solvent used (CDCl₃, CD_3CN_1 DMSO- d_6 ; Supporting Information). ¹H-¹H NOESY contacts for each of the conformational isomers in the mixture allowed one to assume the initial structures for the DFT calculations (Figure S6). As shown in Figure 3, when one takes into account the mutual arrangement of the methyl groups of aglycone, 10a has the folded-out structure, in contrast to 10b, which is rather compact and has a folded-in structure.^{22,23} The E1cB elimination of the mycaminose



(overlapped in the lactone region)

Figure 3. Calculated structures of **10a** (folded-out) and **10b** (folded-in) on the basis of assumed contacts found in the NOESY spectrum using an XC functional: BLYP-D3 with basis set TZ2P (ADF package; see the Supporting Information).

saccharide requires the formation of enolate, and hence, the epimerization at C(2) is possible. In NOESY spectra of **10a** and **10b**, the strong contact H(2)…H(5) and the weak one H(2)…H(11) were noted (Figure S6). A comparison of the calculated structures of **10a** and **10b** and their C(2S) epimers (Figure S9) shows that the presence of the above-mentioned ¹H–¹H contacts in the NOESY spectra excludes epimerization at C(2). In the structure of **10b**, the two antiparallel oriented α,β -unsaturated ketone moieties are stabilized via a mutual $\pi-\pi$ stacking interaction, where distances of C(4)…C(11) and C(5)…C(10) are 3.7 and 3.6 Å, respectively, as calculated by the DFT method (Figure 3). In contrast, these α,β -unsaturated ketone moieties in **10a** are much more distant from each other, as shown in Figure 3 (left, top).

In conclusion, we have developed a new synthetic approach leading to modifications of the 14-membered lactone aglycone of known antibiotics. Our approach enables inversion of the configuration at C(12) and formation of doubly α , β unsaturated ketone aglycones of 14-membered macrolide antibiotics. Further studies on the utility of these transformations for structural diversification of known macrolide antibiotic scaffolds are currently underway.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.1c02799.

Experimental procedures, copy of all ¹H and ¹³C NMR spectra with 2D NMR experiments and signal assignments collected in Tables S1 and S2, FT-IR and ATR spectra, details of DFT calculations with $\Delta G^{\circ}s$, *xyz* coordinates and calculated spectra, and X-ray structural details of 2'-acetylated **2**, **5**, and **8** (PDF)

Parallel execution process information for compounds 9 and 10 (PDF)

Accession Codes

CCDC 2111470, 2111471, and 2111770 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/ data_request/cif, or by emailing data_request@ccdc.cam.ac. uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Arsic, B.; Novak, P.; Barber, J.; Rimoli, M. G.; Kragol, G.; Sodano, F. *Macrolides: Properties, Synthesis and Applications*, 1 ed.; De Gruyter: Berlin, Boston, 2018; DOI: 10.1515/9783110515756.

(2) Fernandes, P.; Martens, E.; Pereira, D. Nature Nurtures the Design of New Semi-Synthetic Macrolide Antibiotics. *Journal of Antibiotics* **2017**, 70 (5), 527–533.

(3) Janas, A.; Przybylski, P. 14- and 15-Membered Lactone Macrolides and Their Analogues and Hybrids: Structure, Molecular Mechanism of Action and Biological Activity. *Eur. J. Med. Chem.* **2019**, *182*, 111662.

(4) Agouridas, C.; Denis, A.; Auger, J.-M.; Benedetti, Y.; Bonnefoy, A.; Bretin, F.; Chantot, J.-F.; Dussarat, A.; Fromentin, C.; D'Ambrières, S. G.; Lachaud, S.; Laurin, P.; Le Martret, O.; Loyau, V.; Tessot, N. Synthesis and Antibacterial Activity of Ketolides (6-O-Methyl-3-Oxoerythromycin Derivatives): A New Class of Antibacterials Highly Potent Against Macrolide-Resistant and -Susceptible Respiratory Pathogens. J. Med. Chem. **1998**, 41 (21), 4080–4100.

(5) Song, Q.-L.; Guo, B.-Q.; Zhang, W.; Lan, P.; Sun, P.-H.; Chen, W.-M. Design, Synthesis and Antibacterial Activity of Novel Ketolides Bearing an Aryltetrazolyl-Substituted Alkyl Side Chain. *J. Antibiot.* **2011**, *64* (8), 571–581.

(6) Velvadapu, V.; Paul, T.; Wagh, B.; Glassford, I.; DeBrosse, C.; Andrade, R. B. Total Synthesis of (-)-4,8,10-Tridesmethyl Telithromycin. J. Org. Chem. 2011, 76 (18), 7516-7527.

(7) Undheim, K. Scaffold Modifications in Erythromycin Macrolide Antibiotics. A Chemical Minireview. *Molecules* **2020**, *25* (17), 3941.

(8) Myers, A. G.; Clark, R. B. Discovery of Macrolide Antibiotics Effective against Multi-Drug Resistant Gram-Negative Pathogens. *Acc. Chem. Res.* **2021**, *54* (7), 1635–1645.

(9) Arsic, B.; Barber, J.; Čikoš, A.; Mladenovic, M.; Stankovic, N.; Novak, P. 16-Membered Macrolide Antibiotics: A Review. *Int. J. Antimicrob. Agents* **2018**, *51* (3), 283–298.

(10) Bhadra, P. K.; Magwaza, R. N.; Nirmalan, N.; Freeman, S.; Barber, J.; Arsic, B. Selected Derivatives of Erythromycin B-In Silico and Anti-Malarial Studies. *Materials* **2021**, *14* (22), 6980.

(11) Arsic, B.; Awan, A.; Brennan, R. J.; Aguilar, J. A.; Ledder, R.; McBain, A. J.; Regan, A. C.; Barber, J. Theoretical and Experimental Investigation on Clarithromycin, Erythromycin A and Azithromycin and Descladinosyl Derivatives of Clarithromycin and Azithromycin with 3-O Substitution as Anti-Bacterial Agents. *Med. Chem. Commun.* **2014**, 5 (9), 1347–1354.

(12) Pavlović, D.; Kimmins, S.; Mutak, S. Synthesis of Novel 15-Membered 8a-Azahomoerythromycin A Acylides: Consequences of Structural Modification at the C-3 and C-6 Position on Antibacterial Activity. *Eur. J. Med. Chem.* **2017**, *125*, 210–224.

(13) Wu, Z.; Lu, Y.; Luo, M.; He, X.; Xiao, Y.; Yang, J.; Pan, Y.; Qiu, G.; Xu, Y.; Huang, W.; Long, P.; Li, R.; Hu, X. Synthesis and Antibacterial Activity of Novel 4"-Carbamates of 6,11-Di-O-Methylerythromycin A. J. Antibiot. **2010**, 63 (7), 343–350.

(14) Novak, P.; Barber, J.; Čikoš, A.; Arsic, B.; Plavec, J.; Lazarevski, G.; Tepeš, P.; Košutić-Hulita, N. Free and Bound State Structures of 6-O-Methyl Homoerythromycins and Epitope Mapping of Their Interactions with Ribosomes. *Bioorg. Med. Chem.* **2009**, *17* (16), 5857–5867.

(15) Macher, I.; Souza, D. D. Process for the Production of Telithromycin. WO 2009053259 A1, April 30, 2009.

(16) Seiple, I. B.; Zhang, Z.; Jakubec, P.; Langlois-Mercier, A.; Wright, P. M.; Hog, D. T.; Yabu, K.; Allu, S. R.; Fukuzaki, T.; Carlsen, P. N.; Kitamura, Y.; Zhou, X.; Condakes, M. L.; Szczypiński, F. T.; Green, W. D.; Myers, A. G. A Platform for the Discovery of New Macrolide Antibiotics. *Nature* 2016, 533 (7603), 338-345.

(17) Haas, J.; Häckh, M.; Justus, V.; Müller, M.; Lüdeke, S. Addition of a Polyhistidine Tag Alters the Regioselectivity of Carbonyl Reductase S1 from Candida Magnoliae. *Org. Biomol. Chem.* **2017**, *15* (48), 10256–10264.

(18) Salanouve, E.; Guillou, S.; Bizouarne, M.; Bonhomme, F. J.; Janin, Y. L. 3-Methoxypyrazoles from 1,1-Dimethoxyethene, Few Original Results. *Tetrahedron* **2012**, *68* (15), 3165–3171.

(19) Cao, P.; Deng, C.; Zhou, Y.-Y.; Sun, X.-L.; Zheng, J.-C.; Xie, Z.; Tang, Y. Asymmetric Nazarov Reaction Catalyzed by Chiral Tris(Oxazoline)/Copper(II). *Angew. Chem., Int. Ed.* **2010**, *49* (26), 4463–4466.

(20) Banert, K.; Meier, B.; Penk, E.; Saha, B.; Würthwein, E.-U.; Grimme, S.; Rüffer, T.; Schaarschmidt, D.; Lang, H. Highly Strained 2,3-Bridged 2H-Azirines at the Borderline of Closed-Shell Molecules. *Chemistry* **2011**, *17* (4), 1128–1136.

(21) Klich, K.; Pyta, K.; Przybylski, P. Regio- and Stereoselective Functionalization of 16-Membered Lactone Aglycone of Spiramycin via Cascade Strategy. *J. Org. Chem.* **2015**, *80* (14), 7040–7049.

(22) Everett, J. R.; Tyler, J. W. The Conformational Analysis of Erythromycin A. J. Chem. Soc., Perkin Trans. 1987, 2 (11), 1659–1667.

(23) Everett, J. R.; Tyler, J. W. The Conformational Analysis of Three Derivatives of Erythromycin A: (9S)-9-Hydroxy-9-Deoxoerythromycin A, (9S)-9,11-O-Isopropylidene-9-Deoxoerythromycin A, and (9S)-Erythromycylamine A by Nuclear Magnetic Resonance Spectroscopy and Molecular Modelling. J. Chem. Soc., Perkin Trans. **1988**, 2 (3), 325–337.