MITOGENOME ANNOUNCEMENT

OPEN ACCESS Check for updates

Taylor & Francis

Taylor & Francis Group

Characterization of the complete mitochondrial genome of the nematophagous fungus *Purpureocillium lavendulum*

Lian-Ming Liang^a (b), Ying Zhang^a, Jianping Xu^{a,b}, Ke-Qin Zhang^a and Yan-Ru Cao^c

^aState Key Laboratory for Conservation and Utilization of Bio-Resources in Yunnan and The Key Laboratory for Southwest Microbial Diversity of the Ministry of Education, Yunnan University, Kunming, China; ^bDepartment of Biology, McMaster University, Hamilton, Canada; ^cKey Laboratory of Special Biological Resource Development and Utilization of Universities in Yunnan province, College of Agriculture and Life Sciences, Kunming University, Kunming, China

ABSTRACT

The complete mitochondrial genome of *Purpureocillium lavendulum* was characterized in this study. This mitogenome is a closed circular molecule of 23,567 bp in length with a GC content of 28.46%, including 15 protein-coding genes, 25 transfer RNA genes, 2 ribosomal RNA genes. Phylogenetic analyses based on sequences at the 14 concatenated mitochondrial protein-coding genes showed that *P. lavendulum* was closely related to *Hirsutella minnesotensis*.

ARTICLE HISTORY Received 22 September 2020 Accepted 29 October 2020

KEYWORDS Nematophagous fungus; *Purpureocillium lavendulum*

Plant-parasitic nematodes (PPNs) cause severe damage to agricultural crops worldwide (Liang et al. 2019). The nematophagous fungus Purpureocillium lavendulum (Pezizomy cotina, Sordariomycetes, Hypocreales, Ophiocordycipitaceae) was previously mis-identified as Paecilomyces lilacinus, a commonly used fungus to control agricultural nematodes, such as the root-knot nematode Meloidogyne incognita (Jatala et al. 1980). Recently, the mitochondrial genomes have been reported from a number of nematophagous fungi species (Deng and Yu 2019; Fang et al. 2019; Li and Yu 2019; Wang et al. 2019; Zhang and Yu 2019), which provides novel insights into the evolution of nematophagous fungi and facilitate further investigations of this ecologically and agriculturally important group of fungi (Zhang et al. 2020). The genus Purpureocillium was proposed in 2011 (Luangsa-Ard et al. 2011) and P. lavendulum was proposed as a new species in 2013 (Perdomo et al. 2013). We have created a high efficient genetic manipulation system for this fungus, thus facilitate the characterization of biological functions of nematode infection-related genes (Liu et al. 2019). Based on the system, key regulators in conidiation procedure in P. lavendulum have been characterized (Chen et al. 2020). However, the genetic information of its mitochondria has not been analyzed. In this study, we report the complete genome of P. lavendulum and investigate its phylogenetic relationship with other ascomycetous species. The annotated genomic sequence was submitted to GenBank under accession number MW019427.

The mitogenome was extracted from the whole genome assembly of *P. lavendulum* which we have sequenced using HiSeq4000 PE150 and PacBio RSII (unpublished data). The whole mitochondrial genome was annotated automatically using the Mitos2 tool (http://mitos2.bioinf.uni-leipzig.de/index.py) based on the Reference Ref 63 Fungi and Yeast 3 code (Bernt et al. 2013). tRNAs were annotated using tRNAscan-SE (Schattner et al. 2005). rRNAs were annotated using hmmer3 based on the database Rfam14 (Johnson et al. 2010).

The complete mitogenome of *P. lavendulum* is a closed circular molecule of 23,567 bp in length with a GC content of 28.46%, which contains 42 genes, including 15 mitochondrial protein-coding genes (PCGs), 25 tRNA genes (tRNA-Trp gene lacked), and 2 rRNA genes. Protein-encoding genes include three ATP synthase subunits (atp6, atp8, and atp9), three cytochrome oxidase subunits (cox1, cox2, and cox3), one apocytochrome b (cob), seven NADH dehydrogenase subunits (nad1, nad2, nad3, nad4, nad4L, nad5, and nad6) and one ribosomal protein (rps3). No introns were existed in all the predicted genes. All tRNA genes are encoded on the sense strand. All PCGs initiated with ATG as the start codon.

Phylogenetic analysis of 14 mitochondrial proteins of *P. lavendulum* and other 18 species in Ascomycota was performed by Bayesian inference (BI). As shown in Figure 1, *P. lavendulum* and *Hirsutella minnesotensis* clustered together and related to *Cordyceps brongniartii, Fusarium moniliformis, Colletotrichum acutatum* and *Sclerotinia borealis*, which are all members from Sordariomycetes.

CONTACT Lian-Ming Liang lianglm@ynu.edu.cn State Key Laboratory for Conservation and Utilization of Bio-Resources in Yunnan and The Key Laboratory for Southwest Microbial Diversity of the Ministry of Education, Yunnan University, Kunming, China; Yan-Ru Cao yanrucao3@aliyun.com Key Laboratory of Special Biological Resource Development and Utilization of Universities in Yunnan province, College of Agriculture and Life Sciences, Kunming University, Kunming, China

^{© 2021} The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. Phylogenetic relationships among 19 Ascomycota fungi inferred based on the concatenated amino acid sequences of 14 mitochondrial protein-coding genes (atp6, atp8, atp9, cox1, cox2, cox3, cob, nad1, nad2, nad3, nad4, nad4L, nad5 and nad6). The tree was generated using MrBayes. Values along branches represent statistical support based on 1000 randomizations.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported in part by grants from the National Natural Science Foundation of China [31860016 and 31660002], Joint Special Project of Universities from Yunnan Provincial Department of Science and Technology [2018FH001-003], and Grants from the Young Academic and Technical Leader Raising Foundation of Yunnan Province (to Lian-Ming Liang) and the Ten-thousand Talents Program in Yunnan province(to Lian-Ming Liang and Yan-Ru Cao).

ORCID

Lian-Ming Liang (D) http://orcid.org/0000-0001-5396-3781

Data availability statement

The data that support the findings of this study are openly available in the GenBank (accession no. MW019427) at https://www.ncbi.nlm.nih. gov/genbank/.

References

Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. Mol Phylogenet Evol. 69(2): 313–319.

- Chen M, Yang H-Y, Cao Y-R, Hui Q-Q, Fan H-F, Zhang C-C, Han J-J, Guo Z-Y, Xu J, Zhang K-Q, et al. 2020. Functional characterization of core regulatory genes involved in sporulation of the nematophagous fungus *Purpureocillium lavendulum*. mSphere. 5:e00932–20.
- Deng C, Yu Z. 2019. The complete mitochondrial genomes of *Dactylellina Leptospora* (Orbiliales, Orbiliaceae). Mitochondrial DNA Part B. 4(1): 1615–1616.
- Fang M, Wang S, Xu J, Jiang L, Zhou D, Zhang K-Q, Zhang Y. 2019. Characterization of the complete mitochondrial genome of *Drechslerella brochopaga*, a fungal species trapping nematodes with constricting rings. Mitochondrial DNA Part B. 4(1):858–859.
- Jatala P, Kaltenbach R, Bocangel M, Devaux J, Campos R. 1980. Field application of *Paecilomyces lilacinus* for controlling *Meloidogyne incognita* on potatoes. J Nematol. 12:226.
- Johnson LS, Eddy SR, Portugaly E. 2010. Hidden Markov model speed heuristic and iterative HMM search procedure. BMC Bioinformatics. 11: 431.
- Li W-J, Yu Z-F. 2019. The complete mitochondrial genomes of *Dactylella tenuis*, a fungus phylogenetically close to nematode-trapping fungus. Mitochondrial DNA B. 4(2):2704–2705.
- Liang LM, Zou CG, Xu J, Zhang KQ. 2019. Signal pathways involved in microbe-nematode interactions provide new insights into the biocontrol of plant-parasitic nematodes. Philos Trans R Soc Lond B Biol Sci. 374(1767):20180317
- Liu L, Cao Y-R, Zhang C-C, Fan H-F, Guo Z-Y, Yang H-Y, Chen M, Han J-J, Xu J, Zhang K-Q, et al. 2019. An efficient gene disruption system for the nematophagous fungus *Purpureocillium lavendulum*. Fungal Biol. 123(4):274–282.

- Luangsa-Ard J, Houbraken J, van Doorn T, Hong S-B, Borman AM, Hywel-Jones NL, Samson RA. 2011. Purpureocillium, a new genus for the medically important Paecilomyces lilacinus. FEMS Microbiol Lett. 321(2):141–149.
- Perdomo H, Cano J, Gené J, García D, Hernández M, Guarro J. 2013. Polyphasic analysis of *Purpureocillium lilacinum* isolates from different origins and proposal of the new species *Purpureocillium lavendulum*. Mycologia. 105(1):151–161.
- Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. Nucleic Acids Res. 33(Web Server issue):W686–W689.
- Wang S, Fang M, Xu J, Jiang L, Zhou D, Zhang K-Q, Zhang Y. 2019. Complete mitochondrial genome and phylogenetic analysis of *Orbilia dorsalia*, a species producing mature sexual structures on culture. Mitochondrial DNA Part B. 4(1):573–574.
- Zhang Y, Yang G, Fang M, Deng C, Zhang K-Q, Yu Z, Xu J. 2020. Comparative Analyses of Mitochondrial Genomes Provide Evolutionary Insights Into Nematode-Trapping Fungi. Front Microbiol. 11:617.
- Zhang Y-Q, Yu Z-F. 2019. The complete mitochondrial genomes of the nematode-trapping fungus *Arthrobotrys musiformis*. Mitochondrial DNA B. 4(1):979–980.