γ-Secretase Components as Predictors of Breast Cancer Outcome

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Abstract

y-secretase is a large ubiquitously expressed protease complex composed of four core subunits: presenilin, Aph1, PEN-2, and nicastrin. The function of y-secretase in the cells is to proteolytically cleave various proteins within their transmembrane domains. Presenilin and Aph1 occur as alternative variants belonging to mutually exclusive ysecretase complexes and providing the complexes with heterogeneous biochemical and physiological properties. ysecretase is proposed to have a role in the development and progression of cancer and y-secretase inhibitors are intensively studied for their probable anti-tumor effects in various types of cancer models. Here, we for the first time determined mRNA expression levels of presenilin-1, presenilin-2, Aph1a, Aph1b, PEN-2, and nicastrin in a set of breast cancer tissue samples (N = 55) by quantitative real-time PCR in order to clarify the clinical significance of the expression of different y-secretase complex components in breast cancer. We found a high positive correlation between the subunit expression levels implying a common regulation of transcription. Our univariate Kaplan-Meier survival analyses established low expression level of y-secretase complex as a risk factor for breast cancer specific mortality. The tumors expressing low levels of y-secretase complex were characterized by high histopathological tumor grade, low or no expression of estrogen and progesterone receptors and consequently high probability to fall into the class of triple negative breast cancer tumors. These results may provide novel tools to further categorize breast cancer tumors, especially the highly aggressive and poorly treatable breast cancer type of triple negative cases, and suggest a significant role for γ -secretase in breast cancer.

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Introduction

 γ -secretase is a large ubiquitously expressed protease complex composed of four core subunits: presenilin (PS), Aph1, PEN-2, and nicastrin (NCT). These subunits are necessary and sufficient for the protease activity of γ -secretase [1,2]. γ -secretase cleaves various type I membrane proteins by regulated intramembrane proteolysis [1,3]. The γ -secretasemediated cleavage releases the C-terminal intracellular domain (ICD) of the substrate protein which may then execute important signaling functions inside the cell. The group of the γ secretase substrates is large and constantly growing encompassing already more than 90 members [3]. Many of the identified substrates are intimately involved in tumorigenesis. Examples of these proteins include Notch receptors and their ligands, CD44, ErbB4, E-cadherin, and MUC1. γ -secretase may influence on tumorigenesis also via its role in angiogenesis as many of the γ -secretase substrates (e.g. Notch, VEGFR-1, IGF1R, ErbB4, cadherins, and APP) are shown to regulate the formation and development of new blood vessels [4]. Thus γ -secretase inhibitors are intensively studied for their anti-tumor effects in various types of cancer models [2,5,6]. Several reports have described inhibitory effects of these compounds on breast cancer cell growth via down-

regulation of Notch signaling pathway which is aberrantly activated in breast cancer [6-8]. While previous studies have described the effects of γ -secretase inhibitors on cancer cells especially concentrating on only one of the γ -secretase substrates at the time (for example Notch or E-cadherin), the multiplicity of γ -secretase substrates suggests that the observed effects can be mediated via the inhibited cleavage of multiple substrates and subsequently altered signaling pathways. In addition to abnormal expression and function of many substrate proteins, the expression and/or activity of γ -secretase complex itself can be disturbed during tumorigenesis.

y-secretase subunits presenilin and Aph1 occur as alternative variants: PS1/PS2 and Aph1a/Aph1b [1,3]. Furthermore Aph1a can be alternatively spliced to short or long splice variant: Aph1aS or Aph1aL [9,10]. These variants seem to be differentially expressed among mouse, rat and human tissues [11-16] and to belong to mutually exclusive y-secretase complexes [9,10,17-19]. Consistently, many studies have suggested distinct yet overlapping biochemical and physiological roles for the subunit isoforms [11,13,20-29]. Altogether at least six distinct y-secretase complexes with different subunit composition and with varying enzymatic activities and physiological outcomes can be formed. It is highly possible that perturbations in the equilibrium of y-secretase complex components leading to profound effects on enzyme activity underlie some physiological disturbances. For example, a shift from the predominance of complexes containing PS1 and/or Aph1a towards a greater proportion of v-secretase complexes containing PS2 and/or Aph1b could be one factor leading to the development of Alzheimer's disease [26,28]. We hypothesized that a similar unbalance in the presence of distinct y-secretase complexes might be associated with the development and progression of breast cancer. Thus we wanted to clarify the clinical significance of the expression of ysecretase components in breast cancer. We aimed to resolve whether one of the distinct y-secretase complex types is preferentially expressed in breast cancer and whether the expression levels of different y-secretase components are associated with tumorigenesis, histopathological subtypes of the tumor, or breast cancer outcome. Here, we report a strong positive correlation between the mRNA expression levels of the y-secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT indicating a tight co-regulation of the transcription. We are able to establish low level of y-secretase complex as a risk factor for breast cancer specific mortality and to reveal the association of low level of y-secretase complex with triple negative type of breast cancer.

Materials and Methods

Patients

Fresh frozen tissue samples from 55 breast cancer cases (54 in the case of NCT) in the Kuopio Breast Cancer Project [30-33] were used in this study. Table 1 summarizes the clinicopathological data of the cases. The mean age at the time of breast cancer diagnosis was 62.6 years and all the study subjects were female. The patients were followed up until

Table 1. The clinicopathological data of the patients (N = 55).

Variable	N (%)
Patients age (years)	
≤ 51.9	17 (30.9)
≥ 52	38 (69.1)
Histopathological grade	
1	8 (14.5)
2	28 (50.9)
3	19 (34.5)
Stage	
1	12 (21.8)
2	42 (76.4)
3	0 (0.0)
4	1 (1.8)
Tumor type	
ductal	40 (72.7)
lobular	9 (16.4)
other	6 (10.9)
Estrogen receptor	
negative	14 (25.5)
positive	41 (74.5)
Progesterone receptor	
negative	22 (40.0)
positive	33 (60.0)
Her2 receptor	
0-2	50 (90.9)
3	4 (7.3)
No data	1 (1.8)
Triple negativity	
ER=0 / PR=0 /Her2=0-2	10 (18.2)
positive	45 (81.8)
Patient status	
Dead, breast cancer	20 (36.4)
Dead, other cause	21 (38.2)
Alive, no recurrence	11 (20.0)
Alive, recurrence	3 (5.5.)
Mean follow-up time (days)	3371.2 [38-6713]

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death or November 2009. The Kuopio Breast Cancer Project has been approved by the joint ethics committee of Kuopio University and Kuopio University Hospital (written consents 1/1989 and 61/2010). Each patient gave informed written consent for participation in the study.

RNA extraction, cDNA preparation, and quantitative real-time RT-PCR

Tissue specimens were stored at -70°C. RNA extraction and cDNA preparation were done essentially as described by Nykopp et al. (2009) [34]. TaqMan® Gene Expression Assays (Applied Biosystems) for the genes studied were Hs00997789_m1 (PS1), Hs01577197_m1 (PS2), Hs00211268_m1 (Aph1a), Hs00229911_m1 (Aph1b), Hs01033961_g1 (PEN-2), Hs00950933_m1 (NCT), and

Hs99999904_m1 (PPIA), which was used as the endogenous control [35]. Standard curves were established by cDNA obtained by reverse transcription of 2 µg of Human Breast Total RNA (Ambion®). Each sample and each point of the standard curve was performed in triplicate reactions. The maximum deviation between the expression values of each triplicate sample was allowed to be 0.3. The mean value of the triplicates was used as the raw expression value. Relative gene expression values were calculated as the ratio between the target gene and the endogenous control (cyclophilin A, PPIA) and were used in the statistical analyses.

In silico databases of human transcriptomes

The GeneSapiens database (http://www.genesapiens.org) was used to analyse previously published results of the gene expression levels of y-secretase subunits in human breast cancer [36]. The database contains 1,504 different human breast carcinoma samples from publicly available Affymetrix microarray experiments. In order to study gene expression levels of y-secretase subunits in triple negative subtype of breast cancer, mRNA expression (Agilent microarray) data of Genome Atlas (TCGA, The Cancer http:// cancergenome.nih.gov) were downloaded from the cBioPortal for Cancer Genomics (http://www.cbioportal.org) [37,38]. The selected TCGA dataset [39] contains 81 basal-like breast tumors and 445 tumors of other subtypes. 80 % of basal-like tumors were characterized as triple negative breast cancers.

Statistical methods

Statistical analyses were carried out using SPSS Statistics 17.0 for Windows (SPSS Inc.). P \leq 0.05 was considered significant in all analyses. Correlation between the expression levels of y-secretase subunits determined in this study were analysed by the Spearman's non-parametric test of correlation. When comparing the expression levels of single v-secretase subunits with known clinicopathological characteristics of the tumors (Table 1), differences between groups were analysed by non-parametric Mann-Whitney U-test in the case of two groups and by non-parametric Kruskal-Wallis test when multiple groups were included in the same comparison. Binary variables of individual y-secretase subunits were created by dividing the samples in two groups based on the mean of the relative gene expression values of the specific subunit (Table S1). The expression values below the mean were designated as 0 and above the mean as 1. Binary variables were used to describe the division of low and high expressing samples between various sample groups and in univariate survival analyses with the Kaplan-Meier method and log-rank test. Breast cancer survival was defined as the time between the date of diagnosis and the date of death due to breast cancer. Deaths by other causes were censored. The descriptive values (sample size, mean, and standard deviation) of low and high expressing sample groups are presented in Table S1. A common variable (named y-secretase) to describe the overall expression level of y-secretase complex in the samples was created by summarizing the zeros and ones of the binary variables of individual subunits. This variable with six ranks was used to calculate mean values, standard deviations and pTable 2. Association of mRNA expression level of γ -secretase complex with clinicopathological characteristics of the tumors.

	γ-secretase					
Variable	Low (%)	High (%)	Mean ± SD ^a	P-value ^b		
Histopathological grade						
1	4 (14.3)	4 (15.4)	3.25 ± 2.32	0.136		
2	13 (46.4)	14 (53.8)	2.74 ± 1.95			
3	11 (39.3)	8 (30.8)	1.79 ± 1.87			
Estrogen receptor						
negative	12 (42.9)	2 (7.7)	0.93 ± 1.33	<0.001**		
positive	16 (57.1)	24 (92.3)	3.03 ± 1.94			
Progesterone receptor						
negative	16 (57.1)	6 (23.1)	1.55 ± 1.77	0.004**		
positive	12 (42.9)	20 (76.9)	3.13 ± 1.95			
Her2 receptor						
0-2	25 (89.3)	24 (96.0)	2.59 ± 2.04	0.144		
3	3 (10.7)	1 (4.0)	1.00 ± 1.41			
Triple negativity						
yes	8 (28.6)	2 (7.7)	1.00 ± 1.49	0.006**		
no	20 (71.4)	24 (92.3)	2.82 ± 1.98			

a. Mean and standard deviation of $\gamma\mbox{-secretase}$ complex expression values of the samples belonging to each separate sample group

b. P-values of γ -secretase variable with six ranks by non-parametric Mann-Whitney U-test (or by non-parametric Kruskal-Wallis test in the case of histopathological grade)

**. Association is significant at the 0.01 level

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values presented in Table 2. Further, a binary γ -secretase variable was established by dividing the variable into low (ranks 0, 1, and 2) and high (ranks 3, 4, 5, and 6) expressing sample groups and used in Table 2 and in Kaplan-Meier analysis.

Results

Expression levels of γ -secretase subunits have a significant mutual correlation

mRNAs of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT were analysed by real-time quantitative PCR utilizing TaqMan® technology. Gene expression levels varied considerably between the samples. We found no preference for any of the subunit variants (PS1/PS2 and Aph1a/Aph1b) over the other in our sample set of breast cancer tissues. Instead, a significant positive correlation between the expression levels of γ -secretase subunits was observed (Table 3). In order to analyse previously published data on the gene expression of γ -secretase subunits in human breast cancer, we used the GeneSapiens *in silico* database of human transcriptomes [36]. The analysis revealed a similar significant positive correlation between the expression levels of γ -secretase subunits in order to analyse previously published data on the gene expression of γ -secretase subunits in human breast cancer, we used the GeneSapiens *in silico* database of human transcriptomes [36]. The analysis revealed a similar significant positive correlation between the expression levels of γ -secretase subunits (Table S2) as observed in our sample set.

Table 3. Correlation between mRNA expression levels of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT determined by Spearman's non-parametric correlation test.

-	PS2	Aph1a	Aph1b	PEN-2	NCT
PS1	0.726**	0.759**	0.737**	0.150	0,576**
PS2		0.709**	0.692**	0.237	0.671**
Aph1a			0.695**	0.343*	0.577**
Aph1b				0.388**	0.569**
PEN-2					0.350**

*. Correlation is significant at the 0.05 level

**. Correlation is significant at the 0.01 level

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Expression of γ -secretase subunits associates with tumor grade and hormone receptor status

We compared the expression levels of single γ -secretase subunits with known clinicopathological characteristics of the tumors (Table 1). The main findings are presented in Tables S3, S4, S5, S6, S7 and S8. The mRNA expression of all ysecretase subunits was lower in the sample group with tumor grade 3 than in lower grade tumors. The association between subunit expression and tumor grade was significant for Aph1b (p = 0.033; Table S6), PEN-2 (p = 0.005; Table S7), and NCT subunits (p = 0.043; Table S8). We found a significant association between mRNA expression of y-secretase subunits and protein expression of the estrogen (ER) and progesterone (PR) hormonal receptors. Low expression of y-secretase subunits was associated with low expression of the receptors. Aph1b (p = 0.032; Table S6) and PEN-2 subunits (p = 0.005; Table S7) had a significant association with human epidermal growth factor receptor 2 (Her2), but in this case the expression level of the subunits was lower in the sample group with high Her2 protein expression than in other tumors.

Triple negative breast cancer subtype is characterized by low expression of γ -secretase subunits

We compared the expression levels of γ -secretase subunits in the sample group of triple negative breast cancer cases (N = 10) to the expression in cancer tissues expressing at least one of the receptors ER, PR, or Her2 and found a significant association with PS2 (p = 0.002; Table S4), Aph1a (p = 0.038; Table S5), Aph1b (p = 0.002; Table S6), and PEN-2 subunits (p = 0.025; Table S7). Interestingly, the expression levels of γ secretase subunits were lower in triple negative breast cancer cases than in other samples. In order to validate this finding in larger sample set, we utilized publicly available TCGA dataset [39] containing mRNA expression levels of PS1, PS2, and Aph1b were significantly lower in the sample group of basal-like tumors (N = 81) than in tumors of other subtypes (N = 445; p < 0,001; Table S9).

Expression of $\boldsymbol{\gamma}\mbox{-secretase}$ subunits associates with clinical outcome

We performed Kaplan-Meier survival analyses in order to detect the possible prognostic role of the expression of γ -secretase subunits in breast cancer specific survival. For the analysis, we divided the samples into low and high expressing groups based on the mean of the relative gene expression values of the specific subunit. The descriptive values (sample size, mean, and standard deviation) of these groups are presented in Table S1. The minimum and maximum follow-up times were 38 days and over 18 years, respectively (Table 1). In addition to follow-up survival, also the 5 year survival was established. The survival curves are presented in Figures 1–3. There was a significant association between the low expression levels of PS1, Aph1a, Aph1b, and NCT and poor breast cancer specific survival.

γ-secretase ensemble has clinical significance

Because of the significant positive correlation of the expression levels of γ -secretase subunits, we created one common variable (named γ -secretase) to describe the overall expression level of γ -secretase complex in the samples (see Materials and Methods for details). The values of γ -secretase variable were compared with known clinicopathological characteristics of the tumors (Table 1). The results of the comparisons (Table 2) reinforce our findings with individual subunits showing a strong association of γ -secretase complex with ER and PR and a decreased expression of the enzyme complex in triple negative breast cancer cases.

The possible involvement of γ -secretase complex in the breast cancer specific survival was examined by Kaplan-Meier survival analysis (Figure 4). Low expression of γ -secretase complex predicted significantly poorer survival than the higher expression levels.

 γ -secretase subunit variants PS1/PS2 and Aph1a/Aph1b are suggested to belong to mutually exclusive γ -secretase complexes [9,10,17-19]. Thus with the subunit variants explored in this study, four distinct γ -secretase complexes can be formed. Therefore we created four novel variables to describe the overall expression level of each complex type. This was performed substantially with the same procedure which was used to create γ -secretase variable but excluding the subunit variants not involved in a specific complex type. Complex type variables showed mutual significant positive correlations (data not shown). Further analyses of the novel variables resulted in highly similar outcomes as the ones of γ -secretase variable (data not shown).

Exceptionally low or high level of γ -secretase complex characterizes specific tumor subtypes

Finally, we wanted to more closely study the characteristics of the tumors with very low (rank 0) or high (rank 6) expression of the γ -secretase complex (Table 4). There were 13 cases with the rank 0 (low expression of all the individual subunits of γ -secretase) in γ -secretase variable. The majority of these tumors (62 %) were classified to have a grade 3. The expression of ER was significantly lower in these tumors than in other cases (p = 0.002). 62 % of the tumors did not express



Figure 1. Breast cancer specific survival in Kaplan-Meier univariate analysis according to the expression levels of presenilins. Low (N = 28) and high (N = 27) mRNA levels of PS1 (A and B) and low (N = 32) and high (N = 23) mRNA levels of PS2 (C and D) at 5 years (A and C) and the whole follow-up time (B and D). doi: 10.1371/journal.pone.0079249.g001

ER. There were significantly more triple negative breast cancer cases in this subgroup of the samples than in other samples (p



Figure 2. Breast cancer specific survival in Kaplan-Meier univariate analysis according to the expression levels of Aph1 variants. Low (N = 34) and high (N = 21) mRNA levels of Aph1a (A and B) and low (N = 34) and high (N = 21) mRNA levels of Aph1b (C and D) at 5 years (A and C) and the whole follow-up time (B and D). doi: 10.1371/journal.pone.0079249.g002



Figure 3. Breast cancer specific survival in Kaplan-Meier univariate analysis according to the expression levels of PEN-2 and nicastrin. Low (N = 33) and high (N = 22) mRNA levels of PEN-2 (A and B) and low (N = 33) and high (N = 21) mRNA expression levels of NCT (C and D) at 5 years (A and C) and the whole follow-up time (B and D). doi: 10.1371/journal.pone.0079249.g003



Figure 4. Breast cancer specific survival in Kaplan-Meier univariate analysis according to the expression of γ -secretase complex. Low (N = 28) and high (N = 26) expression levels of γ -secretase complex at 5 years (A) and the whole follow-up time (B). doi: 10.1371/journal.pone.0079249.g004

= 0.008). In Kaplan-Meier survival analysis, low level of γ -secretase complex was associated with poor prognosis. The association was statistically significant at 5 years (p = 0.040).

The 4 samples with high expression of γ -secretase complex (rank 6 in γ -secretase variable) were characterized by exceptionally low histopathological grade, high expression of ER and PR, and low expression of Her2. None of the cases was of grade 3 or triple negative or expressed high levels of Her2, but all of them expressed ER. Because of the small number of highly expressing samples, it was not possible to show any significance in statistical tests for differences between the groups defined by specific clinicopathological characteristics of the tumors or in Kalpan-Meier survival analysis. All the 4 patients with tumors expressing high levels of γ -secretase subunits were still alive or had died because of other causes than breast cancer in the end of the follow-up time.

Discussion

The potential role of γ -secretase in the development and progression of cancer has been widely accepted [5,40]. To date, most studies have concentrated on investigating the expression levels and function of γ -secretase substrates [41-48] or the effects of γ -secretase inhibitors in breast cancer [49-53]. Limited attention has been given to investigating the expression and function of individual γ -secretase components.

Table 4. The characteristics of the tumors expressing exceptionally low (rank 0) or high (rank 6) levels of γ -secretase complex.

γ-secretase	Ν	Histopathol	ogical g	rade E	R	PR	HER2	Trip	ole negativity
0	13	3		-*	'	-	+	yes	*
6	4	1/2		+		+	-	no	
* designates		statistically sig	nificant	difference		dete	rminer	1 hv	non-narametri

 designates a statistically significant difference determined by non-parametric Mann-Whitney U-test

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To our knowledge this is the first report describing mRNA expression levels of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT in breast cancer tissues and establishing low level of γ -secretase complex as a risk factor for breast cancer specific mortality. Our data provides novel tools to characterize and categorize the diversity of breast cancer tumors. Of special importance is our finding of the association of low expression level of γ -secretase complex with triple negative type of breast cancer.

In this study, we first aimed to investigate the mRNA expression levels of γ -secretase subunits in human breast cancer specimens in order to observe possible predominance of some subunit variants over the others in this cancer type. Our results demonstrate a strong positive correlation between

the mRNA expression levels of the γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT (Table 3) indicating a tight co-regulation of the expression of these transcripts. In silico transcriptome analysis utilizing the GeneSapiens database gave further support to our observation. This finding was unexpected in the light of previous studies showing differential expression of PS1/PS2 and Aph1a/Aph1b among various tissues [11,12,14-16] and the compensatory expression of other members of PS or Aph1 families when the endogenous expression of their counterparts have been artificially suppressed [10,23,29]. However, our results are in line with the studies showing joint expression levels for the various y-secretase components [12,18,54]. It seems that all four proteins closely regulate each other. Knocking out or overexpressing one of the y-secretase subunits decreases and increases the expression levels of other components, respectively [16.18.21.25.27.29.55-57]. Previous studies have been conducted at protein level and the regulation mechanism has been suggested to involve stabilization, maturation, or degradation of the proteins. Our results at mRNA level indicate that also the level of transcription is tightly co-regulated. It is possible that y-secretase complex controls the transcription of its own subunits via a feedback loop [58,59].

Next we wanted to untangle whether there was association between the expression levels of y-secretase subunits and the clinicopathological characteristics of the tumors in breast cancer (Tables S3, S4, S5, S6, S7 and S8). We obtained very similar results with all the tested y-secretase components (PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT) and with the common ysecretase variable (Table 2) which we used to describe the expression level of the whole complex. This was not surprising as the expression levels of the individual subunits closely followed those of each other. Filipovic et al. (2011) studied protein expression of NCT and found significant association with histopathological tumor grade and with hormonal receptor (ER and PR) expressions [60]. Our results are in a full agreement with those findings: The mRNA expression of Aph1b, PEN-2, and NCT were significantly lower in breast cancer cases with high tumor grade and there was a significant association between high expression level of y-secretase complex and hormonal receptors. Previous research has established a cross-talk between Notch and ER in breast cancer [61-63]. Activation of Notch or inhibition of v-secretase has differential effects on ER negative and positive breast cancer cells [61]. Estradiol acting on ER inhibits Notch and amyloid-ß precursor protein signaling [63,64]. We can only speculate whether the cross-talk of Notch and ER is mediated via y-secretase activity. One tempting explanation for the observed high y-secretase complex level in ER expressing tumors is that down-regulation of Notch signaling in ER-positive cells induces a compensatory effect increasing the expression of y-secretase complex. Triple negative breast cancer type is characterized by the lack of expression of ER and PR. Furthermore Her2 is not over-expressed. These tumors tend to be high grade and the disease is aggressive with high recurrence, metastatic, and mortality rates [65]. Hormonal receptor and Her2 antagonists are ineffective in the treatment of this breast cancer type and therefore there is an urgent need

for better therapeutics for this form of cancer [65]. Previous studies have demonstrated a heavy dependence of triple negative breast cancers on Notch signaling and suggested ysecretase inhibitors as effective drugs for this breast cancer type [61-63,66,67]. Unfortunately our study does not give clear support to the idea of utilizing y-secretase inhibitors to treat triple negative breast cancer as the expression of y-secretase subunits was significantly lower in this breast cancer type than in the other cases. Our in silico analysis of publicly available TCGA dataset [39] supported this finding (Table S9). However, it is highly possible that already a small amount of y-secretase complexes exhibits significant activity and that the low level of y-secretase complex expression observed in triple negative breast cancer tumors is sufficient to produce elevated levels of activated Notch species in the conditions of high expression of Notch receptors and ligands typical for triple negative tumors [62.68.69]. Our further studies with larger sample set of triple negative breast cancer tissue samples will elucidate this matter.

Our results indicate that the tumors expressing low levels of v-secretase complex are characterized by higher histopathological tumor grade, low or no expression of hormonal receptors and consequently higher probability to fall into the class of triple negative tumors (Table 4). They seem to be more aggressive and poorly treatable and probably more fatal. Thus it was reasonable to investigate whether the components of y-secretase complex had prognostic value in breast cancer. Kaplan-Meier survival analysis revealed that low expression level of y-secretase complex was associated with poor breast cancer specific survival. Interestingly, the same trend was also observed with triple negative breast cancer cases (N = 10) only (data not shown), although the effect of the expression of y-secretase complex was not significant. The finding was unexpected from the point of view of multiple previous studies suggesting y-secretase inhibitors alone or in combination with other therapeutics as efficient drugs for breast cancer [6-8]. The principal rationale behind this therapeutic intervention is the aberrant Notch signaling in breast cancer, which leads to increased proliferation, restricted differentiation, impaired apoptosis and enhanced maintenance of putative cancer stem cells [43,52,70,71]. However, multiple lines of evidence suggest that the effect of y-secretase or Notch inhibition on cancer cells is far from straightforward. The cytotoxicity of y-secretase inhibitors to breast cancer cells might not be mediated via inhibition of Notch signaling but via proteasome inhibition [72]. Notch signaling itself is highly context-dependent and there is some evidence that Notch homologues may have opposite effects in breast cancer [73,74]. Although Notch deregulation appears to have oncogenic effects in numerous solid tumors, important exceptions exist. Notch-1 has been shown to play an important tumor-suppressive role in epidermal keratinocytes [75-77] and studies in cervical, prostate, lung, brain and liver cancers have also suggested tumor-suppressive function for Notch signaling [78]. It is possible that both the tumor-suppressive and oncogenic properties of Notch are taking place at the same time, and the final outcome is dependent on the cellular context [78,79]. Accordingly, Notch signaling may have multiple

divergent roles also in breast cancer cells. If we further consider γ -secretase with dozens of identified substrates besides Notch receptors [3] and possible hundreds of downstream targets, it becomes evident that the enzyme may play multiple and even opposite roles in the development and progression of cancer. Underlining this notion, γ -secretase has been reported to function as a tumor suppressor in epithelia via Notch signaling as well as via epidermal growth factor receptor and β -catenin [80-85]. In our further studies, we will aim to untangle the main pathways utilized by γ -secretase in breast cancer and the possible correlation between the expression of γ -secretase complex and the activity of these pathways.

Overall, a tempting explanation for many of our results may be a feedback mechanism used to compensate the potentially reduced activity of y-secretase or Notch by increasing transcription of y-secretase subunits. In this case, already a small level of v-secretase complexes would achieve required level of activity in the cells. The increased expression levels, on the other hand, would reflect reduced activity. This idea receives some support from a recent study on the biological activity of y-secretase inhibitor PF-03084014 in breast cancer xenograft models. The inhibitor was shown to induce significant tumor growth inhibition, robust impairment of Notch signaling, and significant upregulation of the mRNA expression level of NCT in HCC1599 model [86]. It has to be kept in mind that only a small percentage of PS is engaged in catalytically active complexes [24,87] and the same is probably true for the other subunits as well. Thus it is indeed highly possible that the expression of a certain subunit protein does not reflect the amount of active complex in the cells. This leads to the conclusion that the expression levels of y-secretase subunits observed in this study might not result in the increased expression at protein level and eventually in the increased activity of the enzymatic complex. However, many previous studies have described altered mRNA expression with a direct consequence of aberrant enzymatic activity [25,55,60,88-91].

All the interesting results introduced here were achieved by a sample set of 55 breast cancer tissues. This number of samples was clearly sufficient for the present study giving firm answers to our research problems. We obtained highly similar results with all of the y-secretase subunits tested and with our y-secretase variable which greatly increases the reliability of the results. Our in silico analyses gave further support to the findings. The results implicate an independent additional effect of low mRNA expression of y-secretase complex along with the other known risk factors on breast cancer specific survival. However, further studies will naturally benefit from larger sample size. As we cannot completely exclude the possibility of unspecific effects and synergy of other tumor characteristics (ER, PR, tumor grade) having an effect on our survival results, further studies using multivariate survival analyses with larger sample groups are needed to clarify the independence of the ysecretase effect.

In conclusion, this is to our knowledge the first report describing mRNA expression levels of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT in breast cancer tissues. We demonstrated a high positive correlation between the expression levels of γ -secretase subunits implying a

common regulation of transcription. We discerned a firm association of γ -secretase with ER and PR, a finding nicely in line with previous results obtained studying expression of NCT in breast cancer [60] and certainly deserving further investigation. We designated γ -secretase complex expression as a potential tool to categorize breast cancer tumors: Tumors with low γ -secretase complex expression typically lack hormone receptors and have a poor prognosis based on higher histopathological tumor grade and lower breast cancer specific survival. Furthermore, we show the association between γ -secretase complex expression and triple negativity of the breast cancer cases. These findings thus pave the way for exploring the role of the γ -secretase complex in triple negative breast cancer and for further categorizing this severe cancer type.

Supporting Information

Table S1. Sample size, mean and standard deviation of the relative gene expression values of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2 and NCT (All) and the same descriptive values of the low and high expressing sample groups categorized based on the mean above (Low and High).

(DOCX)

Table S2. Correlation between mRNA expression levels of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2, and NCT using the GeneSapiens *in silico* database (<u>http://www.genesapiens.org</u>) in human breast carcinoma samples (N = 757 - 953).



Table S3. Association of mRNA expression of presenilin 1 (PS1) with clinicopathological characteristics of the tumors. (DOCX)

Table S4. Association of mRNA expression of presenilin 2(PS2) with clinicopathological characteristics of the
tumors.

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(DOCX)
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Table S5. Association of mRNA expression of Aph1a with clinicopathological characteristics of the tumors. (DOCX)

Table S6. Association of mRNA expression of Aph1b with clinicopathological characteristics of the tumors. (DOCX)

Table S7. Association of mRNA expression of PEN-2 with clinicopathological characteristics of the tumors. (DOCX)

 Table S8. Association of mRNA expression of nicastrin (NCT) with clinicopathological characteristics of the tumors.

(DOCX)

Table S9. Association of mRNA expression of γ -secretase subunits PS1, PS2, Aph1a, Aph1b, PEN-2 and NCT with basal-like disease subtype in publicly available TCGA (<u>http://cancergenome.nih.gov</u>) dataset of 526 breast cancer tumor samples [39].

(DOCX)

References

- De Strooper B (2003) Aph-1, pen-2, and nicastrin with presenilin generate an active γ-secretase complex. Neuron 38: 9-12. doi:10.1016/ S0896-6273(03)00205-8. PubMed: 12691659.
- Selkoe DJ, Wolfe MS (2007) Presenilin: Running with scissors in the membrane. Cell 131: 215-221. doi:10.1016/j.cell.2007.10.012. PubMed: 17956719.
- Haapasalo A, Kovacs DM (2011) The many substrates of presenilin/γsecretase. J Alzheimers Dis 25: 3-28. PubMed: 21335653.
- Boulton ME, Cai J, Grant MB (2008) γ-secretase: a multifaceted regulator of angiogenesis. J Cell Mol Med 12: 781-795. doi:10.1111/j. 1582-4934.2008.00274.x. PubMed: 18266961.
- Shih leM, Wang TL (2007) Notch signaling, γ-secretase inhibitors, and cancer therapy. Cancer Res 67: 1879-1882. doi: 10.1158/0008-5472.CAN-06-3958. PubMed: 17332312.
- Groth C, Fortini ME (2012) Therapeutic approaches to modulating notch signaling: Current challenges and future prospects. Semin Cell Dev Biol 23: 465-472. doi:10.1016/j.semcdb.2012.01.016. PubMed: 22309842.
- Guo S, Liu M, Gonzalez-Perez RR (2011) Role of notch and its oncogenic signaling crosstalk in breast cancer. Biochim Biophys Acta 1815: 197-213. PubMed: 21193018.
- Al-Hussaini H, Subramanyam D, Reedijk M, Sridhar SS (2011) Notch signaling pathway as a therapeutic target in breast cancer. Mol Cancer Ther 10: 9-15. doi:10.1158/1535-7163.MCT-10-0677. PubMed: 20971825.
- Lee SF, Shah S, Li H, Yu C, Han W et al. (2002) Mammalian APH-1 interacts with presenilin and nicastrin and is required for intramembrane proteolysis of amyloid-β precursor protein and notch. J Biol Chem 277: 45013-45019. doi:10.1074/jbc.M208164200. PubMed: 12297508.
- Gu Y, Chen F, Sanjo N, Kawarai T, Hasegawa H et al. (2003) APH-1 interacts with mature and immature forms of presenilins and nicastrin and may play a role in maturation of presenilin.nicastrin complexes. J Biol Chem 278: 7374-7380. doi:10.1074/jbc.M209499200. PubMed: 12471034.
- Coolen MW, van Loo KM, van Bakel NN, Ellenbroek BA, Cools AR et al. (2006) Reduced aph-1b expression causes tissue- and substratespecific changes in γ-secretase activity in rats with a complex phenotype. FASEB J 20: 175-177. PubMed: 16249316.
- Hébert SS, Serneels L, Dejaegere T, Horré K, Dabrowski M et al. (2004) Coordinated and widespread expression of γ-secretase in vivo: Evidence for size and molecular heterogeneity. Neurobiol Dis 17: 260-272. doi:10.1016/j.nbd.2004.08.002. PubMed: 15474363.
- Herreman A, Hartmann D, Annaert W, Saftig P, Craessaerts K et al. (1999) Presenilin 2 deficiency causes a mild pulmonary phenotype and no changes in amyloid precursor protein processing but enhances the embryonic lethal phenotype of presenilin 1 deficiency. Proc Natl Acad Sci U S A 96: 11872-11877. doi:10.1073/pnas.96.21.11872. PubMed: 10518543.
- Ilaya NT, Evin G, Masters CL, Culvenor JG (2004) Nicastrin expression in mouse peripheral tissues is not co-ordinated with presenilin and is high in muscle. J Neurochem 91: 230-237. doi:10.1111/j. 1471-4159.2004.02718.x. PubMed: 15379903.
- Lee MK, Slunt HH, Martin LJ, Thinakaran G, Kim G et al. (1996) Expression of presenilin 1 and 2 (PS1 and PS2) in human and murine tissues. J Neurosci 16: 7513-7525. PubMed: 8922407.
- Saito S, Araki W (2005) Expression profiles of two human APH-1 genes and their roles in formation of presenilin complexes. Biochem Biophys

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Author Contributions

Conceived and designed the experiments: AH MH AM V-MK VK. Performed the experiments: HMP. Analyzed the data: HMP. Wrote the manuscript: HMP. Supervised data analysis: AM. Participated in the interpretation of the results: HMP AH MH VK V-MK AM. Revised the manuscript: AH MH VK V-MK AM.

Res Commun 327: 18-22. doi:10.1016/j.bbrc.2004.11.130. PubMed: 15629423.

- 17. Yu G, Chen F, Levesque G, Nishimura M, Zhang DM et al. (1998) The presenilin 1 protein is a component of a high molecular weight intracellular complex that contains β-catenin. J Biol Chem 273: 16470-16475. doi:10.1074/jbc.273.26.16470. PubMed: 9632714.
- Steiner H, Winkler E, Edbauer D, Prokop S, Basset G et al. (2002) PEN-2 is an integral component of the γ-secretase complex required for coordinated expression of presenilin and nicastrin. J Biol Chem 277: 39062-39065. doi:10.1074/jbc.C200469200. PubMed: 12198112.
- Saura CA, Tomita T, Davenport F, Harris CL, Iwatsubo T et al. (1999) Evidence that intramolecular associations between presenilin domains are obligatory for endoproteolytic processing. J Biol Chem 274: 13818-13823. doi:10.1074/jbc.274.20.13818. PubMed: 10318786.
- Bentahir M, Nyabi O, Verhamme J, Tolia A, Horré K et al. (2006) Presenilin clinical mutations can affect γ-secretase activity by different mechanisms. J Neurochem 96: 732-742. doi:10.1111/j. 1471-4159.2005.03578.x. PubMed: 16405513.
- Chen F, Tandon A, Sanjo N, Gu YJ, Hasegawa H et al. (2003) Presenilin 1 and presenilin 2 have differential effects on the stability and maturation of nicastrin in mammalian brain. J Biol Chem 278: 19974-19979. doi:10.1074/jbc.M210049200. PubMed: 12646573.
- Culvenor JG, Evin G, Cooney MA, Wardan H, Sharples RA et al. (2000) Presenilin 2 expression in neuronal cells: Induction during differentiation of embryonic carcinoma cells. Exp Cell Res 255: 192-206. doi:10.1006/excr.1999.4791. PubMed: 10694435.
- Jayadev S, Case A, Eastman AJ, Nguyen H, Pollak J et al. (2010) Presenilin 2 is the predominant γ-secretase in microglia and modulates cytokine release. PLOS ONE 5: e15743. doi:10.1371/journal.pone. 0015743. PubMed: 21206757.
- Lai MT, Chen E, Crouthamel MC, DiMuzio-Mower J, Xu M et al. (2003) Presenilin-1 and presenilin-2 exhibit distinct yet overlapping γsecretase activities. J Biol Chem 278: 22475-22481. doi:10.1074/ jbc.M300974200. PubMed: 12684521.
- Ma G, Li T, Price DL, Wong PC (2005) APH-1a is the principal mammalian APH-1 isoform present in γ-secretase complexes during embryonic development. J Neurosci 25: 192-198. doi:10.1523/ JNEUROSCI.3814-04.2005. PubMed: 15634781.
- Placanica L, Tarassishin L, Yang G, Peethumnongsin E, Kim SH et al. (2009) Pen2 and presenilin-1 modulate the dynamic equilibrium of presenilin-1 and presenilin-2 γ-secretase complexes. J Biol Chem 284: 2967-2977. PubMed: 19036728.
- Serneels L, Dejaegere T, Craessaerts K, Horré K, Jorissen E et al. (2005) Differential contribution of the three Aph1 genes to γ-secretase activity in vivo. Proc Natl Acad Sci U S A 102: 1719-1724. doi:10.1073/ pnas.0408901102. PubMed: 15665098.
- Serneels L, Van Biervliet J, Craessaerts K, Dejaegere T, Horré K et al. (2009) γ-secretase heterogeneity in the Aph1 subunit: Relevance for alzheimer's disease. Science 324: 639-642. doi:10.1126/science. 1171176. PubMed: 19299585.
- Shirotani K, Edbauer D, Prokop S, Haass C, Steiner H (2004) Identification of distinct γ-secretase complexes with different APH-1 variants. J Biol Chem 279: 41340-41345. doi:10.1074/jbc.M405768200. PubMed: 15286082.
- Männistö S, Pietinen P, Pyy M, Palmgren J, Eskelinen M et al. (1996) Body-size indicators and risk of breast cancer according to menopause and estrogen-receptor status. Int J Cancer 68: 8-13. doi:10.1002/ (SICI)1097-0215(19960927)68:1. PubMed: 8895532.

- Mitrunen K, Jourenkova N, Kataja V, Eskelinen M, Kosma VM et al. (2000) Steroid metabolism gene CYP17 polymorphism and the development of breast cancer. Cancer Epidemiol Biomarkers Prev 9: 1343-1348. PubMed: 11142420.
- Pellikainen MJ, Pekola TT, Ropponen KM, Kataja VV, Kellokoski JK et al. (2003) p21WAF1 expression in invasive breast cancer and its association with p53, AP-2, cell proliferation, and prognosis. J Clin Pathol 56: 214-220. doi:10.1136/jcp.56.3.214. PubMed: 12610102.
- 33. Hartikainen JM, Tuhkanen H, Kataja V, Dunning AM, Antoniou A et al. (2005) An autosome-wide scan for linkage disequilibrium-based association in sporadic breast cancer cases in eastern Finland: Three candidate regions found. Cancer Epidemiol Biomarkers Prev 14: 75-80. PubMed: 15668479.
- Nykopp TK, Rilla K, Sironen R, Tammi MI, Tammi RH et al. (2009) Expression of hyaluronan synthases (HAS1-3) and hyaluronidases (HYAL1-2) in serous ovarian carcinomas: Inverse correlation between HYAL1 and hyaluronan content. BMC Cancer 9: 143. doi: 10.1186/1471-2407-9-143. PubMed: 19435493.
- de Kok JB, Roelofs RW, Giesendorf BA, Pennings JL, Waas ET et al. (2005) Normalization of gene expression measurements in tumor tissues: Comparison of 13 endogenous control genes. Lab Invest 85: 154-159. doi:10.1038/labinvest.3700208. PubMed: 15543203.
- Kilpinen S, Autio R, Ojala K, Iljin K, Bucher E et al. (2008) Systematic bioinformatic analysis of expression levels of 17,330 human genes across 9,783 samples from 175 types of healthy and pathological tissues. Genome Biol 9: R139. doi:10.1186/gb-2008-9-9-r139. PubMed: 18803840.
- Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO et al. (2012) The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov 2: 401-404. doi:10.1158/2159-8290.CD-12-0095 PubMed: 22588877.
- Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B et al. (2013) Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6: pl1 PubMed: 23550210.
- Cancer Genome Atlas Network (2012) Comprehensive molecular portraits of human breast tumours. Nature 490: 61-70. doi:10.1038/ nature11412. PubMed: 23000897.
- Purow B (2012) Notch inhibition as a promising new approach to cancer therapy. Adv Exp Med Biol 727: 305-319. doi: 10.1007/978-1-4614-0899-4_23. PubMed: 22399357.
- Howard EM, Lau SK, Lyles RH, Birdsong GG, Umbreit JN et al. (2005) Expression of e-cadherin in high-risk breast cancer. J Cancer Res Clin Oncol 131: 14-18. doi:10.1007/s00432-004-0618-z. PubMed: 15459769.
- 42. Jeschke U, Mylonas I, Kuhn C, Shabani N, Kunert-Keil C et al. (2007) Expression of E-cadherin in human ductal breast cancer carcinoma in situ, invasive carcinomas, their lymph node metastases, their distant metastases, carcinomas with recurrence and in recurrence. Anticancer Res 27: 1969-1974. PubMed: 17649807.
- Stylianou S, Clarke RB, Brennan K (2006) Aberrant activation of notch signaling in human breast cancer. Cancer Res 66: 1517-1525. doi: 10.1158/0008-5472.CAN-05-3054. PubMed: 16452208.
- Mittal S, Subramanyam D, Dey D, Kumar RV, Rangarajan A (2009) Cooperation of notch and Ras/MAPK signaling pathways in human breast carcinogenesis. Mol Cancer 8: 128. doi: 10.1158/1535-7163.TARG-09-C128. PubMed: 20030805.
- 45. Zardawi SJ, Zardawi I, McNeil CM, Millar EK, McLeod D et al. (2010) High Notch1 protein expression is an early event in breast cancer development and is associated with the HER-2 molecular subtype. Histopathology 56: 286-296. doi:10.1111/j.1365-2559.2009.03475.x. PubMed: 20459529.
- 46. Auvinen P, Tammi R, Tammi M, Johansson R, Kosma VM (2005) Expression of CD 44 s, CD 44 v 3 and CD 44 v 6 in benign and malignant breast lesions: Correlation and colocalization with hyaluronan. Histopathology 47: 420-428. doi:10.1111/j. 1365-2559.2005.02220.x. PubMed: 16178897.
- Junttila TT, Sundvall M, Lundin M, Lundin J, Tanner M et al. (2005) Cleavable ErbB4 isoform in estrogen receptor-regulated growth of breast cancer cells. Cancer Res 65: 1384-1393. doi: 10.1158/0008-5472.CAN-04-3150. PubMed: 15735025.
- Thor AD, Edgerton SM, Jones FE (2009) Subcellular localization of the HER4 intracellular domain, 4ICD, identifies distinct prognostic outcomes for breast cancer patients. Am J Pathol 175: 1802-1809. doi: 10.2353/ajpath.2009.090204. PubMed: 19808643.
- 49. Rasul S, Balasubramanian R, Filipović A, Slade MJ, Yagüe E et al. (2009) Inhibition of γ-secretase induces G2/M arrest and triggers apoptosis in breast cancer cells. Br J Cancer 100: 1879-1888. doi: 10.1038/sj.bjc.6605034. PubMed: 19513078.

- Efferson CL, Winkelmann CT, Ware C, Sullivan T, Giampaoli S et al. (2010) Downregulation of notch pathway by a y-secretase inhibitor attenuates AKT/mammalian target of rapamycin signaling and glucose uptake in an ERBB2 transgenic breast cancer model. Cancer Res 70: 2476-2484. doi:10.1158/0008-5472.CAN-09-3114. PubMed: 20197467.
- Debeb BG, Cohen EN, Boley K, Freiter EM, Li L et al. (2012) Preclinical studies of notch signaling inhibitor RO4929097 in inflammatory breast cancer cells. Breast Cancer Res Treat 134: 495-510. doi: 10.1007/s10549-012-2075-8. PubMed: 22547109.
- Kondratyev M, Kreso A, Hallett RM, Girgis-Gabardo A, Barcelon ME et al. (2012) Gamma-secretase inhibitors target tumor-initiating cells in a mouse model of ERBB2 breast cancer. Oncogene 31: 93-103. doi: 10.1038/onc.2011.212. PubMed: 21666715.
- 53. Séveno C, Loussouarn D, Bréchet S, Campone M, Juin P et al. (2012) γ-secretase inhibition promotes cell death, noxa upregulation, and sensitization to BH3 mimetic ABT-737 in human breast cancer cells. Breast Cancer Res 14: R96. doi:10.1186/bcr3214. PubMed: 22703841.
- Madsen LB, Thomsen B, Larsen K, Bendixen C, Holm IE et al. (2007) Molecular characterization and temporal expression profiling of presenilins in the developing porcine brain. BMC Neurosci 8: 72. doi: 10.1186/1471-2202-8-72. PubMed: 17854491.
- 55. Seo SJ, Hwang DY, Cho JS, Chae KR, Kim CK et al. (2007) PEN-2 overexpression induces γ -secretase protein and its activity with amyloid β -42 production. Neurochem Res 32: 1016-1023. doi:10.1007/s11064-006-9262-0. PubMed: 17401676.
- Takasugi N, Tomita T, Hayashi I, Tsuruoka M, Niimura M et al. (2003) The role of presenilin cofactors in the γ-secretase complex. Nature 422: 438-441. doi:10.1038/nature01506. PubMed: 12660785.
- 57. Zhang YW, Luo WJ, Wang H, Lin P, Vetrivel KS et al. (2005) Nicastrin is critical for stability and trafficking but not association of other presenilin/γ-secretase components. J Biol Chem 280: 17020-17026. doi:10.1074/jbc.M409467200. PubMed: 15711015.
- Checler F, Dunys J (2012) p53, a pivotal effector of a functional crosstalk linking presenilins and pen-2. Neurodegener Dis 10: 52-55. doi: 10.1159/000332935. PubMed: 22205087.
- Lleó A, Berezovska O, Ramdya P, Fukumoto H, Raju S et al. (2003) Notch1 competes with the amyloid precursor protein for γ-secretase and down-regulates presenilin-1 gene expression. J Biol Chem 278: 47370-47375. doi:10.1074/jbc.M308480200. PubMed: 12960155.
- Filipović A, Gronau JH, Green AR, Wang J, Vallath S et al. (2011) Biological and clinical implications of nicastrin expression in invasive breast cancer. Breast Cancer Res Treat 125: 43-53. doi:10.1007/ s10549-010-0823-1. PubMed: 20224929.
- Lee CW, Raskett CM, Prudovsky I, Altieri DC (2008) Molecular dependence of estrogen receptor-negative breast cancer on a notchsurvivin signaling axis. Cancer Res 68: 5273-5281. doi: 10.1158/0008-5472.CAN-07-6673. PubMed: 18593928.
- Lee CW, Simin K, Liu Q, Plescia J, Guha M et al. (2008) A functional notch-survivin gene signature in basal breast cancer. Breast Cancer Res 10: R97. doi:10.1186/bcr2200. PubMed: 19025652.
- Rizzo P, Miao H, D'Souza G, Osipo C, Song LL et al. (2008) Cross-talk between notch and the estrogen receptor in breast cancer suggests novel therapeutic approaches. Cancer Res 68: 5226-5235. doi: 10.1158/0008-5472.CAN-07-5744. PubMed: 18593923.
- 64. Bao J, Cao C, Zhang X, Jiang F, Nicosia SV et al. (2007) Suppression of β-amyloid precursor protein signaling into the nucleus by estrogens mediated through complex formation between the estrogen receptor and Fe65. Mol Cell Biol 27: 1321-1333. doi:10.1128/MCB.01280-06. PubMed: 17130235.
- Griffiths CL, Olin JL (2012) Triple negative breast cancer: A brief review of its characteristics and treatment options. J Pharm Pract 25: 319-323. doi:10.1177/0897190012442062. PubMed: 22551559.
- Haughian JM, Pinto MP, Harrell JC, Bliesner BS, Joensuu KM et al. (2012) Maintenance of hormone responsiveness in luminal breast cancers by suppression of notch. Proc Natl Acad Sci U S A 109: 2742-2747. doi:10.1073/pnas.1106509108. PubMed: 21969591.
- Chen JQ, Russo J (2009) ERalpha-negative and triple negative breast cancer: Molecular features and potential therapeutic approaches. Biochim Biophys Acta 1796: 162-175. PubMed: 19527773.
- Reedijk M, Odorcic S, Chang L, Zhang H, Miller N et al. (2005) Highlevel coexpression of JAG1 and NOTCH1 is observed in human breast cancer and is associated with poor overall survival. Cancer Res 65: 8530-8537. doi:10.1158/0008-5472.CAN-05-1069. PubMed: 16166334.
- Reedijk M, Pinnaduwage D, Dickson BC, Mulligan AM, Zhang H et al. (2008) JAG1 expression is associated with a basal phenotype and recurrence in lymph node-negative breast cancer. Breast Cancer Res Treat 111: 439-448. doi:10.1007/s10549-007-9805-3. PubMed: 17990101.

- Yamaguchi N, Oyama T, Ito E, Satoh H, Azuma S et al. (2008) NOTCH3 signaling pathway plays crucial roles in the proliferation of ErbB2-negative human breast cancer cells. Cancer Res 68: 1881-1888. doi:10.1158/0008-5472.CAN-07-1597. PubMed: 18339869.
- Harrison H, Farnie G, Howell SJ, Rock RE, Stylianou S et al. (2010) Regulation of breast cancer stem cell activity by signaling through the Notch4 receptor. Cancer Res 70: 709-718. doi: 10.1158/0008-5472.CAN-09-1681. PubMed: 20068161.
- 72. Han J, Ma I, Hendzel MJ, Allalunis-Turner J (2009) The cytotoxicity of γ-secretase inhibitor I to breast cancer cells is mediated by proteasome inhibition, not by γ-secretase inhibition. Breast Cancer Res 11: R57. doi:10.1186/bcr2347. PubMed: 19660128.
- O'Neill CF, Urs S, Cinelli C, Lincoln A, Nadeau RJ et al. (2007) Notch2 signaling induces apoptosis and inhibits human MDA-MB-231 xenograft growth. Am J Pathol 171: 1023-1036. doi:10.2353/ajpath.2007.061029. PubMed: 17675579.
- 74. Rizzo P, Osipo C, Pannuti A, Golde T, Osborne B et al. (2009) Targeting notch signaling cross-talk with estrogen receptor and ErbB-2 in breast cancer. Adv Enzyme Regul 49: 134-141. doi:10.1016/ j.advenzreg.2009.01.008. PubMed: 19344631.
- Nicolas M, Wolfer A, Raj K, Kummer JA, Mill P et al. (2003) Notch1 functions as a tumor suppressor in mouse skin. Nat Genet 33: 416-421. doi:10.1038/ng1099. PubMed: 12590261.
- Thélu J, Rossio P, Favier B (2002) Notch signalling is linked to epidermal cell differentiation level in basal cell carcinoma, psoriasis and wound healing. BMC Dermatol 2: 7. doi:10.1186/1471-5945-2-7. PubMed: 11978185.
- Proweller A, Tu L, Lepore JJ, Cheng L, Lu MM et al. (2006) Impaired notch signaling promotes de novo squamous cell carcinoma formation. Cancer Res 66: 7438-7444. doi:10.1158/0008-5472.CAN-06-0793. PubMed: 16885339.
- Leong KG, Karsan A (2006) Recent insights into the role of notch signaling in tumorigenesis. Blood 107: 2223-2233. doi:10.1182/ blood-2005-08-3329. PubMed: 16291593.
- Radtke F, Raj K (2003) The role of notch in tumorigenesis: Oncogene or tumour suppressor? Nat Rev Cancer 3: 756-767. doi:10.1038/ nrc1186. PubMed: 14570040.
- Kang DE, Soriano S, Xia X, Eberhart CG, De Strooper B et al. (2002) Presenilin couples the paired phosphorylation of β-catenin independent of axin: Implications for β-catenin activation in tumorigenesis. Cell 110: 751-762. doi:10.1016/S0092-8674(02)00970-4. PubMed: 12297048.
- Li T, Wen H, Brayton C, Das P, Smithson LA et al. (2007) Epidermal growth factor receptor and notch pathways participate in the tumor

suppressor function of $\gamma\text{-secretase.}$ J Biol Chem 282: 32264-32273. doi:10.1074/jbc.M703649200. PubMed: 17827153.

- Li T, Wen H, Brayton C, Laird FM, Ma G et al. (2007) Moderate reduction of γ-secretase attenuates amyloid burden and limits mechanism-based liabilities. J Neurosci 27: 10849-10859. doi:10.1523/ JNEUROSCI.2152-07.2007. PubMed: 17913918.
- Rocher-Ros V, Marco S, Mao JH, Gines S, Metzger D et al. (2010) Presenilin modulates EGFR signaling and cell transformation by regulating the ubiquitin ligase Fbw7. Oncogene 29: 2950-2961. doi: 10.1038/onc.2010.57. PubMed: 20208556.
- 84. Zhang YW, Wang R, Liu Q, Zhang H, Liao FF et al. (2007) Presenilin/ γ -secretase-dependent processing of β -amyloid precursor protein regulates EGF receptor expression. Proc Natl Acad Sci U S A 104: 10613-10618. doi:10.1073/pnas.0703903104. PubMed: 17556541.
- 85. Xia X, Qian S, Soriano S, Wu Y, Fletcher AM et al. (2001) Loss of presenilin 1 is associated with enhanced β-catenin signaling and skin tumorigenesis. Proc Natl Acad Sci U S A 98: 10863-10868. doi: 10.1073/pnas.191284198. PubMed: 11517342.
- 86. Zhang CC, Pavlicek A, Zhang Q, Lira ME, Painter CL et al. (2012) Biomarker and pharmacological evaluation of the γ-secretase inhibitor PF-03084014 in breast cancer models. Clin Cancer Res 18: 5008-5019. doi:10.1158/1078-0432.CCR-12-1379. PubMed: 22806875.
- 87. Beher D, Fricker M, Nadin A, Clarke EE, Wrigley JD et al. (2003) In vitro characterization of the presenilin-dependent γ -secretase complex using a novel affinity ligand. Biochemistry 42: 8133-8142. doi:10.1021/bi034045z. PubMed: 12846562.
- Coolen MW, Van Loo KM, Van Bakel NN, Pulford DJ, Serneels L et al. (2005) Gene dosage effect on γ-secretase component aph-1b in a rat model for neurodevelopmental disorders. Neuron 45: 497-503. doi: 10.1016/j.neuron.2004.12.054. PubMed: 15721236.
- Coolen MW, van Loo KM, Ellenbroek BA, Cools AR, Martens GJ (2006) Ontogenic reduction of aph-1b mRNA and γ-secretase activity in rats with a complex neurodevelopmental phenotype. Mol Psychiatry 11: 787-793. doi:10.1038/sj.mp.4001846. PubMed: 16718279.
- Edbauer D, Winkler É, Haass C, Steiner H (2002) Presenilin and nicastrin regulate each other and determine amyloid β-peptide production via complex formation. Proc Natl Acad Sci U S A 99: 8666-8671. PubMed: 12048259.
- 91. Francis R, McGrath G, Zhang J, Ruddy DA, Sym M et al. (2002) Aph-1 and pen-2 are required for notch pathway signaling, γ -secretase cleavage of β APP, and presenilin protein accumulation. Dev Cell 3: 85-97. doi:10.1016/S1534-5807(02)00189-2. PubMed: 12110170.